

Enhanced YAP expression leads to EGFR TKI resistance in lung adenocarcinomas

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Supplementary information

Material and Methods

Chemicals

Verteporfin and fluvastatin were purchased from Sigma (St. Louis, MO, USA). Afatinib, gefitinib and dasatinib (Selleckchem, Houston, TX, USA) 0.001 to 10 μ M were used to investigate cell viability. Verteporfin, fluvastatin, afatinib, gefitinib or dasatinib were all dissolved in dimethyl sulfoxide (DMSO) with a final concentration of DMSO < 0.1 %. The vehicle control group contained 0.1 % DMSO.

shRNA expression

Lentivirus containing short hairpin RNAs (shRNAs) expressed in a lentiviral vector (pLKO.1-puro) were generated in 293 T cells. Various pLKO plasmids to knockdown YAP, and scrambled control were provided by National RNAi Core Facility of Academia Sinica, Taipei, Taiwan. For lentivirus production, 293T cells in 6-cm dish were transfected with 2.5 μ g pLKO.1-puro lentiviral vectors expressing different shRNAs along with 0.25 μ g of envelope plasmid pVSVg and 2.5 μ g of packaging plasmid pCMV Δ R8.91. Virus was collected 48 h after transfection. To prepare various knockdown cells, HCC827^{GR}, HCC827^{AR}, H1975, or H1975^{AR} cells were infected with lentivirus for 24 h, and followed with 2 μ g/ml puromycin selection. The sequences of the lentiviral YAP shRNAs were CAGGTGATACTATCAACCAAA and CCCAGTTAAATGTTCCACCAAT.

Immunoblotting

Cells were lysed using RIPA buffer with the addition of proteases and phosphatases inhibitors (Roche, Indianapolis, IN, USA). Quantified lysates were separated on an SDS polyacrylamide gel. The blots were probed with the indicated antibodies as follows: EGFR, phospho-EGFR (Y1068), YAP (all from Cell Signaling, Beverly, MA, USA); and actin, GAPDH (Sigma).

Quantitative RT-PCR

RNA was harvested from cells using Trizol. Complementary DNA was prepared using Transcriptor Reverse Transcriptase (Roche). Quantitative PCR was performed using SYBR Green (Applied Biosystems, Foster City, CA, USA). The primers used are listed in Table S2.

Statistical analyses

The data were analyzed using Student's t test or one-way ANOVA. P values <0.05 were considered significant.

Supplementary figures

Figure S1. Development of TKI-resistant lines

HCC827 cells long-termly exposed to sub-toxic doses of gefitinib or afatinib. Compared to the parental HCC827 cells, the resistant lines (A)HCC827^{GR} and (B) HCC827^{AR} were not sensitive to the treatment of gefitinib or afatinib, respectively. (C) H1975^{AR} cells were more resistant to the treatment of afatinib compared to H1975 cells.

Figure S2. Single cell clones isolated from TKI-resistant lines

Protein expression of YAP from single cell clones isolated from TKI-resistant lines (A)HCC827^{GR}, (B) HCC827^{AR} or (C) H1975^{AR} cells. Moreover, the mRNA levels of EGFR, Met or HER2 were detected in the single clones from (A) HCC827^{GR}, (B) HCC827^{AR} or (C) H1975^{AR} cells.

Figure S3. Reduced YAP expression in the presence of YAP inhibitors

Reduced cell proliferation was detected in the YAP knockdowns (A-C). Reduced YAP expression detected in the presence of verteporfin (D-F) or fluvastatin (G-I).

Figure S4. Reduced tumor size in combined therapy group

Xenograft tumors in vehicle, afatinib, fluvastatin, and co-treated groups.

Figure S5. Original blots displayed in Figures 1 and 2

Original blots of (A) figures 1 D & E and (B) figures 2 A, D & G.

Figure S6. Original blots displayed in Figure 3

Original blots of (A) figures 3 B, F & J and (B) figures 3 C, G & K.

Figure S7. Original blots displayed in Figures 4 and 5

Original blots of (A) figures 4E-H, (B) figures 5E-H and (C) figures 5I-L.

