

Combined use of EpCAM and FR α enables the high-efficiency capture of circulating tumor cells in non-small cell lung cancer

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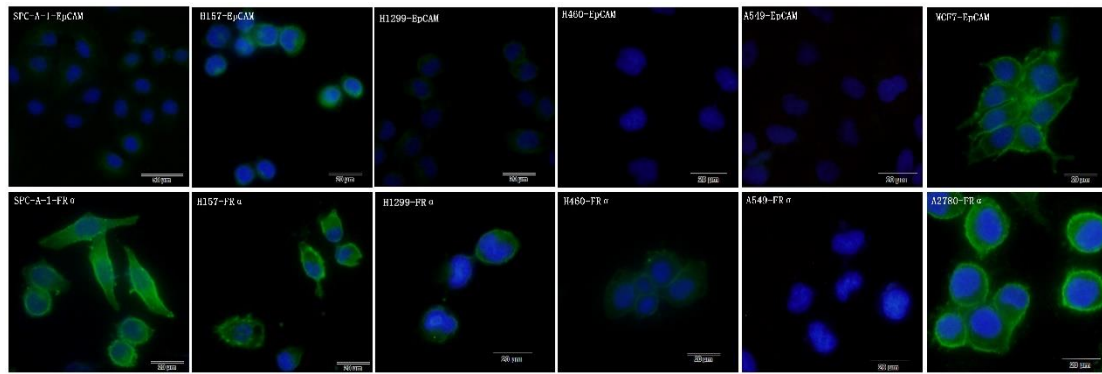


Fig.S1 Immunofluorescent detection of EpCAM and FR α expression in 5 types of NSCLC cells.

EpCAM and FR α stained with Alexa Fluor[®] 488 are green at an excitation of 488 nm, and the nuclei stained with DAPI are blue at an excitation of 405 nm. As a positive control, we used EpCAM-stained MCF7 cells, which are highly expressed EpCAM, and FR α -stained A2780 cells, which are highly expressed FR α .

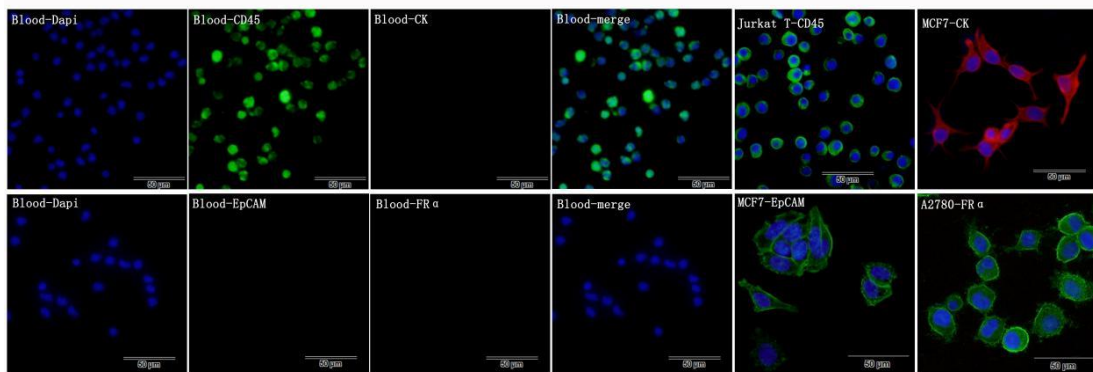


Fig. S2 Immunofluorescent detection of the expressions of CK19, CD45, EpCAM, and FR α in peripheral blood mononuclear cells: CD45s stained with Alexa Fluor[®] 488 are green at an excitation of 488 nm; CK19s stained with Alexa Fluor[®] 568 are red at an excitation of 568 nm; EpCAM and FR α stained with Alexa Fluor[®] 488 are green at an excitation of 488 nm; and the nuclei stained with DAPI are blue at an excitation of 405 nm. As a positive control, we used CD45-stained Jurkat cells, which highly express CD45, CK19-stained MCF7 cells, which highly express CK19, EpCAM-

stained MCF7 cells, which highly express EpCAM, and FR α -stained A2780 cells, which highly express FR α .

