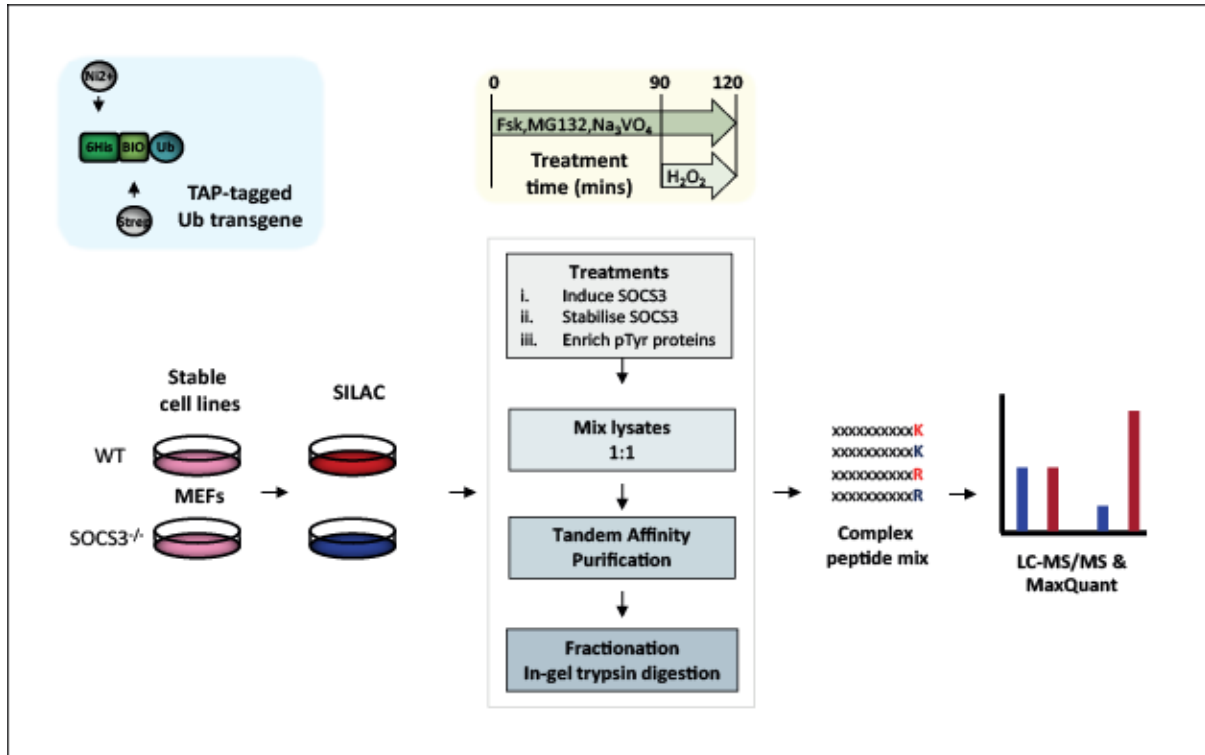
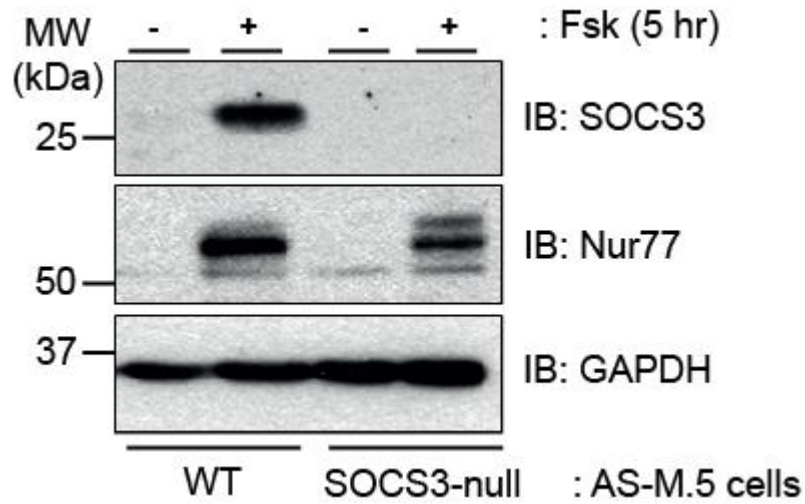


## SUPPLEMENTARY INFORMATION

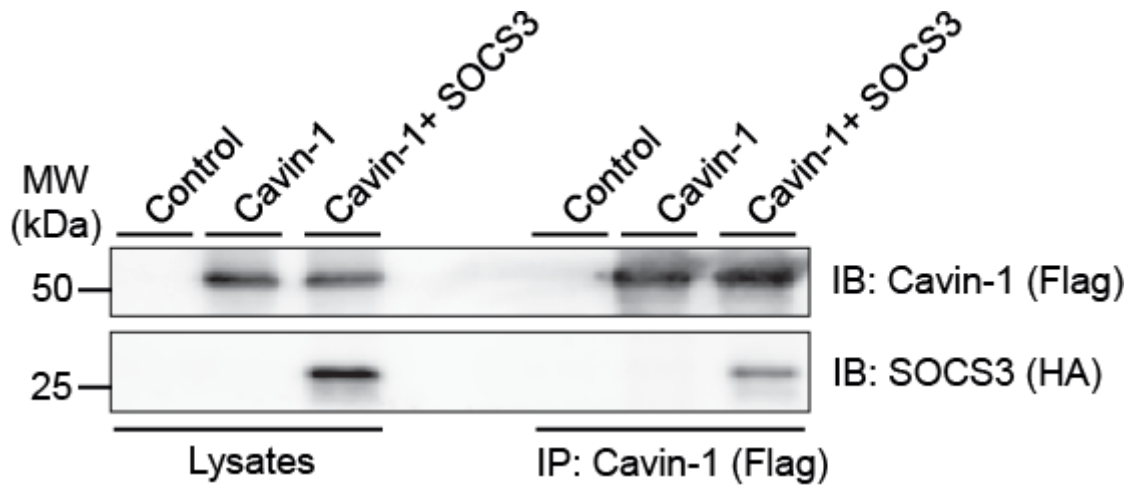


### Supplementary Figure 1:

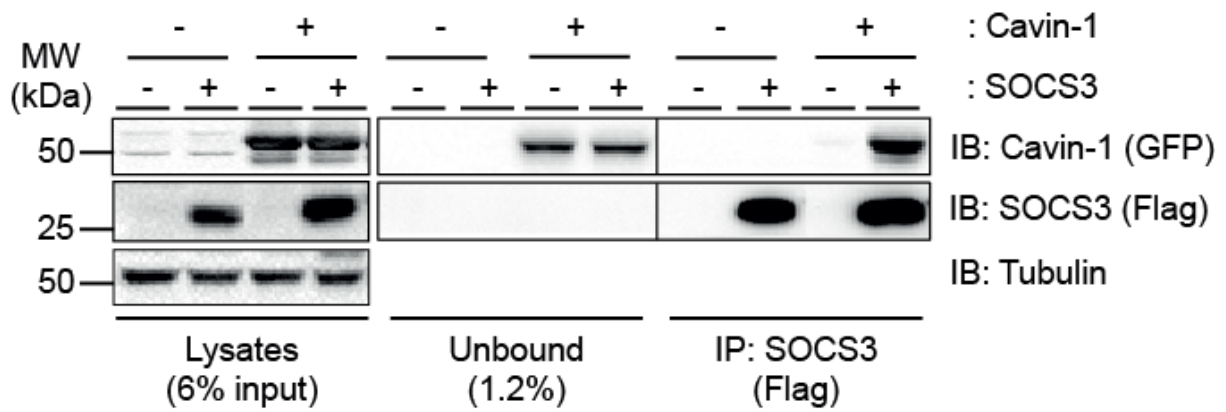
Differentially SILAC-labelled, tandem affinity purified ubiquitinomes isolated from WT MEFs and *SOCS3*<sup>-/-</sup> MEFs each expressing tagged ubiquitin transgenes (blue box) were compared following induction SOCS3 by treatment with forskolin (50  $\mu$ M) for 2 hrs. Proteasomal degradation of SOCS3 and potential ubiquitinated substrates was inhibited by co-incubation with proteasome inhibitor MG132 (6  $\mu$ M) while Tyr phosphorylation status was preserved through inhibition of protein Tyr phosphatases by incubation with sodium orthovanadate (Na<sub>3</sub>VO<sub>4</sub>: 1 mM) for 1.5 hrs then hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>: 0.2 mM) for a further 30 mins prior to harvesting. Tandem-affinity-purified proteins were digested with trypsin and peptides processed by sequential liquid chromatography and mass spectrometry.



**Supplementary Figure 2:** Detergent-soluble whole cell lysates from WT and SOCS3-null AS-M.5 human angiosarcoma-derived ECs treated with either vehicle or 50  $\mu$ M Fsk for 5 hr were equalised for protein content for SDS-PAGE for immunoblotting with the indicated antibodies. Data shown are representative of multiple experiments.