Table S1. Culture conditions for developing TKI resistant cell lines

TKI resistant clones were developed by treating EGFR active mutant cell lines, H1975 or HCC827, with subtoxic doses of TKI for totally 60 days.

Cell line	Drug treated	Conc (μ M)/duration (day)	Developed line
H1975	Afatinib	0.1/15 → 0.5/20 → 1.0/25	H1975 ^{AR}
HCC827	Gefitinib	0.2/10 → 2/10 → 5/10 → 10/30	HCC827 ^{GR}
HCC827	Afatinib	0.01/10 → 0.1/20 → 0.5/30	HCC827 ^{AR}

Table S2. Sequences of QPCR primers

Gene	Forward	Reverse
GAPDH	GCATTGCCCTCAACGAC	GTCTCTCTCTCCTCTTGTGC
YAP	CTCCCCAGTGACGAGAGAGC	CTAGGTCTGCGACCTCGAC
CTGF	CCTGGTCCAGACCACAGAGT	TGGAGATTTTGGGAGTACGG
ANKRD1	AGAAGTGTGCTGGGAAGACG	GCCATGCCTTCAAAATGCCA
EGFR	ACTCATGCTCTACAACCC	CCAATACCTATCCGTTACAC
MET	CCATCCAGTGTCTCCAGAAGTGAT	ATGGTCAGCCTTGTCCCTC
HER2	CCATAACACCCACCTCTGCT	ACTGGCTGCAGTTGACACAC

Fig S1

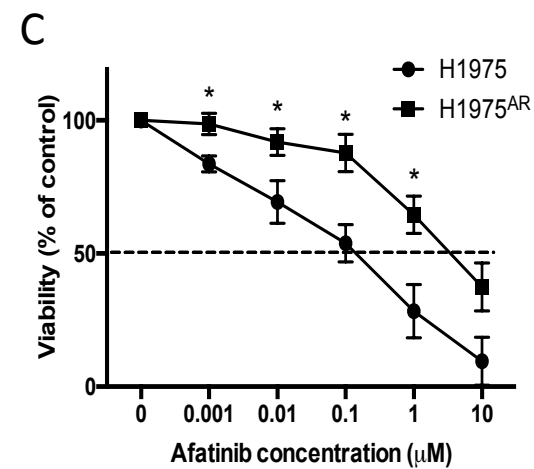
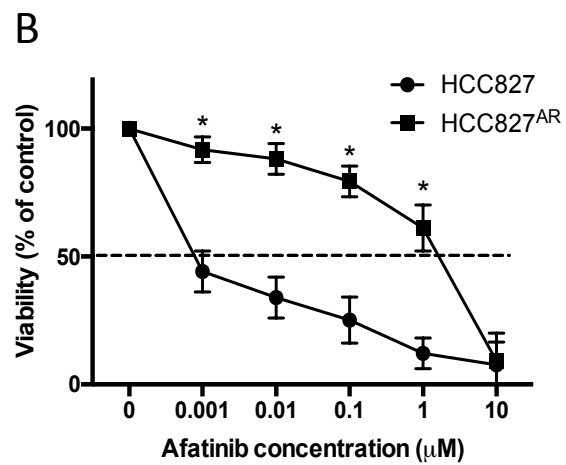
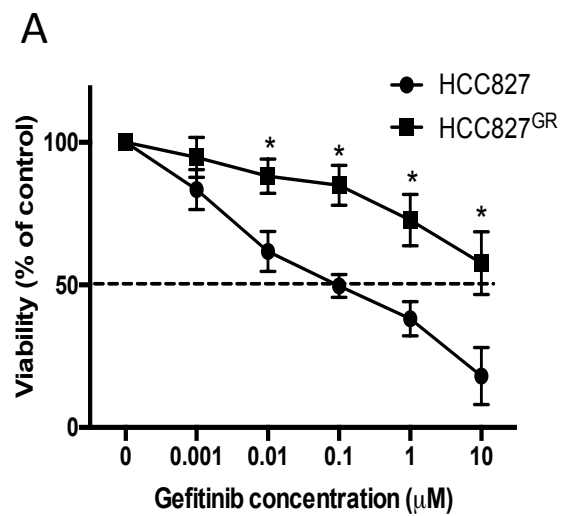


