Supplementary Figures



Supplementary Figure 1. SEC of recombinant NT*-Bri2 BRICHOS and PAGE analysis of the fractions. (a) NT*-Bri2 BRICHOS was separated on a Superdex 200 PG column, and oligomers (oli), dimers (dim) and monomers (mon) were collected and analyzed with native PAGE (b), SDS-PAGE under reducing conditions (c), and non-reducing conditions (d). Lane M shows migration of protein size markers with masses indicated to the left. T stands for unresolved NT*-Bri2 BRICHOS proteins before SEC.



Supplementary Figure 2. Characterization of rh Bri2 BRICHOS species. Electrospray mass spectra of rh Bri2 BRICHOS monomers (a), and oligomers at 50 V (b) or 175 V (c) collision voltage. (d) CD spectra at 25°C of rh Bri2 BRICHOS oligomer (blue), dimer (green), monomer (red), and proSP-C BRICHOS (black). (e) Fluorescence emission of 10 μ M bis-ASN in sodium phosphate buffer (black) or in the presence of 2 μ M rh Bri2 BRICHOS oligomer (blue), dimer (green), or monomer (red).



Supplementary Figure 3. Electron micrograph and single particle analysis of Bri2 oligomers. (a) Refinement with full sized data (1.038 Å/pixel) shows significant similarities between the 3D density maps with C2 symmetry (left) and D2 symmetry (rigth). Convergence plots for the maps with (b) C2 and (c) D2 symmetries (1.038 Å/pixel).



Supplementary Figure 4. Different assembly states of rh Bri2 BRICHOS species after

incubation. The Bri2 BRICHOS oligomers, dimers and monomers at concentrations that are 2-3 times higher than the ones used in the ThT assay are incubated in buffer as the same as the ThT assay at 37° C. Samples were analyzed for assembly state by SDS-PAGE after 0, 1, 4 and 24 hours under reducing (**a**) and non-reducing conditions (**b**).



Supplementary Figure 5. Global fits of aggregation traces of A β 42 in the presence and absence of rh Bri2 BRICHOS species. Global fits (solid lines) of aggregation traces (dots) at different peptide concentrations from 1.6 (red) to 9 (dark violet) μ M A β 42 in the absence or in the presence of 0.9 μ M Bri2 BRICHOS. The fitting parameters are summarized in Supplementary Table 2.



Supplementary Figure 6. Seeded aggregation traces of A β 42 in the presence and absence of rh Bri2 BRICHOS species. Seeded aggregation traces of 3 μ M A β 42 with 0.6 μ M preformed A β 42 fibrils in the presence of 0 (black), 10 (violet), 30 (cyan), 50 (green), 70 (yellow) and 100 % (red) Bri2 BRICHOS (molar percentage referred to monomeric subunits). The inserted graphs show a zoom for the first two hours where the initial slope is determined by a linear fit to the first 30 min.



Supplementary Figure 7. Aggregation kinetics of A β 42 in the presence of different Bri2 BRICHOS oligomer, dimer and monomer concentrations. Aggregation kinetics of 3 μ M A β 42 in the presence of different Bri2 BRICHOS concentrations (molar percentage referred to monomeric subunits): 0 (black), 10 (violet), 30 (cyan), 50 (green), 70 (yellow) or 100 % (red). The global fits (solid lines) of the aggregation traces (dots) were constrained such that only one single rate constants is the sole free fitting parameter, indicated at the top of each column. χ^2 values describing the quality of the fits are listed in **Supplementary Table 3**.



Supplementary Figure 8. Immuogold TEM of Aβ42 fibrils with and without Bri2

BRICHOS. 5 μ M A β 42 was incubated with and without 50% molar ratio of rh Bri2 BRICHOS monomer or dimer overnight at 37°C, or the same concentrations of Bri2 BRICHOS were incubated alone. The samples were treated with a Bri2 BRICHOS antibody and a gold-labeled secondary antibody and characterized by TEM. The sizes of the scale bars are indicated underneath each panel. (a) 50% Bri2 BRICHOS dimer alone; (b) 50% Bri2 BRICHOS monomer alone; (c) A β 42 alone; (d, f and h) A β 42+50% Bri2 BRICHOS dimer; (e, g and i) A β 42+50% Bri2 BRICHOS monomer. Black arrows in d-g identify A β 42 fibril ends covered by Bri2 BRICHOS.



