

## Supporting Information

# Encapsulation of Autoinducer Sensing Reporter Bacteria in Reinforced Alginate-based Microbeads

*Ping Li<sup>1</sup>, Mareike Müller<sup>1,\*</sup>, Matthew Wook Chang<sup>2</sup>, Martin Frettlöh<sup>3</sup>, and Holger  
Schönherr<sup>1,\*</sup>*

1. Physical Chemistry I and Research Center of Micro and Nanochemistry and Engineering (*Cμ*), Department of Chemistry and Biology, University of Siegen, Adolf-Reichwein-Str. 2, 57076, Siegen, Germany.

2. Department of Biochemistry, Yong Loo Lin School of Medicine, National University of Singapore, 14 Medical Drive, Singapore 117599, Singapore & NUS Synthetic Biology for Clinical and Technological Innovation (SynCTI), Life Sciences Institute, National University of Singapore, 28 Medical Drive, Singapore 117456, Singapore.

3. Quh-Lab Food Safety, Siegener Str. 29, 57080, Siegen, Germany.

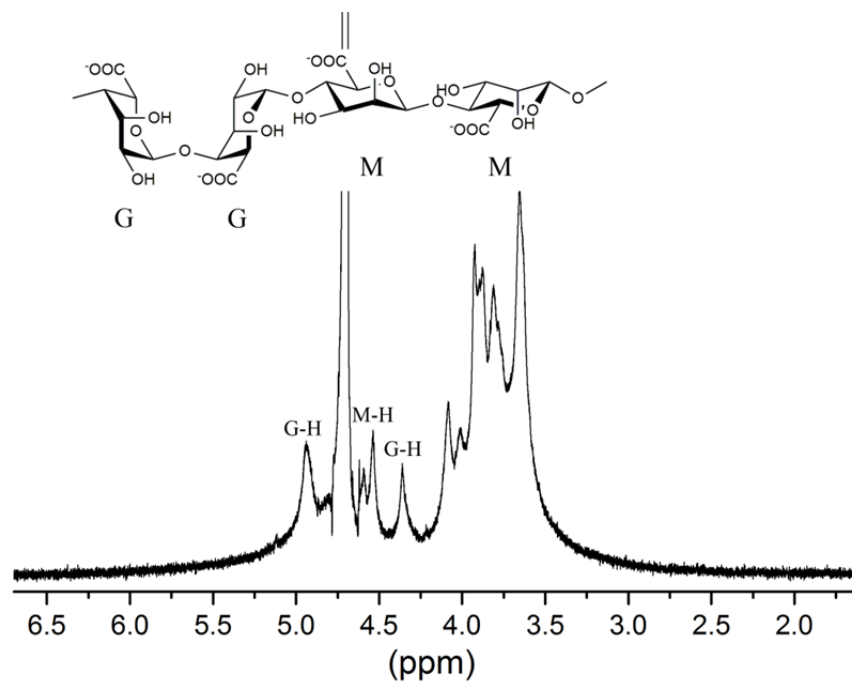
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\* Corresponding authors: Dr. M. Müller and Prof. Dr. H. Schönherr, Fax: +49(0)271 740 2805;  
E-mail: [m.mueller@chemie-bio.uni-siegen.de](mailto:m.mueller@chemie-bio.uni-siegen.de); [schoenherr@chemie.uni-siegen.de](mailto:schoenherr@chemie.uni-siegen.de)

