

Supplemental Material: Epistasis between TIFAB and miR-146a, neighboring genes in del(5q) MDS

Supplemental Methods

Mice and bone marrow transplantation.

Tifab^{-/-} C57Bl/6 mice were described previously¹. miR-146a^{-/-} C57BL/6 mice were obtained from Dr. David Baltimore as previously described². All animals were bred and housed in the Association for Assessment and Accreditation of Laboratory Animal Care-accredited animal facility of Cincinnati Children's Hospital Medical Center. For non-competitive BM transplantation, BM mononuclear cells (0.3-3x10⁶) from C7BL/6 wild-type, *Tifab*^{-/-}, miR-146a^{-/-}, or *Tifab*^{-/-};miR-146a^{-/-} mice were injected *i.v.* into lethally-radiated (7.0 Gy and 4.75 Gy after 3 hours) syngeneic recipient Boy/J mice.

BM and PB analysis

For cytopins, 2.5-5x10⁵ BM cells were spun onto slide at 300 rpm for 5 minutes and then stained with Wright-Giemsa. For BM sections, tibias were fixed in formalin and then stained with hematoxylin and eosin. Complete blood counts were performed on peripheral blood isolated from the tail vein every 4 weeks and analyzed using the Drew Scientific Hemavet 950.

Quantitative PCR analysis

RNA was extracted from cells and purified with Trizol Reagent (Life Technologies 15596-026) or Quick-RNA MiniPrep Kit (Zymo Research R1055). cDNA was generated using a high capacity RNA to cDNA kit (Life Technologies 4387406 or 4368814). qPCR was carried out using Taqman Master mix and probes (Life Technologies): *Tifab* (Mm0421026_m1) and *Gapdh* (Mm99999915_g1). qPCR was performed on an Applied Biosystems StepOne Plus Real-Time PCR System. miR-146a expression was performed as previously described³.

Clonogenic progenitor assays

Hematopoietic clonogenic progenitor frequencies were determined by plating 2x10⁵ BM cells/ml isolated from transplanted wild-type, *Tifab*^{-/-}, miR-146a^{-/-}, or *Tifab*^{-/-};miR-146a^{-/-} mice in methylcellulose media containing human erythropoietin, murine SCF, murine IL-3, and human IL-6 (Methocult M3434; Stem Cell Technologies). Colonies were scored after 10 days.

Flow cytometry analysis

Antibodies used for flow cytometric analysis of BM and PB cells are described previously¹. All FACS analyses were performed on BD FACS Aria or Canto machines. FACS data analysis was performed using BD FACSDiva or FlowJo software.

Patient samples

Informed consent was obtained according to protocols approved by the review board of Cleveland Clinic. Diagnoses were reviewed at Cleveland Clinic and adapted, when required, to WHO 2008 criteria. For microarray and SNP analysis, BM mononuclear cells were isolated from MDS/AML patients, as previously described².

RNA sequencing analysis

Transcriptomes of lineage⁻cKit⁺ BM cells from wild-type (WT, n=3), *Tifab*^{-/-} knockout (TIFAB-KO, n=3), *miR-146a*^{-/-} (miR146a-KO, n=3), *Tifab*^{-/-}; *miR-146a*^{-/-} mice (dKO, n=4) were compared by standard RNA-seq analysis methods. Total RNA was amplified using the Ovation RNA-Seq System v2 (NuGEN) according to the manufacturer's protocol. The libraries were prepared with the Nextera XT DNA Sample Preparation kit (Illumina Technologies). 1 ng of cDNA was suspended in Tagment DNA Buffer, and tagmentation (fragmentation and tagging with the adaptors) was performed with the Nextera enzyme (Amplicon Tagment Mix), incubating at 55°C for 10 min. NT Buffer was then added to neutralize the samples. Libraries were prepared by PCR with the Nextera PCR Master Mix, and 2 Nextera Indexes (N7XX, and N5XX) according to the following program: one cycle of 72°C for 3min, one cycle of 98°C for 30s, 12 cycles of 95°C for 10s, 55°C for 30s, and 72°C for 1min, and one cycle of 72°C for 5min. The purified cDNA was captured on an Illumina flow cell for cluster generation. Libraries were sequenced on the Illumina HiSeq2500 following the manufacturer's protocol. Paired-end reads were aligned to UCSC mm10 genome (downloaded from Illumina's iGenomes repository; https://support.illumina.com/sequencing/sequencing_software/igenome.html) using TopHat⁴ and mouse mm10-refseq gene GTF file. FeatureCounts was used for read counting⁵. Differentially expressed genes were predicted by three independent methods including limma/voom^{6,7}, edgeR⁸, and DESeq2⁹. FDR < 0.05 and fold change > 2x were used for default cutoffs.