Fig S2

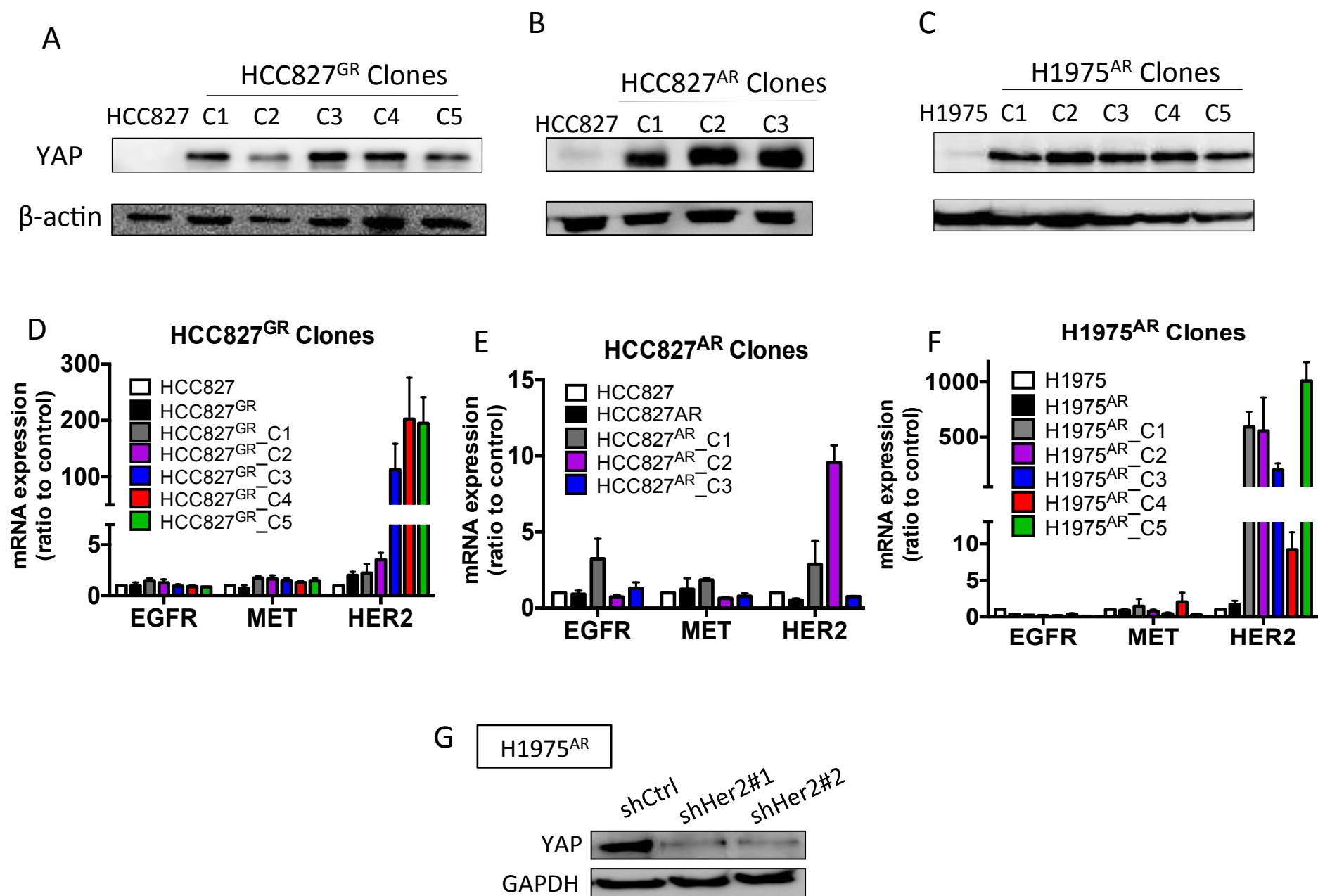


Fig S3