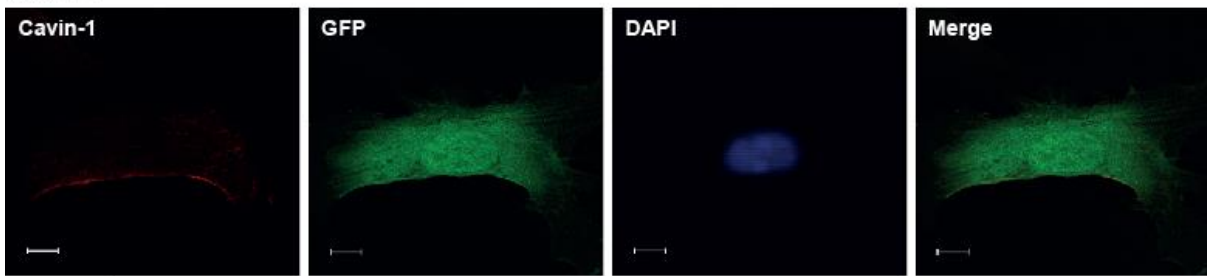
**Supplementary Figure 3:** Protein-equalised soluble cell extracts from HEK293 cells transfected with expression constructs encoding HA-tagged SOCS3 and Flag-tagged cavin-1 as indicated were processed by immunoprecipitation (IP) with anti-Flag M2-agarose beads prior to SDS-PAGE and immunoblotting with the indicated antibodies. Whole cell lysates from the samples used in the IP were also analysed in parallel.



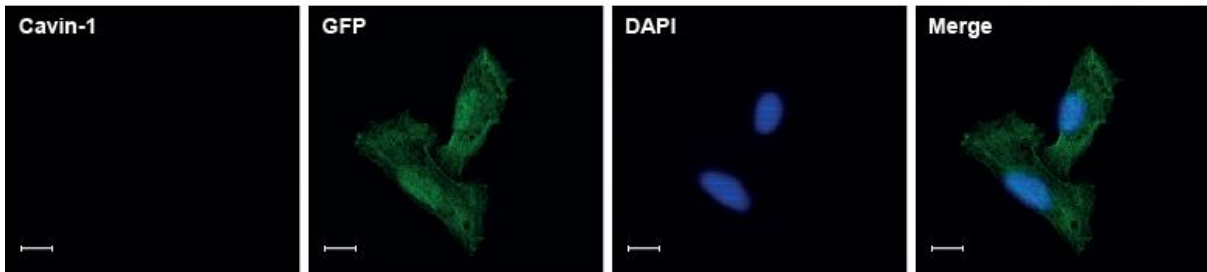
**Supplementary Figure 4:** Protein-equalised soluble cell extracts from HEK293 cells transfected with expression constructs encoding Flag-SOCS3 and GFP-tagged cavin-1 as indicated were processed by immunoprecipitation (IP) with anti-Flag M2-agarose beads prior to SDS-PAGE and immunoblotting with the indicated antibodies. Fractions of the whole cell lysates from the samples and unbound supernatants from the IP were also fractionated by SDS-PAGE for immunoblotting in parallel. Data shown are representative of three experiments.

# A

*Cavin-1*<sup>+/+</sup>

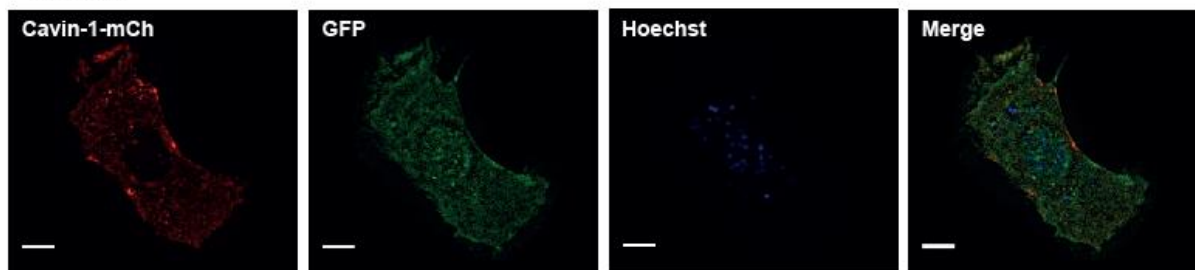


*Cavin-1*<sup>-/-</sup>



# B

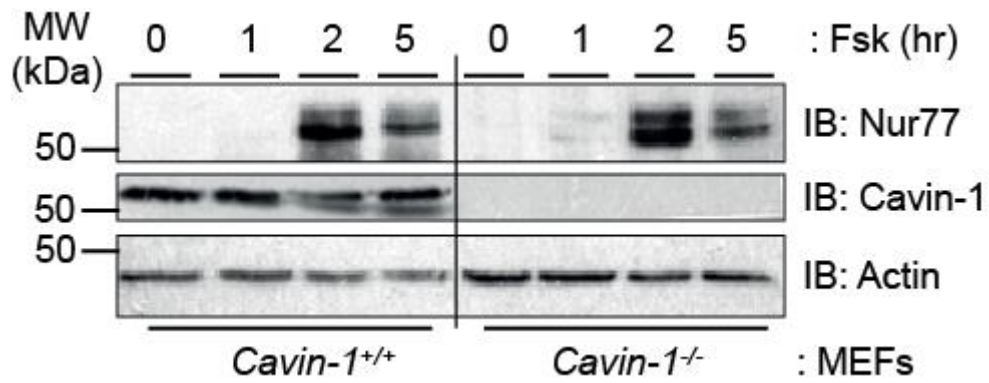
*Cavin-1*<sup>-/-</sup>



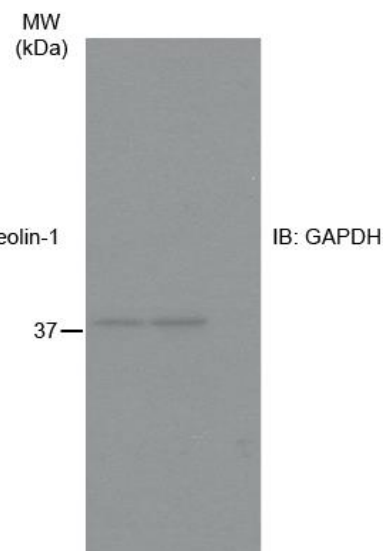
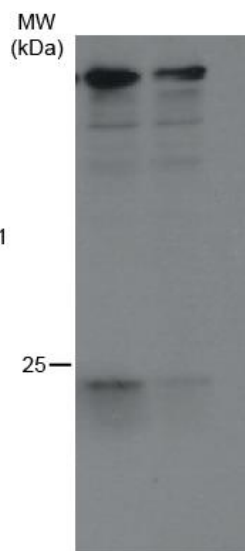
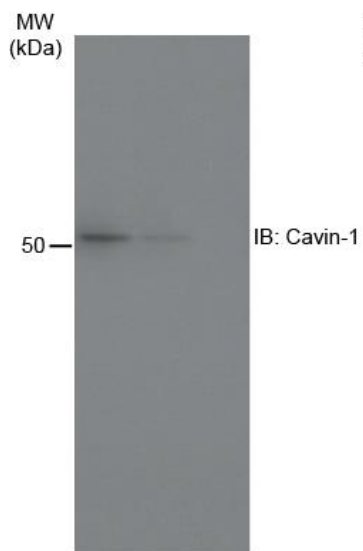
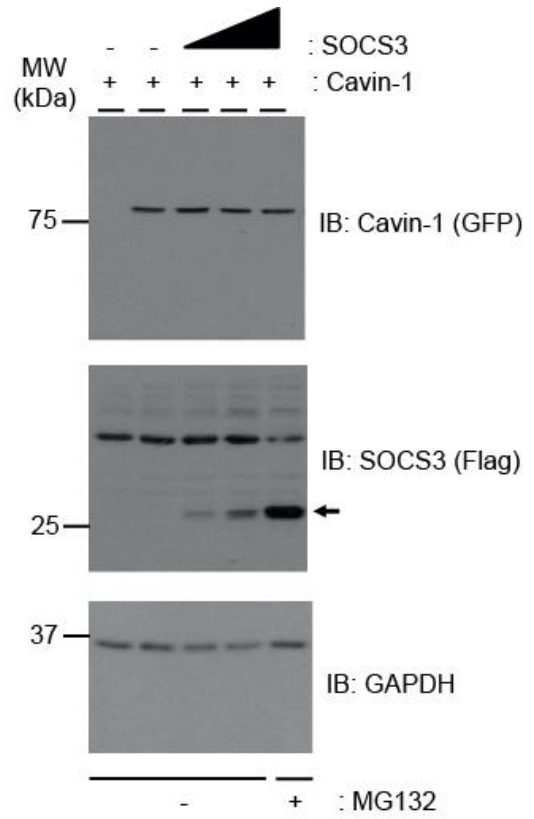
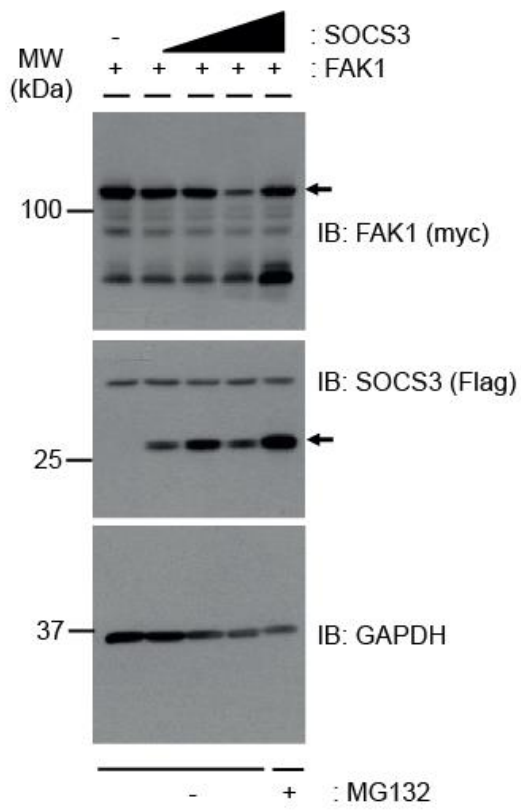
## Supplementary Figure 5:

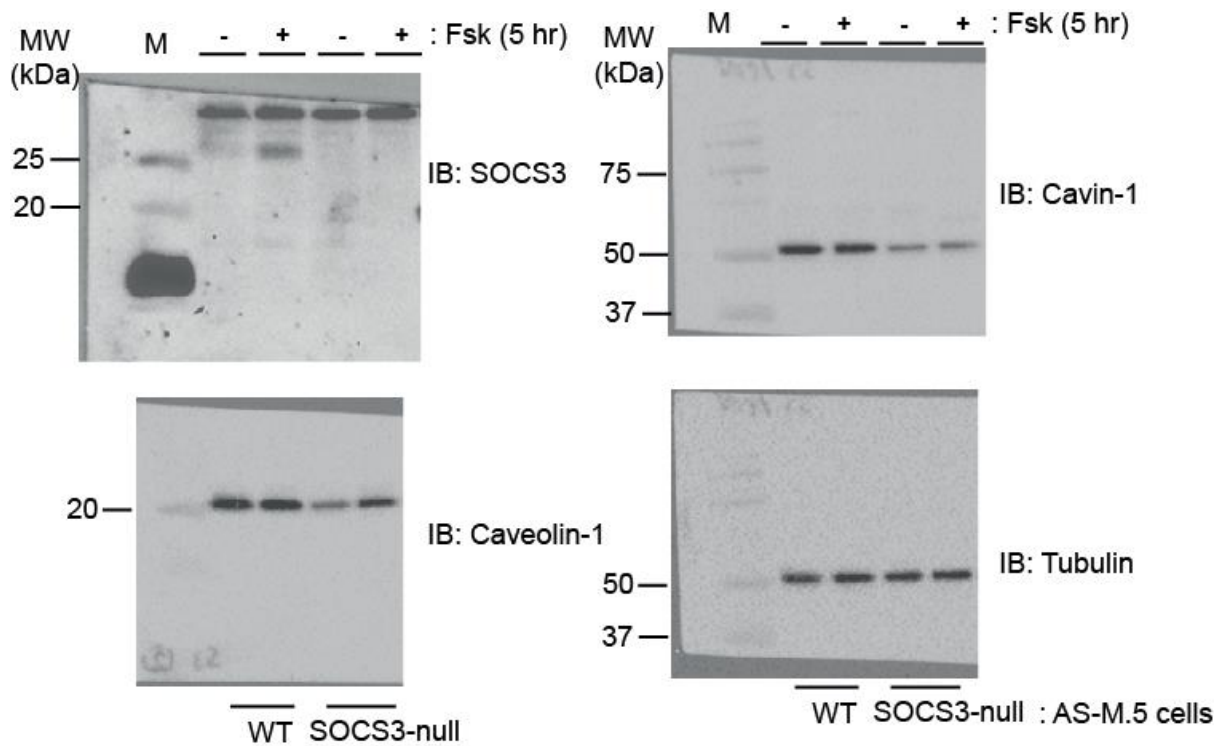
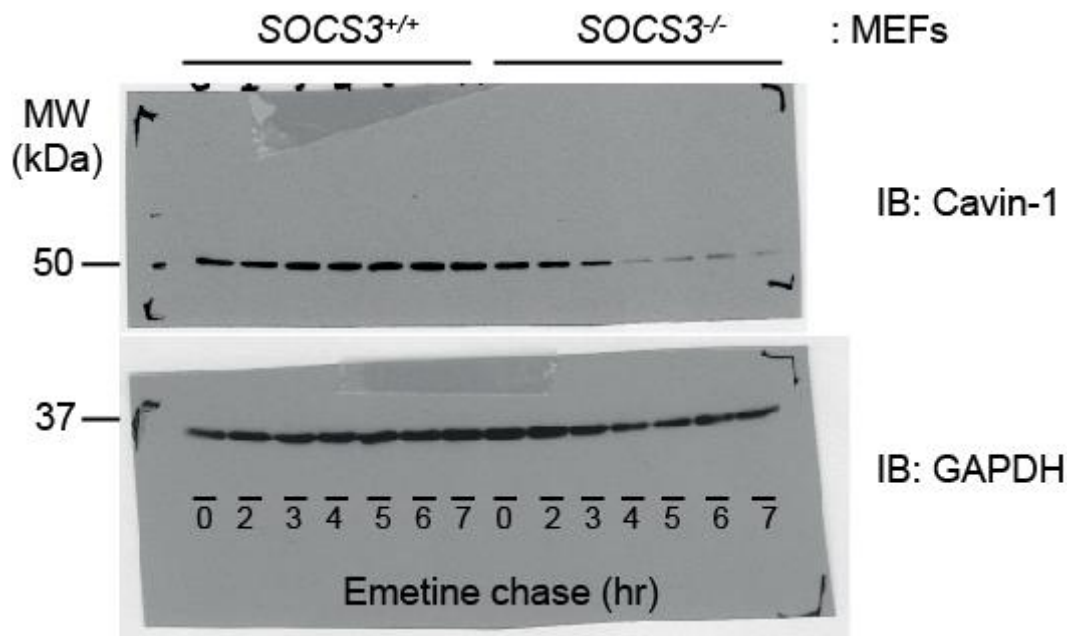
(A) WT (*cavin-1*<sup>+/+</sup>) and *cavin-1*<sup>-/-</sup> MEFs transiently expressing GFP (green) were stained with DAPI prior to being fixed, solubilised and stained with anti-cavin-1 antibody (red) before mounting for imaging by confocal microscopy. Scale bar = 10  $\mu$ m.

(B) *Cavin-1*<sup>-/-</sup> MEFs transiently co-expressing GFP (green) and cavin-1-mCh (red) were stained with Hoechst 33342 prior to being fixed for imaging by confocal microscopy. Scale bar = 10  $\mu$ m.