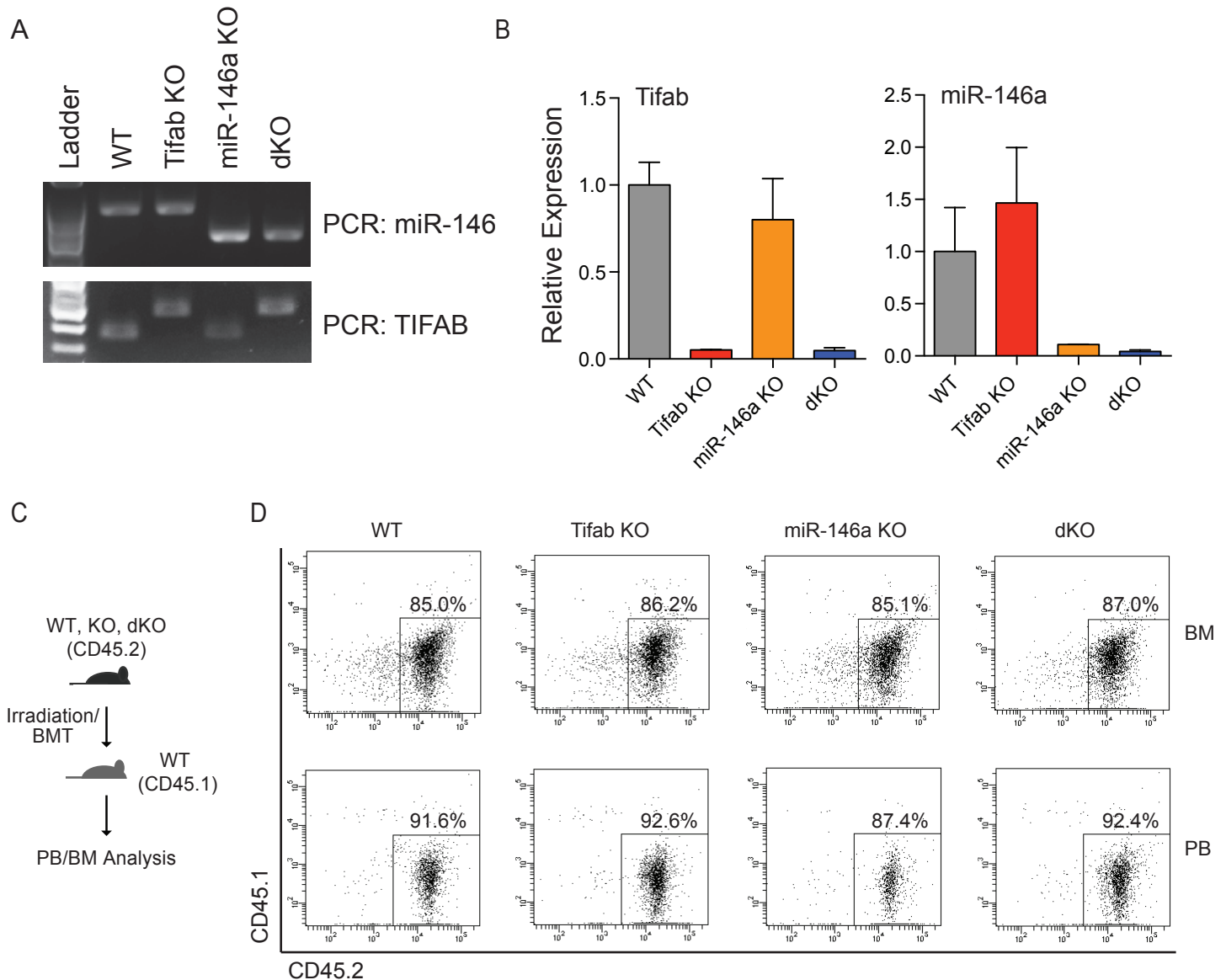
Synergy gene analysis

To examine synergistic effects, we adopted a concept used in McMurray *et al*¹⁰. Briefly, *a* = fold change value for a given gene in the *miR-146a*^{-/-} group, *b* = fold change value for the same gene in the *Tifab*^{-/-} group, and *d* = mean expression value for this gene in the dKO group. Then synergy score was calculated using $(a+b)/d$ if $d > 1$ and $d/a + d/b$ if $d < 1$.

Supplemental References

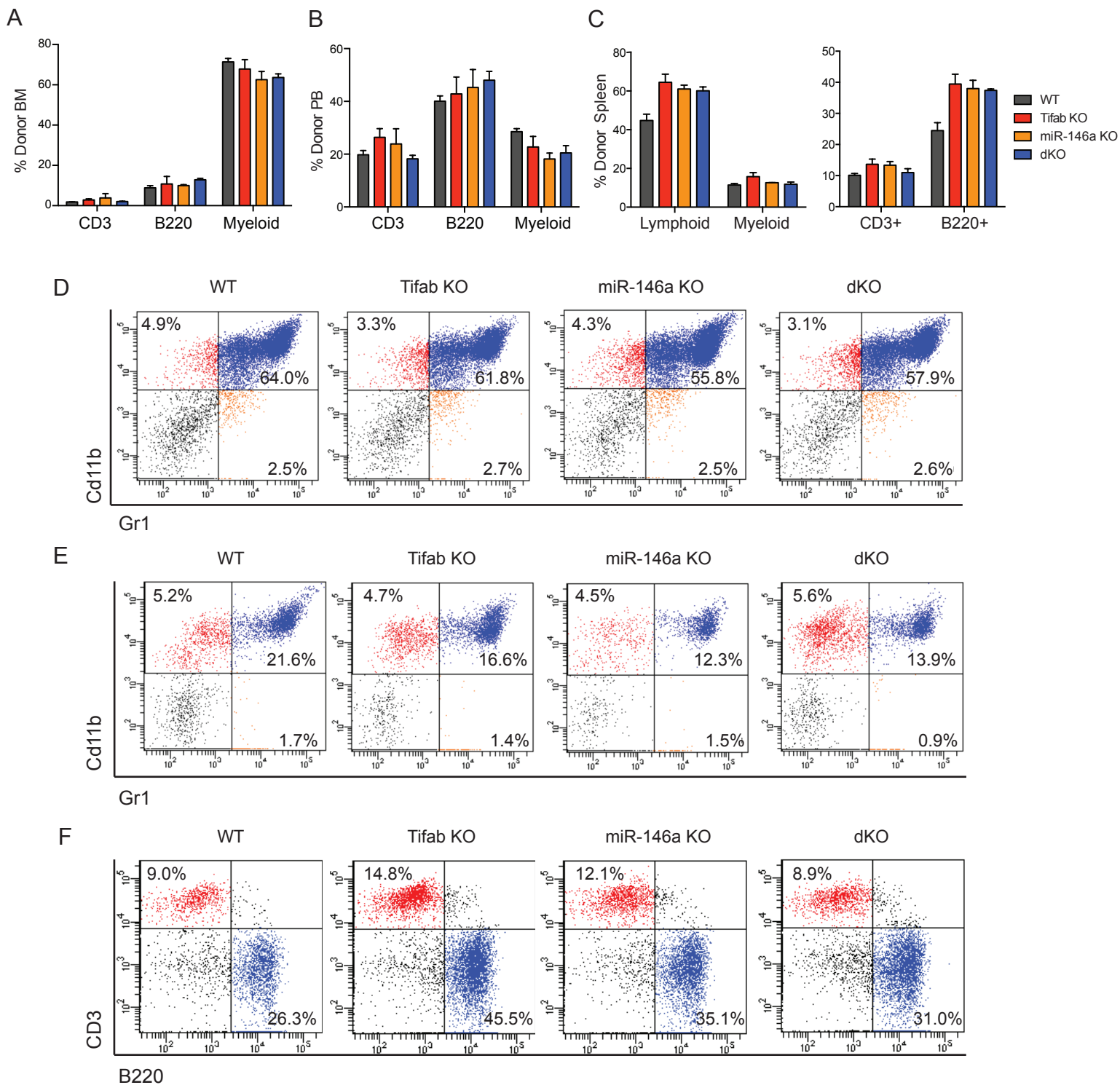
1. Varney, M.E. *et al.* Loss of Tifab, a del(5q) MDS gene, alters hematopoiesis through derepression of Toll-like receptor-TRAF6 signaling. *J Exp Med* **212**, 1967-85 (2015).
2. Fang, J. *et al.* Myeloid malignancies with chromosome 5q deletions acquire a dependency on an intrachromosomal NF-kappaB gene network. *Cell Rep* **8**, 1328-38 (2014).
3. Starczynowski, D.T. *et al.* MicroRNA-146a disrupts hematopoietic differentiation and survival. *Exp Hematol* **39**, 167-178 e4 (2011).
4. Trapnell, C., Pachter, L. & Salzberg, S.L. TopHat: discovering splice junctions with RNA-Seq. *Bioinformatics* **25**, 1105-11 (2009).
5. Liao, Y., Smyth, G.K. & Shi, W. featureCounts: an efficient general purpose program for assigning sequence reads to genomic features. *Bioinformatics* **30**, 923-30 (2014).
6. Law, C.W., Chen, Y., Shi, W. & Smyth, G.K. voom: Precision weights unlock linear model analysis tools for RNA-seq read counts. *Genome Biol* **15**, R29 (2014).
7. Ritchie, M.E. *et al.* limma powers differential expression analyses for RNA-sequencing and microarray studies. *Nucleic Acids Res* **43**, e47 (2015).
8. Robinson, M.D., McCarthy, D.J. & Smyth, G.K. edgeR: a Bioconductor package for differential expression analysis of digital gene expression data. *Bioinformatics* **26**, 139-40 (2010).
9. Love, M.I., Huber, W. & Anders, S. Moderated estimation of fold change and dispersion for RNA-seq data with DESeq2. *Genome Biol* **15**, 550 (2014).
10. McMurray, H.R. *et al.* Synergistic response to oncogenic mutations defines gene class critical to cancer phenotype. *Nature* **453**, 1112-6 (2008).

