Supplementary Materials

Table S1 Primers used in this study

-F and -R suffixes indicate forward and reverse primers, respectively. NdeI and HindIII restrictions sites in primer sequences are underlined.

Primers	Sequence
chsH1-F	CGCGCATATGACGGTTGTTGGTGCTGTGCTG
chsH1-R	GGCC <u>AAGCTT</u> CTACGAATCCCTCATCGAAAG
chsH2-F	GGGC <u>CATATG</u> ACCGGGGTGAGCGACATTC
chsH2-R	CCGG <u>AAGCTT</u> CATTCGTCAGGCTCCCATGCG
chsH2∆duf35-R	CCGC <u>AAGCTT</u> CAACGCATCATAGCGTCGGGATCCAAATCG
duf35-F	CGGC <u>CATATG</u> CGTCCCTCGTCGTCGC
ltp2-F	GCG <u>CATATG</u> TTATCGGGTCAGGCGGCC
ltp2-R	GGCC <u>AAGCTT</u> AACCCAGGATCAGCCCGG
ltp2nostopcodon-R	AAGCTTACCCAGGATCAGCCCGGACGTAG



Figure S1 Coomassie blue stained 12% SDS-PAGE of fractions from purification of Nterminal His-tagged Ltp2 from Recombinant *R. jostii* RHA1 using Ni²⁺-NTA chromatography. BenchmarkTM protein ladder (Biorad) was used as a molecular weight marker. Samples loaded on gel are (1) insoluble cell pellet (2) crude lysate (3) flow though (4) 20 mM imidazole wash (5) 150 mM imidazole elution (6) 250 mM imidazole wash and (7) concentrated protein from 150 mM imidazole elution. Bands corresponding to the expected sizes of Ltp2 is shown on the right margin of the gel.



Figure S2 Coomassie blue stained 12% SDS-PAGE of fractions from purification of Cterminal His-tagged Ltp2 and untagged ChsH1-ChsH2ΔDUF35 from Recombinant *R. jostii* **RHA1 using Ni²⁺-NTA chromatography.** BenchmarkTM protein ladder (Biorad) was used as a molecular weight marker. Samples loaded on gel are (1) insoluble cell pellet (2) crude lysate (3) flow though (4) 20 mM imidazole wash (5) 150 mM imidazole elution (6) 250 mM imidazole wash and (7) concentrated protein from 150 mM imidazole elution. (8) Purified His-tagged Ltp2-ChsH1-ChsH2 complex loaded as control to show the expected positions of the various proteins on the gel.