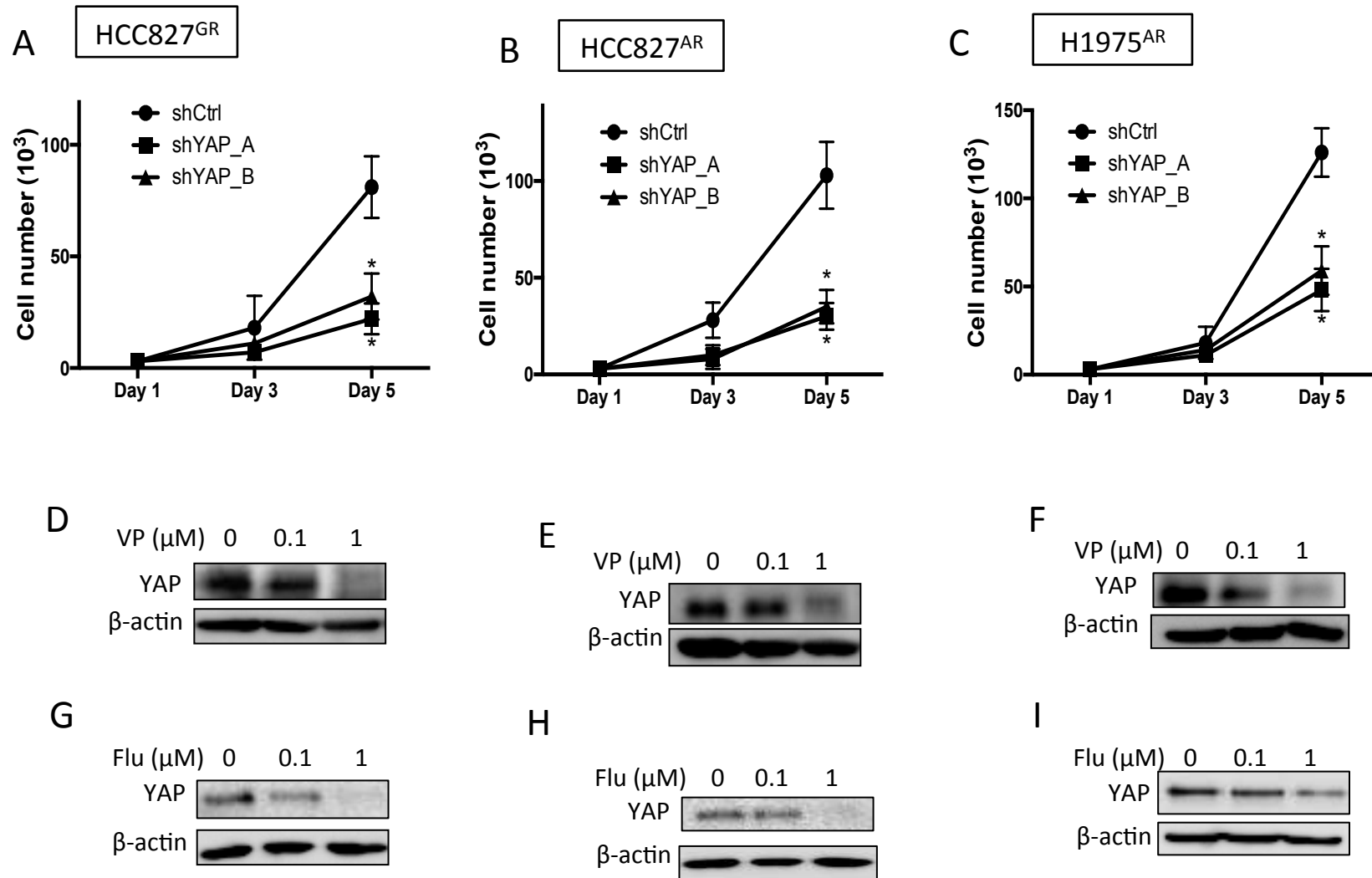
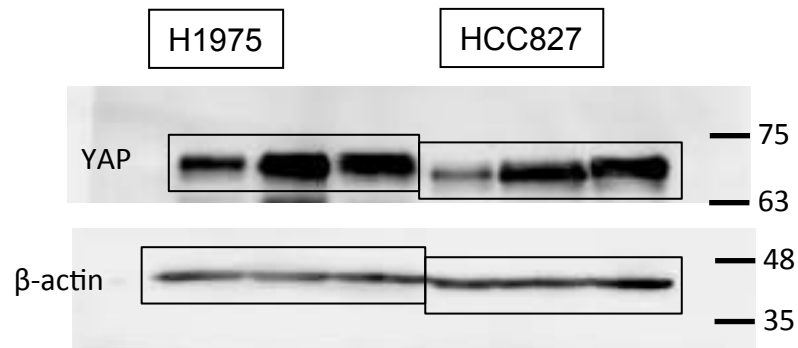