Supplemental Figure 1



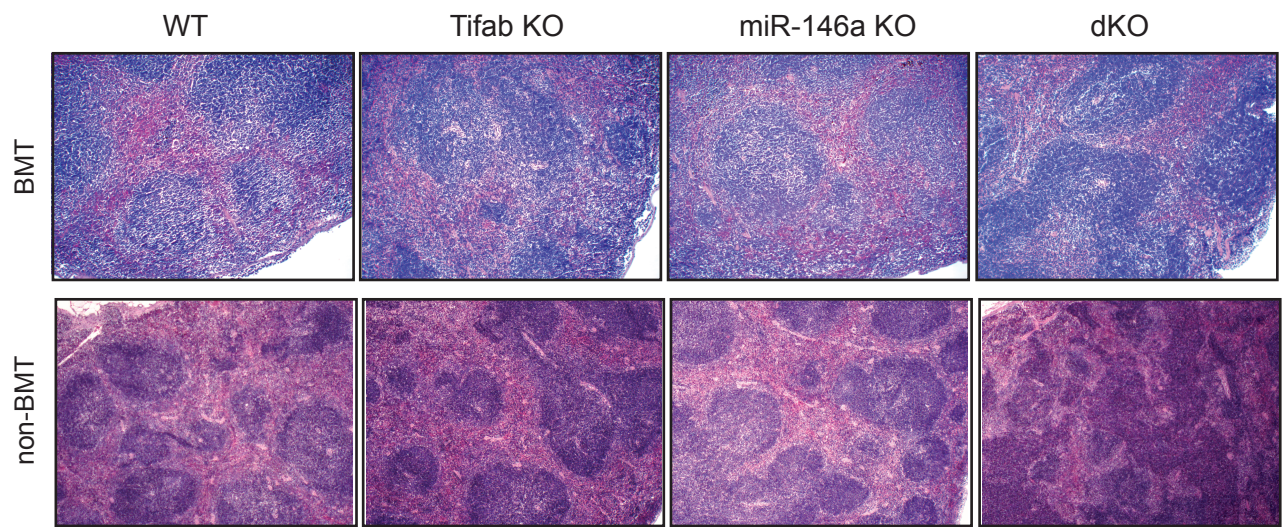
Generation of Tifab and miR-146a double knockout mice. (A) PCR was performed using primers to detect wild-type or deleted miR-146a and Tifab alleles on genomic DNA. (B) Quantitative RT-PCR was performed to measure Tifab mRNA or miR-146a from BM mononuclear cells. (C) Overview of transplantation of BM cells from WT, Tifab KO, miR-146a KO, and dKO (CD45.2) mice into lethally-irradiated WT CD45.1 mice. (D) Proportion of donor cells in the peripheral blood and BM of recipient mice (n = 3 mice evaluated per group).

