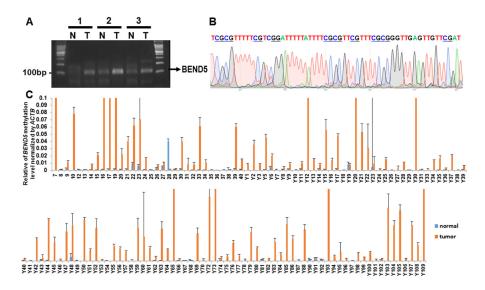
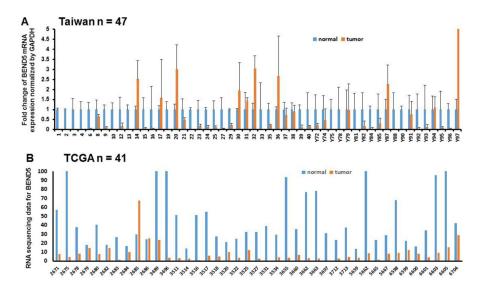
## Hypermethylation of *BEND5* contributes to cell proliferation and is a prognostic marker of colorectal cancer

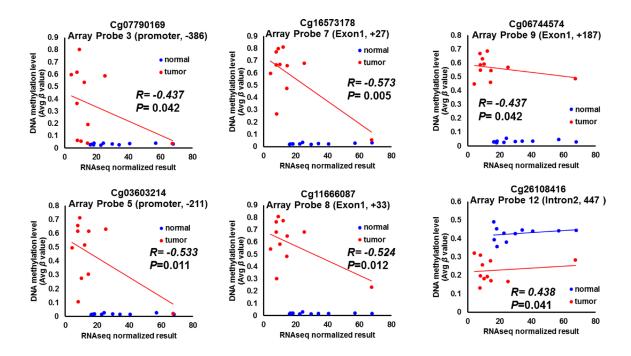
#### SUPPLEMENTARY MATERIALS



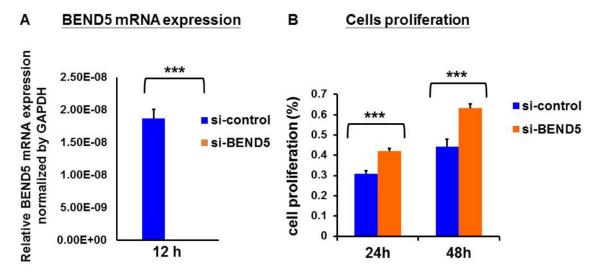
**Supplementary Figure 1: Hypermethylation of the** *BEND5* **promoter gene was analyzed in CRC patients. (A)** The end products of BEND5 QMSP were run in the gel shown on the right molecular weight. (B) Bisulfite sequencing was performed to verify the specificity of the BEND5 QMSP assay. (C) Raw data for the BEND5 QMSP assay in 133 CRC patients. The data are presented as means ± s.d. The experiments were performed with at least three technical replicates.



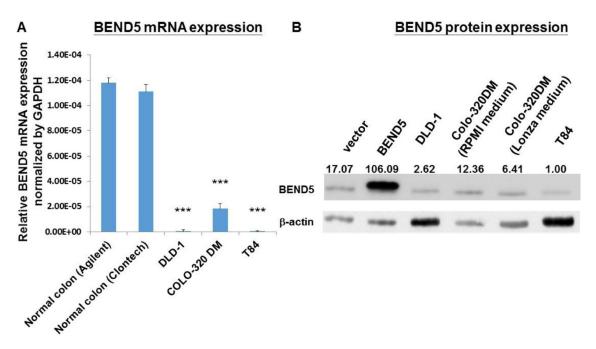
Supplementary Figure 2: The BEND5 mRNA expression levels were analyzed in CRC tumors and paired normal colorectal tissues. (A) The BEND5 mRNA expression levels were analyzed using real-time RT-PCR in 47 patients in Taiwan. The data are presented as means  $\pm$  s.d. The experiments were performed with at least three technical replicates. (B) The BEND5 mRNA expression levels were analyzed through the RNA sequencing of paired tissues from normal controls and 41 paired CRC patients from TCGA.



