

Supporting information

Peptide Ligands for Targeting the Extracellular Domain of EGFR: Comparison Between Linear and Cyclic Peptides.

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Table 1. Chemical shift, coupling constant and temperature dependence of amide resonances of ^1H NMR resonances for CycloL1 in DMSO- d_6 .

Sequence	Chemical shift in ppm (δ)					$\Delta\delta\Delta/T$ ppb/K	$^3J_{\text{HN}\alpha}$
	NH	CH	CH	CH	Other		
K1	7.78	4.26	1.63	1.54		N.D	N.D
L2	8.30	4.22	1.73	1.49	0.88	3.5	↓
A3	8.10	4.06	1.26			4.9	↓
R4	8.42	3.69	1.85	1.47	3.14	4	↓
L5	7.79	4.34	1.62	1.54	0.85	2.6	↓
L6	7.77	4.15	1.63	1.55	0.86	2.3	↓
T7	8.50	4.27	3.90	1.04		4.3	↑

*Upside arrows represent the coupling constant ≥ 6 Hz and downward arrows indicate coupling constants ≤ 5 Hz. N.D not determined.

Table 2. Chemical shift, coupling constant and temperature dependence of amide resonances of of ¹H NMR resonances for CycloL1.1 in DMSO-d₆.

Sequence	Chemical shift in ppm (δ)					$\Delta\delta\Delta/T$ ppb/K	³ J _{Hα}
	NH	CH	CH	CH	Other		
K1	8.04	3.93	1.75	1.49	3.11	5.1	↑
l2	8.22	4.33	1.64	1.50	0.88	4.3	↓
a3	7.59	4.01	1.07			5.1	↓
r4	8.16	4.09	1.81	1.57		5.7	↑
l5	7.90	4.05	1.82	1.60	0.89	4.8	↑
l6	7.90	4.33	1.64	1.50	0.88	0.7	↑
t7	8.23	4.16	1.35			3.3	↓

*Upside arrows represent the coupling constant ≥ 6 Hz and downward arrows indicate coupling constants ≤ 5 Hz. L amino acid is represented by capital letter and D amino acids by small letter.

Table 3. ¹H NMR NOESY data with cross-peak connectivities indicating spatial connectivities between the protons for **Cyclo.L1**.

Connectivity between the protons		Connectivity between the protons	
$\alpha\text{N (i,i)}$		$\beta\text{N (i,i)}$	
K1NH	K1 α H	K1NH	K1 β H
L2NH	L2 α H	L2NH	L2 β H
A3NH	A3 α H	A3NH	A3 β H
R4NH	R4 α H	R4NH	R4 β H
L5NH	L5 α H	L5NH	L5 β H
L6NH	L6 α H	L6NH	L6 β H
T7NH	T7 α H	T7NH	T7 β H
$\alpha\text{N (i,i+1)}$		Other	
K1 α H	L2NH	K1NH	K1 γ H
L2 α H	A3NH	L2NH	L2 γ H
A3 α H	R4NH	L2NH	L2 δ H
R4 α H	L5NH	R4NH	R4 γ H
L6 α H	T7NH	L5NH	L5 γ H
NN (i,i+1)		L5NH	L5 δ H
T7NH	K1NH	L6NH	L6 γ H
R4NH	L5NH	L6NH	L6 δ H
R4 NH(Guanidine)	L5NH	K1 α H	K1 β H
NH-side chain long range		L2 α H	L2 β H
K1NH	T7 β H	L2 α H	L2 γ H
T7NH	L6 β H	L2 α H	L2 δ H
R4NH	A3 β H	R4 α H	R4 β H
A3NH	L2 β,γ,δ H	T7 α H	T7 β H
		T7 α H	T7 γ H
		A3 α H	A3 β H
		L5 α H	L5 β, γ H

Table 4. ¹H NMR NOESY data with cross-peak connectivities indicating spatial connectivities between the protons for **Cyclo.L1.1**.

Connectivity between the protons		Connectivity between the protons	
Nα (i,i)		βN (i,i)	
K1NH	K1 α H	K1NH	K1 β H
l2NH	l2 α H	l2NH	l2 β H
a3NH	a3 α H	a3NH	a3 β H
r4NH	r4 α H	r4NH	r4 β H
l5NH	l5 α H	l5NH	l5 β H
l6NH	l6 α H	l6NH	l6 β H
t7NH	t7 α H		
αN (i,i+1)		Other	
K1 α H	l2NH	K1NH	K1 γ H
a3 α H	r4NH	l2NH	l2 γ H
r4 α H	l5NH	l2NH	l2 δ H
l6 α H	t7NH	r4NH	r4 γ H
t7 α H	K1NH	l5NH	l5 γ H
NN (i,i+1)		l5NH	l5 δ H
t7NH	l6NH	l6NH	l6 γ H
l2NH	K1NH	l6NH	l6 δ H
NN (long range)		r4 α H	r4 β H
l2NH	l5/6NH	r4 α H	r4 γ H
K1NH	K1 ϵ H	l2 α H	l2 β H
NH-side chain long range		l2 α H	l2 γ H
K1NH	t7 β H	l2 α H	l2 δ H
r4NH	a3 β H	t7 α H	t7 γ H
a3NH	l2 β,γ H	l6 α H	l6 β H
		l6 α H	l6 δ H

*L amino acids are represented by capital letters and D amino acids are represented by smallletters.

