2 Supplementary Figure S1:



Supplementary Figure S1: Phenotype of SF200 (ΔlpxR9 ΔpagL7 ΔpagP8 ΔaroA ΔydiV
ΔfliF). (A) Scanning electron microscopy of SF200. ΔfliF results in a non-flagellated
phenotype. White arrows indicate putative OMV formation; insert showing OMV formation
at higher magnification on the bacterial surface. (B) Schematic representation of the Lipid A
molecule. Left: Wt Salmonella is able to modify the Lipid A structure by means of pagP,
pagL and lpxR resulting in heterogeneous mixtures of Lipid A molecules. Right: Deletion of
such genes resulted in the homogeneous hexa-acylated structure of SF200.

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Supplementary Figure S2: Effect of UK-1 and SF200 ($\Delta lpxR9 \ \Delta pagL7 \ \Delta pagP8 \ \Delta aroA$ $\Delta ydiV \ \Delta fliF$) immunization on mouse survival upon secondary infection. Mice were pretreated with PBS (naïve), heat-inactivated UK-1 (UK-1 immunized) or heat-inactivated SF200 (SF200 immunized). Pre-exposed mice were intravenously infected with 10¹ UK-1 (**A**) and 5*10⁶ SF200 (**B**). Mouse survival was monitored for 14 days. Uninfected mice served as control. Displayed are values of mean ± SD of four replicates in each group.



Supplementary Figure S3: TNF- α induction of *Salmonella* in naïve and immunized mice upon infection. Naïve and immunized CT26 tumor-bearing mice were infected intravenously and intratumorally with 5*10⁶ SF200 ($\Delta lpxR9 \ \Delta pagL7 \ \Delta pagP8 \ \Delta aroA \ \Delta ydiV \ \Delta fliF$). TNF- α levels were measured in sera of CT26 tumor bearing mice isolated 1.5 h upon infection with SF200. Displayed are values of mean \pm SD. Results are representative of two independent experiments with six replicates in each group. *, p<0.05; ***, p<0.001.

28 Supplementary Figure S4:



Supplementary Figure S4: Spleen phenotype of naïve and immunized mice upon infection
with Salmonella. Naïve and immunized CT26 tumor-bearing mice were infected
intravenously 5*10⁶ SF200 (ΔlpxR9 ΔpagL7 ΔpagP8 ΔaroA ΔydiV ΔfliF) (A) or SL7207 (B).
Spleen weight was measured with a scale 6 dpi and used as indicator for splenomegaly.
Displayed are values of mean ± SD. Results are representative of two independent
experiments with six replicates in each group. *, p<0.05; **, p<0.01.

37 Supplementary Figure S5:



39 Supplementary Figure S5: Tumor colonization of naïve and immunized mice upon 40 infection with *Salmonella*. Naïve and immunized CT26 tumor-bearing mice were infected 41 intratumorally with $5*10^6$ SF200 ($\Delta lpxR9 \ \Delta pagL7 \ \Delta pagP8 \ \Delta aroA \ \Delta ydiV \ \Delta fliF$). Total CFU 42 values were determined by serial plating at 4 dpi. Displayed are values of mean \pm SD. Results 43 are representative of two independent experiments with six replicates in each group.



45 Supplementary Figure S6:

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Supplementary Figure S6: Pre-exposure reduces the formation of necrosis in the early 47 stages of infection upon intratumoral infection with Salmonella. CT26 tumor-bearing 48 mice were infected with $5*10^6$ SF200 ($\Delta lpxR9 \ \Delta pagL7 \ \Delta pagP8 \ \Delta aroA \ \Delta ydiV \ \Delta fliF$) via 49 50 intratumoral infection. 48 hpi, tumors were isolated and prepared for immune histochemical staining. Immunized mice are less prone to necrosis formation and hypoxia. Dispersion of 51 52 salmonellae in and beyond necrotic center, and presence of neutrophils in immediate proximity to the salmonellae was only clearly visible in naïve mice. "N" denotes areas 53 54 necrosis. Hypoxia was stained with antibodies against metabolites of pimonidazole-HCl, otherwise administered i.v. 30 mins prior to isolation. Myeloperoxidase (MPO) denotes 55 56 presence of neutrophilic granulocytes, and Salmonella was stained using a specific antibody. 57 Differential staining was performed on consecutive sections. Scale bar corresponds to 100 58 µm. Images representative of at least 3 replicates are displayed.