Supplemental Figure 2



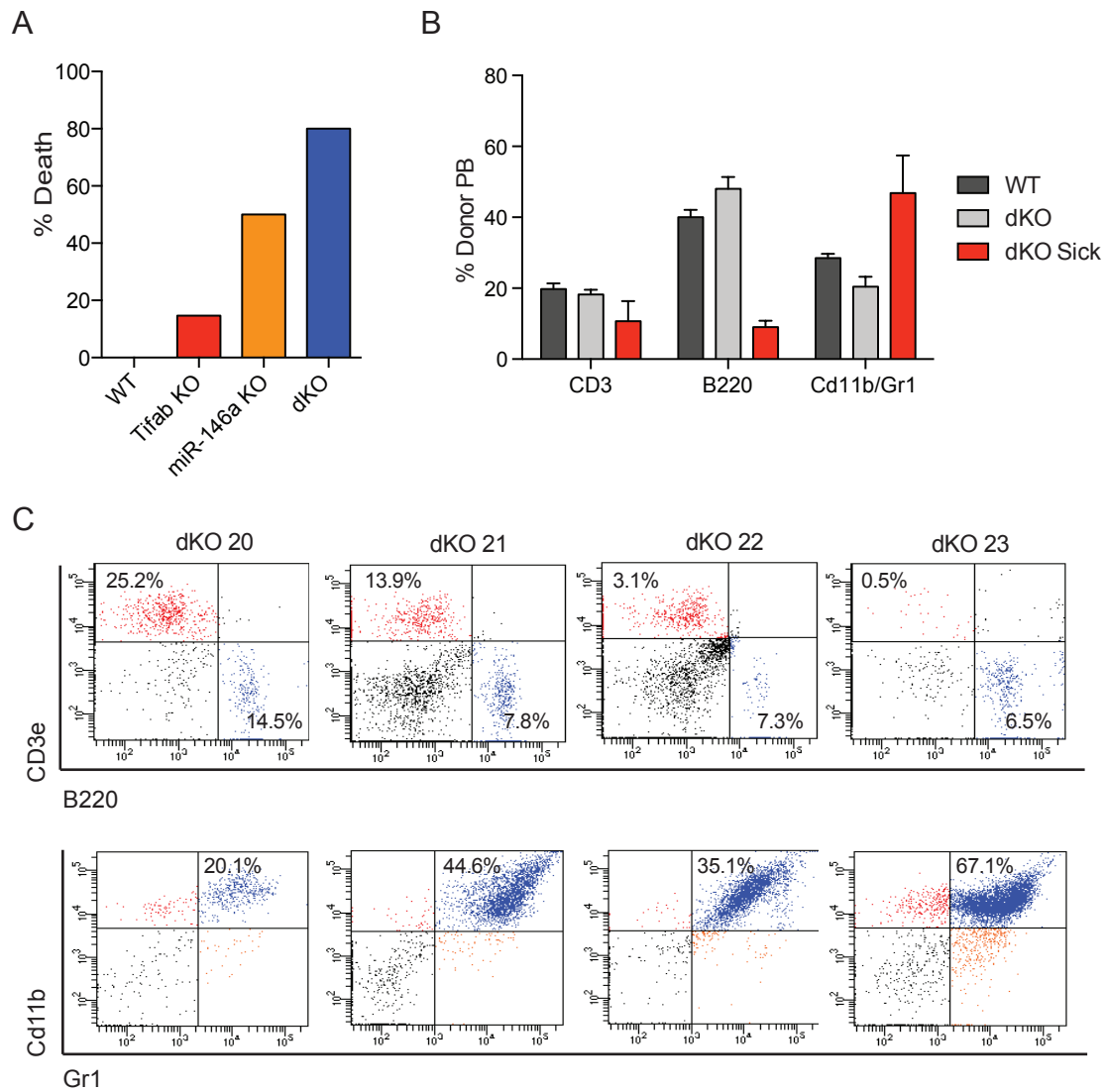
Immunophenotypic analysis of bone marrow, peripheral blood, and spleen. (A-C) Flow cytometric analysis of myeloid (CD11b, Gr1) and lymphoid (CD3, B220) proportions within the BM (A), PB (B), and spleen (C) of transplanted mice. (D-F) Representative flow cytometric plots from BM (D), PB (E), and spleen (F). Percent values within dot blots are calculated from the parent gate.

Supplemental Figure 3



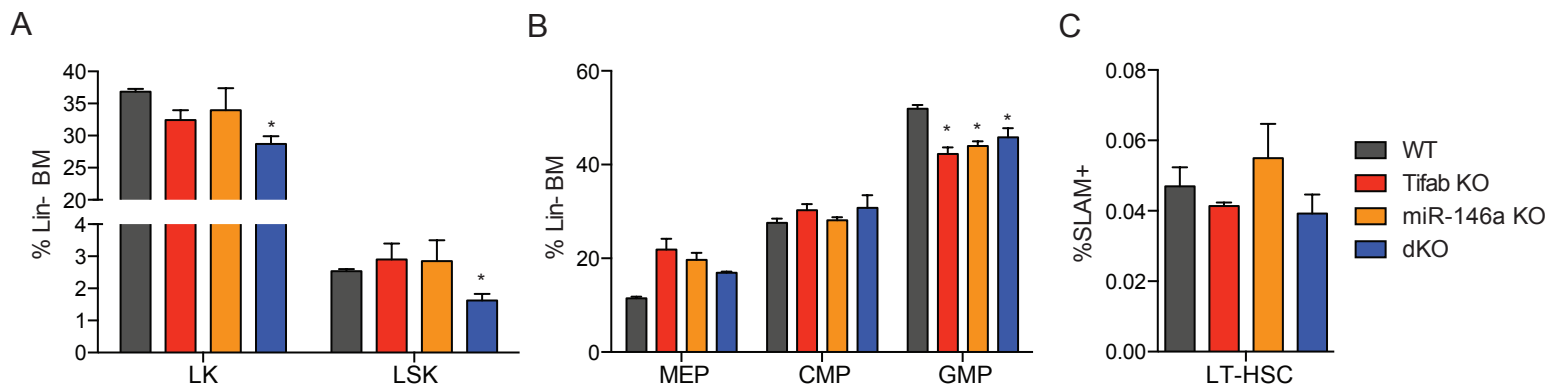
Morphological analysis of spleens. H&E-stained spleen from representative mice prior (non-BMT) or after transplantation (BMT).

Supplemental Figure 4.



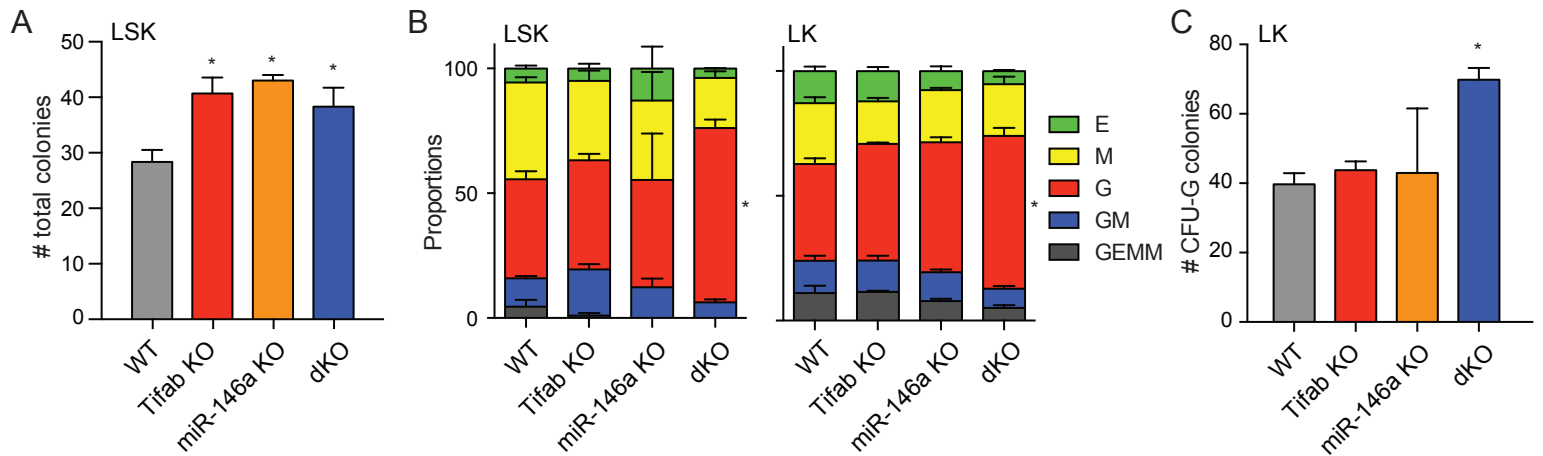
Analysis of moribund mice. (A) Percent of mice became moribund after transplantation. (B) Immunophenotype analysis of PB from WT, dKO, and moribund dKO mice. (C) Representative flow cytometric blots of PB cells from four moribund dKO mice. Percent values within dot blots are calculated from the parent gate.

