

1 **Supplementary Information**

2 **Correlated production and consumption of chloromethane in the *Arabidopsis*** 3 ***thaliana* phyllosphere**

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5 Muhammad Farhan Ul Haque^{1‡}, Ludovic Besaury^{1§}, Thierry Nadalig¹, Françoise Bringel¹, Jérôme
6 Mutterer², Hubert Schaller² and Stéphane Vuilleumier^{1*}

7 8 **Authors and affiliations**

9 ¹ Université de Strasbourg, CNRS, GMGM UMR 7156, Department of Microbiology, Genomics and
10 the Environment, Strasbourg, France

11 ² Institut de Biologie Moléculaire des Plantes, UPR2357 CNRS, Strasbourg, France

12 [‡] Present address: School of Environmental Sciences, University of East Anglia, Norwich, UK

13 [§] Present address: Université de Reims Champagne Ardenne, INRA, FARE UMR A614, Reims,
14 France

15 16 *** Corresponding author:**

17 Stéphane Vuilleumier, Université de Strasbourg, Equipe Adaptations et Interactions Microbiennes
18 dans l'Environnement, Département Microorganismes, Génomes, Environnement, UMR 7156
19 UNISTRA - CNRS, 28 rue Goethe, F-67000 Strasbourg, France, phone: +33-3-68-85-2022; fax: +33-
20 3-68-85-1926; e-mail: vuilleumier@unistra.fr

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22 **Supplementary Information**

23 ***HOL1* genotyping of plants**

24 Wild-type (Col-0) and *HOL1* gene variants of *A. thaliana* were screened by performing two sets of
25 PCR reactions. The first PCR was performed using gene specific primer pair (hol1 and hol2, Table
26 S3) to confirm homozygosity of mutants. The second PCR was performed using primer hol1 and T-
27 DNA specific primer jmlb2 (Supplementary Table S1) to confirm T-DNA insertion. Each PCR
28 reaction mix (25 μ L) contained 2.5 μ L PCR buffer 10x (Tris-HCl 10 mM at pH 9.0, KCl 50 mM, MgCl₂
29 1.5 mM, Triton 0.1% and BSA 0.2 mg mL⁻¹), 0.4 μ M of each primer, 0.12 μ M of each dNTPs, 0.2 μ L
30 Taq DNA polymerase (5 U μ L⁻¹, Q biogene) and ultrapure filtered sterilised water. The PCR program
31 involved initial denaturation at 95°C for 3 mins, followed by 35 cycles of three steps (94°C for 1
32 min, 60°C for 1 min and 72°C for 45 seconds), and final elongation at 72°C for 3 mins. PCR products
33 were visualised under UV light using a Geldoc apparatus (Bio-Rad) after agarose gel
34 electrophoresis.

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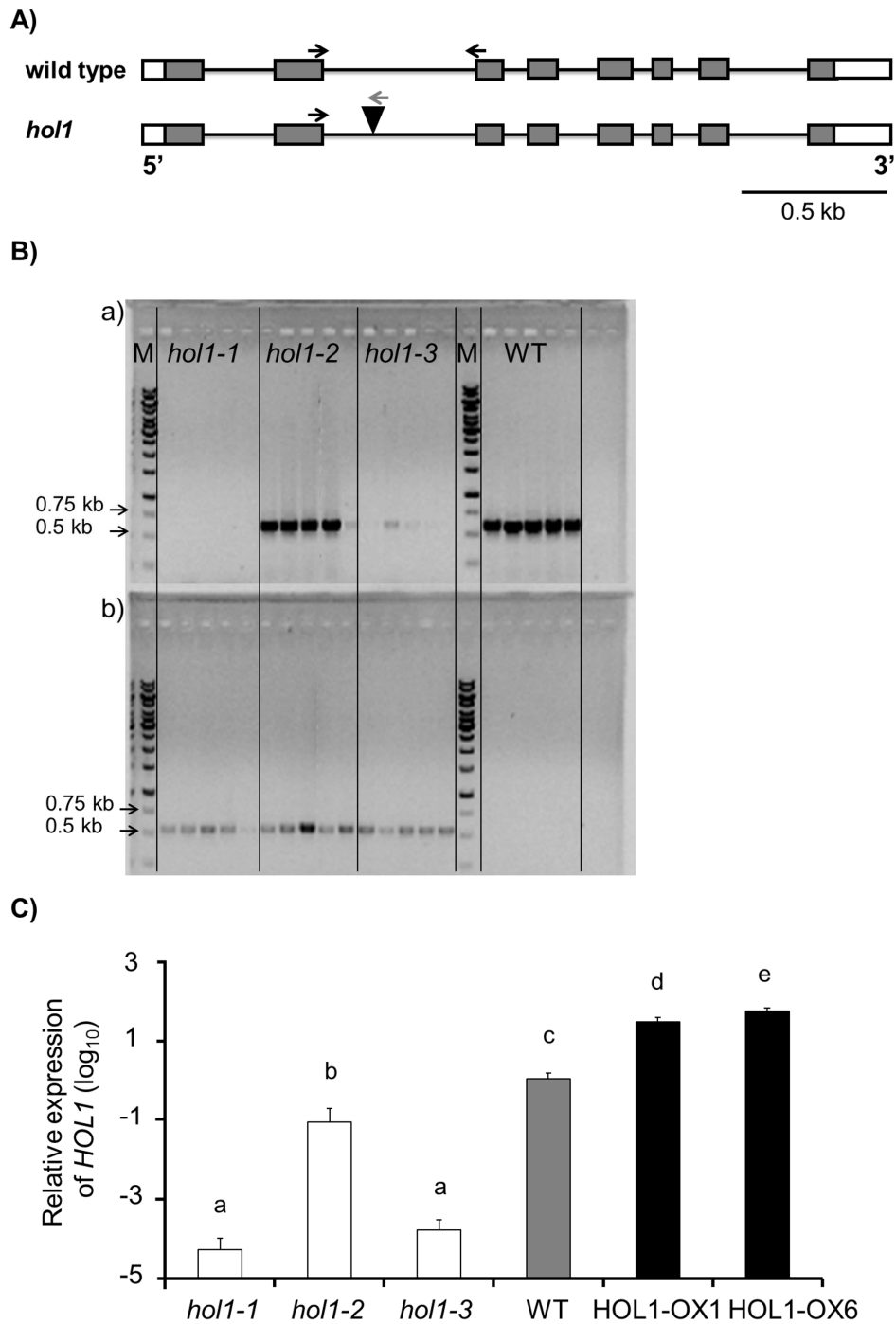
36 **Gene copy number calculation**

