Supplementary Data

Co-produced natural ketolides methymycin and pikromycin inhibit bacterial growth by preventing synthesis of a limited number of proteins

Mashal M. Almutairi, Maxim S. Svetlov, Douglas A. Hansen, Nelli F. Khabibullina, Dorota Klepacki, Han-Young Kang, David H. Sherman, Nora Vázquez-Laslop, Yury S. Polikanov, Alexander S. Mankin

Content:

Supplementary Tables S1 and S2 Supplementary Figures S1 – S3

Crystals	70S-MTM with A-, P- and E-tRNAs	70S-PKM with A-, P- and E-tRNAs						
Diffraction data								
Space Group	P212121	P212121						
Unit Cell Dimensions, Å (a x b x c)	209.62 x 448.78 x 622.53	209.34 x 447.21 x 620.61						
Wavelength, Å	0.9795	0.9795						
Resolution range (outer shell), Å	311-2.70 (2.77-2.70)	200-2.60 (2.67-2.60)						
l/σl (outer shell with l/σl=1)	8.22 (1.06)	8.58 (0.97)						
Resolution at which I/σI=1, Å	2.70	2.60						
Resolution at which I/σI=2, Å	2.87	2.82						
CC(1/2) at which $I/\sigma I=1, \%$	21.8	17.3						
CC(1/2) at which $I/\sigma I=2$, %	45.0	43.3						
Completeness (outer shell), %	98.9 (95.0)	98.3 (96.4)						
R _{merge} (outer shell)%	14.2 (118.5)	13.9 (122.1)						
No. of crystals used	1	1						
No. of Reflections Observed	5,320,880	5,786,516						
Used: Unique	1,561,494	1,729,501						
Redundancy (outer shell)	3.41 (3.39)	3.35 (3.19)						
Wilson B-factor, Å ²	52.2	49.2						
Refinement								
R _{work} /R _{free} , %	22.2/26.4	23.8/28.3						
No. of Non-Hydrogen Atoms								
RNA	200,225	200,247						
Protein	90,976	90,982						
lons (Mg, K, Zn, Fe)	2,879	2,423						
Waters	5,102	3,394						
Ramachandran Plot								
Favored regions, %	94.04	93.93						
Allowed regions, %	5.18	5.30						
Outliers, %	0.78	0.77						
Deviations from ideal values (RMSD)								
Bond, Å	0.004	0.004						
Angle, degrees	0.822	0.838						
Chirality	0.040	0.040						
Planarity	0.005	0.005						
Dihedral, degrees	14.533	14.530						
Average B-factor (overall), Å ²	59.3	52.7						

Table S1. X-ray data collection and refinement statistics.

 $\begin{array}{l} R_{\text{merge}} = \Sigma \left| I - \langle I \rangle \right| / \Sigma I, \text{ where I is the observed intensity and } \langle I \rangle \text{ is the average intensity from multiple measurements.} \\ R_{\text{work}} = \Sigma \left| F_{\text{obs}} - F_{\text{calc}} \right| / \Sigma F_{\text{obs}}. \text{ For calculation of } R_{\text{free}}, 5\% \text{ of the truncated dataset was excluded from the refinement.} \end{array}$

Strain	Mutation	MTM	РКМ	ERY	SOL	CHL
SQ110DTC ^a	WT	16	32	2	ND	1
	G2057U	>256	>256	>256	ND	1
	A2058G	>256	>256	>256	ND	1
	A2059G	>256	>256	>256	ND	1
	C2611U	>256	>256	>256	ND	1
BWDK ^b	WT	4	4	ND	0.5	0.8

Table S2. Minimal inhibitory concentrations (μg/ml) of antibiotics against *E. coli* strains

^{a)} MIC for SQ110DTC cells was determined in LB medium

^{b)} MIC for BWDK cells was determined in minimal M9 medium lacking methionine, but supplemented with the other 19 amino acids.



Figure S1. The checkerboard testing of possible synergy between MTM and PKM reveals them as essentially additive antibiotics. Black circles: the MTM/PKM combinations, open circles: the combinations of the ribosome-targeting synergistic streptogramin antibiotics quinupristin and dalfopristin. The dashed diagonal line approximates the fractional inhibitory concentrations (FIC) values for a hypothetical pair of ideally additive antibiotics.



Figure S2. MTM and PKM inefficiently arrest translation at the motifs problematic for the ketolide-bound ribosome. (A) and (B): In vitro toeprinting analysis comparing the efficiency of ketolides TEL, MTM and PKM in causing ribosomal arrest at the VDK sequence of ermBL (A) or the RLR motif of ermDL (B). Positions of the toeprint bands (which are 16-17 nts downstream from the first nucleotide of the P-site codon) generated by ribosomes stalled due to the presence of MTM, PKM or TEL are indicated on the gels by black arrowheads. The codon in the P site of the ketolide-stalled ribosome is boxed in a black frame in the sequence of the corresponding genes. Ribosomes that escape ketolide-mediated stalling are captured at the Lys codon of ermBL (A) or the Phe codon of ermDL (B) (boxed in grey frames) because of the presence in the reactions of (A) Thr-RS inhibitor (borrelidine) or (B) Pro-RS inhibitor (L-PSA), respectively. The toeprint bands of the captured ribosomes are marked with grey arrowheads. The toeprint bands of the ribosomes arrested at the start codons of *ermBL* and *ermDL* because of the presence of the initiation inhibitor retapamulin (RET), are indicated by an arrow. The control sample labelled as "none" had no ketolides or retapamulin added, but contained the respective RS inhibitor. (C) In vitro translation in the E. coli S30 extract of the fusA gene encoding the 78 kDa protein EF-G. Translation reactions, supplemented with [³⁵S]-Lmethionine, were carried out in the absence of antibiotics ('no drug'), in the presence of 50 μ M of TEL or in the presence of the indicated concentrations of MTM. The black arrow indicates the band corresponding to the full-size EF-G and the gray arrow points to the truncated product generated due to TEL-induced translation arrest at the codon 358 of the *fusA* gene (5).



Figure S3. Reducing of the general translation rate by two-fold does not prevent cell growth and proliferation. (A): At near-MIC concentrations of the protein synthesis inhibitor chloramphenicol, translation rate in *E. coli* cells (strain BWDK) decreases approximately two-fold (dashed line marks a 50% decrease in the translation rate). Nevertheless, cells synthesizing all proteins at approximately 50% level continue to grow and multiply (B). In (B) cells were diluted to the final optical density of A_{600} = 0.025 in the same defined medium that was used for measuring translation rate (see Materials and Methods) and then grown overnight at 37°C in 15 ml tubes. Aliquots of the overnight cultures were placed into a 96-well plate and stained with Alamar Blue (blue color – no cell growth, pink color – live cells).