Fig S4



Fig S5

A



B

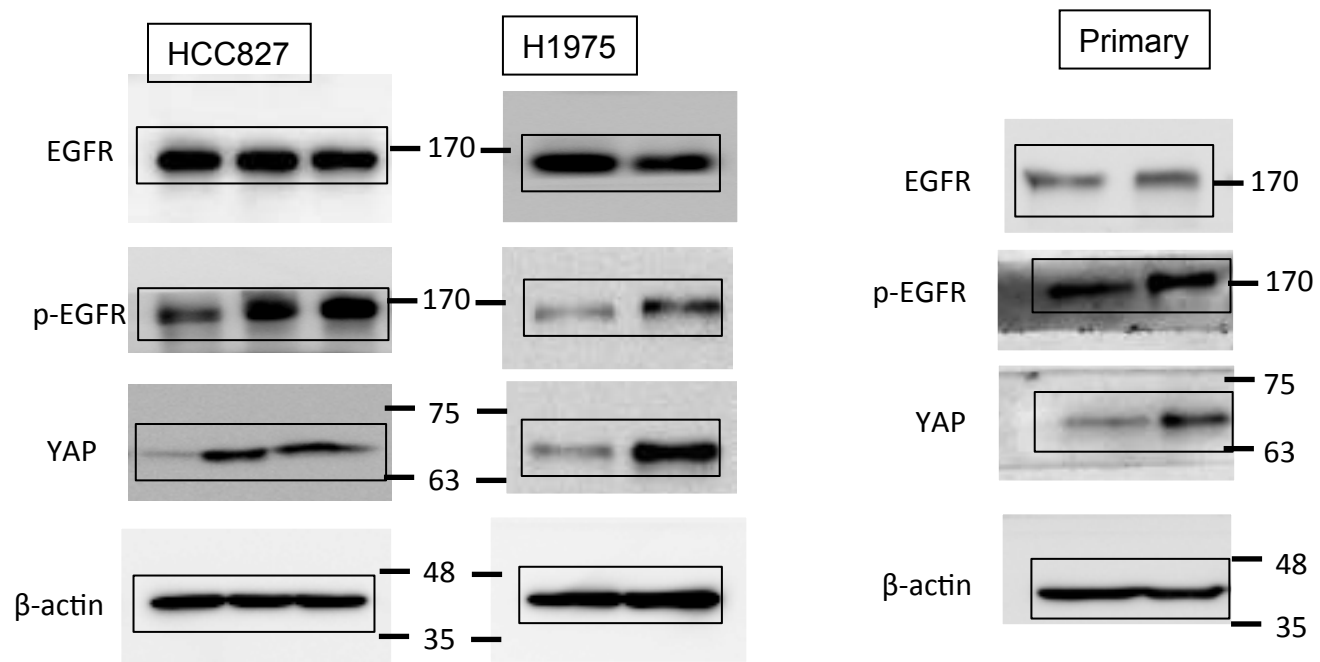
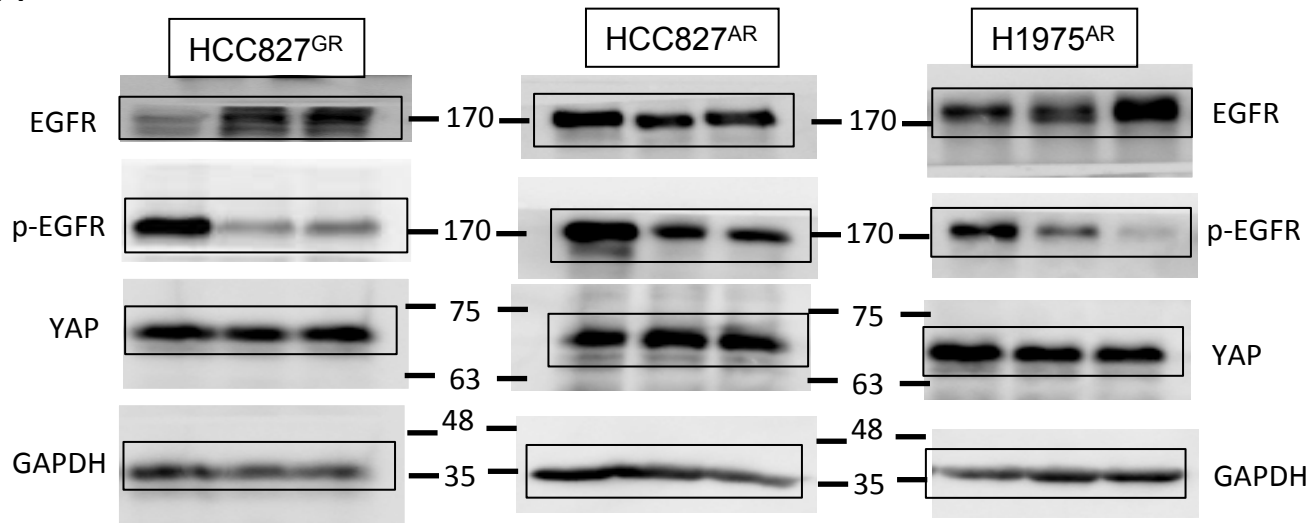


