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## **Supplemental Information**

### **Visualizing the Assembly Pathway**

#### **of Nucleolar Pre-60S Ribosomes**

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**Table S1. Plasmids used in this Study. Related to STAR methods plasmid constructs and yeast strains**

<b>Name</b>	<b>Relevant information</b>	<b>Source</b>
<b>pFA6a-TAP-klURA3</b>	CBP-TEVcleavage site-ProtA, <i>TADH1</i> , for genomic C-terminal tagging	This study
<b>pFA6a-Flag-natNT2</b>	Flag, <i>TCYC1</i> , for genomic C-terminal tagging	This study
<b>pnatNT2 <i>P<sub>YTM1</sub></i> Flag</b>	<i>P<sub>YTM1</sub></i> , Flag tag, for genomic N-terminal tagging	This study
<b>YCplac111-TAP-Flag-<i>YTM1</i></b>	CEN, <i>LEU2</i> , <i>PYTM1</i> , <i>TADH1</i> , N-term. ProtA-TEVcleavage site-CBP-Flag tag (Thoms et al., 2016)	
<b>YCplac111-TAP-Flag-<i>ytm1</i> ΔUBL</b>	CEN, <i>LEU2</i> , <i>PYTM1</i> , <i>TADH1</i> , N-term. ProtA-TEVcleavage site-CBP-Flag tag	This study
<b>YCplac111-TAP-Flag-<i>ytm1</i> E80A</b>	CEN, <i>LEU2</i> , <i>PYTM1</i> , <i>TADH1</i> , N-term. ProtA-TEVcleavage site-CBP-Flag tag	This study
<b>YCplac111-pA-TEV-(His)<sub>6</sub>-<i>RPF1</i></b>	CEN, <i>LEU2</i> , <i>PNOP53</i> , <i>TADH1</i> , N-term. ProtA-TEVcleavage site-(His) <sub>6</sub> tag	This study
<b>pET15b-<i>CtRrp1</i>-(His)<sub>6</sub> *</b>	Amp <sup>r</sup> , T7 promoter, <i>lac</i> operator	(Bassler et al., 2017)
<b>pET15b-<i>Nsa1</i>-(His)<sub>6</sub> *</b>	Amp <sup>r</sup> , T7 promoter, <i>lac</i> operator	This study

E.coli expression plasmids are marked with asterisk.

**Table S2. Yeast Strains used in this Study. Related to STAR methods plasmid constructs and yeast strains**

Name	Relevant genotype	Source
<b>W303</b>	wild-type, MAT	(Thomas and Rothstein, 1989)
<b>DS1-2b *</b>	wild-type	(Nissan et al., 2002)
<b>NSA1-TAP Flag-YTM1 *</b>	<i>NSA1-TAP::klURA3, P<sub>Flag-Ytm1</sub>-YTM1::natNT2</i>	This study
<b>RIX1-TAP RPF2-Flag *</b>	<i>RIX1-TAP::TRP1, RPF2-Flag::natNT2</i>	This study
<b>BRX1-HTpA</b>	<i>BRX1-HTpA::His3MX4</i>	This study
<b>pA-TEV-(HIS)<sub>6</sub>-RPF1</b>	<i>rpf1::HIS3MX4, YCplac111 pA-TEV-(HIS)<sub>6</sub>-RPF1</i>	This study

Strains marked with an asterisk were derived from DS1-2b (Nissan et al., 2002)

**Table S3: Data collection and refinement statistics for ScNsa1. Related to STAR methods protein production, crystallization and structure determination**