Supplemental Figure 5.



Analysis of hematopoietic stem/progenitor proportions in the BM. (A) LK and LSK proportions in BM from mice transplanted with WT, Tifab^{-/-}, miR-146a^{-/-}, or dKO BM cells (n = 3). (B) MEP, CMP, and GMP proportions in BM from mice transplanted with WT, Tifab^{-/-}, miR-146a^{-/-}, or dKO BM cells (n = 3). (C) LT-HSC (LSK⁺ CD150⁺CD48⁻) proportions in BM from mice transplanted with WT, Tifab^{-/-}, miR-146a^{-/-}, or dKO BM cells (n = 3). *, P < 0.05; Student's t-test. In all cases, statistical comparisons were made to WT mice.

Supplemental Figure 6.



Analysis of hematopoietic stem/progenitor colony-forming function. (A) Numbers of colony-forming cells in methylcellulose from LSK cells from WT, Tifab^{-/-}, miR-146a^{-/-}, or dKO transplanted mice (n = 3). (B) Proportion of colony-forming cells in methylcellulose from LSK or LK isolated from WT, Tifab^{-/-}, miR-146a^{-/-}, or dKO transplanted mice (n = 3). GM, granulocyte-monocyte; G, granulocyte; M, monocyte; E, erythroid (BFU-E). (C) Numbers of CFU-G from LK cells from WT, Tifab^{-/-}, miR-146a^{-/-}, or dKO transplanted mice (n = 3).*, P < 0.05; Student's t-test as compared to WT.

Supplemental Table 1. Differentially expressed genes in LK.

Tifab KO vs WT

Gene	Fold Change (Log2)	P value	FDR
Tifab	-1.84	6.34E-15	1.53E-10
Prom1	2.27	1.58E-10	1.90E-06
Doc2	1.13	1.81E-07	0.001
Dnmt	-1.37	1.99E-07	0.001
C3	1.15	2.39E-07	0.001
Sic15a2	1.83	7.26E-07	0.003
H2afy	-1.05	9.67E-07	0.003
Mtus1	1.40	9.95E-07	0.003
Pgfpyp1	1.27	1.33E-06	0.004
Nucb2	1.22	1.69E-06	0.004
Prkcb	1.14	1.76E-06	0.004
Mexa3	1.22	4.32E-06	0.009
Sycp1	-0.11	5.19E-06	0.010
F830028O10Rik	1.12	7.48E-06	0.013
Ncam1	1.49	8.51E-06	0.014
B4galnt6	1.50	1.17E-05	0.018
Fcrl1a	1.17	1.44E-05	0.020
G51	1.24	2.20E-05	0.028
Mrc1	1.90	2.26E-05	0.028
Axl	1.76	2.32E-05	0.028
Vcam1	1.86	3.11E-05	0.034
Gem	-1.19	3.21E-05	0.034
Mras	2.29	3.29E-05	0.034
Ptn3	1.07	3.49E-05	0.035
Cod5a1	-1.07	3.94E-05	0.037
Tex11	5.92	4.04E-05	0.037
Epx	1.31	4.24E-05	0.038
Ear1	1.44	4.61E-05	0.040
Gamb	1.00	5.59E-05	0.045
Igf1	2.62	5.99E-05	0.045
Enah	1.63	5.94E-05	0.046
Gda	1.06	6.19E-05	0.047
Sema1	-3.11	6.36E-05	0.047
Lrpuk	-0.06	6.83E-05	0.048
Igsf6	0.82	7.05E-05	0.049
C1qb	2.80	7.37E-05	0.049
Abcc2	1.85	7.49E-05	0.049
Bksgnt4	0.90	7.70E-05	0.049