37 Calibration curves were obtained to determine the gene copy number of *cmuA* and 16S rDNA per
38 μ g of template in the phyllosphere using tenfold dilution series of standards (10 ng to 10 fg,
39 Supplementary Fig. S1) of *M. extorquens* CM4 genomic DNA. *M. extorquens* CM4 genome contain
40 5 copies of the 16S rRNA gene and 1 copy of the *cmuA* gene¹. Assuming average molecular masses
41 of 650 Daltons (Da) for a nucleotide pair of double-stranded DNA, gene copy number for a given
42 quantity of standard for the calibration curve was calculated by [quantity (ng) / L (bp) x 650 (Da) x
43 10⁻⁹] x 6.02 x 10²³ x [copy number of gene per genome] where L is the sequence length (6180732
44 bp for *M. extorquens* CM4 strain). To calculate *syfp2* copy number, plasmid pME8266 (6381 bp,
45 with 1 copy of *syfp2*)² was used as the standard. Threshold cycle (Ct) values were plotted against

46 copy number of serially diluted DNA standards, and the linear relationship obtained was used to
47 calculate the number of gene copies per μg of phyllosphere DNA template in qPCR.

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49 **Supplementary Figures**



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51 **Fig. S1. Screening of *HOL1* variants of *A. thaliana*.**

52 A) Organisation of gene *HOL1* in *A. thaliana* wild type (Col-0), based on locus data from the
 53 *Arabidopsis* Information Resource (<http://www.arabidopsis.org>). Gene *HOL1* comprises 8 exons
 54 (boxes) and 7 introns (lines) and encodes a 227- amino acids methyltransferase protein. The T-DNA

55 site of insertion in *hol1* mutant plants is shown (black triangle). Position of wild-type sequence
56 *hol1* and *hol2* primers (black) and T-DNA primer *jmlb2* (grey) used for genotyping are shown (see
57 Table S3 for primer sequences).

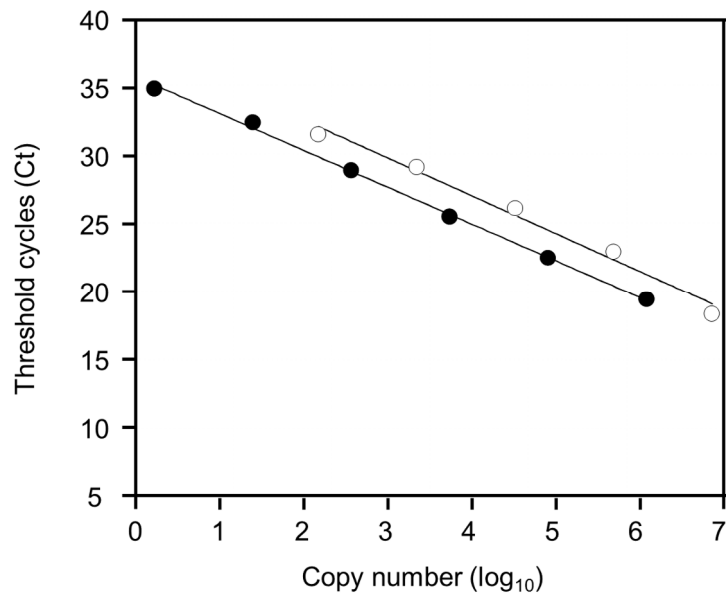
58 B) Genotyping of gene *HOL1* in *A. thaliana* wild type (Col-0) and three independent lines (*hol1-1*,
59 *hol1-2* and *hol1-3*) of *hol1* mutant plants using wild type primer pair *hol1-hol2* (a) and T-DNA
60 primer pair *hol1-jmlb2* (b). M, marker line.