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<b>Data collection</b>	
Wavelength (Å)	1.0
Space group	P 21 21 21
Cell dimensions	
a, b, c (Å)	54.72 81.1 87.57
$\alpha$ , $\beta$ , $\gamma$ (°)	90, 90, 90
Resolution (Å)	59.5 - 2.4 (2.49 - 2.4)
$R_{\text{merge}}$	0.04384(0.1126)
$I/\sigma(I)$	34.78 (14.02)
Reflections total	111874 (8692)
Reflections unique	15750 (1543)
Multiplicity	7.1 (5.6)
<b>Refinement</b>	
$R_{\text{work}}$	0.1592 (0.1724)
$R_{\text{free}}$	0.2027 (0.2799)
No. atoms	
Protein	2955
Water	194
B-factors	
Protein	20.70
Water	22.80
R.m.s deviations	
Bond lengths (Å)	0.011
Bond angles (°)	1.41
Ramachandran plot	
Most favoured (%)	97
Disallowed	0.55

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**Table S4: Data collection and refinement statistics for CtRrp1. Related to STAR methods protein production, crystallization and structure determination**

	Native 1 (C2)	Native 2 (P6 <sub>3</sub> 22)	Pt-derivative
<b>Data collection</b>			
Wavelength (Å)	1.03323		1.07156
Space group	C2	P6 <sub>3</sub> 22	P6 <sub>3</sub> 22
Cell dimensions			
a, b, c (Å)	131.72, 74.67, 68.79	124.26, 124.26, 124.79	214.75, 214.75, 125.95
α, β, γ (°)	90, 90.52, 90	90, 90, 120	90, 90, 120
Resolution (Å)	47.36 – 2.65 (2.74 – 2.65)	49.22 – 2.24 (2.32 – 2.24)	47.72 – 3.3 (3.41 – 3.3)
<i>R</i> <sub>pim</sub>	0.093 (0.3138)	0.021 (0.3559)	0.114 (1.935)
/σ( <i>I</i> )	6.88 (1.67)	24.62 (2.16)	12.61 (1.18)
Reflections total	219854 (17947)	1632833 (166862)	1010978 (96718)
Reflections unique	19400 (1827)	80956 (7988)	26210 (2572)
Multiplicity	11.3 (9.8)	20.2 (20.9)	38.6 (37.6)
<b>Refinement</b>			
<i>R</i> <sub>work</sub>	0.2052 (0.2967)	0.1836 (0.2855)	
<i>R</i> <sub>free</sub>	0.2342 (0.3679)	0.2090 (0.2918)	
No. atoms			
Protein	3746	3723	
Water	117	212	
B-factors			
Protein	40.42	69.65	
Water	38.82	68.03	
R.m.s deviations			
Bond lengths (Å)	0.003	0.003	
Bond angles (°)	0.52	0.68	
Ramachandran plot			
Most favoured (%)	98.66	98.66	
Disallowed	0.45	0.45	

**Table S5. Cryo-EM model and refinement statistics. Related to STAR methods model building and refinements**

	State C	State E
<b>Data collection</b>		
Particles	157,184	115529
Pixel size (Å)	1.084	1.084
Defocus range (µm)	0.9 – 3.5	0.9 – 3.5
Voltage (kV)	300	300
Electron dose (e <sup>-</sup> Å <sup>-2</sup> )	27	27
<b>Model Refinement</b>		
<b>Model composition</b>		
Non-hydrogen atoms	88243	135041
Protein residues	5800	9709
RNA bases	1943	2680
<b>Refinement</b>		
Resolution for refinement (Å)	3.6	3.3
Map sharpening B-factor (Å <sup>2</sup> )	-144	-123
Average B-factor (Å <sup>2</sup> )	174.8	101.1
FSC <sub>average</sub>	0.845	0.805
<b>R.m.s. deviations</b>		
Bond lengths (Å)	0.0129	0.0068
Bond angles (°)	1.35	1.17
<b>Validation &amp; Statistics</b>		
<b>Validation</b>		
Molprobit score	2.86	2.4
Clashscore, all atoms	8.08	6.53
Good rotameres (%)	89.06	95.29
<b>Ramachandran Plot</b>		
Favored (%)	86.66	90.64
Outliers (%)	3.56	1.33
<b>Validation (RNA)</b>		
Correct sugar puckers (%)	97.1	97.1
Good backbone conformations (%)	72.1	66.7