Figure legends for supporting information

Figure S1. HR-MS for Cyclic peptides A) Cyclo(KLARLLT) and B) Cyclo(KN3)larllt).

Figure S2. SPR analysis of linear peptide GLARLLT. EGFR was immobilized on CM5 sensor chip and different concentrations of peptide was injected to obtain the SPR response. Binding of peptide GLARLLT to EGFR was observed with increasing concentration of the peptide.

Figure S3. Curve fitting of SPR sensorgram for **Cyclo.L1.1** using 1:1 Langmuir interaction describing the single exponential of the data. Black lines are experimental data and different colored lines represent fitted lines. K_D values reported were from k_{on} and k_{off} calculations.

Figure S4. Curve fitting of SPR sensorgram for **L1.7** using 1:1 Langmuir interaction describing the single exponential of the data. Black lines are experimental data and different colored lines represent fitted lines.

Figure S5. HPLC for peptide **L1**.

Figure S6. HPLC for peptide **L1.3**.

Figure S7. HPLC for peptide **L1.4**.

Figure S8. HPLC for peptide **L1.5**.

Figure S9. HPLC for peptide **L1.6**.

Figure S10. HPLC for peptide **L1.7**.

Figure S11. HPLC for peptide **L1**

Figure S12. MALDI-TOF of peptide **L1.3**

Figure S13. MALDI-TOF of peptide **L1.5**

Figure S14. MALDI-TOF of peptide **L1.6**

Figure S15. MALDI-TOF of peptide **L1.7**

Figure S16. MALDI-TOF-TOF (MSMS) of peptide **L1**

Figure S17. MALDI-TOF-TOF (MSMS) of peptide **L1.3**

Figure S18. MALDI-TOF-TOF (MSMS) of peptide **L1.5**

Figure S19. MALDI-TOF-TOF (MSMS) of peptide **L1.6**

Figure S20. MALDI-TOF-TOF (MSMS) of peptide **L1.7**

Figure S21. ^1H NMR spectrum for peptide **L1**.

Figure S22. ^1H NMR spectrum for peptide **L1.1**.

Figure S23. ^1H NMR spectrum for peptide **L1.2**.

Figure S24. ^1H NMR spectrum for peptide **L1.3**.

Figure S25. ^1H NMR spectrum for peptide **L1.4**.

Figure S26. ^1H NMR spectrum for peptide **L1.5**.

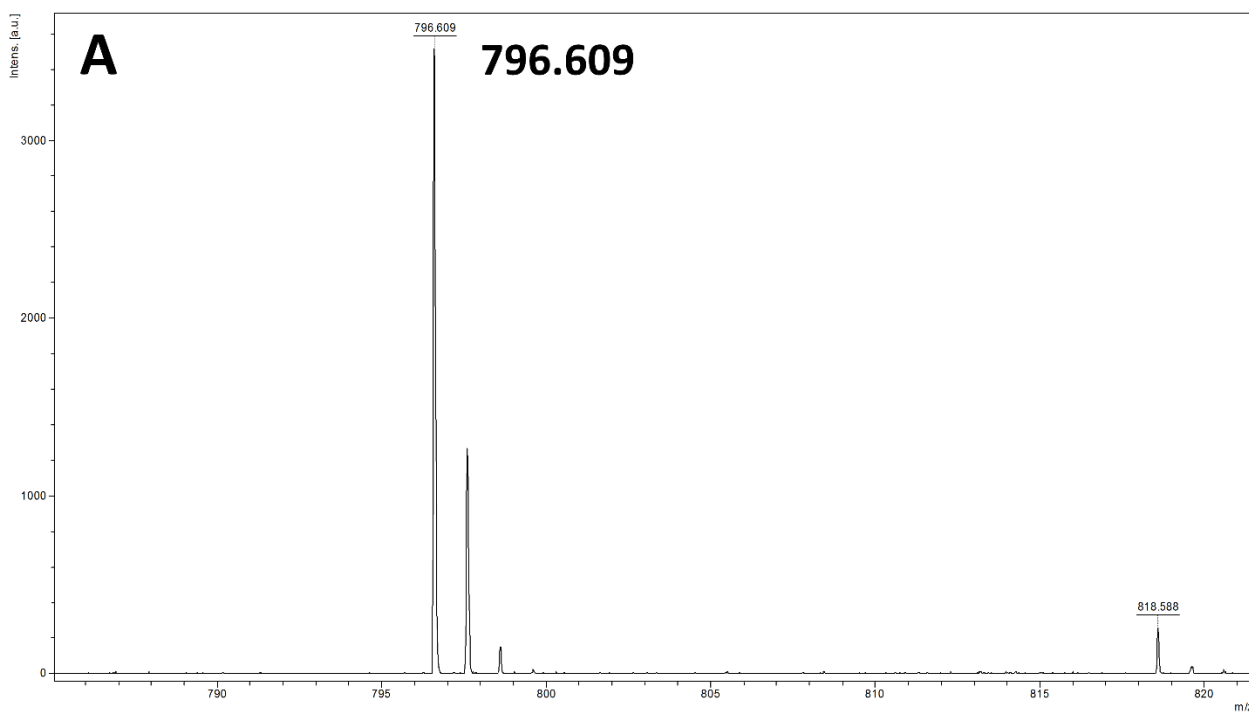
Figure S27. ^1H NMR spectrum for peptide **L1.6**.