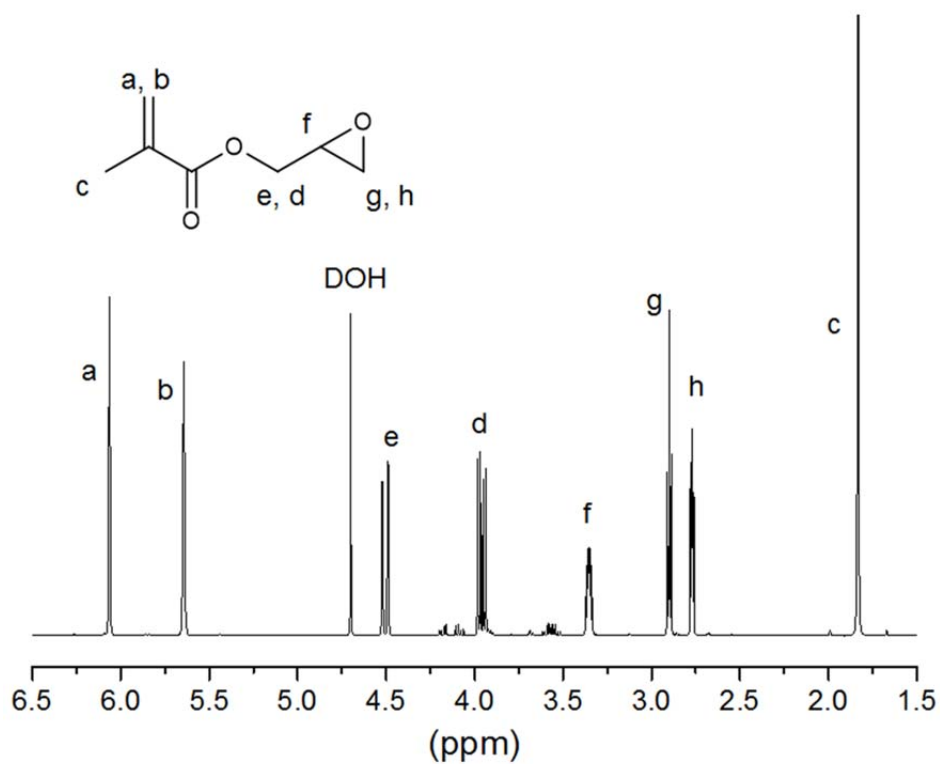
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## 1. $^1\text{H}$ NMR spectra

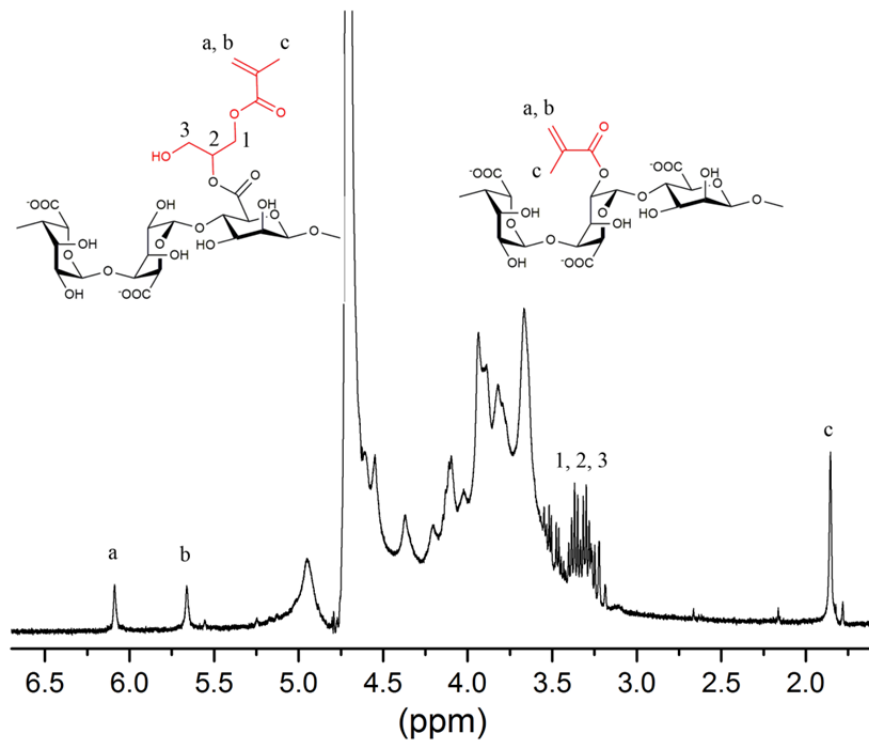
a)  $^1\text{H}$  NMR (400 MHz,  $\text{D}_2\text{O}$ ,  $\delta$ ):  $\delta = 4.95$  (s, G-H),  $4.55$  (s, M-H),  $4.36$  (s, G-H) ppm;



b)  $^1\text{H}$  NMR (400 MHz,  $\text{D}_2\text{O}$ ,  $\delta$ ):  $\delta = 6.07$  (dd, 1H,  $-\text{CH}=\text{CH}_2$ ),  $5.64$  (dd, 1H,  $-\text{CH}=\text{CH}_2$ ),  $4.49$  (m, H,  $\text{CH}_2$ ),  $3.95$  (m, H,  $\text{CH}_2$ ),  $3.35$  (m, H, CH),  $2.90$  (dd, H,  $\text{CH}_2$ ),  $2.77$  (dd, H,  $\text{CH}_2$ ),  $1.83$  (s, 3H,  $\text{CH}_3$ ) ppm;

