Gut	epimerase	Gut	SmedASXL_014050	Gut	A009946 pXW0105	Gut	A013828 pXW0034
	C	-	Com	*99	Contraction of the second	: 393	Kanal
Gut	SmedASXL_008095	Gut	SmedASXL_052019	Gut	SmedASXL_003002	Gut	SmedASXL_019073
-	Car	-	and a state of the	Antonia a	Contraction of the second	· ·	R.
Gut	SmedASXL_004676	Gut	SmedASXL_017839	Gut	SmedASXL_002380	Gut	SmedASXL_077649
•			Contraction of the second		Cield -		C
Gut	SmedASXL_015989	Gut	SmedASXL_016646	Epidermal	SmedASXL_045004	Epidermal	SmedASXL_004972
	and a little and the second		A A				0
Epidermal	SmedASXL_015008	Anterior	SmedASXL_016543	Anterior	SmedASXL_013187	Pharynx	SmedASXL_065556
	$\bigcirc$						KE -
Para-pharynx	SmedASXL_004874	Para-phary	nx SmedASXL_011934	Para-pharyn>	SmedASXL_012302	Para-pharyn	x SmedASXL_008647
-	Carl	•••					C
Neural	SmedASXL_019161	Neural	SmedASXL_009860	Neural	SmedASXL_005730	Other	SmedASXL_012033
			0	:	E		0

#### Figure S1

Figure S1. Expression Patterns of Pigment Cell Marker Candidates.

Expression patterns were determined by whole mount *in situ* hybridization (WISH). All genes shown have down-regulation in depigmented animals in whole worm RNAseq dataset. 14 genes show gut expression patterns, 3 show epidermal expression patterns, 2 show anterior expression patterns, 1 shows pharyngeal expression pattern, 4 show para-pharyngeal expression patterns and 3 show neural expression patterns. Scale bar: 100µm.

#### Figure S2





A. Planarian pigmentation is gradually but completely lost over the course of 9 days. Loss in both dendritic<sup>+</sup> and punctate<sup>+</sup> pigment cell population, detected by WISH, is observed alongside depigmentation. Punctate<sup>+</sup> pigment cell loss happens prior to dendritic<sup>+</sup> cell loss. **B.** During light induced depigmentation, punctate<sup>+</sup> pigment cell population drops first. Significant decreases were observed from D0 to D3, and from D3 to D6, but remained at similar low levels from D6 to D9. Dendritic<sup>+</sup> pigment cell population drops at later time points during depigmentation. No significant changes were observed from D0 to D3. Significant decreases of dendritic<sup>+</sup> pigment cells were observed from D3 to D6, and from D6 to D9. Scale bar: 100µm. p-values, \* p<0.05, \*\* p<0.01, \*\*\* p<0.001, n. s. not significant.</p>





# Figure S3. The effect of *zfp-1* RNAi on the population of dendritic and punctate markers during repigmentation and regeneration.

A. Fluorescent *in situ* hybridization of dendritic<sup>+</sup> (*PBGD-1*) and punctate<sup>+</sup> (*lysoLP-1*) pigment cells during repigmentation (Day 1 to Day 6) in control and *zfp-1* RNAi animals. 24hpi Repig D1: Animals were lethally irradiated at Repig D0 and fixed 24 hours after irradiation, equivalent to Repig D1 time point. Establishing a baseline of dendritic<sup>+</sup> and punctate<sup>+</sup> population during repigmentation in absence of entire neoblast population. B. Quantification of pigment cell populations in repigmenting control and *zfp-1* RNAi animals as shown in A. The difference in the population of dendritic<sup>+</sup> and punctate<sup>+</sup> pigment cells is non-significant between control and *zfp-1* RNAi animals at all time point. C. Whole mount in situ hybridization of PBGD-1<sup>+</sup> and lysoLP-1<sup>+</sup> pigment cells during head and tail regeneration in control and zfp-1 RNAi animals. Scale bars: 250µm. n. s. not significant.



#### Figure S4

## Figure S4. Retention of Dendritic<sup>+</sup> and Punctate<sup>+</sup> Cells After Lethal Irradiation Suggests Slow Turn Over Rates of Planarian Pigment Cells.

**A.** Detected by double fluorescent *in situ* hybridization (dFISH), both dendritic<sup>+</sup> and punctate<sup>+</sup> population remained comparable to that of control group 14 days after exposure to lethal dosage of  $\gamma$ -irradiation. **B.** No significant changes in either dendritic<sup>+</sup> or punctate<sup>+</sup> population density can be observed from animals D10 to D14 after lethal irradiation in comparison to control group. During this time, the animals did not shrink in size significantly, either. Thus, the unchanged pigment cell density is not due to the reduction in animal size, but rather due to the unchanged absolute numbers of pigment cells. Scale bars: 250µm. n. s. not significant.





### Figure S5. Dynamics of pigment cells during repigmentation + lethal irradiation.

**A.** Pigment cell lineage progresses toward punctate-positive stage without presence of stem cells. Animals were lethally irradiated on Day 8 of repigmentation, and pigment cell lineage progression were shown by double fluorescent *in situ* hybridization with dendritic (*PBGD-1*) and punctate (*lysoLP-1*) markers. **B.** Population dynamics of dendritic<sup>+</sup> and punctate<sup>+</sup> cells following repigmentation and irradiation. Statistics: two-tailed, unpaired *t*-test. Scale bars: 250µm. *p*-values, \* *p*<0.05, \*\* *p*<0.01, \*\*\* *p*<0.001, n. s. not significant.





Figure S6. Effects of *hnf4(RNAi)* on repigmentation and regeneration of pigment cells.

**A.** Whole mount *in situ* hybridization of *PBGD-1*<sup>+</sup> pigment cells in repigmenting control and hnf4(RNAi) planarians. Following RNAi of hnf4, the number of *PBDG-1*<sup>+</sup> cells on day 4 of repigmentation remains the same, but the staining intensity is decreased compared to controls. Planarians were exposed to constant red light (625 nm) rather than white light for the final three days of the depigmentation protocol to optimize worm health. **B.** Whole mount *in situ* hybridization of *PBGD-1*<sup>+</sup> pigment cells in regenerating control and hnf4(RNAi) planarians. Scale bars: 250 $\mu$ m.







Each row indicates various markers of cell type and each column is RNAi to a given gene. Stains were performed at the end point of animal health and sometimes on the precipice of head regression and animal lysis. Despite this, we did not find overt effects on any given cell type.

Table S1

Click here to Download Table S1

Table S2

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