60 Supplementary Figure S7:



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62 Supplementary Figure S7: Pre-exposure reduces the formation of necrosis in the early stages of infection upon intravenous infection with Salmonella. CT26 tumor-bearing mice 63 were infected with $5*10^6$ SF200 ($\Delta lpxR9 \Delta pagL7 \Delta pagP8 \Delta aroA \Delta y diV \Delta fliF$) via intravenous 64 infection. 24 hpi, tumors were isolated and prepared for immune histochemical staining. 65 66 Immunized mice are less prone to necrosis formation and hypoxia. Dispersion of salmonellae in and beyond necrotic center, and presence of neutrophils in immediate proximity to the 67 salmonellae was only clearly visible in naïve mice. "N" denotes areas necrosis. Hypoxia was 68 stained with antibodies against metabolites of pimonidazole-HCl, otherwise administered i.v. 69 70 30 mins prior to isolation. Myeloperoxidase (MPO) denotes presence of neutrophilic 71 granulocytes, and Salmonella was stained using a specific antibody. Differential staining was performed on consecutive sections. Scale bar corresponds to 100 µm. Images representative 72 of at least 3 replicates are displayed. 73

75 Supplementary Figure S8:



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Supplementary Figure S8: Pre-exposure reduces the formation of necrosis in the early 77 stages of infection upon intratumoral infection with Salmonella. CT26 tumor-bearing 78 mice were infected with $5*10^6$ SF200 ($\Delta lpxR9 \ \Delta pagL7 \ \Delta pagP8 \ \Delta aroA \ \Delta ydiV \ \Delta fliF$) via 79 intratumoral infection. 24 hpi, tumors were isolated and prepared for immune histochemical 80 staining. Immunized mice are less prone to necrosis formation and hypoxia. Dispersion of 81 salmonellae in and beyond necrotic center, and presence of neutrophils in immediate 82 proximity to the salmonellae was only clearly visible in naïve mice. "N" denotes areas 83 necrosis. Hypoxia was stained with antibodies against metabolites of pimonidazole-HCl, 84 otherwise administered i.v. 30 mins prior to isolation. Myeloperoxidase (MPO) denotes 85 presence of neutrophilic granulocytes, and Salmonella was stained using a specific antibody. 86 Differential staining was performed on consecutive sections. Scale bar corresponds to 100 87 μm. Images representative of at least 3 replicates are displayed. 88

90 Supplementary Figure S9:

O CT26 〔) RenCa



naïve mouse

Re-challenge

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Supplementary Figure S9: Re-challenge (2°) experiment with CT26. Upon successful CT26 tumor clearance in response to SF200 ($\Delta lpxR9 \ \Delta pagL7 \ \Delta pagP8 \ \Delta aroA \ \Delta ydiV \ \Delta fliF$), mice were re-inoculated with CT26 (2°). Concurrent primary inoculation (1°) of RenCa and inoculation on naïve mice served as control. Pictures display tumors 14 days post inoculation.





Supplementary Figure S10: Cytokine, chemokine and growth factor detection in supernatants, sera and lysates upon infection with SF200 and Wt. Cytokine, chemokine and growth factor concentrations in supernatants of (A) 264.7 RAW macrophages cells (6 hpi with MOI 10) and (B) sera (1.5 hpi, 6 hpi and 24 hpi; infection: $5*10^6$ SF200 or $5*10^6$ Wt) were quantified. The cytokine levels were analyzed using a Bio-PlexProTM kit and compared to Wt infections. Displayed are values of mean \pm SD of four replicates in each group. Uninfected macrophages and mice served as control.

Supplementary Table

106 Supplementary Table S1: Bacterial strains and plasmids used in this study

Strain	Description	Source	Ref.
Salmonella Typhimurium strains			
SL7207	hisG ⁻ , ∆aroA	Lab stock	(1)
SF200	ΔlpxR9 ΔpagL7 ΔpagP8 ΔaroA ΔydiV ΔfliF	This study	-
E. coli strains			
Symbioflor-2	Escherichia coli Symbioflor-2	SymbioPharm	-
	(G1/2, G3/10, G4/9, G5, G6/7 and G8, pooled 1:1)		

107 1. Hoiseth SK and Stocker BA. Aromatic-dependent *Salmomella* Typhimurium are non-virulent and effective as

108 live vaccines. Nature. 1981; 291:238-9.