61 C) Relative expression of gene *HOL1* in wild-type *A. thaliana* *hol1* mutant lines, and *HOL1*
62 overexpressor lines (*HOL1-OX1* and *HOL1-OX6*), as measured by qRT-PCR (see Materials and
63 Methods for calculation). Error bars represent the standard deviation of at least five biological
64 replicates, and small letters (a – e) show statistical significance at $p < 0.05$ by Student's t-test. The 2
65 *A. thaliana* lines with lowest (*hol1-1*) and highest (*HOL1-OX6*) expression of *HOL1* were selected for
66 subsequent experiments.

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71 **Fig. S2. Calibration curve for 16S rRNA and *cmuA* genes.** Serial tenfold dilutions of standards (10
72 ng – 1 fg) of *M. extorquens* CM4 total DNA were used as template for qPCR, and the results
73 expressed with DNA as gene copies against threshold cycle (Ct) values. Linear regression lines are
74 $y = 41.206 - 3.225x$ ($R=0.999$) and $y = 38.759 - 3.136x$ ($R=0.992$) for 16S rRNA (white) and *cmuA*
75 (black) respectively.

76 **Supplementary Tables**77 **Table S1. *cmuA* and 16S rRNA gene copies per mg of fresh leaf as measured by qPCR.**

	<i>cmuA</i>	16S rRNA
Leaf surface DNA		
<i>hol1</i>	13.5 (± 3.2)	7.0 (± 5.6) · 10 ⁵
WT	70.5 (± 27.6)	4.9 (± 0.4) · 10 ⁵
HOL1-OX	157.2 (± 43.0)	6.2 (± 2.4) · 10 ⁵
Leaf total DNA		
<i>hol1</i>	22.2 (± 15.9)	4.9 (± 0.7) · 10 ⁷
WT	85.8 (± 9.6)	6.9 (± 3.1) · 10 ⁷
HOL1-OX	164.7 (± 36.1)	3.2 (± 0.5) · 10 ⁷

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80 **Table S2. *syfp2* gene copies per mg of fresh leaves inoculated with bioreporter as measured by**
81 **qPCR.**

Plant type	<i>syfp2</i> copies per mg of fresh leaves
<i>hol1</i>	$5.3 (\pm 0.2) \cdot 10^5$
WT	$4.4 (\pm 0.2) \cdot 10^5$
HOL1-OX	$6.1 (\pm 0.3) \cdot 10^5$
not inoculated	$1.2 (\pm 0.6) \cdot 10^0$

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83 **Table S3. Primers used in the study**

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85	Target (purpose)		Primer sequence 5' – 3' ^a	Reference
86	<hr/>			
87	<i>Primers for bacterial genes</i>			
88	<i>cmuA</i>	cmuA802F	TTCAACGGCGAYATGTATCCYGG	3
89	(qPCR)	cmuA968R	CCRCCRTTRTAVCCVACYTC	4
90	16S rRNA	Bact1369F	CGGTGAATACGTTTCYCGG	5
91	(qPCR)	Prok1492R	GGWTACCTTGTTACGACTT	5
92	<i>syfp2</i>	MF34	ACAAGCAGAAGAACGGCATC	2
93	(qPCR)	MF35	GCTTGGACTGGTAGCTCAGG	2
94	16S rRNA	Gray28F	GAGTTTGATCNTGGCTCAG	6
95	(454)	Gray519R	GTNTTACNGCGGCKGCTG	7
96	<i>cmuA</i>	cmuA802F	TTCAACGGCGAYATGTATCCYGG	3
97	(454)	cmuA1244R	TABTCCATDATGGCYTCGAC	this study
98	<hr/>			
99	<i>Primers for plant genes</i>			
100	<i>HOL1</i>	hol1	GGAGTCAGTCTTCTTAGCTTACC	8
101	(genotyping)	hol2	GTGCGCTTTCGGAAATATCCAATCC	8
102	T-DNA	jmb12	TTGGGTGATGGTTCACGTAGTGGG	8
103	insertion			
104	<i>HOL1</i>	holqPCR_F	GAAAGCGCACTCGCGAAAGCTAAT	this study
105	(qPCR)	holqPCR_R	ATGCAGGTCTCATCTCCGGTTCAA	this study
106	<i>ACTIN2</i>	SS77	TTCAATGTCCCTGCCATGTATG	9
107	(qPCR)	SS78	AATACCGGTTGTACGACCAC	99

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109 ^a Equimolar mixtures at degenerate positions: N (G,A,T,C); V (G,A,C); D (G,A,T); B (G,T,C); R (G,A); K
 110 (G,T); W (A,T); Y (T,C).

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