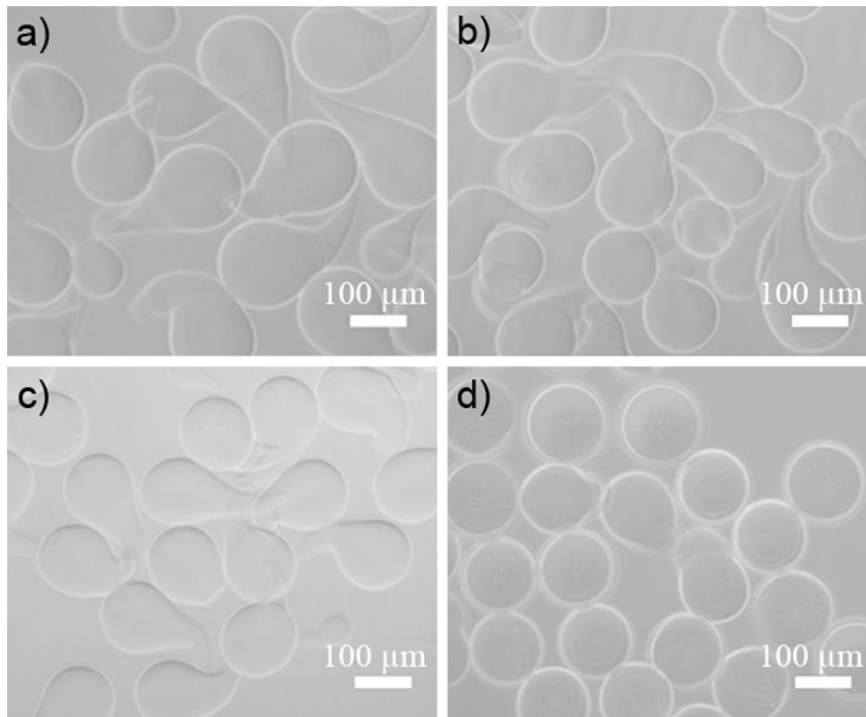


c)  $^1\text{H}$  NMR (400 MHz,  $\text{D}_2\text{O}$ ,  $\delta$ ):  $\delta = 6.09$  (dd, 1H,  $-\text{CH}=\text{CH}_2$ ),  $5.65$  (dd, 1H,  $-\text{CH}=\text{CH}_2$ ),  $\delta=4.94$  (s, 10H, G-H),  $4.54$  (s, M-H),  $4.34$  (s, 10H, G-H),  $4.49$  (m,  $\text{CH}_2$ ),  $3.95$  (m,  $\text{CH}_2$ ),  $3.35$  (m, CH),  $1.83$  (s, 3H,  $\text{CH}_3$ ) ppm;



**Figure S1.**  $^1\text{H}$ -NMR spectra of a) alginate, b) glycidyl methacrylate, and c) alginate-MA.

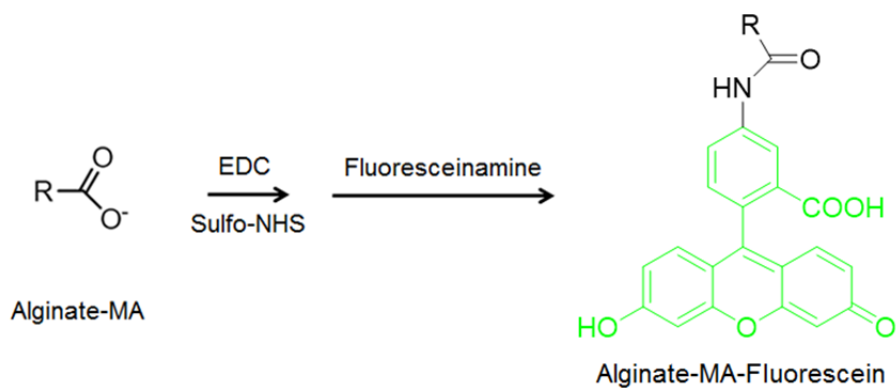
## 2. Alginate-MA beads obtained at different electric potential



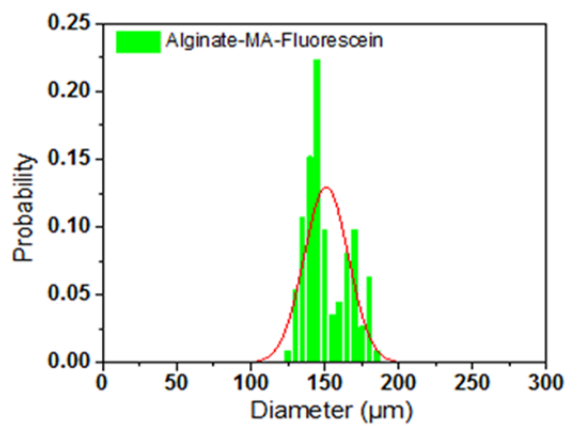
**Figure S2.** Optical microscopy images of 2 wt% alginate-MA beads formed under electric potentials of: a) 4.0 kV; b) 5.0 kV; c) 5.5 kV; and d) 6.0 kV, respectively, the beads were illuminated for 5 min with UV and were hardened 15 min in  $\text{CaCl}_2$  gelling solution.