miR-146a KO vs WT

Gene	Fold Change (Log2)	P value	FDR
Lym7	3.10	3.19E-32	7.69E-28
Cmas1	-2.46	9.72E-22	1.17E-17
Gzmb	1.66	5.90E-17	4.73E-13
Olfms5	-8.25	1.65E-16	9.94E-13
Gm2a	-1.30	1.71E-15	8.21E-12
Doc2	1.34	2.79E-15	1.12E-11
Wdfy1	1.51	4.76E-14	1.51E-10
Vps13c	1.12	5.34E-14	1.51E-10
F830028O10Rik	1.26	5.64E-14	1.51E-10
1700112E06Rik	-2.66	9.81E-13	2.36E-09
Prkcb	1.43	1.75E-12	3.84E-09
Prk35	1.42	1.17E-11	2.36E-08
Cyp11a1	2.64	5.34E-10	9.89E-08
Mtus1	1.45	1.34E-10	2.31E-07
Irgm2	1.86	2.09E-10	3.35E-07
Nkx41	-1.54	8.01E-10	1.20E-06
Ctcah7	1.37	1.33E-09	1.84E-06
Egr3	-1.82	8.00E-09	1.07E-05
Erd1	1.11	1.55E-08	1.96E-05
Igfp1	2.52	2.87E-08	3.46E-05
Dnt1	-1.23	3.62E-08	4.19E-05
Serpina3g	0.98	5.46E-08	5.97E-05
Zbtb16	1.05	6.85E-08	9.26E-05
Nirc5	0.94	1.01E-07	0.0001
Th2	2.15	1.12E-07	0.0002
Gm12250	1.51	1.60E-07	0.0001
Skp2	1.02	1.73E-07	0.0002
Smm3	-1.04	1.76E-07	0.0002
C3	0.99	2.45E-07	0.0002
Alm	0.75	3.00E-07	0.0002
Tec2d3	-1.25	3.07E-07	0.0002
Irgm1	1.03	3.74E-07	0.0003
Fcrl1a	1.02	4.11E-07	0.0003
Rplp1	-0.83	6.05E-07	0.0004
Z810417H13Rik	-0.77	6.45E-07	0.0004
Nrg1	2.57	9.18E-07	0.0006
Clec5a	1.37	9.46E-07	0.0006
Doc2a	0.99	9.89E-07	0.0007
Stk17b	-0.82	1.14E-06	0.0007
Ppbb	-1.21	1.30E-06	0.0008
Clec4e3	4.99	1.33E-06	0.0008
Mir394	1.70	1.79E-06	0.0010
Lgcat1	0.80	1.87E-06	0.0010
Tgfp1	2.58	1.93E-06	0.0011
HistH2bc	-1.50	2.02E-06	0.0011
Hmg5	-0.80	2.07E-06	0.0011
Camk2b	-3.82	2.21E-06	0.0011
Rab6b	-1.33	2.41E-06	0.0012
Sic31a2	1.08	2.55E-06	0.0013
B4galnt6	1.31	2.69E-06	0.0014
Gstm5	-0.84	3.01E-06	0.0014
H1f0	-0.81	3.15E-06	0.0015
Gm1966	0.69	3.45E-06	0.0016
Sypx1	-0.99	3.87E-06	0.0017
Alp11a	0.81	4.10E-06	0.0018
App	0.86	4.18E-06	0.0018
Sdr21	0.82	4.75E-06	0.0020
P4	0.88	4.84E-06	0.0020
Tmem9	-1.00	5.00E-06	0.0020
Znf41-ps	1.02	6.03E-06	0.0024
Sowahc	1.02	6.05E-06	0.0024
Rpl19	-0.81	6.29E-06	0.0028
Igf1	0.91	7.50E-06	0.0029
Mras	2.15	7.68E-06	0.0030
Tlil4	0.97	8.04E-06	0.0030
Fhd3a	0.88	8.23E-06	0.0030
Icxf5	1.67	9.95E-06	0.0036
Gpr84	2.50	1.03E-05	0.0036
Rps4x	-0.92	1.12E-05	0.0039
Alas1	0.74	1.14E-05	0.0039
R3hdm4	-0.69	1.19E-05	0.0040
H2-Q5	0.85	1.24E-05	0.0042
Bpic	-0.97	1.27E-05	0.0042
Ccdc69	-1.05	1.29E-05	0.0042
Rpl33	-0.87	1.32E-05	0.0042
Gpr35	2.68	1.33E-05	0.0042
Tmem50b	0.77	1.45E-05	0.0045
R3hnc1	-1.25	1.51E-05	0.0046
HistH3d	1.42	1.59E-05	0.0048
Slamf7	1.40	1.74E-05	0.0052
Scpp5	0.80	1.80E-05	0.0053
H3fb	-0.72	1.83E-05	0.0053
Rpl14	-0.94	1.87E-05	0.0053
Pdse2a	0.98	1.89E-05	0.0053
Adpgk	0.78	1.91E-05	0.0053
Tlratp3	-0.85	1.91E-05	0.0053
Lyl1	0.63	2.04E-05	0.0056
Ahnak	-0.81	2.14E-05	0.0058
Ear1	1.65	2.14E-05	0.0058
Parp14	0.74	2.22E-05	0.0060
Sifn5	0.82	2.26E-05	0.0060
Ebf1	2.06	2.39E-05	0.0062
Rps14	-0.70	2.40E-05	0.0062
Rhhd2	1.11	2.46E-05	0.0063
Dusp8	-2.31	2.71E-05	0.0068
Emt5	0.82	2.72E-05	0.0068
Cont2	0.65	2.88E-05	0.0072
Igll1	2.13	2.96E-05	0.0073
Tmem10	-0.76	3.08E-05	0.0075
Per1	-1.03	3.21E-05	0.0077
Olfrl1396	-6.14	3.63E-05	0.0087
Srafs	-0.73	3.72E-05	0.0088
Gpr128	0.67	3.77E-05	0.0088
Pou2af1	1.77	3.97E-05	0.0092
Neo1	3.34	4.11E-05	0.0093
Kcna2	4.11	4.12E-05	0.0093
Acvr1b	0.97	4.15E-05	0.0093
Wdr33os	-0.70	4.38E-05	0.0097
Fam46a	0.61	4.38E-05	0.0097
4930503L19Rik	-0.82	4.68E-05	0.0102
Gm12185	2.03	4.74E-05	0.0102
Abca13	1.21	4.74E-05	0.0102
Tnfrsf15	3.46	5.03E-05	0.0107
Stat1	0.63	6.11E-05	0.0127
G2e3	0.76	6.12E-05	0.0127
Foab	-0.99	6.13E-05	0.0127
Cp	1.72	6.22E-05	0.0128
Igfb6	0.75	6.52E-05	0.0132
Tifab	0.66	6.54E-05	0.0132
Cdkn1a	-0.89	7.11E-05	0.0147
Dmat2	0.74	7.41E-05	0.0147
Rps10	-0.80	7.90E-05	0.0156
Camp1	-0.94	7.99E-05	0.0156
Cers6	0.61	8.06E-05	0.0156
Zfp518b	0.87	8.20E-05	0.0158
Flx	0.87	8.36E-05	0.0160
Uoua4	0.59	8.46E-05	0.0160
Rasgef1b	-0.99	8.53E-05	0.0160
Atp8b4	0.68	8.76E-05	0.0163
1110038B12Rik	0.65	8.86E-05	0.0163
Crip1	-0.67	8.91E-05	0.0163
Sivn3	0.65	8.95E-05	0.0163
Rplk1	0.60	9.10E-05	0.0164
Tnfrsf12a	-1.35	9.13E-05	0.0164
S1pr3	0.93	9.35E-05	0.0167
Rrm2	0.58	9.50E-05	0.0168
Bhlhe40	-1.28	9.75E-05	0.0171
Rpl32	-0.74	9.80E-05	0.0174

