

SUPPLEMENTAL DATA for the manuscript:

**OVEREXPRESSION OF SPHINGOSINE-1 PHOSPHATE LYASE PROTECTS
INSULIN-SECRETING CELLS AGAINST CYTOKINE TOXICITY**

Claudine Hahn^{1*}, Karolina Tyka^{1*}, Julie D. Saba², Sigurd Lenzen¹ and Ewa Gurgul-Convey¹

From ¹Institute of Clinical Biochemistry, Hannover Medical School, 30625 Hannover, Germany, ² Children's Hospital Oakland Research Institute, University of California, San Francisco, CA, USA

*equally contributed to this work

This file contains the following supplementary data:

Table S1, Table S2, Table S3 and Table S4

Table S1. Effects of cytokines on the mRNA level of the most abundant S1P receptor and transporter types in insulin-secreting INS1E cells.

Gene		untreated	IL-1 β	Cytokine mixture
<i>S1Pr2</i>	6 h	100 \pm 20 (4)	157 \pm 33 (4)	268 \pm 62 (4)
	24 h	100 \pm 22 (3)	463 \pm 127 (3)*	375 \pm 107 (3)*
<i>S1Pr3</i>	6 h	100 \pm 22 (4)	33 \pm 12 (4)**	33 \pm 15 (4)**
	24 h	100 \pm 20 (6)	204 \pm 71 (6)*	190 \pm 51 (6)**
<i>S1Pr5</i>	6 h	100 \pm 10 (4)	95 \pm 24 (4)	116 \pm 12 (4)
	24 h	99 \pm 10 (6)	175 \pm 24 (6)**	194 \pm 45 (6)*
<i>abca1</i>	6 h	100 \pm 7 (4)	71 \pm 7 (4)*	53 \pm 5 (4)**
	24 h	100 \pm 10 (6)	236 \pm 28 (6)**	289 \pm 27 (6)***

Total RNA was isolated and after reverse transcription real-time RT-PCR was performed. Expression was normalized to the house-keeping gene β -actin. Shown are values as % of untreated. Data are means \pm SEM, with the number of experiments indicated in parentheses. *p<0.05, **p<0.01, ***p<0.001 vs. untreated, ANOVA followed by Bonferroni.

Table S2. Concentration-dependent effects of S1P on cell viability in insulin-secreting INS1E cells

S1P [μ M]	Cell viability
	24 h
0	100 \pm 2 (5)
0.5	94 \pm 3 (4)
5	92 \pm 3 (4)
10	72 \pm 8 (4)
25	47 \pm 13 (4)

Cell viability was measured after a 24 h incubation with various concentrations of S1P by means of the MTT assay. Shown are values as % of untreated. Data are means \pm SEM, with the number of experiments indicated in parentheses. * p <0.05 vs. untreated, ANOVA followed by Bonferroni.

Table S3. Gene expression of enzymes of the sphingolipid pathway in insulin-secreting INS1E cells.

Gene	relative gene expression			
<i>CerS1</i>	1.6E-04	±	4.2E-05	(7)
<i>CerS2</i>	1.9E-02	±	2.6E-03	(8)
<i>CerS3</i>	5.4E-06	±	2.2E-06	(8)
<i>CerS4</i>	9.3E-08	±	4.2E-08	(7)
<i>CerS5</i>	3.4E-02	±	3.8E-03	(8)
<i>spp1</i>	2.0E-04	±	3.6E-05	(8)
<i>spp2</i>	1.2E-03	±	2.6E-04	(8)
<i>sk1</i>	1.0E-04	±	1.5E-05	(11)
<i>sk2</i>	6.3E-03	±	1.5E-03	(8)
<i>CDase acid</i>	1.4E-02	±	2.9E-03	(12)
<i>CDase neutral</i>	5.5E-03	±	1.2E-03	(9)
<i>spl</i>	6.9E-03	±	6.5E-04	(10)
<i>SPT lc1</i>	2.2E-02	±	3.1E-03	(3)
<i>SPT lc2</i>	8.0E-03	±	2.1E-03	(4)
<i>SPT sc</i>	2.8E-05	±	1.0E-05	(4)
<i>SMS acid</i>	1.5E-02	±	2.8E-03	(4)
<i>SMS alkaline</i>	1.7E-05	±	1.3E-06	(3)
<i>SMase</i>	1.2E-03	±	3.0E-04	(7)
<i>Aldh3a2</i>	2.6E-03	±	2.2E-04	(15)

Gene expression was measured in untreated insulin-secreting INS1E cells by means of real-time RT-PCR and normalized to the house-keeping gene β -actin, using the QuantiTect SYBR GreenTM technology (QIAGEN, Hilden, Germany) based on a fluorescent dye that binds only double-stranded DNA. The reactions were performed using the DNA Engine OpticonTM Sequence Detection System (Biozym Diagnostik, Hess. Oldendorf, Germany). Data are means \pm SEM, with the number of experiments indicated in parentheses.