### 3. Characterization of fluorescein-labelled Alginate-MA beads

a)



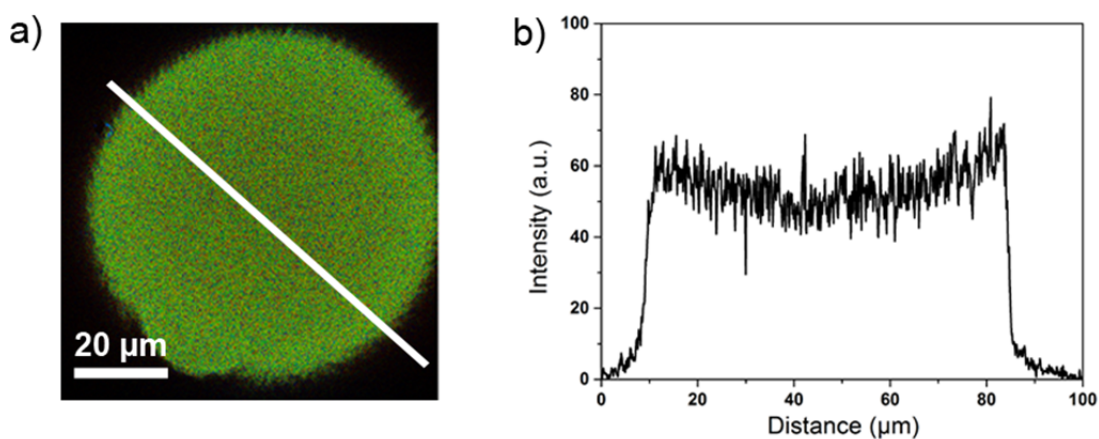
b)



**Figure S3.** a) Scheme of conjugation of alginate-MA conjugated with fluoresceinamine; b) Diameter distribution of fluorescein-labelled alginate-MA beads formed under the following conditions: 2 wt% fluorescein-labelled alginate-MA, 6.0 kV.

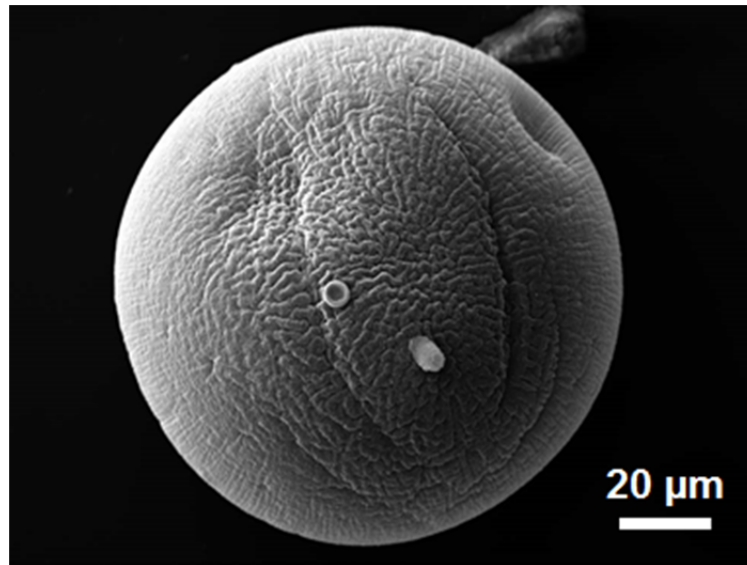
#### 4. Polymer distribution inside Fluorescein-labelled Alginate-MA beads

The concentration of fluorescein-labelled alginate-MA in the beads was investigated using a confocal laser scanning microscope (PicoQuant, Germany), comprising an OLYMPUS IX-71 frame (Olympus, Germany) and a commercial main optical unit (Microtime 200, PicoQuant, Germany). The measurements were done with a water immersion objective (60× magnification, Olympus, Germany). A pulsed diode laser (LDH-D-C-485, PicoQuant, Germany) with an excitation wavelength of 485 nm was used. The images (80  $\mu\text{m}$   $\times$  80  $\mu\text{m}$ ) were recorded with a resolution of 512 pixels  $\times$  512 pixels and the data were analyzed using software SymPhoTime (PicoQuant, Germany).