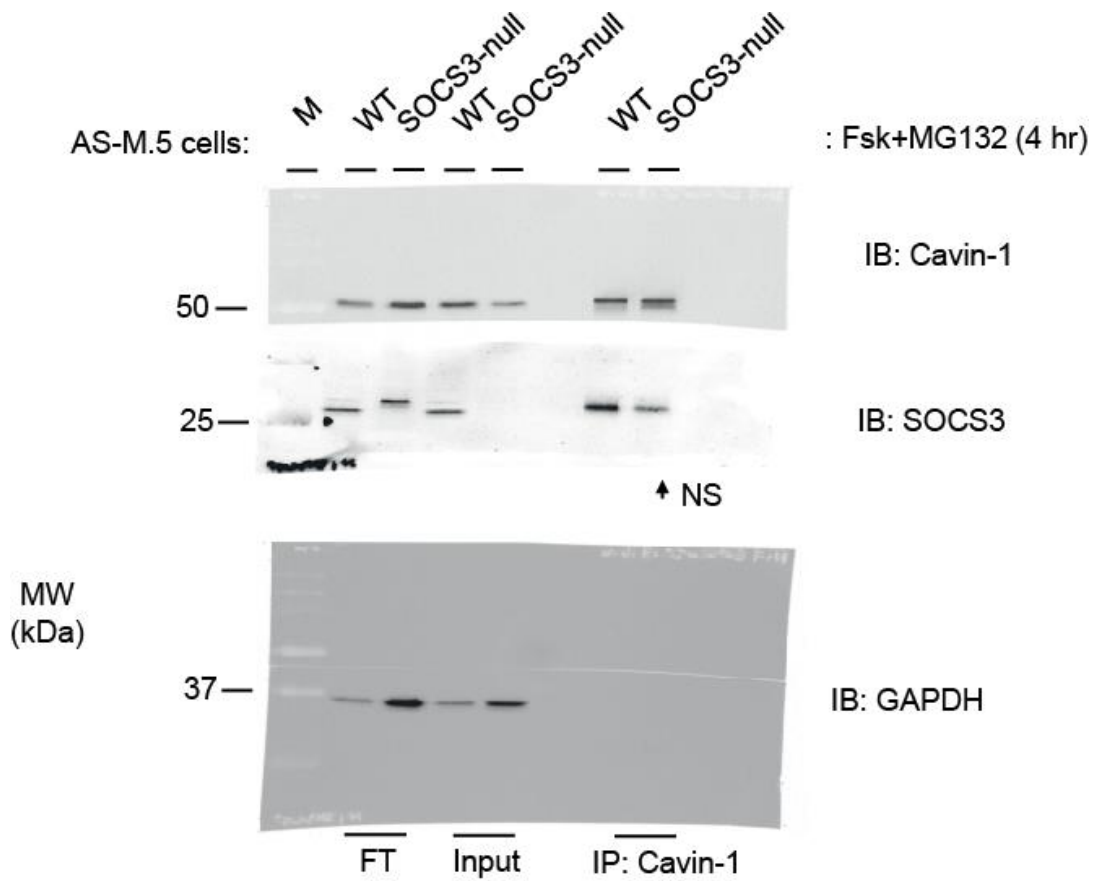
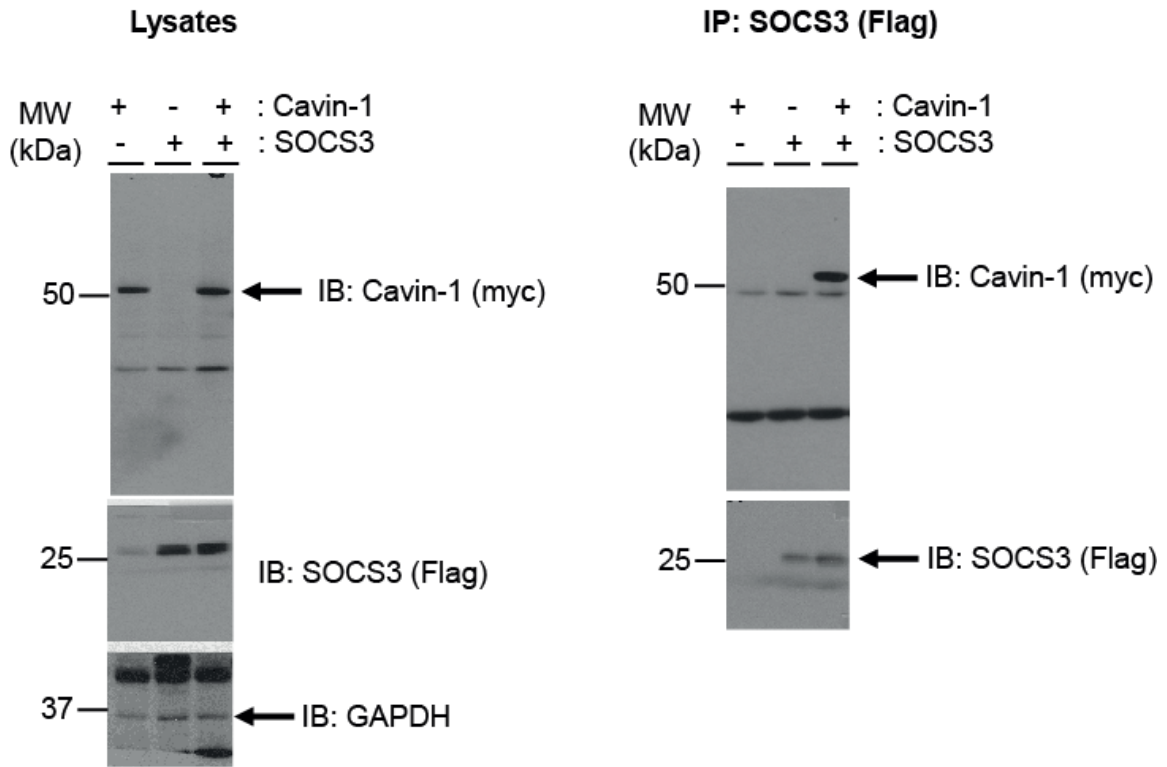


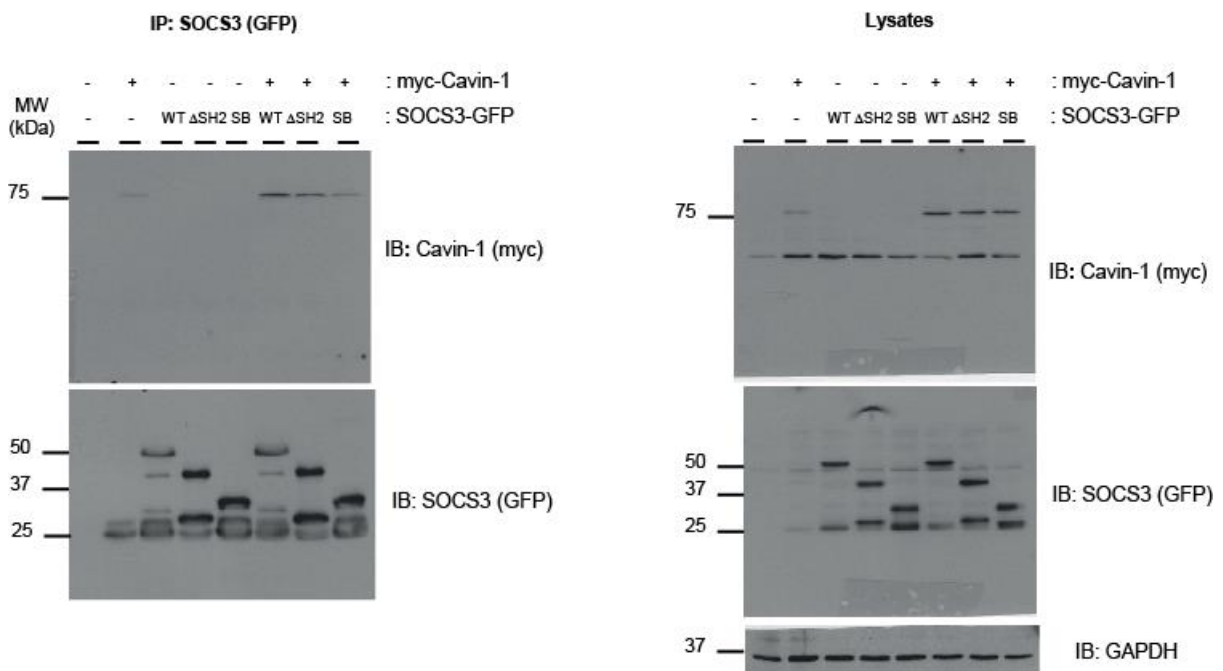
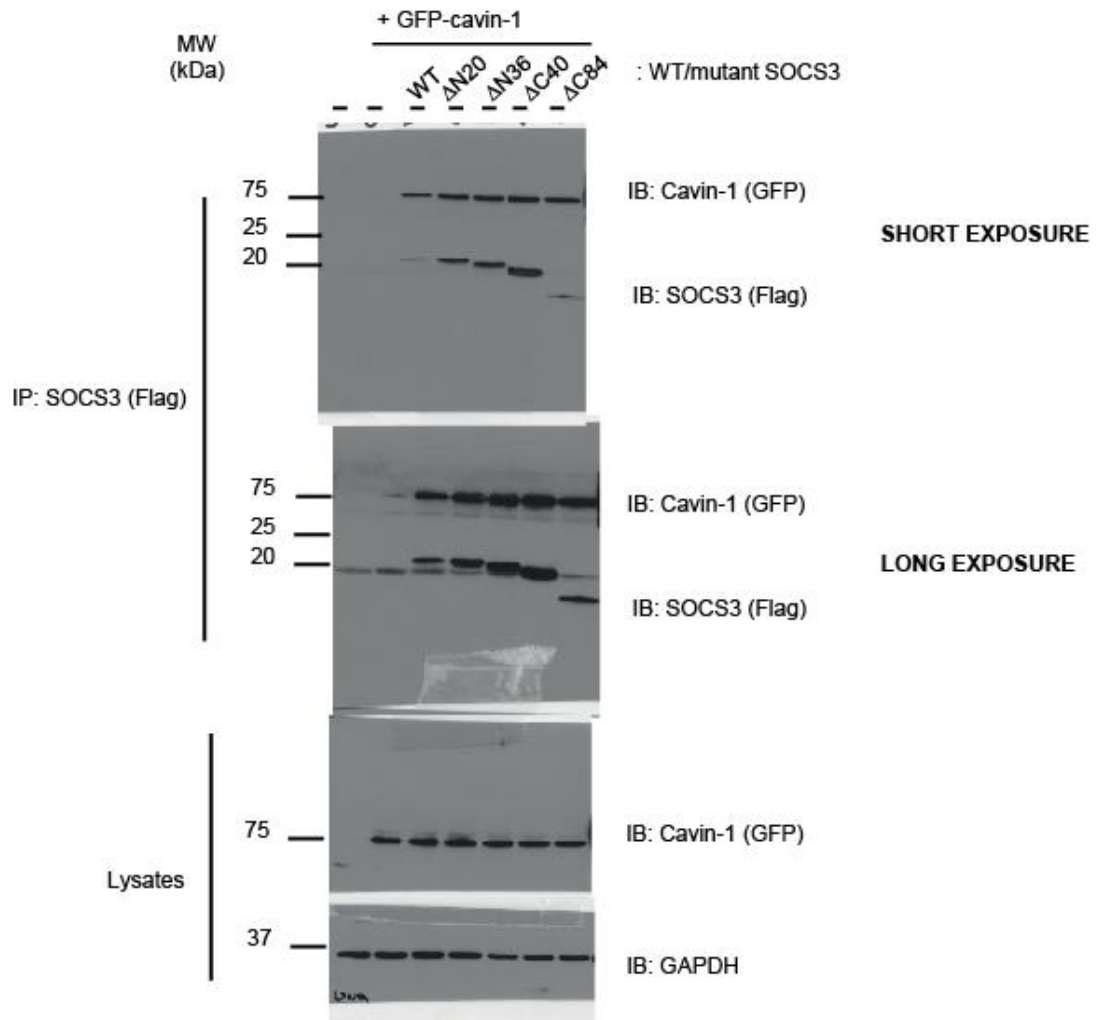
**Supplementary Figure 6:** Detergent-soluble whole cell lysates from *cavin-1*<sup>+/+</sup> and *cavin-1*<sup>-/-</sup> MEFs treated with either vehicle or 50  $\mu$ M Fsk for the indicated times were equalised for protein content for SDS-PAGE for immunoblotting with the indicated antibodies. Data shown are representative of three experiments.