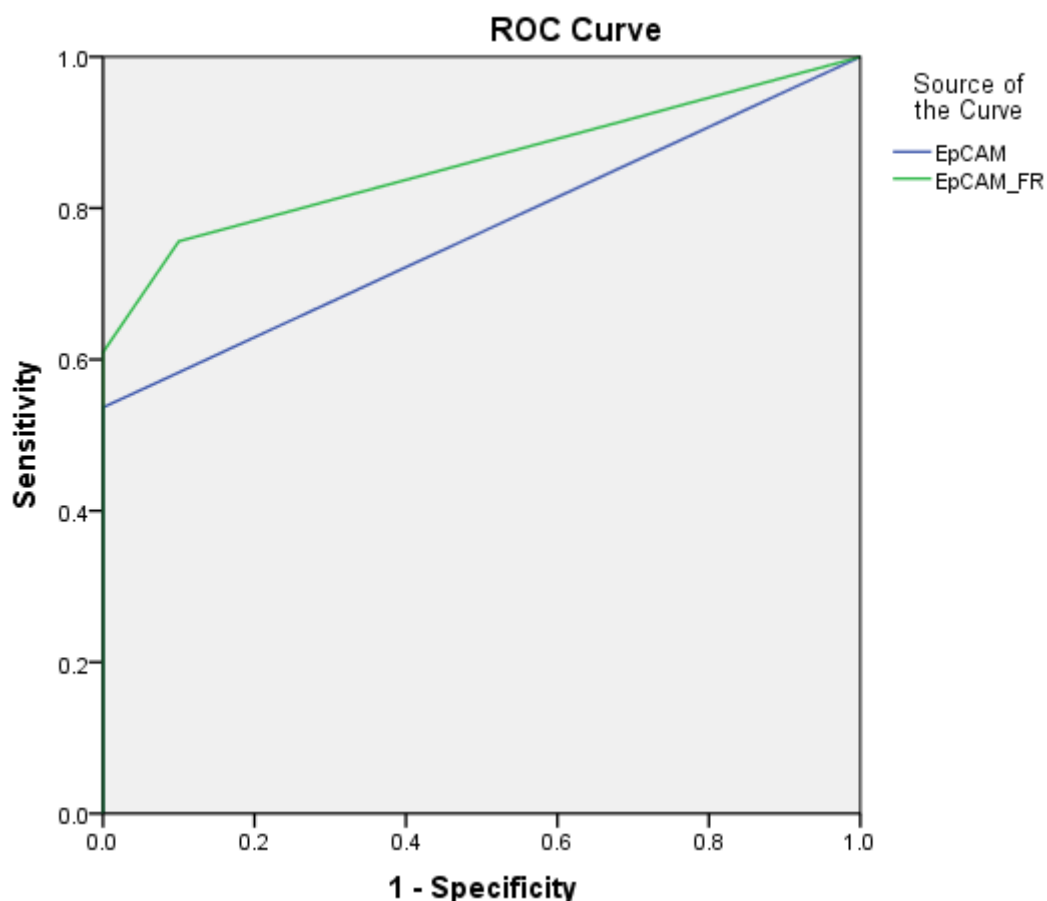


Fig. S3 ROC plots of using anti-EpCAM-MNs plus anti-FR α -MNs & anti-EpCAM-MNs to detect CTCs from 41 NSCLC patients and 10 healthy donors. The area under the receiver operating characteristic curve (AUC-ROC) analysis showed that the combination of anti-EpCAM-MNs and anti-FR α -MNs had a higher AUC (0.8585, 95% CI: 0.7579-0.9592, $p=0.0005$) than that of anti-EpCAM-MNs alone (0.7683, 95% CI: 0.6384-0.8982, $p=0.0091$).