Figure S28. ^1H NMR spectrum for peptide **L1.7**.

Figure S29. HSQC spectrum for peptide **L1**.

Figure S30. HSQC spectrum for peptide **L1.5**.

Figure S31. CD spectra of linear and cyclic peptides in methanol. 1 mg/mL of peptide was dissolved in methanol and CD spectra was acquired at 25 °C, and the spectra were average of 4 scans.



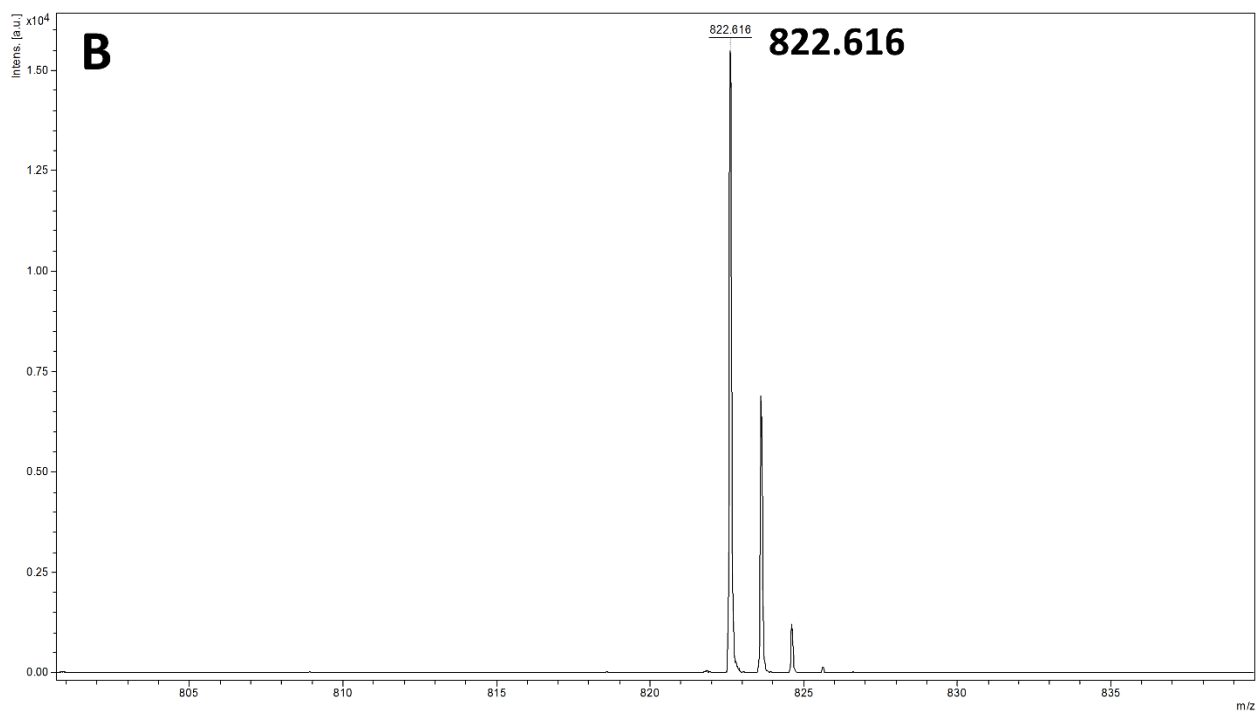


Figure S1. HR-MS for Cyclic peptide A) Cyclo(KLARLLT) and B) Cyclo(KN3)larllt).