Table S4 Primers used in real-time RT-PCR analysis (all from Invitrogen)

Gene	FW	REV
<i>S1Pr1</i>	CATCTGCTGCTTGATCATCC	AGCCTCCTGCTAACAGGT
<i>S1Pr2</i>	CCTTCGTGGCCAACACCTT	ATGGCCAGGAGGCTGAA
<i>S1Pr3</i>	ATCTTGCCCCTCTACTCCAA	GGATCTCTCGGAGTTGTGGT
<i>S1Pr4</i>	TATGGCTGCATCGGTCTGT	AGAGCACATAGCCCTTGGAG
<i>S1Pr5</i>	TGTGCGCTTTCATCGTGC	AGGATGTTGGTGGCGTAGG
<i>Abca1</i>	CCCTCCTGGAGAGTGCTTTG	CTGGTCACAGCGGTATCTCC
<i>Abcc1</i>	GATGGGGCCTTTGCTGAGTT	CCCTAAACCACTGACACCATTC
<i>Abcg2</i>	ATGTTAGGACTGAAGAGGACGG	AGACACTACGCTTTGGCCTG
<i>Aldh3a2</i>	CTCCCAACAGCGAGTCCAAG	ATCACAGCTGATCCTTGACGA
<i>Spns2</i>	CCTCATGCTGTGCCCTTTTG	CTCGTCAGCCTGCTGCTC
<i>Cer Synthase1</i>	TACCCTTTCTTCCATGACCC	CGGTGGCATAGACGGAAT
<i>Cer Synthase2</i>	GCTTGCTTTCTACTGGTCCC	CTGCTCGGACATAAATTGGCA
<i>Cer Synthase3</i>	TACGGACTGGCAAAGAAGTG	CAGCCACCGTCATCATGAAA
<i>Cer Synthase4</i>	TGCTGCCTTGCCAATTG	GCACTGGGTTGGGCTTTATC
<i>Cer Synthase5</i>	GCTGGCAGCAGGTCTCTTCT	CCCTTCAGCCTTTTCTCATC
<i>Alkaline CDase</i>	ACAACAACACTTGGGTGGTG	ATCCTGTCAACATGCCAC
<i>Neutral CDase</i>	ACTCTGGCCCAGCAGGATT	GGACTTTGCCCGGTTTAAGA
<i>Alkaline SMS</i>	TGACTGGGGGTCTGTCCAA	GCAGGAGTCATGTAGCGAGC
<i>Acid SMS</i>	TCTTTGAGGACGATGTGGTG	GCACTGATGGCAAAGAGATG
<i>sk1</i>	CTTCTGGAGGAGGCTGAGGT	TCAGACCGTCACCGGACAT
<i>sk2</i>	CAAGCCCTACACATACAGCG	GCCACGTGGGTAGGTGTAGA
<i>spp1</i>	CGAGCTGGCCAAAGTGAGCA	AGATGATCACCAGCCTCCGG
<i>spp2</i>	TTCTCCCATTCACCCACTG	TCACAACGGGAGGAGAGGAG
<i>spl</i>	GCCATTCCTGAAGTTGGACA	CCTGAGGCTTTCCCTTCTTG
<i>huSPL</i>	ACGGCCTGGTGGCATT	CTGACAATTGGGGATTCCC

<i>SpMn</i>	GGCTGCTGGTGGCTCCATCTA	TGGATGAACTGGGCCAGTT
<i>SPTlc1</i>	TTGAAGAATGGCAGCCAGAG	CCGAGGAAATTAAGGAGGC
<i>SPTlc2</i>	GAAAAGTGCCACCATGCAAC	ACAGATGGGTCGATTCCAGT
<i>SPTsc</i>	GAAGGAGTATTTTGCCTGGC	GGGGATGAAGACATAGGCAG
<i>bip</i>	CCACCAGGATGCAGACATTG	AGGGCCTCCACTTCCATAGA
<i>chop</i>	CCAGCAGAGGTCACAAGCAC	CGCACTGACCACTCTGTTC
<i>β-actin</i>	GAACACGGCATTGTAACCAACTGG	GGCCACACGCAGCTCATTGTA