



S2. Caveolin-1 is not involved in DC differentiation. (A),(B) DC percentage in spleen (A) or lymph nodes (B). WT and CAV1^{-/-} mice were sacrificed and spleen and inguinal lymph nodes were obtained and digested in RPMI plus collagenase IV (5 mg/ml) and DNase I (5 mg/ml) for 45 min. Then, cells were stained (30 min) with anti-CD11c, anti-MHC-II and Zombie Aqua, washed and fixed with a PBS/paraformaldehyde 4% solution and then evaluated by flow cytometry. The percentage of cells CD11c⁺MHC-II^{high}ZA^{neg} was assessed. Each dot represents one animal and the bar is the mean (n = 5 for spleen, n=3 for lymph node). (C) Skin from WT and CAV1^{-/-} mice skin was processed to obtain a single cell

suspension. Briefly, WT and *CAV1*^{-/-} mice were sacrificed, shaved and skin pieces of 1 cm² were digested using a solution containing collagenase IV (5 mg/ml) and DNase I (5 mg/ml) for 1 h. Then, cells were stained (30 min) with anti-CD45, anti-CD11c, anti-MHC-II and Zombie Aqua, washed and fixed with a PBS/paraformaldehyde 4% solution and then evaluated by flow cytometry. The percentage of cells CD45⁺CD11c⁺MHC-II^{high}ZA^{neg} was assessed. Each dot represents one animal and the bar is the mean (n = 3). **(D)** Bone marrow cells from WT or *CAV1*^{-/-} mice were cultured for 6 days in medium containing GM-CSF (20 ng/ml) to generate BM-DCs. Then, cell viability (Zombie Aqua) and CD11c⁺MHC-II⁺ was assessed by flow cytometry. No differences were observed in CD11c expression between BM-DCs from both mouse strains (n=5).