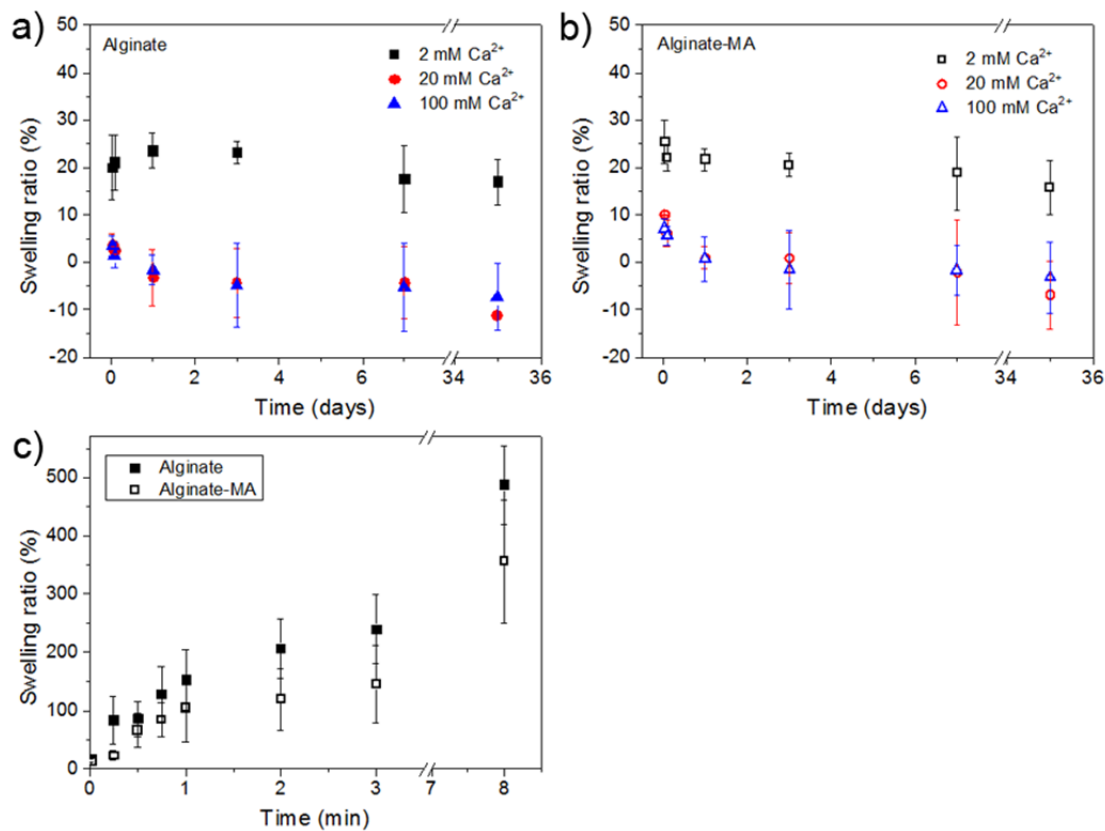
**Figure S4.** a) Confocal laser scanning microscopy image of 2 wt% alginate-MA beads formed at 6.0 kV, 5 min UV irradiation and 15 min hardening in  $\text{CaCl}_2$  gelling solution; b) Corresponding cross sectional fluorescence intensity plot of Figure S4a).

## 5. Scanning electron microscopy (SEM) image



**Figure S5.** SEM image of an alginate-MA bead (diameter: 100  $\mu\text{m}$ ), 2 wt% alginate-MA beads were formed at 6.0 kV, followed by 5 min UV irradiation and 15 min hardening. The size of beads in the hydrated state (before water/ethanol exchange) was  $184 \pm 6 \mu\text{m}$ .

