

c

b

e

d

Sox10+/tdTomato+ cells p75+/tdTomato+ cells GFAP+/tdTomato+ cells

Activated glia outside NB

Supplementary Figure 1 *Plp-CreERT2* lineage tracing in intact and injured skin. **a** tdTomato reporter expression in nerve bundles (NB) and around hair follicles (HF). **b** Immunofluorescence staining of *Plp-CreERT2* traced cells in granulation tissue for $tdTomato$ (red) and $Sox10⁺$ (green) and quantification of the percentage of $Sox10⁺$ per tdTomato+ cells in the granulation tissue of injured skin at D14 (N=2, n=6). **c** Immunofluorescence staining of tdTomato⁺ *Plp-CreER^{T2}*-traced cells in granulation tissue (red) with $p75^+$ (green) and quantification of the percentage of $p75^+$ per tdTomato+ cells in the granulation tissue of injured skin at D7 (N=4, n=10). **d** Immunofluorescence staining of D14 tdTomato⁺ *Plp-CreER^{T2}* traced cells (red) with glial fibrillary acidic protein (GFAP) (green) and quantification of the percentage of GFAP⁺ per tdTomato⁺ cells in the granulation tissue of injured skin at D7 (N=3, n=10) and D14 (N=2, n=6). **e** Immunofluorescence and corresponding quantification of proliferative (Ki67+), activated (c-Jun+), de-differentiated (p75+) glial cells outside NBs (N=6, n=9). Data are represented as mean±SEM. Scale bars, 50µm (**a**), 25µm (**b**,**c**,**d**).

a Plp-CreER^{T2} tdTomato 30h Tamoxifen pulse **b** Quantification of Plp-CreER^{T2} traced hair

follicular melanocytes

^c Immunolabeling of melanocytic lineage in D7 wounds

d

Dct+/tdTomato+ cells

α-SMA+/tdTomato+ cells

Supplementary Figure 2 *Plp-CreERT2*-mediated recombination of adult melanocytes in anagen hair follicles and absence of melanocytic lineage contribution to the wound bed of injured skin. **a** *Plp-CreERT2* driven recombination of anagen murine skin shows only partial tdTomato reporter expression in hair follicles (HFs), 30h post TM injection. Co-labelling of tdTomato (red) and the melanocytic marker Dopachrome tautomerase Dct (green). **b** Quantification of percentage of HFs displaying no, partial or complete *Plp-CreER^{T2}* mediated recombination of their respective melanocytic population, 30h post-TM injection (n=220 HFs, N=1). **c** Immunolabelling for tdTomato reporter (red) and Dct (green) of *Plp-CreER^{T2}* and *Tyr-CreER^{T2}* traced melanocytic cells in D7 wound bed and in an adjacent HF fails to reveal traced melanocytes in the granulation tissue upon injury. Sections were counter stained with Hoechst 33258 (blue). **d** Immunofluorescence staining of tdTomato⁺ *Plp-CreER^{T2}* traced cells (red) in granulation tissue with melanocyte marker Dopachrome tautomerase (Dct) (green) and quantification of the percentage of Dct⁺ per tdTomato⁺ cells in the granulation tissue of injured skin at D7 (N=3, n=8) and D14 (N=2, n=5). **e** Immunofluorescence staining of tdTomato+ *Plp-CreERT2* traced cells (red) with myofibroblast marker alpha smooth muscle actin (α-SMA) (green) and quantification of the percentage of α-SMA+ per $tdTomato⁺$ cells in the granulation tissue of injured skin at D7 (N=3 animals, n=11 wounds) and D14 (N=2, n=6). Scale bars, 50µm.

Supplementary Figure 3 Fate of *Plp-CreERT2*-traced cells after *Sox10* cKO. **a** Schematic of the murine model used to trace glial cells in skin of *Sox10* cKO mice. **b** Immunolabelling of Sox10 (green) and tdTomato tracer (red) of D14 *Sox10* cKO wound bed. Black arrowhead, tdTomato⁺ Sox10⁻ cell; white arrowhead, tdTomato⁺ Sox10⁺ cell **c**,**d** Corresponding quantification of Sox10 expression among *Plp-CreERT2*-traced tdTomato+ cells (**c**) within nerve bundles (NBs) and (**d**) in the wound bed (N=2 animals, n=4 wounds for each condition). Scale bar, 20µm.

Supplementary Figure 4

Supplementary Figure 4 Neovascularization, immune cell infiltration and collagen maturation are not affected upon loss of glial contribution to tissue repair. **a** Immunolabelling of CD31 (red) in granulation tissue at D7 and related quantification of number of blood vessels mm⁻² and percentage of CD31⁺ area per total granulation tissue surface in skin wounds at D7 did not display noticeable differences upon loss of *Sox10*. **b** Immunolabelling of CD45 (red) in granulation tissue at D7 and quantification of CD45⁺ cells in the skin wounds at D7 and D14 did not show any difference upon *Sox10* depletion. **c** Immunolabelling of CD206 (red) in granulation tissue at D7 and quantification of CD206+ area in skin wounds at D7 showed no significant changes upon *Sox10* cKO. **d** Herovici staining for collagen maturation performed on D14 sections of Ctrl and *Sox10* cKO wounds. Data are represented as mean±SEM of N=3, n=11 (Ctrl), N=2, n=6 (*Sox10* cKO) (**a** – blood vessels mm-2); N=3, n=7 (Ctrl), N=6, n=9 (*Sox10* cKO) (**a** – blood vessel area); N=3, n=10 (Ctrl D7), N=5, n=8 (Ctrl D14); N=5, n=9 (*Sox10* cKO D7), N=4, n=7 (*Sox10* cKO D14) (**b**); N=5, n=8 (Ctrl), N=8, n=12 (*Sox10* cKO) (**c**) Scale bars, 25µm (**a**-**c**), 100µm (**d**).

a Western blot analysis of α-SMA in total D7 wound protein

b Western blot analysis of Periostin in total D7 wound protein

β-actin

Periostin

Supplementary Figure 5 Full scans of western blots performed on lysed wound samples. **a** Full scans of α-SMA western blots corresponding to quantification in Figure 5j (N=3, n=6) Human fibroblasts treated with TGF-β1 were used as control. **b** Full scans of Periostin western blots corresponding to quantification in Figure 5k (N=3, n=6) Samples were standardized to β-actin expression.

Supplementary Table 1 Top upregulated growth factor and cell signalling related genes with reported functional relevance.

Supplementary Table 2 Top upregulated TGF-β signalling related genes with reported functional relevance.

Supplementary Table 3 Top upregulated chemotaxis and inflammation related genes with reported functional relevance.

Supplementary Table 4 Top upregulated migration and adhesion related genes with

reported functional relevance.

Supplementary Table 5 Top upregulated ECM related genes with reported functional

relevance.

Supplementary Table 6 Top upregulated angiogenesis related genes with reported functional relevance.

Supplementary Table 7 Genotyping primers.

Supplementary Table 8 Primary antibodies.

Supplementary Table 9 Secondary antibodies.

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