dKO vs WT

Gene	Fold Change (Log2)	P value	FDR
Lym7	3.15	1.87E-28	4.03E-24
Wdfy1	2.23	2.89E-26	3.48E-22
Tifab	-2.79	2.36E-18	1.89E-14
Gm2a	-1.34	1.17E-15	7.01E-12
Foab	-1.31	5.48E-14	2.84E-10
Tec2d3	-1.65	7.32E-13	2.94E-09
Olfms5	-5.38	2.38E-12	8.19E-09
H2afy	-1.03	1.73E-10	5.19E-07
Rplp1	-1.16	3.82E-09	1.02E-05
Emp1	-2.06	1.93E-08	4.66E-05
Vps13c	0.90	4.14E-08	9.05E-05
G2e3	1.11	5.42E-08	0.0001
Crip1	-1.19	8.04E-08	0.0001
Ahnak	-0.90	1.71E-07	0.0003
Lyp1a1	3.23	2.28E-07	0.0004
Gdppp1	1.61	3.11E-07	0.0005
Ncam1	1.82	3.47E-07	0.0005
Skp2	0.93	5.06E-07	0.0007
4930433N12Rik	5.09	6.69E-07	0.0008
F830028O10Rik	1.03	6.74E-07	0.0008
Smm3	-1.02	9.77E-07	0.0011
Kf10	-0.93	1.11E-06	0.0012
Cdkn1a	-1.24	1.22E-06	0.0013
Klh42	1.13	1.91E-06	0.0019
Rplp2	-0.82	2.16E-06	0.0019
G53001106Rik	1.11	2.33E-06	0.0022
Perp	-1.77	3.22E-06	0.0029
Doc2	1.08	5.11E-06	0.0043
Zfp354a	4.73	5.29E-06	0.0043
Bhlhe40	-1.58	5.33E-06	0.0043
Sic2e2a	1.07	6.51E-06	0.0051
Rps15	-0.67	6.94E-06	0.0052
Kl4	-0.91	7.07E-06	0.0052
Lbc	-1.02	8.09E-06	0.0057
Kcnj2	1.26	9.09E-06	0.0062
Rpl37a	-0.78	9.39E-06	0.0062
Erd1	1.05	9.47E-06	0.0062
Vcam1	2.11	1.67E-05	0.0074
Clec4a3	6.07	1.48E-05	0.0089
Cernp1	-0.90	1.49E-05	0.0089
Rps26	-0.73	1.99E-05	0.0092
Sic31a2	1.01	1.99E-05	0.0094
Per1	-1.22	1.78E-05	0.0100
Rps14	-0.92	1.97E-05	0.0108
Vamp6	-0.93	2.02E-05	0.0108
Olfrl1396	-6.16	2.19E-05	0.0114
Tmem9	-0.95	2.32E-05	0.0119
R3hdm4	-0.77	2.39E-05	0.0120
Mtus1	1.29	2.65E-05	0.0129
Pdse3p3	0.66	2.67E-05	0.0134
Codc107	-1.46	2.74E-05	0.0129
Kl8	-0.77	2.87E-05	0.0134
Adpgk	0.85	3.06E-05	0.0134
Rpl33	-0.76	3.14E-05	0.0134
Rc3h2	0.76	3.14E-05	0.0134
Irgm1	1.13	3.14E-05	0.0134
Rasa2	0.99	3.18E-05	0.0134
Kcnj2	1.22	3.26E-05	0.0136
Aoah	-1.13	3.32E-05	0.0135
Stard6	1.50	3.42E-05	0.0137
Tgfp1	3.70	3.56E-05	0.0141
Igfb1	-1.20	4.17E-05	0.0152
Nrg1	3.13	3.73E-05	0.0143
Clec5a	1.69	4.05E-05	0.0152
Dusp16	-0.81	4.11E-05	0.0152
Th2	2.60	4.23E-05	0.0152
Cyp11a1	2.60	4.23E-05	0.0152
Emt5	0.90	4.29E-05	0.0152
Abca13	2.23	4.68E-05	0.0163
Tlil1	-0.68	5.30E-05	0.0162
Phk2a2a	0.82	5.49E-05	0.0186
Mit1	0.85	5.64E-05	0.0189
Pcdha8	5.70	5.73E-05	0.0189
Cmas1	-0.99	6.01E-05	0.0196
HistH3d	1.79	6.49E-05	0.0208
Rpl36	-0.82	6.74E-05	0.0213
Alm	0.79	7.17E-05	0.0224
Gpr128	0.78	7.60E-05	0.0232
Bpic	-1.01	7.62E-05	0.0232
Sifn9	0.83	7.87E-05	0.0237
Zfp16	0.90	8.17E-05	0