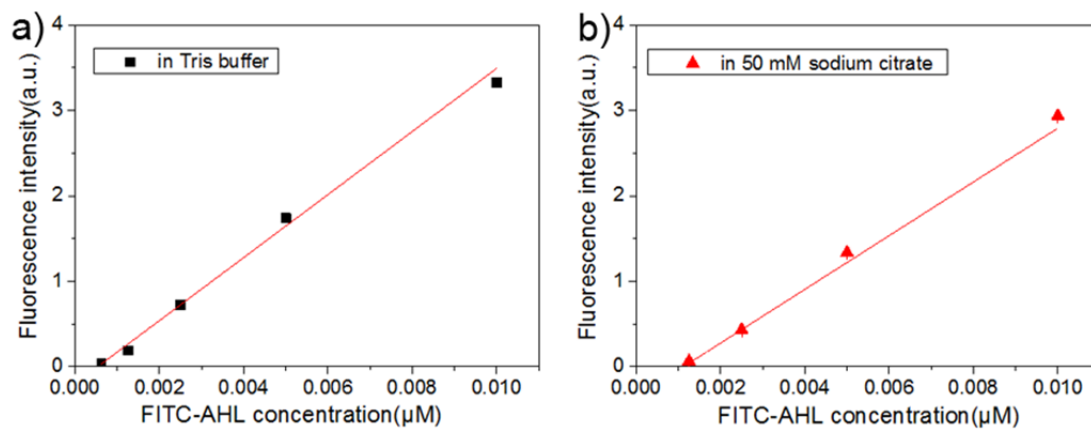
## 6. Hydrogel swelling



**Figure S6.** a) Swelling ratio of 2 wt% alginate beads in 2 mM, 20 mM, 100 mM  $\text{CaCl}_2$  solution vs. time; b) Swelling ratio of 2 wt% alginate-MA beads in 2 mM, 20 mM, 100 mM  $\text{CaCl}_2$  solution vs. time; c) Swelling ratio of 2 wt% alginate and 2 wt% alginate-MA beads in 50 mM sodium citrate solution vs. time; 2 wt% alginate beads were hardened in 100 mM  $\text{CaCl}_2$  solution for 15 min, 2 wt% alginate-MA beads were exposed to 5 min UV irradiation and hardened for 15 min.



## 7. FITC-AHL standard curves



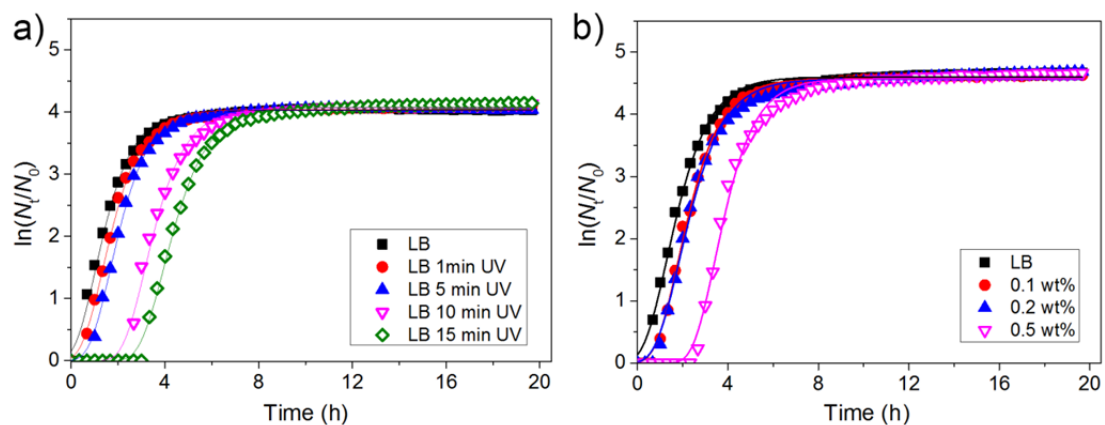
**Figure S7.** Standard curves and linear regression fit of fluorescence intensity vs. concentration of FITC-AHL in a) Tris buffer (10 mM, pH = 8.5) and b) 50 mM sodium citrate solution.

## 8. Bacteria growth curve and fitting

To investigate the influence of alginate, alginate-MA, initiator Irgacure 2959, and UV irradiation time on the bacteria viability, the proliferation of *Escherichia coli* TOP10 pTetR-LasR-pLuxR-GFP (*E. coli* pLuxR-GFP) in each solution was analyzed by measuring the absorbance at a wavelength of 600 nm ( $OD_{600}$ ) using a microplate reader. The absorbance of the culture media (without bacteria) was subtracted for each time point. The growth curves were characterized by plotting  $\ln(N_t/N_0)$  vs. time  $t$  and fitting the data using the Gompertz equation<sup>1</sup>:

$$\ln(N_t/N_0) = a \exp[-\exp(b - ct)] \quad (1)$$

From equation 1, the stationary phase bacteria population, lag time and maximum growth rate, generation time were calculated (according to Zwietering, M. H.; Jongenburger, I.; Rombouts, F. M.; van 't Riet, K., Modeling of the Bacterial Growth Curve. *Applied and Environmental Microbiology* **1990**, 56 (6), 1875-1881): the asymptotic value of a growth curve in the logarithmic form equals  $a$ ; the lag time  $\lambda = (b-1)/c$ ; the maximum specific growth rate  $\mu_m = ac/e$ ,  $e$  is Euler number; and the generation time equals  $(\ln 2)/\mu_m$ .