Fig S6

A



B

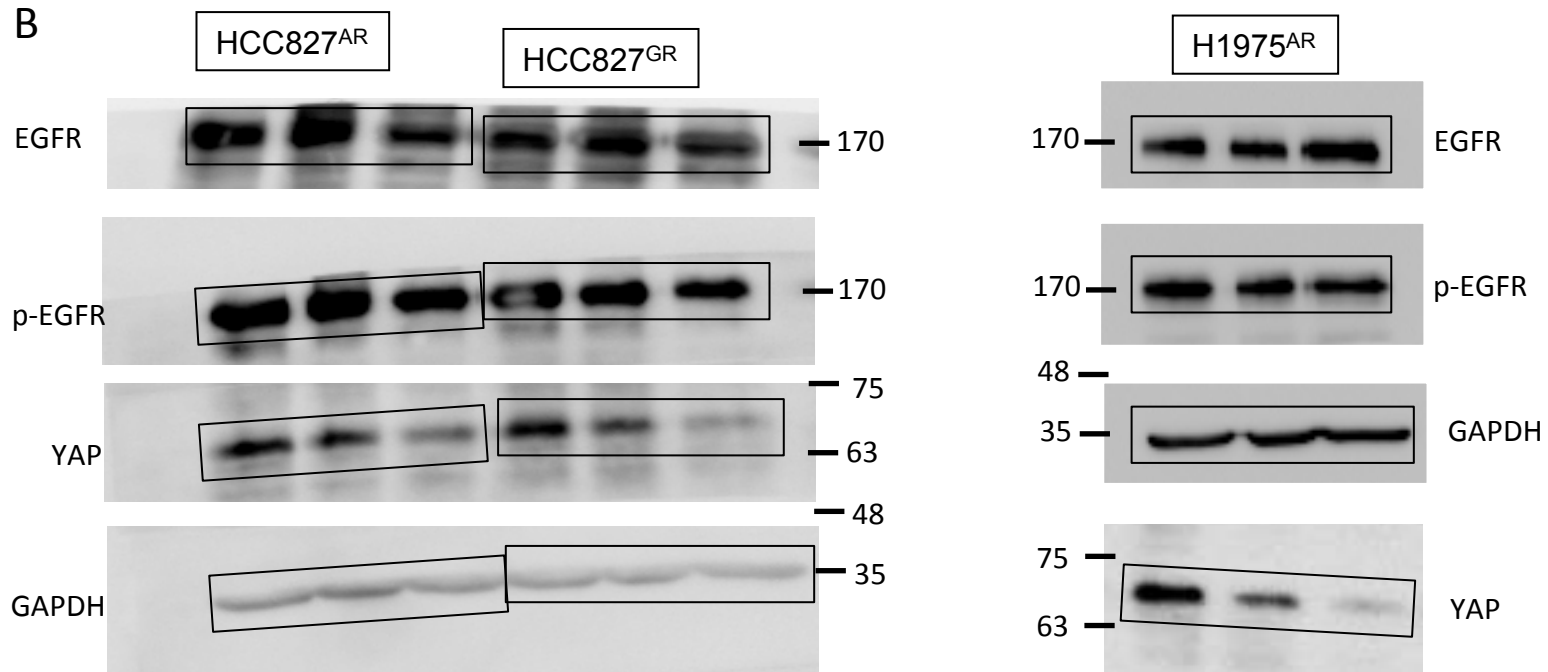