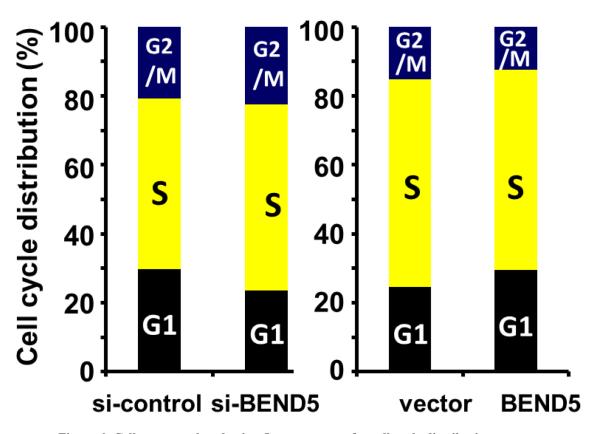
Supplementary Figure 3: Pearson correlation analysis of tissues between DNA methylation and RNA sequencing in 11 normal controls and 11 paired CRC patients.



Supplementary Figure 4: Knockdown of endogenous BEND5 mRNA expression induced proliferation of DLD-1 CRC cells. (A) BEND5 mRNA expression analyzed using real-time RT-PCR. (B) Cell proliferation was analyzed using the SRB assay. The data are presented as means  $\pm$  s.d., \*\*\* $P \le 0.001$ . The experiments were performed with at least three technical replicates.



Supplementary Figure 5: BEND5 mRNA and protein expression analysis in normal tissue and cells. (A) BEND5 mRNA was highly expressed in normal colon tissues but was not expressed in CRC cell lines. The data are presented as means  $\pm$  s.d., \*\*\* $P \le 0.001$ . The experiments were performed with at least three technical replicates. (B) The BEND5 protein expression annualized by Western Blot in CRC cell lines and BEND5 plasmid transfected DLD-1 cell lines.



Supplementary Figure 6: Cells were analyzed using flow cytometry for cell cycle distribution.

# Supplementary Table 1: The clinical parameters of 26 paired CRC patients who analyzed by Illumina Infinium HumanMethylation450 BeadChip array assays

Characteristics	Total  n (%)		
Character issues			
Total	26		
Clinicopathological parameters			
Age < 60	8 (30.8)		
> 60	18 (69.2)		
Sex Male	18 (69.2)		
Female	8 (30.8)		
Tumor type			
Adeno	21 (84.0)		
Mucinous	1 (4.0)		
Others	3 (12.0)		
Tumor stage			
Early (stage I and II)	16 (61.5)		
Late (stage III and IV)	10 (38.5)		
Primary tumor			
Submucosa	1 (3.8)		
Muscularis propria	5 (19.2)		
Subserosa	17 (65.4)		
Penetrate the visceral	3 (11.5)		
Regional lymph nodes			
No regional lymph node metastasis	17 (65.4)		
Metastasis in regional lymph nodes	9 (34.6)		
Distant metastasis			
No distant metastasis	24 (92.3)		
Distant metastasis	2 (7.7)		
Differentiation grade			
Well	3 (11.5)		
Moderate	18 (69.2)		
Poor	5 (19.2)		
Location			
colon	20 (76.9)		
Rectal	6 (23.1)		

Supplementary Table 2: BEND5 protein expression in relation to clinical parameters for CRC<sup>a</sup>

		Total	BEND5 protein			
Characteristics		Total	Low High		<i>P</i> -value	
		$n^{\mathrm{b}}$	n (%)	n (%)		
Overall		94	82 (87.2)	12 (12.8)	'	
Clinicop	pathological parameters					
Age	< 65	66	58 (87.9)	8 (12.1)	0.740	
	> 65	27	23 (85.2)	4 (14.8)		
Sex	Male	68	58 (85.3)	10 (14.7)	0.500	
	Female	26	24 (92.3)	2 (7.7)		
Tumor t	ype					
Adeno		83	71 (85.5)	12 (14.5)	0.402	
Mucinous		10	10 (100.0)	0 (0.0)		
Small c	eell	1	1 (100.0)	0 (0.0)		
Tumor l	ocation					
Colon		74	63 (85.1)	11 (14.9)	0.450	
Rectal		20	19 (95.0)	1 (5.0)		
Differen	tiation grade					
Well		19	17 (89.5)	2 (10.5)	0.845	
Modera	ate	57	48 (84.2)	9 (15.8)		
Poor		6	5 (83.3)	1 (16.7)		
Tumor s	stage					
I		3	3 (100.0)	0 (0.0)	0.407	
II		42	34 (81.0)	8 (19.0)		
III		34	31 (91.2)	3 (8.8)		
IV		15	14 (93.3)	1 (6.7)		
Primary	tumor					
Submi	ıcosa	0	0 (0.0)	0 (0.0)	0.417	
Muscu	ılaris propria	7	6 (85.7)	1 (14.3)		
Subse	rosa	80	71 (88.8)	9 (11.3)		
Penetr	ate the visceral	7	5 (71.4)	2 (28.6)		
Regiona	l lymph nodes					
No reg	gional lymph node metastasis	50	41 (82.0)	9 (18.0)	0.130	
Metas	tasis in regional lymph nodes	44	41 (93.2)	2 (6.8)		
Distant	metastasis					
No distant metastasis		79	68 (86.1)	10 (13.9)	0.683	
Distan	t metastasis	15	14 (93.3)	1 (6.7)		

<sup>&</sup>lt;sup>a</sup>These results were analyzed by the Fisher's exact test.