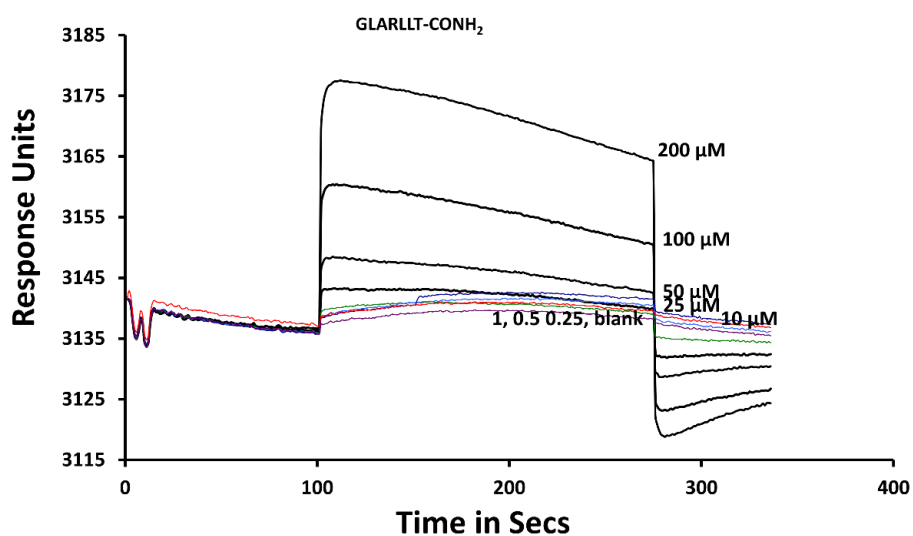


Figure S2. SPR analysis of linear peptide GLARLLT. EGFR was immobilized on CM5 sensor chip and different concentrations of peptide was injected to obtain the SPR response. Binding of peptide GLARLLT to EGFR was observed with increasing concentration of the peptide.

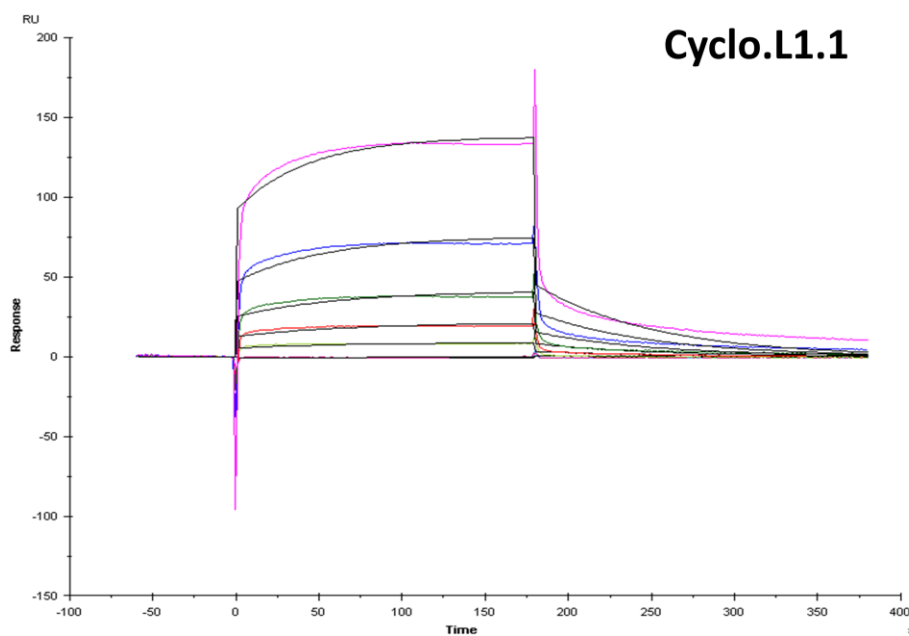


Figure S3. Curvefitting of SPR sensorgram for **Cyclo.L1.1** using 1:1 Langmuir interaction describing the single exponential of the data. Black lines are experimental data and different colored lines represent fitted lines. K_D values reported were from k_{on} and k_{off} calculations.

L1.7

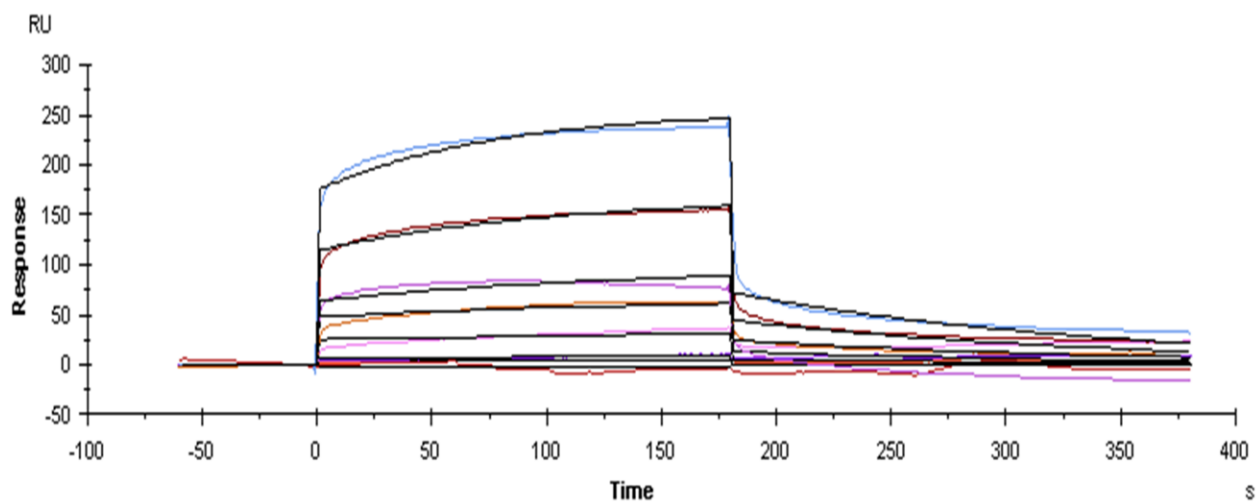


Figure S4. Curvefitting of SPR sensorgram for **L1.7** using 1:1 Langmuir interaction describing the single exponential of the data. Black lines are experimental data and different colored lines represent fitted lines.

