

Table S1. Summary of three different types of DNA unzipping assays, all of which are at near base pair resolution. Related to STAR Methods.

	Applications	Method	Place used (with trapping mode)	References
unzipping tracker	Real-time tracking of DNA translocase without tagging	Constant force (e.g., 18 pN) 	Fig.1, Fig. S1B (single trap)	This study
unzipping staller	Real-time stalling of DNA translocase without tagging	Constant extension 	Fig. S6 (single trap) Fig. 4, Fig. S4A, Fig. 5, Fig. S5A, Fig. S6 (dual trap)	This study
unzipping mapper	Mapping both locations and strengths of interactions within a DNA-protein complex	Constant loading rate (e.g., 8-16 pN/s) 	Figs. 2B-D, Fig. 3, Fig. S2C, Fig. S3, Fig. S4B, Fig. 6, Figs. S7A-B (single trap) Fig. 2A, Figs. S2A-B & S2D (dual trap)	This study. Brennan et al., 2016 Li et al., 2015 Jin et al., 2010 Hall et al., 2009

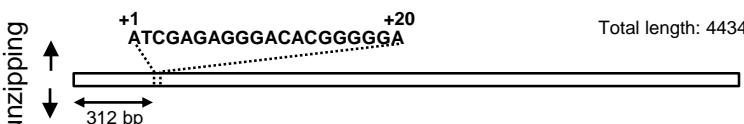
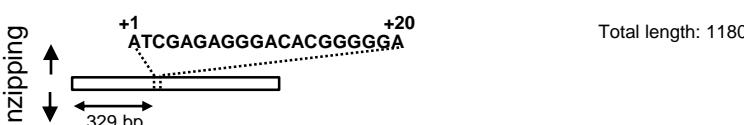
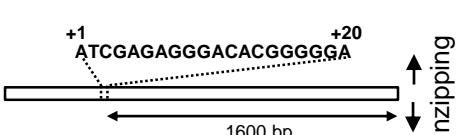
Table S2. Summary of translocation properties of Mfd. Experiments were carried out under a constant unzipping force of 18 pN. Shown are means and SEMs with the total numbers of traces indicated in parentheses. Within measurement uncertainties, Mfd's mean distance, time, and speed are similar under 2 mM ATP and 1 mM NTPs when translocating either away from ("unzip") or towards ("rezip") the DNA fork. Related to Figures 1 and 2.

	mean distance (bp)	mean time (s)	mean speed (bp/s)
2 mM ATP (unzip)	200 ± 37 (48)	28.2 ± 5.5 (48)	7.2 ± 0.4 (48)
2 mM ATP (rezip)	247 ± 47 (53)	44.5 ± 8.3 (53)	5.8 ± 0.2 (53)
1 mM NTPs (unzip)	223 ± 49 (25)	38.4 ± 11.4 (25)	7.3 ± 0.5 (25)
1 mM NTPs (rezip)	218 ± 57 (29)	39.2 ± 11.0 (29)	6.5 ± 0.5 (29)

Table S3. Oligonucleotides used in this study. All oligos were purchased from Integrated DNA Technologies (IDT). ‘HEX’ indicates the HEX fluorophore labeled at the 5'-end. Related to Key Resources Table and STAR Methods.

pRL574_rfaQ ops_27bpafterA20_F	GGTAGCGTGCTTTTCCTCTAGAGTGCTGGCGAAC
pRL574_rfaQ ops_27bpafterA20_F	GCCCCATGGATGAGGCTAACCCCCGTGCTCCCTCTC
pRL574_rfaQ ops_276afterA20_F	GGTAGCGTGCTTTCCGTCTCTTGACTCCGACAAAGG
pRL574_rfaQ ops_276afterA20_R	GCCCCATGGATGAGGCTACCGGACTACGGTGCAGCT
pRL574_F_2kb	GCTGGCGAAAGGGGGATGT
pRL574_R_2kb_AlwNI_CTA	GGAACAGCTACTGGGAGTCTCAAGGAAGCCGTATTG
pRL574_F_1kb_inversed	TGGGGTCGAGGTGCCGTAAAG
pRL574_R_1kb_inversed_AlwNI	GGAACAGCTACTGCCAGGAACCACGGTAAGGGATG
pRL574_ops_275aA20_4kb_F_DraIII	GCAACACCTAGTGTATGCTGCAAGGCGATT
pRL574_ops_275aA20_4kb_R	AGACTTAACGGTGAGCATTCT
new_pRL574_F_DraIII	GACACCTAGTGAATTGAGCTCGGTACC
new_P1_R_pRL574	GGAAACAGCTATGACCATGATTAC
PRL574_F_AlwNI_CTA	CGCAGCTACTGTAAAACGACGGCCAGTGAAT
PRL574_R_800afterA20_Bsal	GATGGTCTCACGGTGATCCAGATCGTTGGTGAAC
Mfd_R685A_fwd	CGAAATGATCTCCGCTTCCGCAGCGCCAAG
Mfd_R685A_rev	CTTGGCGCTGCGGAAAGCGGAGATCATTG
Mfd_N817A_fwd	GGACAGGTTATTATCTTACGCTGATGTGGAAAACATTGAG
Mfd_N817A_rev	CTGAATGTTTCCACATCAGCGTAGAGATAATAACCTGTCC
40-mer DNA.oligo 1	HEX- AATGTCAGGCCGGAATAACTTCCTATAATGCGCCACCACT
40-mer DNA.oligo 2	AGTGGTGGCGCATTATAGGAAGTTATTCCGGCCTGACATT

Table S4. DNA unzipping templates used in this study. +1 indicates transcription start site (TSS). +20 indicates the last nucleotide incorporated into the transcript of a TEC paused at +20 position. Related to STAR Methods.

Name	Place used	Description
4.4 kb, co-directional	Figures 1, S1C, 2B-D, S3	 Total length: 4434 bp
1.2 kb, co-directional	Figures 2A, 6A-D, S7A-B	 Total length: 1180 bp
1.1 kb, reversed	Figure 6A-B	 Total length: 1105 bp
2.0 kb, reversed	Figures 4, 5, S4, S5A, S6	 Total length: 1980 bp
4.1 kb, co-directional, ops 276 bp after +20	Figures 3C, 3E, 3F	 Total length: 4118 bp
1.2 kb, co-directional, ops 27 bp after +20	Figures 3B, 3D, 3F	 Total length: 1214 bp