**Figure S8.** Plots of the logarithm of normalized bacteria number vs. time: a) *E. coli* pLuxR-GFP proliferation in LB medium after 0, 1, 5, 10, 15 min UV irradiation; b) *E. coli* pLuxR-GFP proliferation in LB medium after 5 min UV irradiation in presence of 0.1, 0.2, 0.5 wt% Irgacure 2959.

**Table S1:** Parameters of growth curves

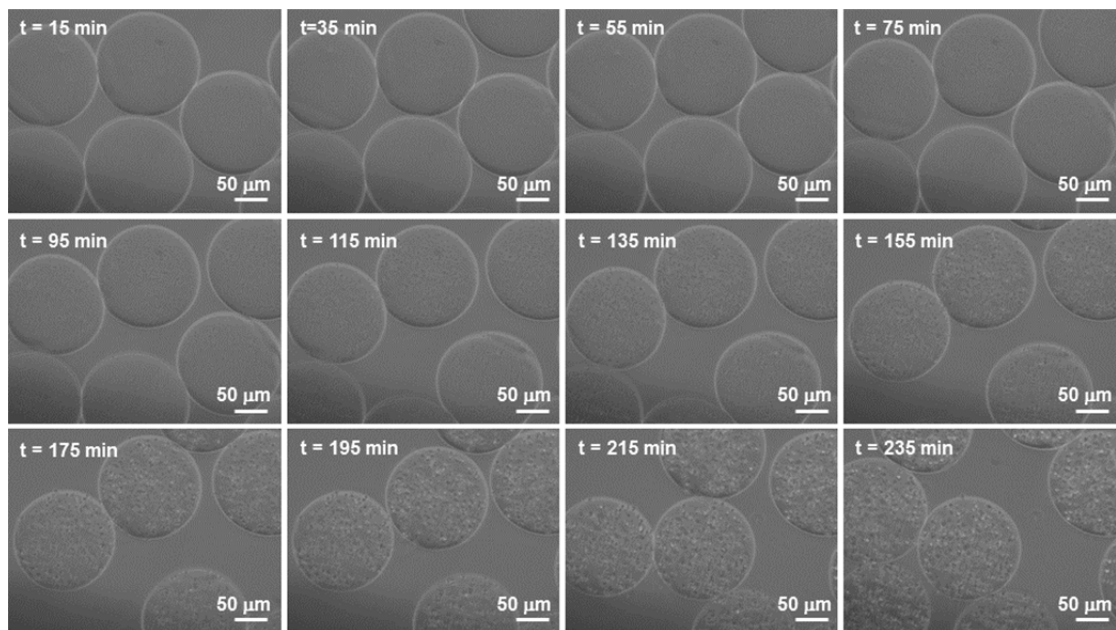
	Asymptotic value	Lag time (min)	Generation time (min)
LB medium	$4.7 \pm 0.6$	$11 \pm 9$	$25 \pm 3$
2% Alginate-MA/LB solution	$4.7 \pm 0.8$	$10 \pm 9$	$26 \pm 4$
LB, 1 min UV	$4.6 \pm 0.7$	$11 \pm 8$	$25 \pm 4$
LB, 5 min UV	$4.5 \pm 0.6$	$26 \pm 11$	$27 \pm 5$
LB, 10 min UV	$4.6 \pm 0.8$	$132 \pm 9$	$26 \pm 4$
LB, 15 min UV	$4.6 \pm 0.5$	$174 \pm 10$	$26 \pm 4$
LB, 5 min UV, 0.1 wt% Irgacure 2959	$4.6 \pm 0.6$	$47 \pm 15$	$25 \pm 5$
LB, 5 min UV, 0.2 wt% Irgacure 2959	$4.5 \pm 0.8$	$47 \pm 17$	$26 \pm 4$
LB, 5 min UV, 0.5 wt% Irgacure 2959	$4.7 \pm 0.7$	$147 \pm 14$	$27 \pm 4$

## 9. Encapsulation efficiency

**Table S2.** Encapsulation efficiency of *E. coli* pLuxR-GFP

Bacteria count in LB broth (CFU/mL)	Bacteria count in hydrogel beads (CFU/cm <sup>3</sup> )	Encapsulation efficiency
$7.0 \times 10^7$	$5.3 \times 10^7$	76%
$1.3 \times 10^8$	$1.1 \times 10^8$	82%
$1.5 \times 10^8$	$1.3 \times 10^8$	89%
$1.9 \times 10^8$	$1.1 \times 10^8$	56%

