Supplementary information

Artemisia annua extract prevents ovariectomy-induced bone loss by blocking receptor activator of nuclear factor kappa-B ligand-induced differentiation of osteoclasts

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Supplementary methods

Cell viability assay. MC3T3-E1 osteoblastic cells (5×10^3 cells/well) were seeded onto 6-well plates with 10% FBS- α -MEM for 24 h. The cells were incubated in serum-free α -MEM with AaE, artemsinin, artemisinic acid, or arteannuin B at the indicated concentrations for 24 h and 72 h. The cell viability was measured using an MTT assay.

Alkaline phosphatase activity. MC3T3-E1 cells (5×10^4 cells/well) were seeded onto 12-well plates with 10% FBS- α -MEM (growth medium) or 10% FBS- α -MEM containing 10 nM dexamethasone, 50 µg/mL ascorbic acid, and 2 mM β -glycerophosphate (osteogenic medium). The cells were treated with the indicated concentrations of AaE, artemisinin, artemisinic acid, or arteannuin B for 72 h. To assess the activity of alkaline phosphatase, the cells were washed with PBS and then suspended in 0.5 mL PBS containing 0.1 M glycine, 1 mM MgCl₂, and 0.05% Triton X-100. The lysate (100 µL was incubated with 250 µL of *p*-nitrophenyl phosphate solution at 37°C for 30 min. Enzymatic activity was terminated by adding 100 µL of ice cold 3 M NaOH, and the amount of liberated *p*-nitrophenol was measured by monitoring absorbance at 405 nm.



Figure S1. The effects of AaE, artemisinin, and arteannuin B on body weight and normal tissues in OVX mice. Eight-week-old female OVX mice (N=10) were administered vehicle (PBS containing 1% DMSO and 1% Tween-20), 1 or 10 mg/kg BW of AaE, or 0.1 mg/kg BW of E2 by oral gavage 5 times per week for 12 weeks. In addition, OVX mice were divided into 6 groups of 10 mice and were administered vehicle, 10 or 20 mg/kg BW of artemisinin, 20 mg/kg BW of arteannuin B, or 0.1 mg/kg BW of E2 by oral gavage 5 times per week for 12 weeks. Sham-operated mice (N=10) received the vehicle alone. The body weights were measured in mice treated (a) AaE, (b) artemisinin (ART) or arteannuin B (AB) at 12 weeks. Serum levels of (c) ALT and AST to assess liver damage and (d) BUN and creatinine to assess kidney damage were quantified using their respective commercially available assay kits. The data are expressed as the mean \pm SE. [#]*P* < 0.01 *versus* sham-operated mice (S), ^{*}*P* < 0.05, ^{**}*P* < 0.01 *versus* OVX mice.



Figure S2. The effects of AaE and its active components on ALP activity of osteoblastic cells. (a) MC3T3-E1 cells were incubated with the indicated concentration of AaE, artemisinin (ART), artemisinic acid (AA), or arteannuin B (AB) for 24 h and 72 h. Cell viability was analyzed using an MTT assay. The data are expressed as the mean \pm SE. **P* < 0.05, ***P* < 0.01 *versus* cells without compounds. (b) MC3T3-E1 cells were cultured in growth medium (GM) or osteogenic medium (OM) treated with the indicated concentration of AaE, ART, AA, or AB for 72 h. ALP activity was assessed as described in the *Supplementary methods* section. The data are expressed as the mean \pm SE. **P* < 0.01 *versus* cells with OM alone.



Figure S3. The effects of AaE on RANKL and OPG secretion from proinflammatory cytokinestimulated osteoblastic cells. hFOB 1.19 and MG-63 cells were incubated with (a) TNF- α (10 ng/mL) or (b) IL-1 β (10 ng/mL) and the indicated concentration of AaE for 24 h. RANKL and OPG levels in culture media from the osteoblastic cells were measured using commercially available RANKL and OPG ELISA kits. The data are expressed as the mean \pm SE. [#]P < 0.05 versus cells treated with media alone (C), ^{*}P < 0.05, ^{**}P < 0.01 versus TNF- α -treated cells or IL-1 β -treated cells.



Figure 5b





Figure 6a