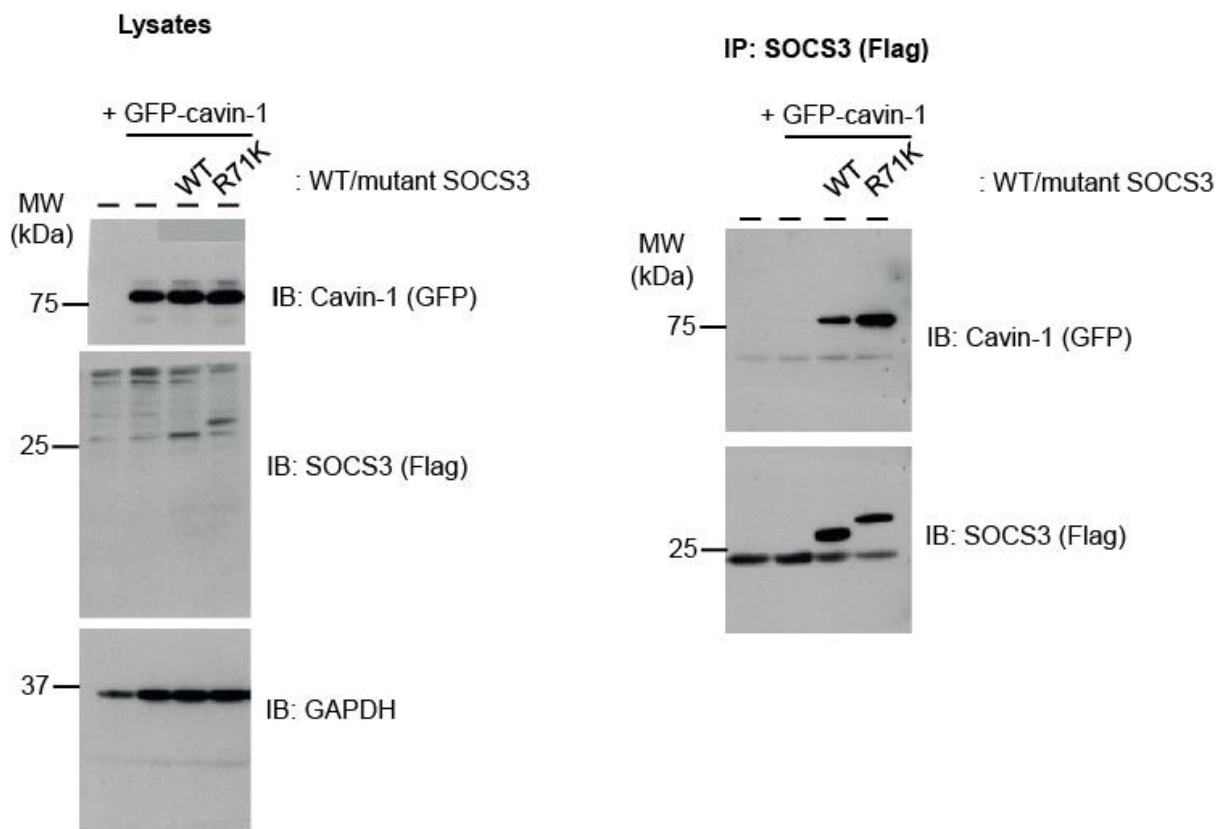
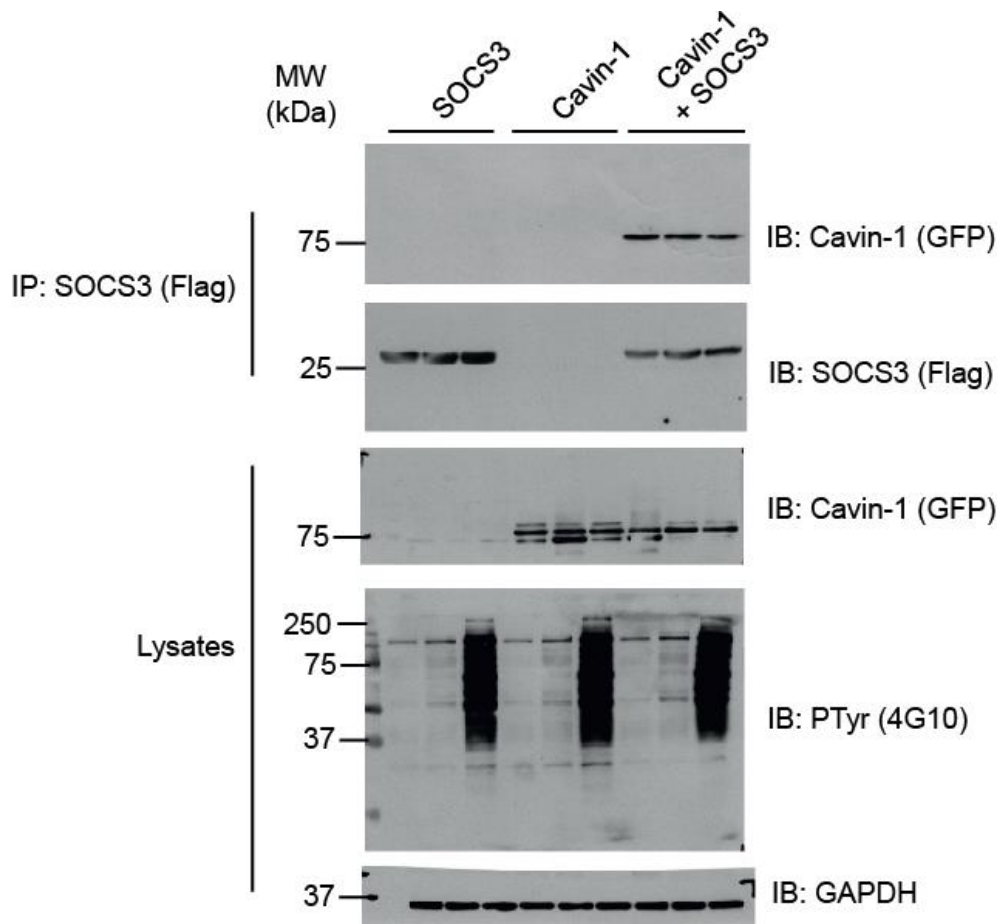