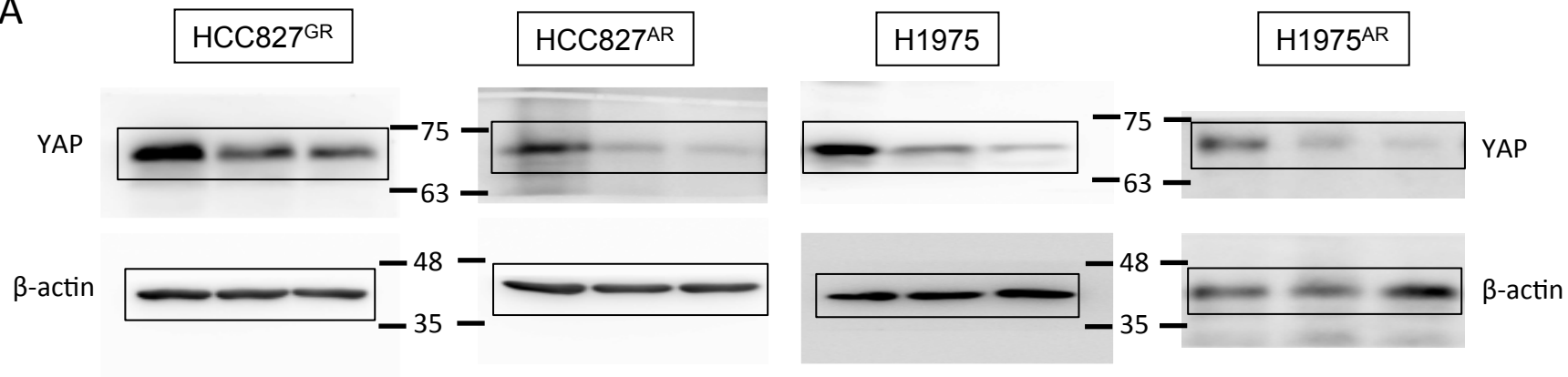
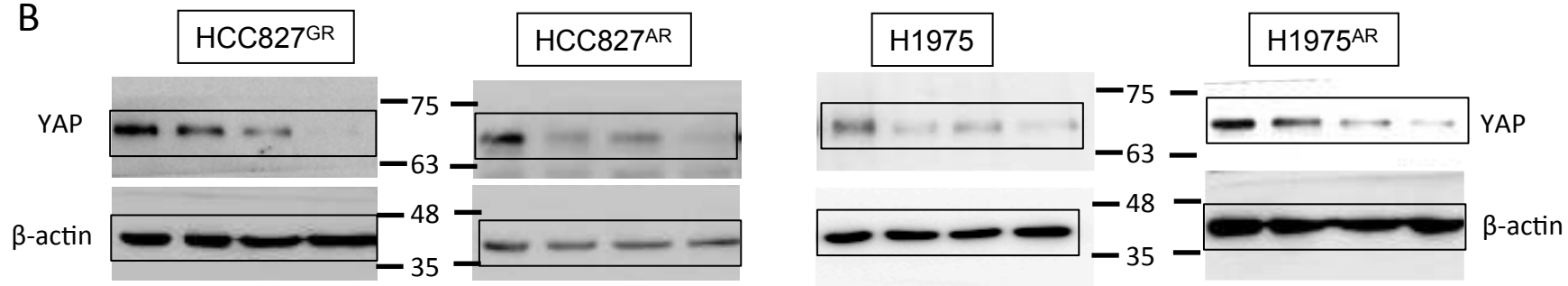