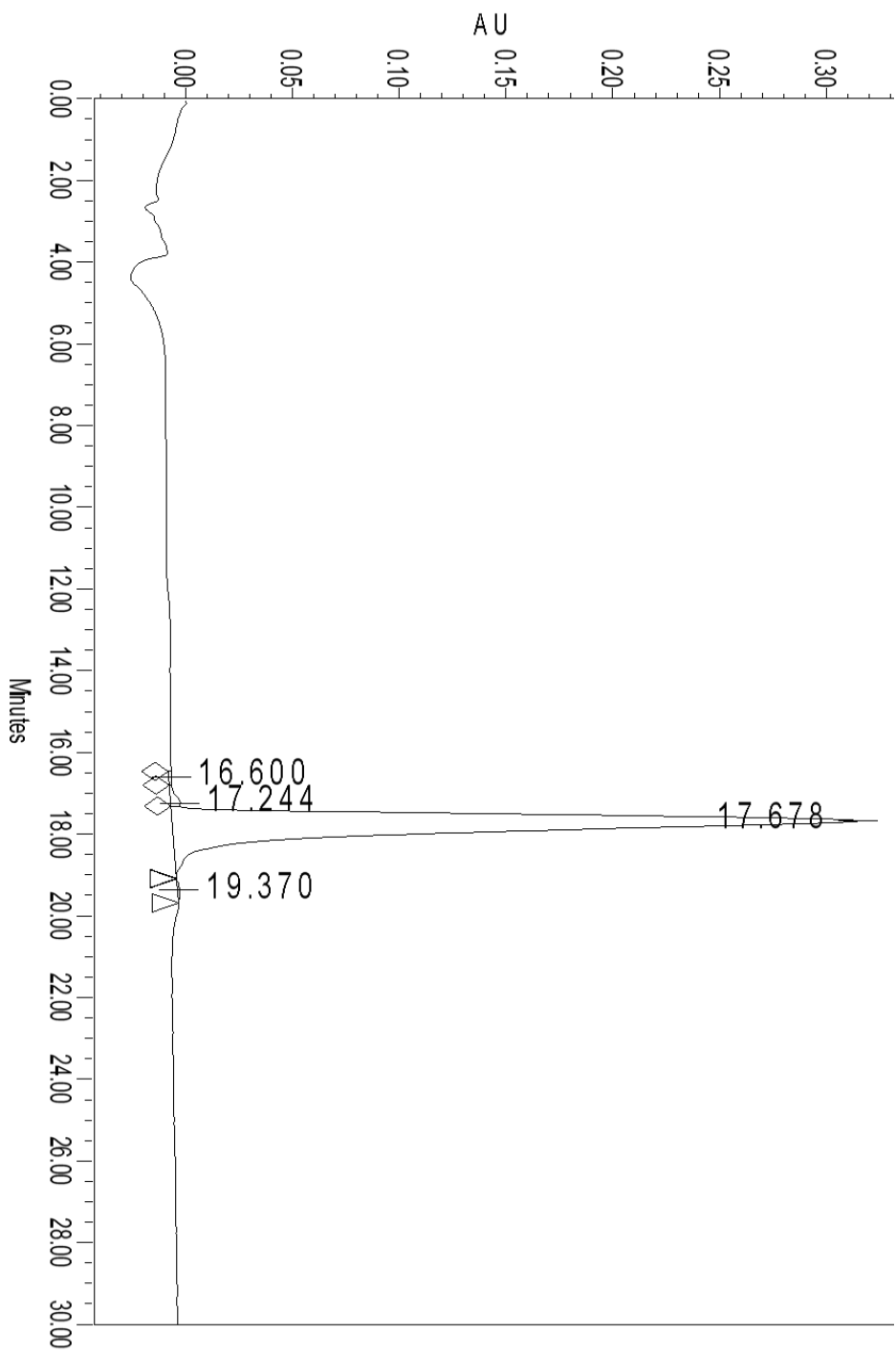


Figure S5: HPLC for peptide L1.