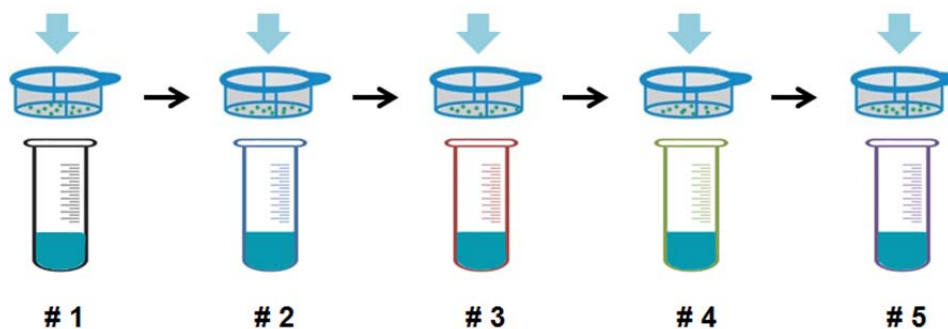
## 10. Bacteria proliferation inside beads



**Figure S10.** Time-lapse optical microscopy images of alginate-MA beads incubated in LB medium at 37°C. The increase amount of black dots indicates the proliferation of the entrapped *E. coli* pLuxR-GFP, the time interval between subsequent images is 20 min and the last picture was taken after around 240 min incubation.

## 11. Bacteria leakage test

Before bacteria leakage experiment, the beads were thoroughly washed with an excess volume (25 mL each time) of CaCl<sub>2</sub> solution to remove free bacteria attached on the bead surface, as shown in Scheme S1.



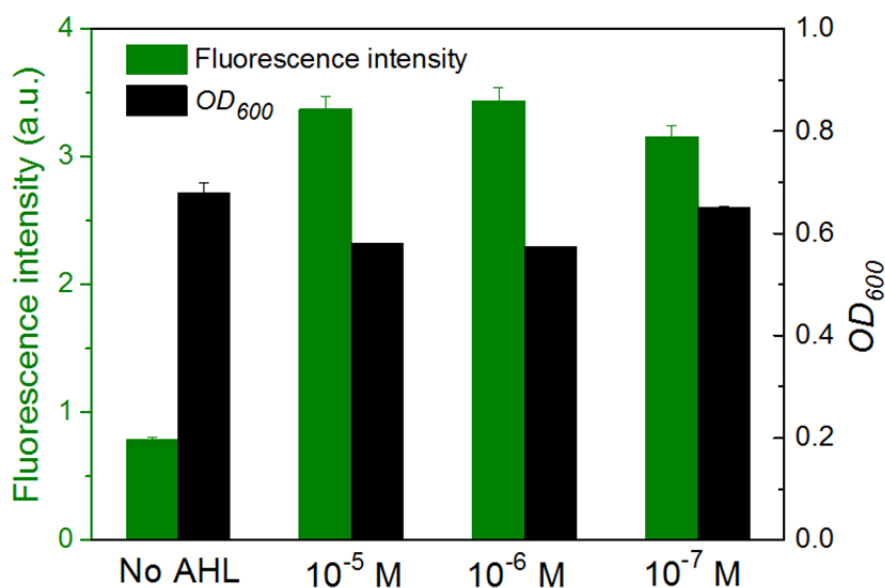
**Scheme S1.** Removal of free bacteria attached on the bead surface.

The number of bacteria in the washing solution is summarized in Table S3.

Table S3. The number of *E. coli* pLuxR-GFP in the washing solution.

Washing time / Sample name	Alginate beads washing solution CFU/mL	Alginate-MA beads washing solution CFU/mL
# 1	22	17
# 2	9	8
# 3	4	2
# 4	0	0
# 5	0	0

## 12. Fluorescence intensity and $OD_{600}$



**Figure S11.** Fluorescence intensity and  $OD_{600}$  of *E. coli* pLuxR-GFP encapsulated in alginate-MA beads at  $1.0 \times 10^{-5}$ ,  $1.0 \times 10^{-6}$ ,  $1.0 \times 10^{-7}$  mol/L 3OC<sub>12</sub>HSL after 4 h incubation. Error bars: standard deviation of three technical replicates from one exemplary data set out of a three times repeated experiment (biological replicate).

Reference:

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1 Zwietering, M. H.; Jongenburger, I.; Rombouts, F. M.; van 't Riet, K. *Modeling of the Bacterial Growth Curve. Appl. Environ. Microbiol.* **1990**, *56*, 1875-1881.