Fig S7

A



B



C

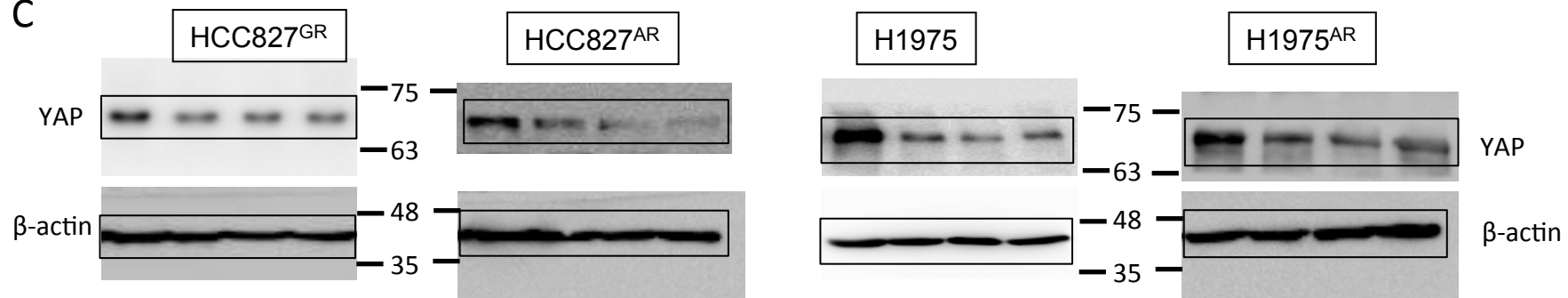
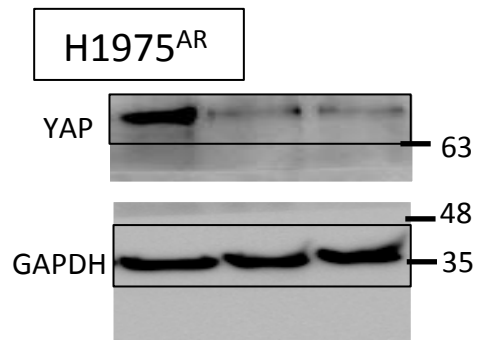
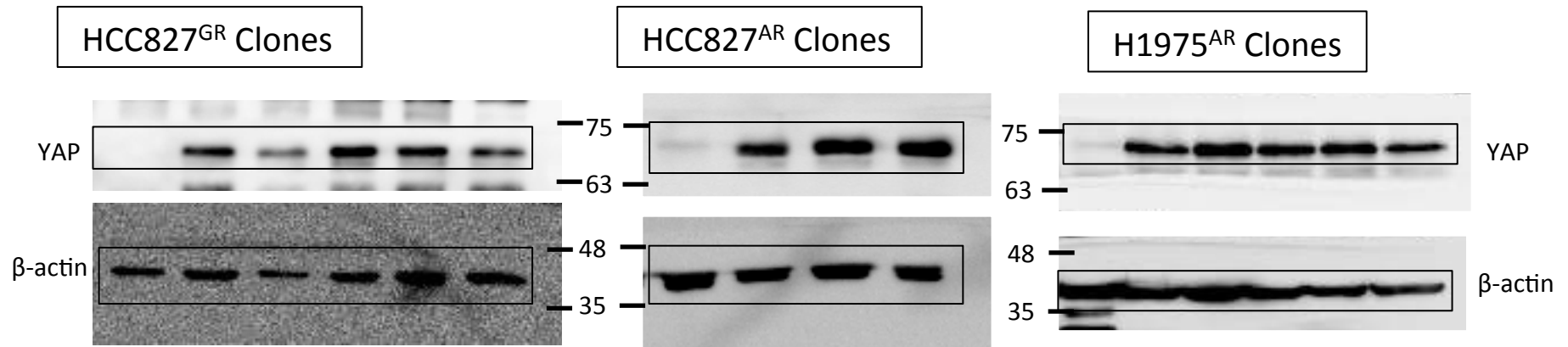


Fig S8

A. Original blots of FigS2



B. Original blots of FigS3