Supplementary Figure 9. Analyses of rh Bri2 BRICHOS expressed in mammalian cells, and rh Bri2 BRICHOS incubation in buffer and serum. (a) Lysates of HEK293 cells expressing rh Bri2 BRICHOS were incubated with (+) and without (-) *N*-Ethylmaleimide (NEM) overnight at 37°C, and the samples were then analyzed by Western blotting. Lane C shows analyses of non-transfected cells. (b) Lysates of HEK293 cells expressing rh Bri2 BRICHOS were incubated with (+) and without (-) 5 U PNGase F overnight at 37°C, and then the samples were analyzed by SDS-PAGE under reducing conditions and western blotting. (c) Monomeric rh Bri2 BRICHOS incubated at 4°C or 37°C overnight and then analyzed by SDS-PAGE under reducing conditions. (d) Monomeric rh Bri2 BRICHOS incubated in mouse serum at 37°C, or serum only, analyzed by western blotting under reducing conditions at different time points (in hours) as indicated above each lane. (e) Recombinant proSP-C BRICHOS was kept at -20°C (fresh) or incubated at 37°C overnight and then analyzed by SDS-PAGE under reducing conditions. (f) Fibril formation of 3 μ M A β 42 alone (black), or in the presence of 50% molar ratio of proSP-C BRICHOS kept at -20°C (red) or incubated at 37°C overnight (blue).

Supplementary Tables

Supplementary Table 1. Free thiols/molecule of Bri2 BRICHOS monomer under different conditions.

Non-reduced	Denatured	Reduced
-0.01 ± 0.06	0.03 ± 0.02	2.00 ± 0.04

Data represents mean values \pm sd. of three measurements

Supplementary Table 2. Fitting parameters from global fit in Supplementary Figure 5.

Fitting parameter	Aβ42 alone	Monomer	Dimer	Oligomer
$\sqrt{k_n k_+} \left[M^{-1} s^{-1} \right]$	7.9 ± 0.3	7.9 [§]	7.9 [§]	7.9 [§]
$\sqrt{k_+k_2} \left[M^{-3/2} s^{-1} \right]$	$1.32 \cdot 10^5 *$	$4.15\cdot 10^4$	$3.03\cdot 10^4$	$6.41\cdot 10^4$
Relative $\sqrt{k_+k_2}$	100 %	31 %	23 %	48 %
Relative k ₊ k ₂	100 %	10 %	5 %	24 %

* Fitting error $< 2 \times 10^{1}$, [§] Fixed value to A β 42 alone

Supplementary	Table 3.	Assessment of c	juality of	fits shown	in Sup	plementary	Figure	7.
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χ ²	${m k}_n$ free	k ₂ free	k_+ free
Monomer	33.3	7.4	7.2
Dimer	52.5	18.2	16.6
Oligomer	18.0	1.6	6.1

Supplementary Table 4. Gamma oscillations power in mouse hippocampal slice

preparations under different conditions shown in Figure 5.

Experiment	Mean ± sem [10 ⁻⁹ V2]	Number
KA control gamma	4.96 ± 0.41	24
KA gamma after 50 nM Aβ42	1.28 ± 0.17	17
KA gamma after Aβ42 and 10 nM Bri2 BRICHOS oligomers	1.65 ± 0.47	7
KA gamma after Aβ42 and 50 nM Bri2 BRICHOS oligomers	2.78 ± 0.13	7
KA gamma after Aβ42 and 100 nM Bri2 BRICHOS oligomers	2.47 ± 0.42	7
KA gamma after Aβ42 and 10 nM Bri2 BRICHOS dimers	0.98 ± 0.17	7
KA gamma after Aβ42 and 50 nM Bri2 BRICHOS dimers	3.45 ± 0.44	9
KA gamma after Aβ42 and 100 nM Bri2 BRICHOS dimers	2.94 ± 0.53	7
KA gamma after Aβ42 and 10 nM Bri2 BRICHOS monomers	1.21 ± 0.53	5
KA gamma after Aβ42 and 20 nM Bri2 BRICHOS monomers	2.69 ± 0.44	5
KA gamma after Aβ42 and 50 nM Bri2 BRICHOS monomers	5.68 ± 0.09	8
KA gamma after 50 nM Bri2 BRICHOS oligomers	5.69 ± 0.04	3
KA gamma after 50 nM Bri2 BRICHOS dimers	5.92 ± 0.1	3
KA gamma after 50 nM Bri2 BRICHOS monomers	5.81 ± 0.05	3

Supplementary Table 5. Primer sequences for NT*-Bri2 BRICHOS cloning

Primers	Sequence (from 5'to 3')
Sense	TATTGAATTCCCTGGTGCCACGCGGTTCTCAGACAATTGAAGAAAATATT
Antisense	GGATCGGGTACCAAGCTTACAGTTTGTAAGTTTCCTTGT