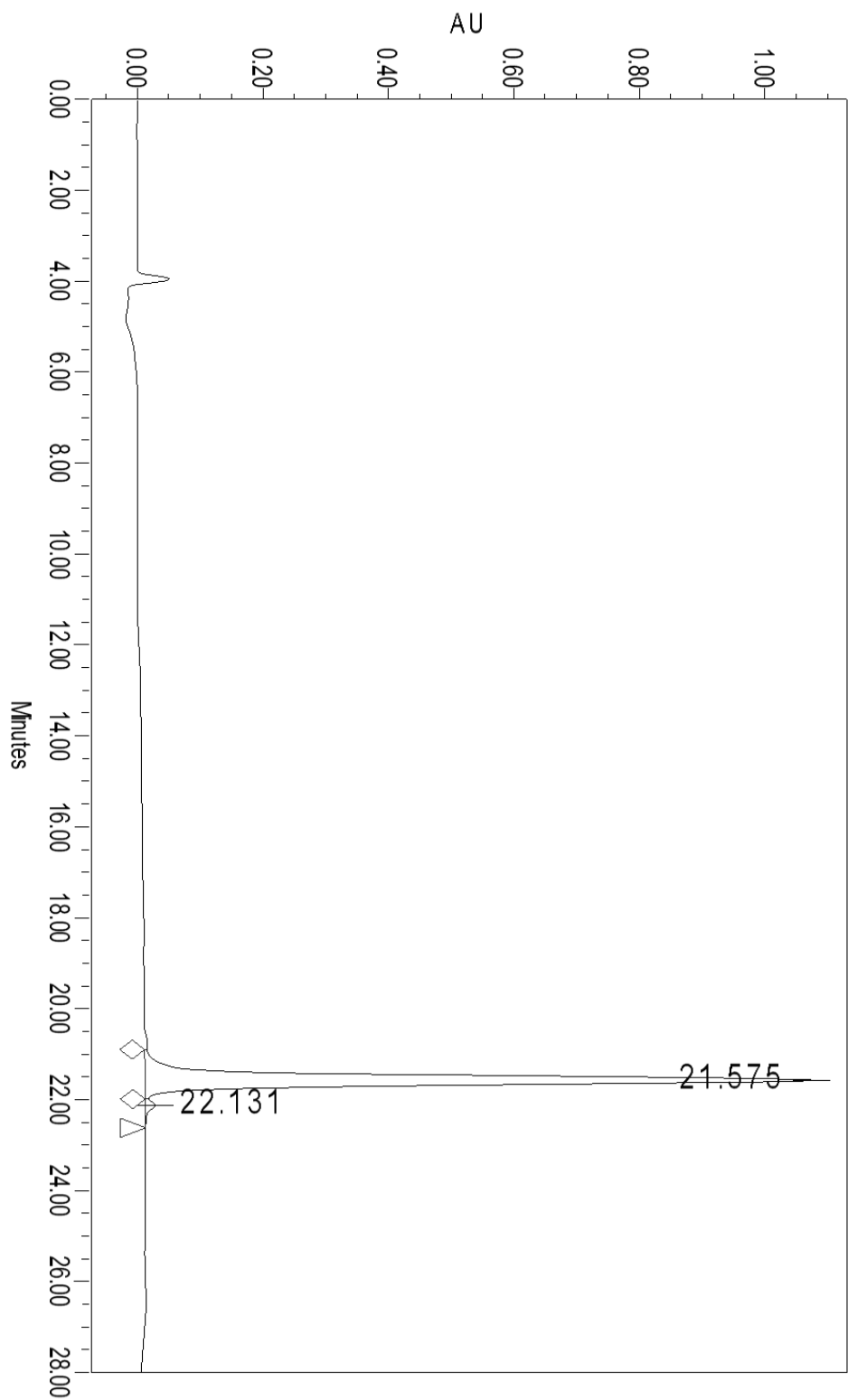


Figure S6. HPLC of peptide **L1.3**.