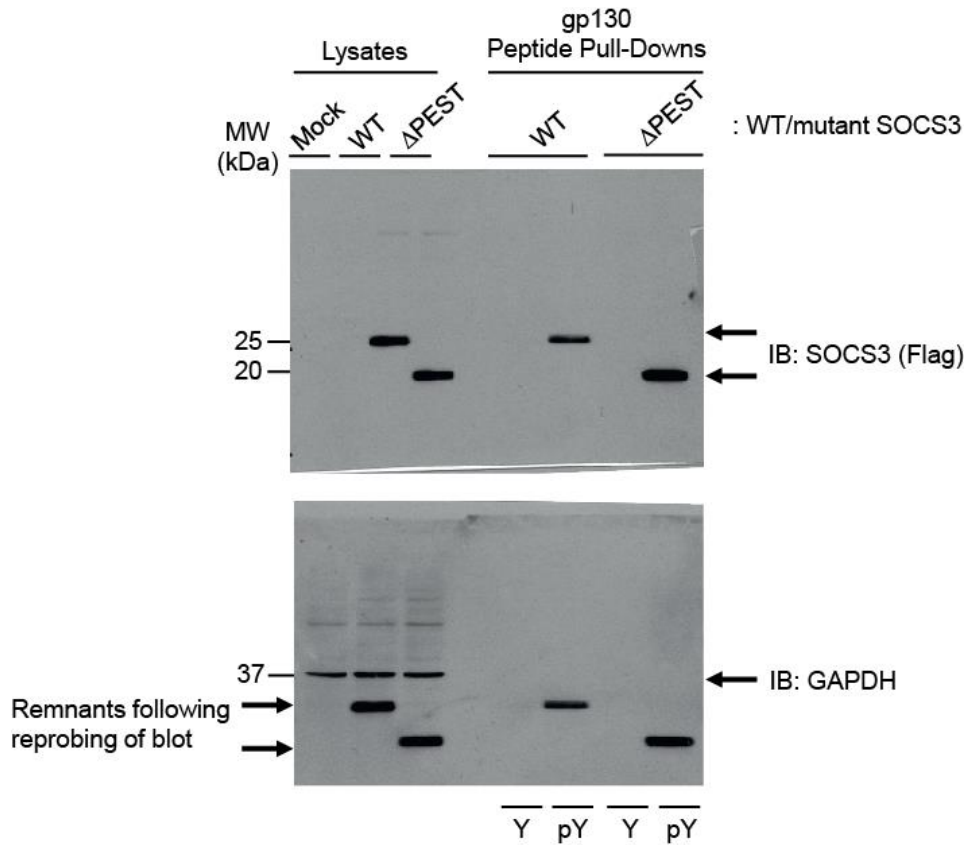
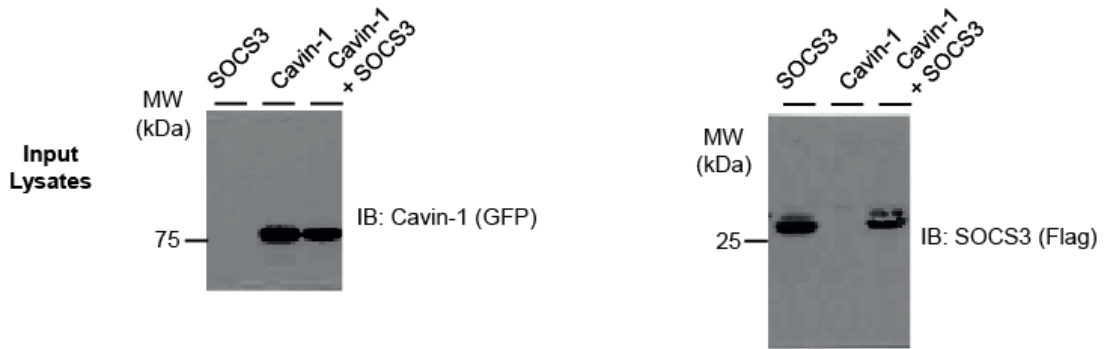
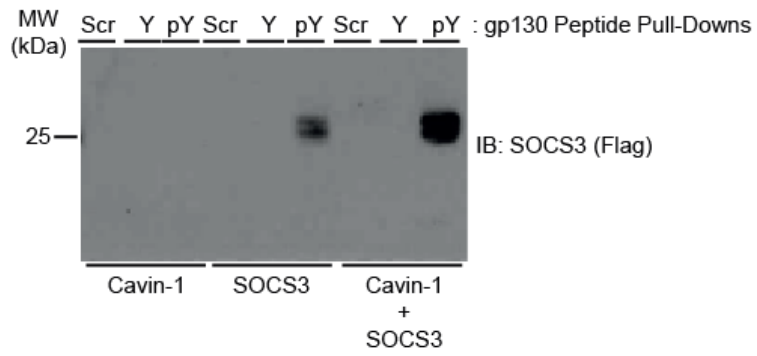


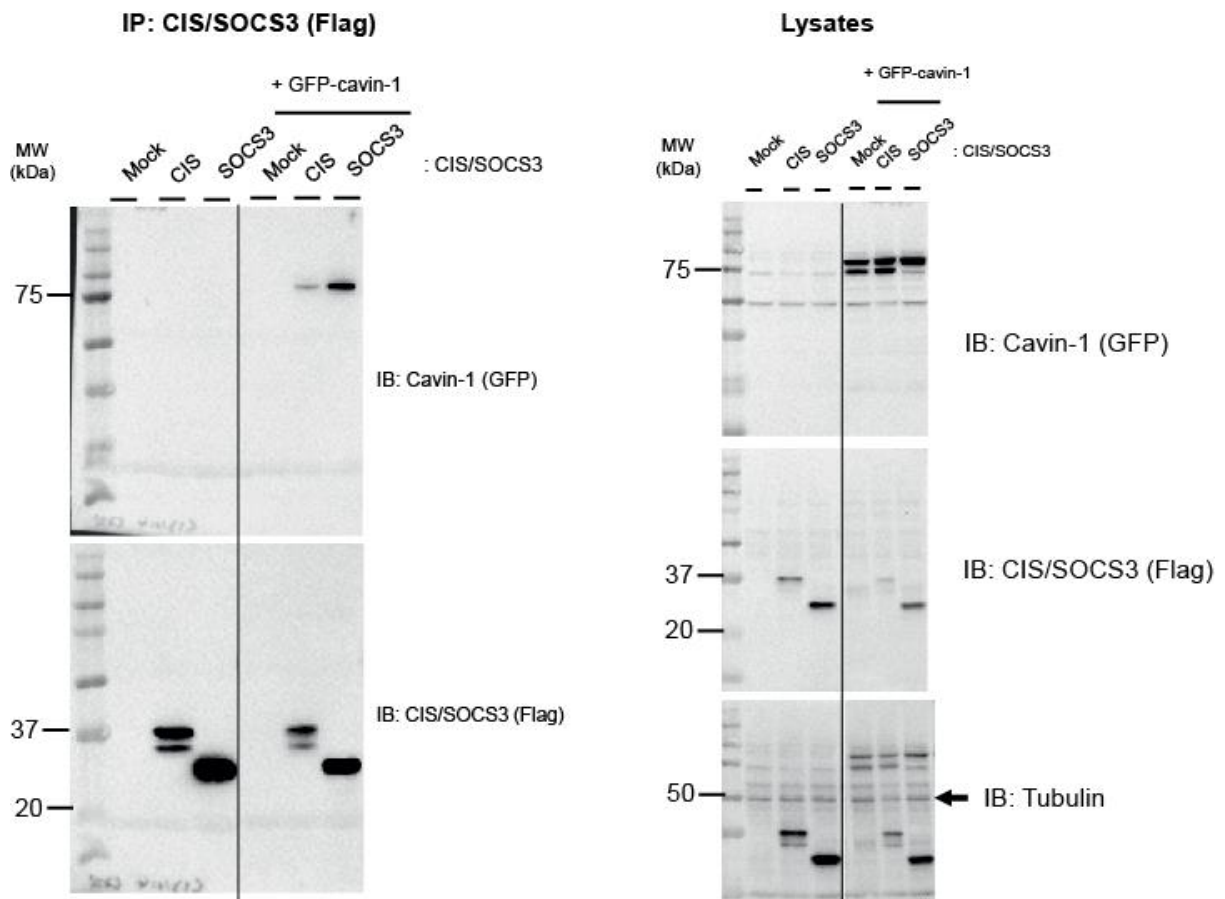
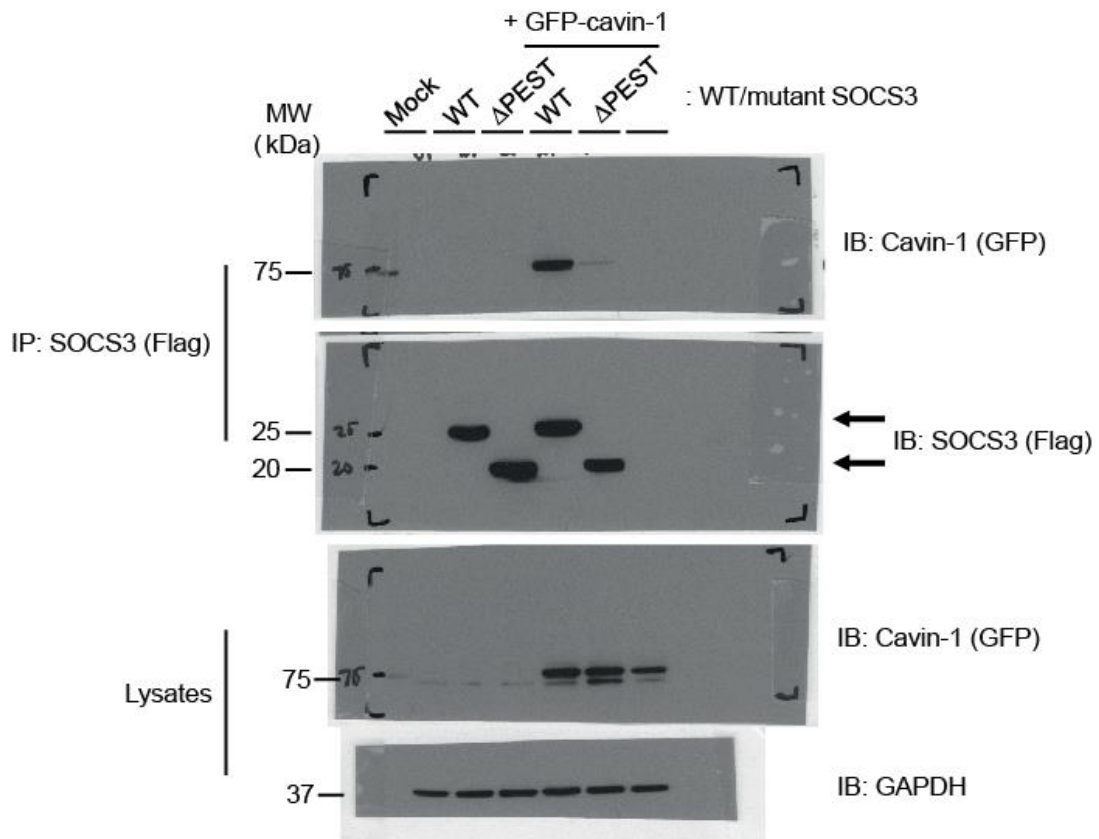


