The small molecule luteolin inhibits *N*-acetyl-α-galactosaminyltransferases and reduces mucin-type *O*-glycosylation of amyloid precursor protein

Feng Liu¹, Kai Xu⁵, Zhijue Xu¹, Matilde de las Rivas², Xing Li¹, Jishun Lu¹, Ignacio Delso³, Pedro Merino², Ramon Hurtado-Guerrero^{2, 4, *} and Yan Zhang^{1, *}

¹Key Laboratory of Systems Biomedicine (Ministry of Education) and Collaborative Innovation Center of Systems Biomedicine, Shanghai Center for Systems Biomedicine, Shanghai Jiao Tong University, 800 Dongchuan Road, Shanghai 200240, China.

²Instituto de Biocomputación y Fisica de Sistemas Complejos (BIFI), BIFI-IQFR (CSIC) Joint Unit, Universidad de Zaragoza, 50009, Zaragoza, Spain.

³Instituto de Síntesis Química y Catálisis Homogénea (ISQCH). Universidad de Zaragoza. CSIC. E-50009 Zaragoza. Aragón, Spain.

⁴Fundación ARAID, 50018 Zaragoza, Spain.

⁵Key Laboratory of Exploration and Utilization of Aquatic Genetic Resources, Ministry of Education, College of Fisheries and Life Science, Shanghai Ocean University, Shanghai 201306, China.

Running title: Luteolin inhibits ppGalNAc-Ts

*To whom correspondence should be addressed:

Yan Zhang, Shanghai Center for Systems Biomedicine, Shanghai Jiao Tong University, 800 Dongchuan Rd., 200240, Shanghai, China; Tel./Fax: 86-21-34206778; E-mail: yanzhang2006@sjtu.edu.cn.

Ramon Hurtado-Guerrero, Instituto de Biocomputación y Fisica de Sistemas Complejos (BIFI), BIFI-IQFR (CSIC) Joint Unit, Universidad de Zaragoza, 50009, Zaragoza, Spain; Tel.: +34 976 762997, fax: + 34 976 762990; E-mail: rhurtado@bifi.es.

Key words: *O*-glycosylation, ppGalNAc-T, glycosylation inhibitor, glycoprotein, glycosyltransferase, luteolin, amyloid precursor protein (APP), crystal structure

TABLE OF CONTENTS

1. SUPPLEMENTAL TABLE

Table S1. Data collection and refinement statistics.

2. SUPPLEMENTAL FIGURES

Figure S1. Aggregation-based activity of luteolin with ppGalNAc-T2.

Figure S2. Dose-dependent inhibition of ppGalNAc-Ts by luteolin in the presence of EA2 substrate and inhibition of ppGalNAc-T2 by luteolin in the presence of various substrates. Figure S3. Sequence of ppGalNAc-T2.

Figure S4. Surface representation of one of the dimers present in the asymmetric unit.

Figure S5. Stacked ¹H NMR spectra of luteolin in deuterated buffer.

	ppGalNAc-T2 in complex with the luteolin and UDP/Mn ⁺
Space group	P2 ₁ 2 ₁ 2 ₁
Wavelength (Å)	0.97
Resolution (Å)	20-2.30
	(2.42-2.30)
Cell dimensions	a = 116.98
a, b, c (Å)	b = 123.11
	c = 248.15
α, β, γ (°)	90, 90, 90
Mn(I) half-set correlation CC (1/2)	0.997 (0.590)
Unique reflections	151903
Completeness	95.5 (99.5)
R _{pim}	0.050 (0.535)
Ι/σ(Ι)	10.3 (1.6)
Redundancy	6.7 (6.9)
R _{work} / R _{free}	0.213/0.251
RMSD from ideal geometry, bonds (Å)	0.013
RMSD from ideal geometry, angles (°)	1.710
 GalNAc-T2 (Å²)	54.66
 UDP (Å²)	49.23
 luteolin (Å²)	103.02
 solvent (Å²)	44.38
 ethylenglycol(Å²)	64.17
Ramachandran plot:	
Most favoured (%)	96.50
Additionally allowed (%)	3.03
Disallowed (%)	0.48
PDB ID	5NDF

Table S1. Data collection and refinement statistics. Values in parentheses refer to the highest resolution shell. Ramachandran plot statistics were determined with PROCHECK.



Fig. S1 Aggregation-based activity of luteolin with ppGalNAc-T2. (A-C) The particle spectrum change of luteolin in different concentration of TritonX-100 (v/v). The particles spectrum was measured by dynamic light scattering (DLS, Particle Size Analyzer, Z90s, Malvern Instruments Ltd., UK). (D) The change of ppGalNAc-T2 catalytic velocity with different concentrations of TritonX-100 (v/v). (E) The inhibitory potency of luteolin (15 mM) under the different concentrations of TritonX-100 (v/v). 10 mM EDTA was used as 100% inhibitor control. Data are shown as the mean \pm SD (n=3). Two-tailed Student's t-test, **, P<0.01; ***, P<0.001; n.s., no significant difference.



Fig. S2 Dose-dependent inhibition of ppGalNAc-Ts by luteolin in the presence of EA2 substrate and inhibition of ppGalNAc-T2 by luteolin in the presence of various substrates. The IC_{50} values of luteolin for ppGalNAc-Ts were determined under the standard conditions of the HPLC-based assay with EA2, Muc1a, Muc5AC or Muc7 as the substrate (Experimental Procedures). The data are expressed as percentages of the control (DMSO, 100%) and shown as the mean \pm SD (n=3).



Fig. S3 Sequence of ppGalNAc-T2. Secondary structure elements from the ppGalNAc-T2 structure are shown, with a-helices, 3₁₀-helices and b-strands in red.



Fig.S4 Surface representation of one of the dimers present in the asymmetric unit. The surface of one of the monomers is depicted in grey whereas the other one is in orange. The flexible loop is shown in yellow in both ppGalNAc-T2 molecules. The two circles in blue and the red square point out the lectin domain position of both molecules, and the position of luteolin in one of the monomers, respectively.



Fig.S5 Stacked ¹H NMR spectra of luteolin in deuterated buffer: A. Normal-scale spectrum. HDO is used as standard for chemical shift (4.70 ppm), B. Spectrum with increased intensity (300-fold), where luteolin signals arise, C. Aromatic region of the solvent suppression spectrum.