Exx	1.20	0.0001003	0.0174
Fndc7	3.31	0.0001023	0.0176
Rps3a1	-0.61	0.0001044	0.0177
Vwf	-0.68	0.0001045	0.0177
Atap1	0.86	0.0001057	0.0178
Car12	0.90	0.0001065	0.0178
Car1	-0.62	0.0001076	0.0178
Gpatch4	-0.86	0.0001077	0.0178
Gbp7	0.60	0.0001111	0.0181
Gm1821	-1.39	0.0001116	0.0181
Msp1	-0.59	0.0001178	0.0189
Ier2	-0.75	0.0001178	0.0189
Gdppg1	1.08	0.0001185	0.0189
Comm4	-0.88	0.0001196	0.0189
Zfp346	0.76	0.0001259	0.0197
Homez	0.93	0.0001259	0.0197
Rpl37a	-0.62	0.0001281	0.0199
Calr	0.67	0.0001292	0.0199
Gda	0.79	0.0001358	0.0208
Lmna	-1.04	0.0001408	0.0214
Rpl13	-0.62	0.0001451	0.0220
Vamp8	-0.67	0.0001458	0.0220
Finem	0.65	0.0001478	0.0220
Trc	1.60	0.0001486	0.0220
Neur3	-0.74	0.0001491	0.0220
Zbtb10	-0.84	0.0001502	0.0220
Fhb	0.56	0.0001631	0.0238
Ubs52	-0.80	0.0001677	0.0243
Chordc1	0.61	0.0001701	0.0245
Uqcrq	0.66	0.0001716	0.0246
Wdr65	3.21	0.0001738	0.0247
Gm6297	1.46	0.0001742	0.0247
Gm10451	3.10	0.0001762	0.0247
Tsc22d1	-0.56	0.0001766	0.0247
Far2	0.79	0.0001814	0.0252
Zfp780b	0.86	0.0001858	0.0257
Pik3c2a	0.69	0.000188	0.0258
Nav2	0.73	0.0001885	0.0258
Frit2	-0.58	0.0001894	0.0259
Hdc1	-0.80	0.0001916	0.0259
Dok	0.96	0.000195	0.0262
Prom1	1.16	0.0001956	0.0262
Zfp780	0.95	0.0002002	0.0266
Bag1	-0.67	0.0002045	0.0270
Chdh	0.56	0.0002056	0.0270
Il18bp	1.04	0.0002085	0.0273
Papss2	0.64	0.0002099	0.0273
Syr1	0.61	0.0002107	0.0273
Megf6	5.64	0.0002132	0.0274
Kcnj2	0.98	0.000217	0.0278
Pfms	-1.12	0.0002187	0.0278
Sgpl1	1.39	0.0002278	0.0288
Cd300e	5.49	0.0002295	0.0289
H2-Ab1	0.58	0.0002313	0.0290
Gm10248	-3.82	0.0002337	0.0291
Ubb	-1.54	0.0002374	0.0293
Rps5	-0.73	0.0002376	0.0293
Sic35a5	0.99	0.0002417	0.0296
Zscan29	0.67	0.0002426	0.0296
Cow11	-0.38	0.0002445	0.0297
Tet2	0.57	0.0002454	0.0297
Vcam1	1.23	0.0002485	0.0298
D230029D16Rik	0.64	0.0002487	0.0298
Spr1a	6.89	0.0002558	0.0299
Irfm2	-0.74	0.0002562	0.0304
Itripl2	0.66	0.0002621	0.0308
B230219D22Rik	0.59	0.0002627	0.0308
Zfp184	-0.62	0.0002699	0.0314
Rc3h2	0.62	0.00027	0.0314
A630072M18Rik	0.85	0.0002786	0.0322
Srz2	0.63	0.0002901	0.0331
E230008N13Rik	-1.67	0.0002912	0.0331
Ser	0.64	0.0002913	0.0331
Anp32b	-0.60	0.0002921	0.0331
Zdhc21	0.57	0.000293	0.0331
Srbp1	2.43	0.0002963	0.0333
Serinc4	1.22	0.0002978	0.0333
Rpl9	-0.60	0.000325	0.0362
Dyrk3	-0.68	0.0003265	0.0362
Zfp783	0.86	0.0003307	0.0364
Faim3	5.42	0.0003316	0.0364
Med23	0.61	0.000338	0.0370
Cyp2r1	1.59	0.0003403	0.0371
Gm15915	-0.66	0.0003486	0.0378
Rbm5	0.55	0.0003566	0.0386
Rint1	0.73	0.0003668	0.0394
Stat2	0.75	0.0003733	0.0399
Gbp5	0.92	0.0003755	0.0400
Cas7	0.92	0.0003856	0.0403
Zfp324	0.90	0.0003893	0.0411
Z810403A07Rik	0.60	0.000392	0.0412
Rpl36a	-0.57	0.0003952	0.0413
Zbtb40	0.73	0.0004021	0.0419
Sic5a3	0.69	0.0004061	0.0419
Sik1	-0.64	0.0004064	0.0419
Rassf4	0.59	0.0004076	0.0419
Hsp1	0.59	0.0004147	0.0425
Mtr	0.58	0.0004174	0.0426
Klf10	-0.71	0.0004232	0.0429
Rab4a	-0.76	0.0004248	0.0429
Trmpc	0.76	0.0004267	0.0429
BC055324	0.63	0.0004277	0.0429
Cybb	0.71	0.0004347	0.0434
Lis4h	0.54	0.0004409	0.0438
Rpl24	-0.57	0.000444	0.0440
Pfme2b	1.43	0.0004458	0.0444
Sic7a8	0.60	0.000455	0.0447
Csf2rb2	0.79	0.0004577	0.0448
Hhrs	5.86	0.0004609	0.0449
Hesr15b	0.63	0.0004707	0.0457
Met	0.62	0.0004735	0.0458
Rnt1	0.72	0.000477	0.0459
Lg4	1.56	0.0004843	0.0464
Lbp1	0.92	0.0004856	0.0464
Palb2	0.79	0.0004903	0.0465
A530099J19Rik	1.94	0.000492	0.0465
Lz2	1.79	0.0004924	0.0465
Srrm40	-0.59	0.0005215	0.0487
Ear6	1.83	0.0005219	0.0487
Lsm5	-0.91	0.000522	0.0487
Cdcb	0.59	0.0005295	0.0487
Neur1	0.72	0.000529	0.0487
Pla2g15	1.20	0.0005398	0.0498

Supplemental Table 2. Synergy Genes in dKO LK.

Gene	Relative Expression			SynergyEffect
	miR-146a KO vs WT	Tifab KO vs WT	dKO vs WT	
Dgkk	14.84	13.42	52.88	0.53
Ltf	3.64	5.85	16.93	0.56
Ngp	2.17	3.44	9.78	0.57
Cd300e	45.04	12.87	82.53	0.70
Lypd6	15.53	13.63	40.54	0.72
Mir677	14.84	11.06	28.55	0.91
Gpr84	5.64	4.66	10.30	1.00
Timm8a2	20.60	32.50	52.46	1.01
Sfrp1	14.56	30.72	41.74	1.08
Nrg1	5.93	3.57	8.74	1.09
Abca13	2.32	2.92	4.71	1.11
Gdpgp1	2.12	1.54	3.05	1.20
Neo1	10.16	6.63	13.69	1.23
Acpp	2.42	2.91	4.24	1.26
Ncam1	1.64	2.80	3.53	1.26
Spint1	31.74	12.74	32.83	1.35
Vcam1	2.35	3.63	4.33	1.38
Itga1	1.46	1.80	2.30	1.41
Gramd3	1.65	1.68	2.32	1.43
Stc2	5.78	6.02	8.20	1.44
Csf2ra	1.44	1.47	2.01	1.45
Stard5	1.70	1.54	2.22	1.46
Scgb3a1	12.37	21.75	22.95	1.49
Ifi204	2.67	3.12	3.89	1.49
Clca1	1.58	1.69	2.18	1.50
Kcnj2	1.97	1.69	2.39	1.53
Gbp8	9.97	8.33	11.92	1.53
Higd1b	12.37	11.46	15.37	1.55
Mlxipl	18.86	25.93	28.52	1.57
Slfn2	2.76	3.01	3.66	1.58
Papss2	1.56	1.64	2.02	1.58
Clec5a	2.59	2.70	3.22	1.64
Prkcb	2.69	2.21	2.95	1.66
Ceacam1	1.49	1.43	1.75	1.66

Supplemental Table 3. Del(5q) MDS/AML patient characteristics.

Disease	Risk	TIFAB deletion	miR-146a deletion
sAML	1	Yes	No
MDS	0	Yes	No
MDS	0	Yes	No
sAML	1	Yes	Yes
MDS	1	Yes	Yes
MDS	0	Yes	Yes
MDS	1	Yes	Yes
MDS	1	Yes	Yes
MDS	0	Yes	Yes

Risk: 1, high-risk; 0, low-risk
sAML, secondary AML.