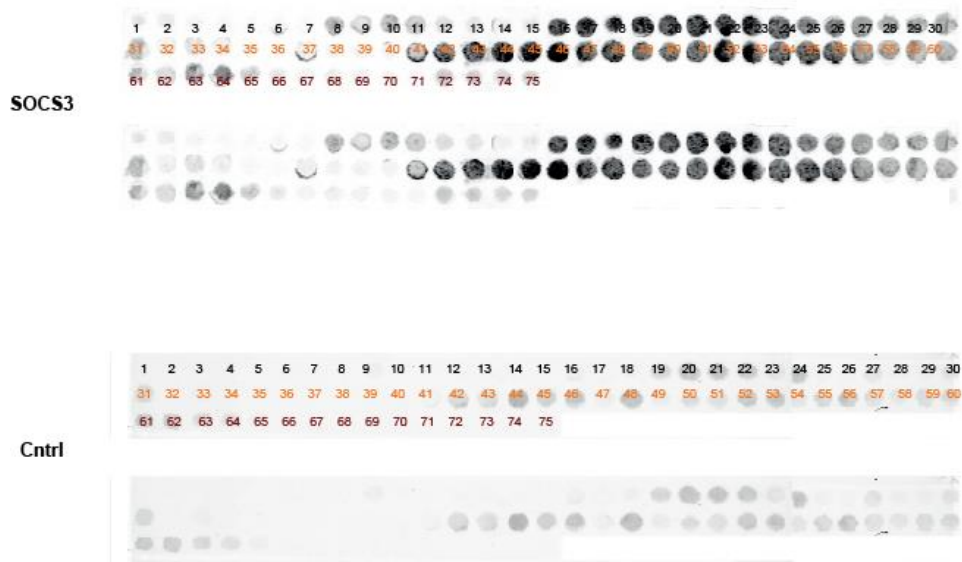
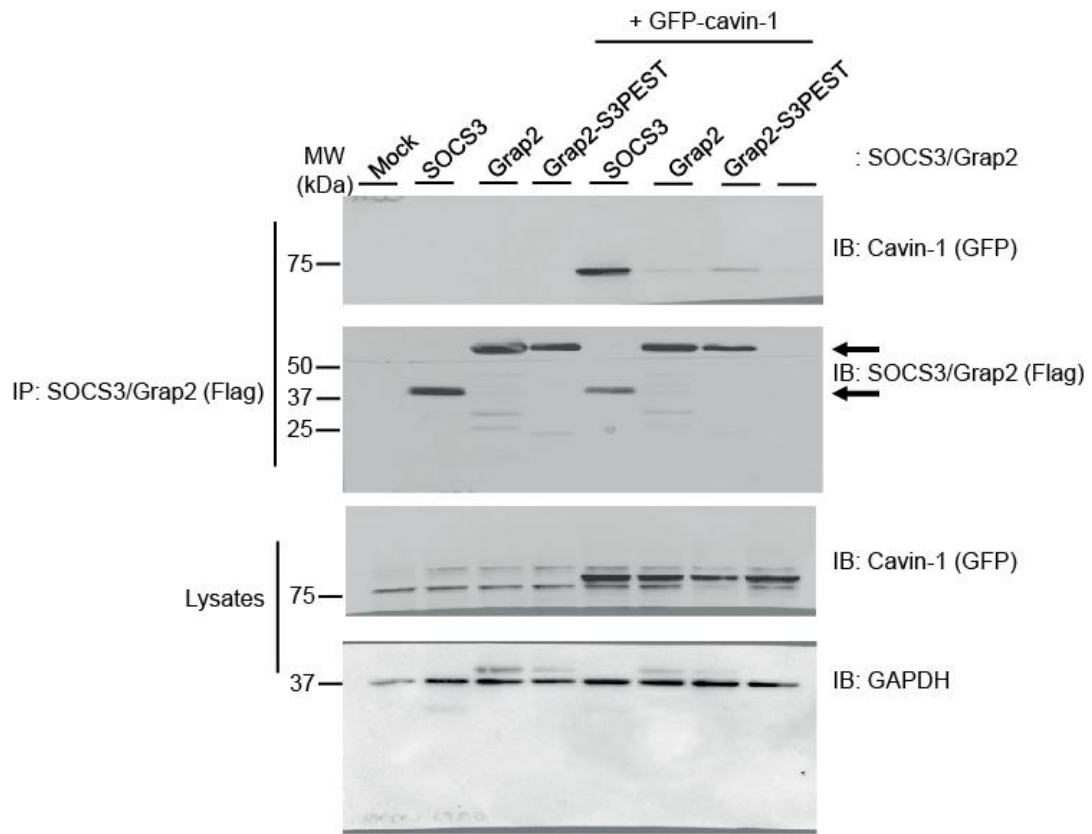


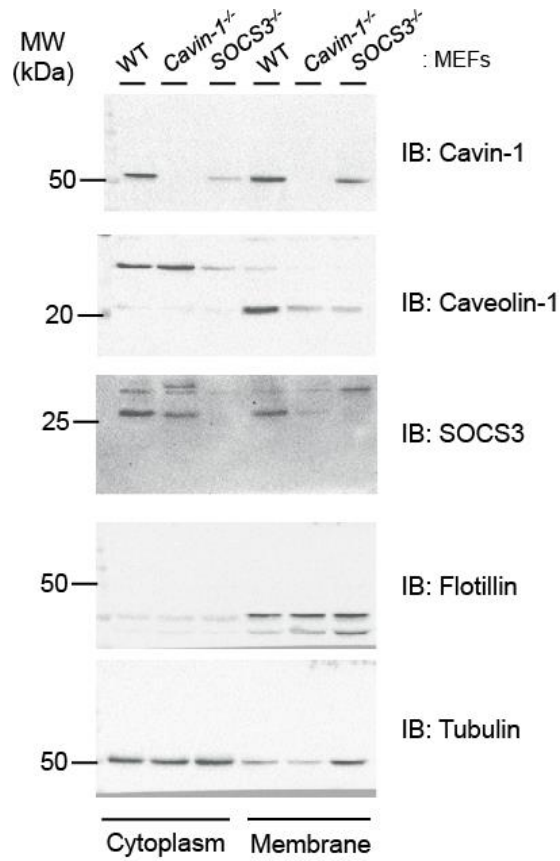
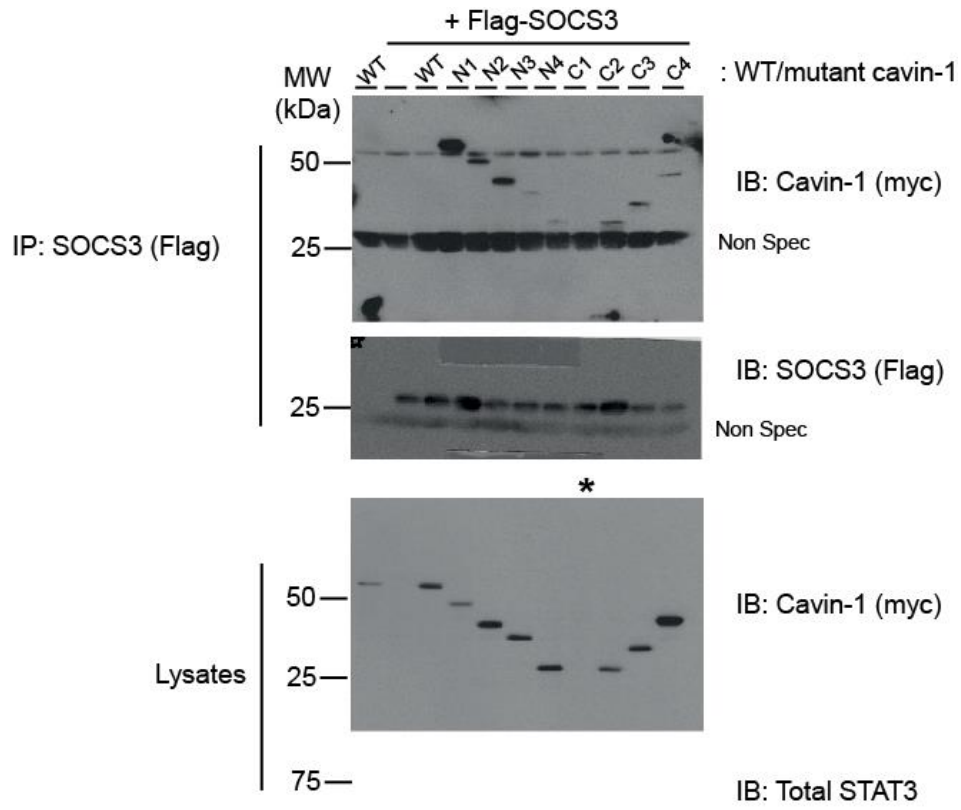


