## Dengue virus-induced ER stress is required for autophagy activation, viral replication, and pathogenesis both *in vitro* and *in vivo*

Ying-Ray Lee<sup>1#</sup>, Szu-Han Kuo<sup>2#</sup>, Ching-Yen Lin<sup>1</sup>, Po-Jung Fu<sup>2</sup>, Yee-Shin Lin<sup>2</sup>, Trai-Ming Yeh<sup>3</sup>, and Hsiao-Sheng Liu<sup>2\*</sup>

<sup>1</sup>Department of Medical Research, Chiayi Christian Hospital, 600 Chiayi, Taiwan. (Ying-Ray Lee: yingray.lee@gmail.com; Ching-Yen Lin: jouyuan22@gmail.com) 
<sup>2</sup>Department of Microbiology and Immunology, College of Medicine, National Cheng Kung University, 701 Tainan, Taiwan. (Szu-Han Kuo: s46004022@mail.ncku.edu.tw; Po-Jung Fu: U9807038@cmu.edu.tw; Yee-Shin Lin: yslin1@mail.ncku.edu.tw; Hsiao-Sheng Liu: a713@mail.ncku.edu.tw)

**Running title:** Dengue virus induces ER stress, autophagy, viral replication and pathogenesis

**Key Words:** Dengue virus; ER stress; Unfolded protein response; Autophagy, viral replication

\*Corresponding author: Hsiao-Sheng Liu
Department of Microbiology and Immunology,
College of Medicine,
National Cheng Kung University,
1 University Road,
Tainan, 70101,
Taiwan.

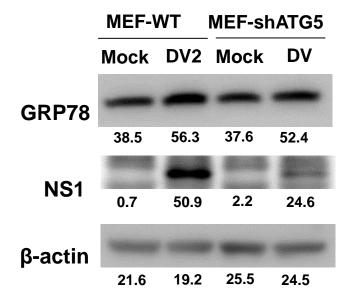
Telephone number: 886-2353535, ext. 5630.

E-mail: a713@mail.ncku.edu.tw

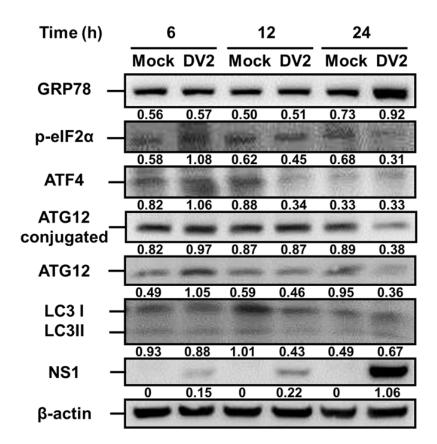
# These authors contributed equally to this work.

<sup>&</sup>lt;sup>3</sup> Department of Medical Laboratory, Science and Biotechnology, College of Medicine, National Cheng Kung University, 701 Tainan, Taiwan. (Trai-Ming Yeh: today@mail.ncku.edu.tw)

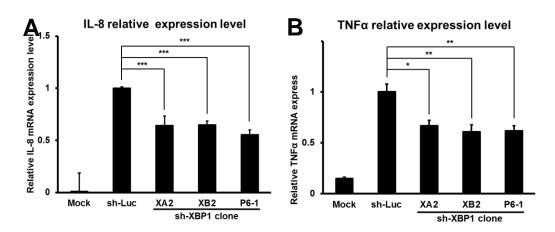
## **Supplementary Data**



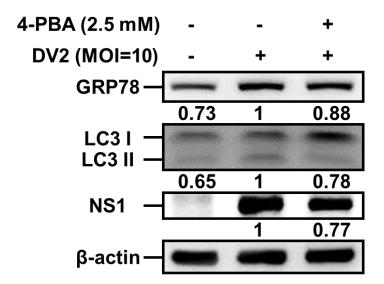
S1 appendix. Autophagy does not affect DENV2-induced ER stress. MEF cells were mock or shATG5 transfected for 18 h, and then with or without DENV2 infection (MOI=20). The protein levels of GRP78 and DENV2-NS1 were investigated by Western blotting using specific antibodies at 24 h p.i..  $\beta$ -actin was used as the loading control. The number under the band is the quantification of band intensity after normalization with  $\beta$ -actin.



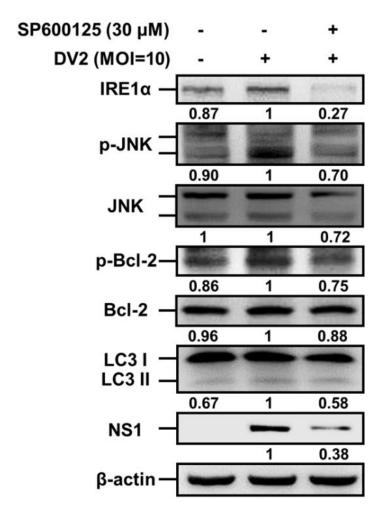
S2 appendix. PERK-eIF2 $\alpha$ -ATF4-ATG12 signaling pathway is involved in DENV2-induced autophagy activation. Huh7 cells were infected with DENV2 (MOI=10) and the samples were collected at 6 h, 12 h and 24 h p.i.. The protein levels of GRP78, p-eIF2 $\alpha$ , ATF4, ATG12 free form and conjugated form, LC3 II/LC3 I and NS1 were detected by Western blotting at the indicated time points. The number under the band is the quantification of band intensity after normalization with  $\beta$ -actin.



S3 appendix. Silencing of XBP1 reduces IL-8 and TNF $\alpha$  mRNA expression during DENV2 infection. Huh7 cells were transfected with sh-XBP1 and the cells were infected with DENV2 (MOI=10). The mRNA expressions of (A) IL-8 and (B) TNF $\alpha$  were determined by real-time PCR after 36 h p.i.. Expression of  $\beta$ -actin mRNA was used as an internal control, and the relative levels were quantification after normalization with  $\beta$ -actin.



**S4** appendix. ER stress inhibitor suppressed autophagic activity and DENV2 NS1 protein expression in SK-N-SH cells. SK-N-SH cells were infected with DENV2 (MOI=10) with or without the treatment of 4-PBA (2.5 mM), and the expressions of GRP78, LC3 and DENV2-NS1 proteins were determined by Western blotting at 36 h p.i. using specific antibodies.



S5 appendix. The inhibitor of JNK signaling pathway suppressed autophagic activity and DENV2 NS1 protein expression in SK-N-SH cells. SK-N-SH cells were infected with DENV2 (MOI=10) with or without the treatment of SP600125 (30  $\mu$ M), the expressions of IRE1 $\alpha$ , p-JNK, LC3 and NS1 proteins were determined with Western blotting at 36 h p.i..