<sup>&</sup>lt;sup>b</sup>For some categories, the number of samples (*n*) was lower than the overall number analyzed because clinical data were unavailable for these samples.

### Supplementary Table 3: Cox proportional hazard model of clinical parameters and BEND5 methylation levels associated with 5-year overall survival in patients with colorectal cancer from TCGA data set

	Multivariate analysisa			
Variable	HR	95% CI	<i>P</i> -value <sup>C</sup>	
Sex	1.216	0.635–2.327	0.556	
Age	1.036	1.008-1.065	0.012*	
Tumor type	2.674	1.145-6.244	0.023*	
Race	1.103	0.454–2.679	0.829	
Stage	2.836	1.864–4.314	< 0.001***	
BEND5 <sup>b</sup> methylation	2.042	1.061-3.933	0.033*	

<sup>&</sup>lt;sup>a</sup>These results were analyzed by the Cox regression model.

#### Supplementary Table 4: The predicted BEND5 binding sites in promoters of genes

Supplementary Tuble to the producted BELLIE Shirting Steel in producted of genes		
Gene name	Sequence (5' to 3')	
BCL2	TTAATCAGAAGA	
CCNB1	TCCAATTGCTGA	
CCND1	CCAATTAGGAA	
CCND2	TCCTTAATAACGAGA	
PCNA	CCAATGGGAG	
CDKN1A	TATTCAGGAGA	

 $<sup>^{</sup>b}$ BEND5 was considered hypermethylated at an average  $\beta$  value of > 0.5.

 $<sup>^{</sup>c*}P < 0.05$ ;  $^{**}P < 0.01$ ;  $^{***}P < 0.001$ .

## Supplementary Table 5: List of primers sequences and their reaction conditions used in the present study

Real-t	time RT-PCR <sup>a</sup>	Sequence (5' to 3')	Probe	Tm (°C)	Size (bp)
CARRIA	Forward	AGCCACATCGCTCAGACAC	W.C.		
<i>GAPDH</i>	Reverse	GCCCAATACGACCAAATCC	#60	60	66
BEND5	Forward	AGCCACATCGCTCAGACAC		60	95
	Reverse	GCCCAATACGACCAAATCC	#1		
BCL2	Forward	ACAGAGGATCATGCTGTACTTAAAAA		60	96
	Reverse	TTATTTCATGAGGCACGTTATTATTAG	#6		
	Forward	ACATGGTGCACTTTCCTCCT		60	103
CCNB1	Reverse	AGGTAATGTTGTAGAGTTGGTGTCC	#18		
	Forward	GACCTTCGTTGCCCTCTGT	"10	60	105
CCND1	Reverse	CTCAGGTTCAGGCCTTGC	#49		
	Forward	GGACATCCAACCCTACATGC		60	71
CCND2	Reverse	CGCACTTCTGTTCCTCACAG	#49		
	Forward	TGGAGAACTTGGAAATGGAAA	4.50	60	95
PCNA	Reverse	GAACTGGTTCATTCATCTCTATGG	#69		
an muli	Forward	TCACTGTCTTGTACCCTTGTGC	//22	60	128
CDKN1A	Reverse	GGCGTTTGGAGTGGTAGAAA	#32		
	Forward	CCAACCGCGAGAAGATGA		60	97
ACTIN	Reverse	CCAGAGGCGTACAGGGATAG	#64		
Quantitativ	e methylation-spec	cific PCR	l l		
	Forward	TGGTGATGGAGGAGGTTTAGTAAGT		60	132
ACTB	Reverse	AACCAATAAAACCTACTCCTCCCTTAA			
	TagMan probe	ACCACCACCAACACACAATAACAAACACA			
	M-Forward	GTTTGGGTTTTGGGGAGTC		63	109
BEND5	M-Reverse	GATCGAACAACTCAACCCG			
	TagMan probe	CGAAAATAAAAATCCGACGA		1	
GAPDH	Forward	GGT AGT TGT TAG GAA AGT TTG TTG G			120
	Reverse	TAA TCT AAA AAA AAC ATC ACC CAA A		60	138

<sup>&</sup>lt;sup>a</sup>RT-PCR: reverse-transcriptase polymerase chain reaction.