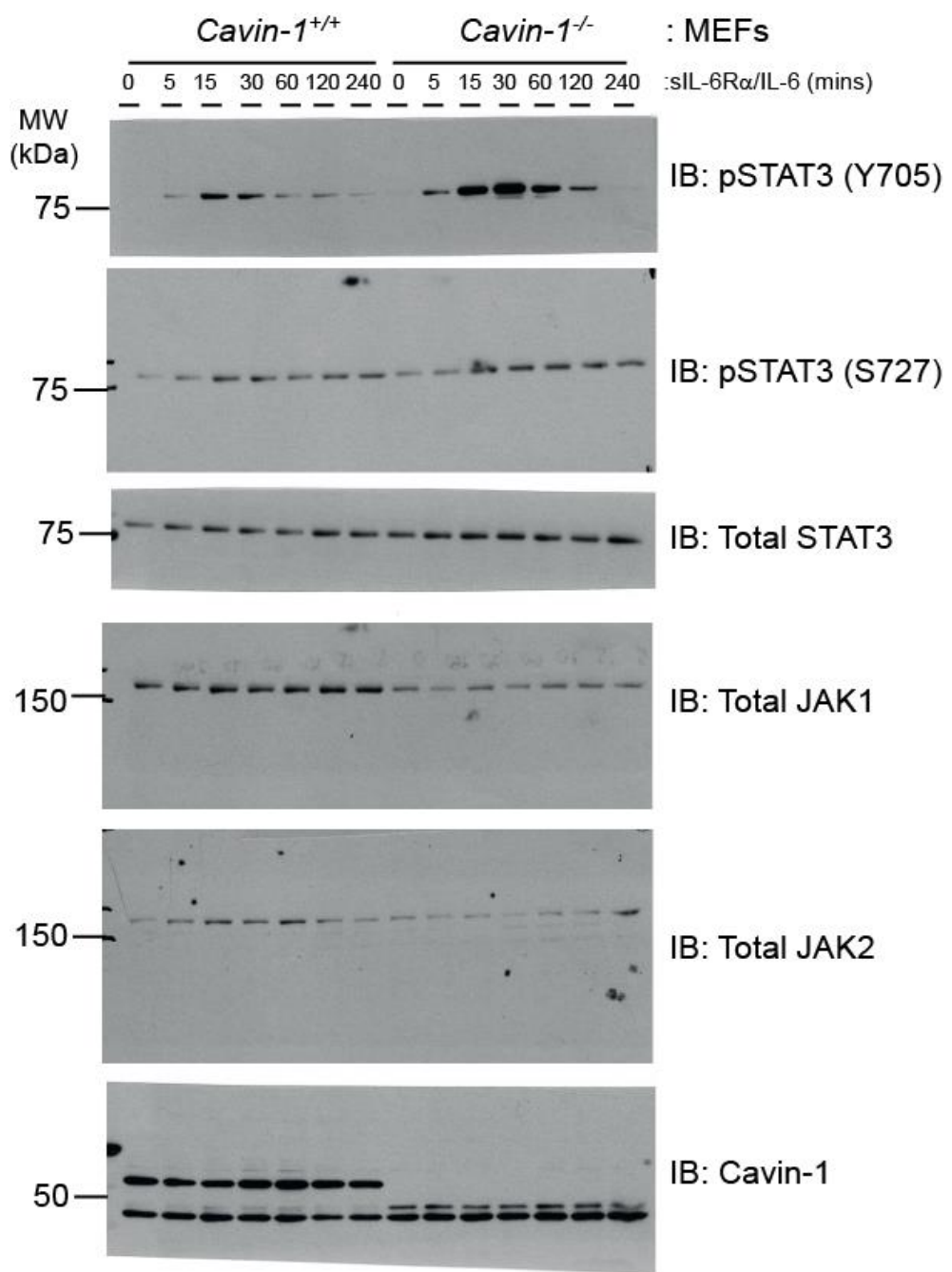




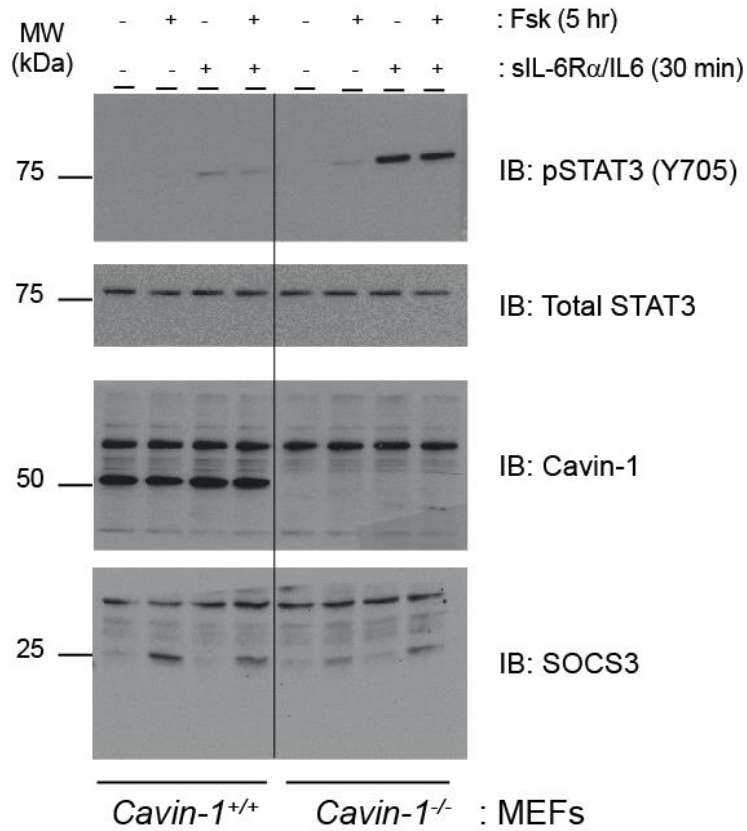
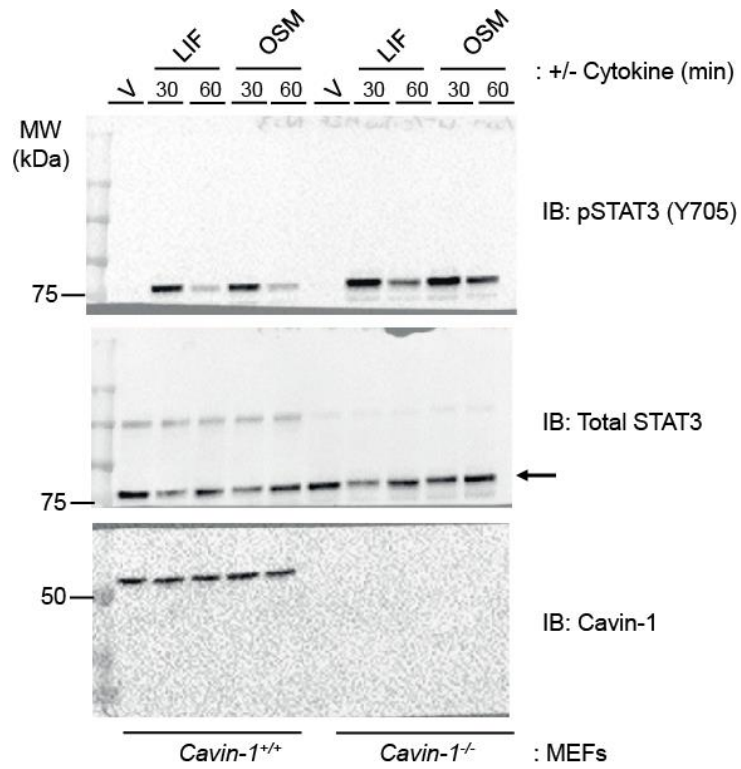












**Supplementary Figure 7:**

Uncropped blots used to generate data presented in Figures 1-9.