Auto-Scaled Chromatogram

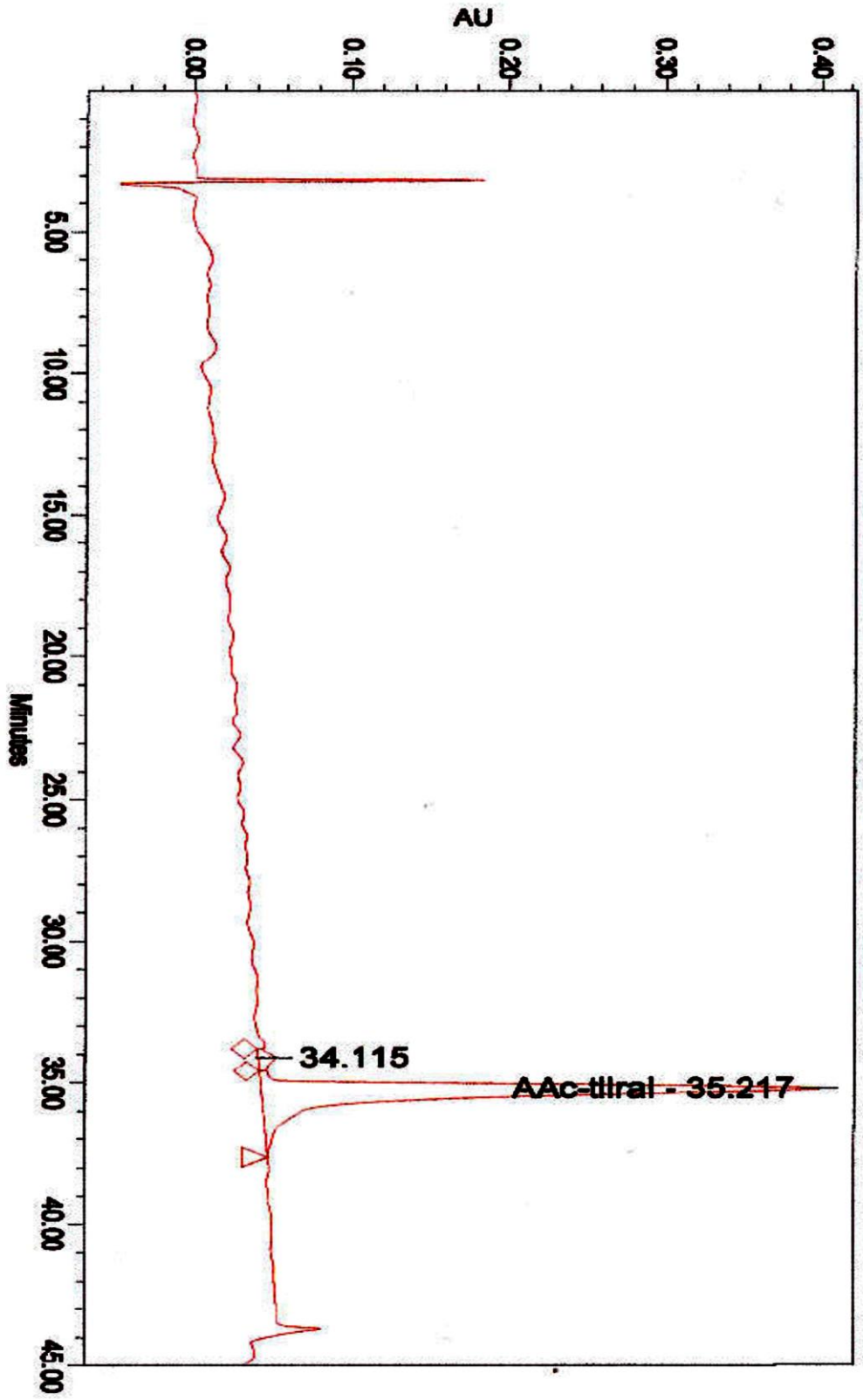


Figure S7. HPLC for peptide L1.4.

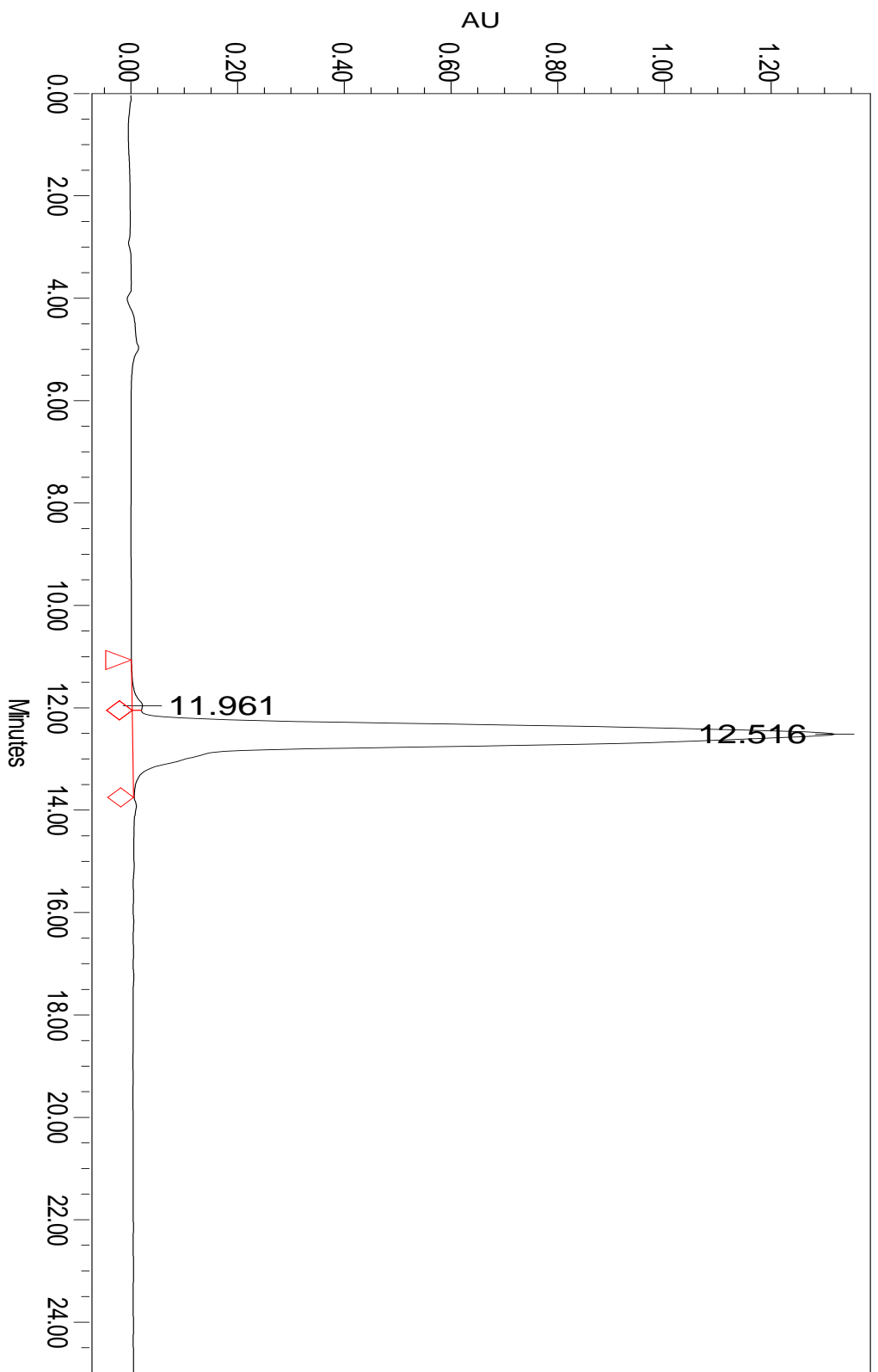


Figure S8. HPLC for peptide **L1.5**.

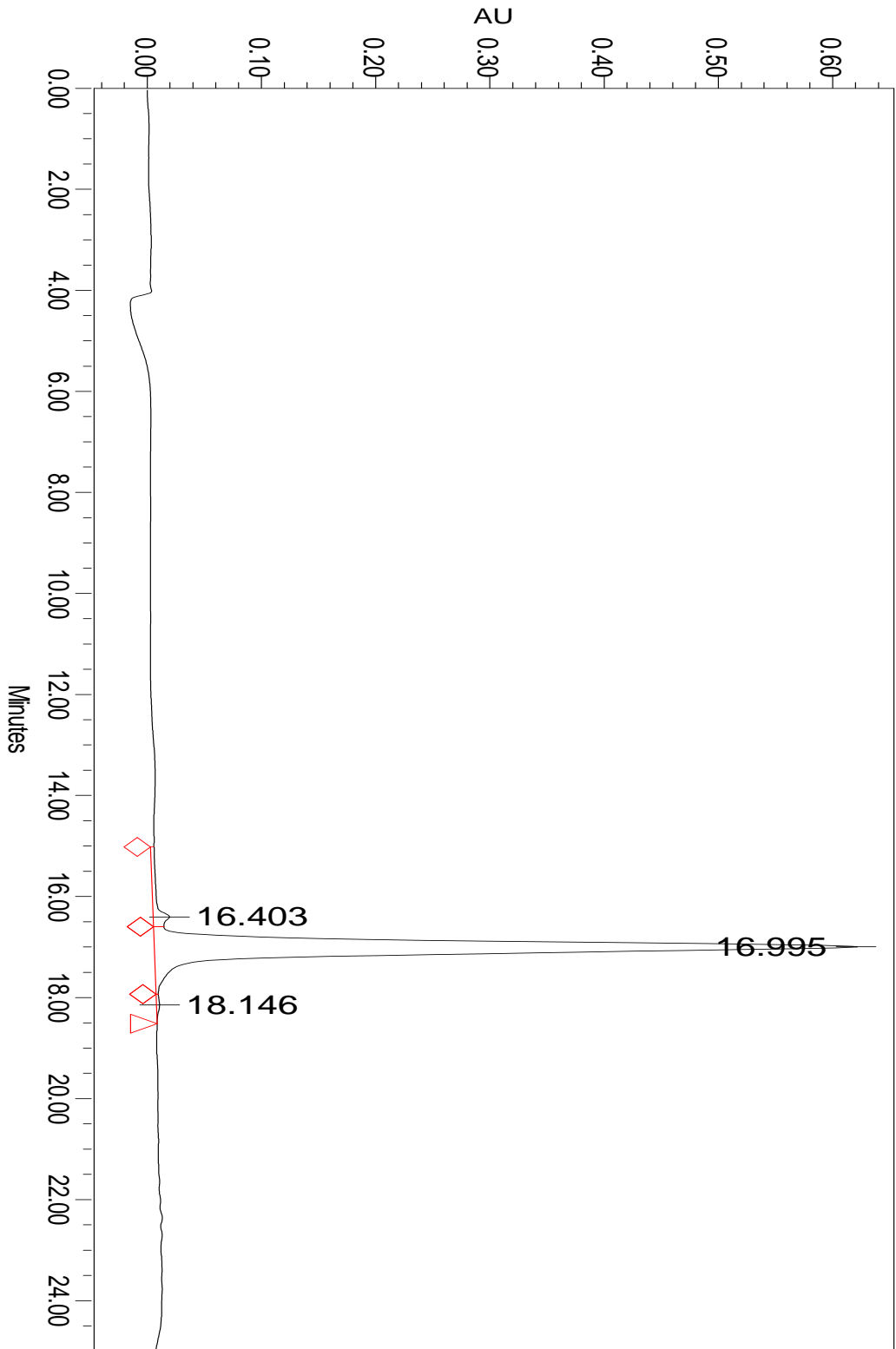


Figure S9. HPLC for peptide L1.6.

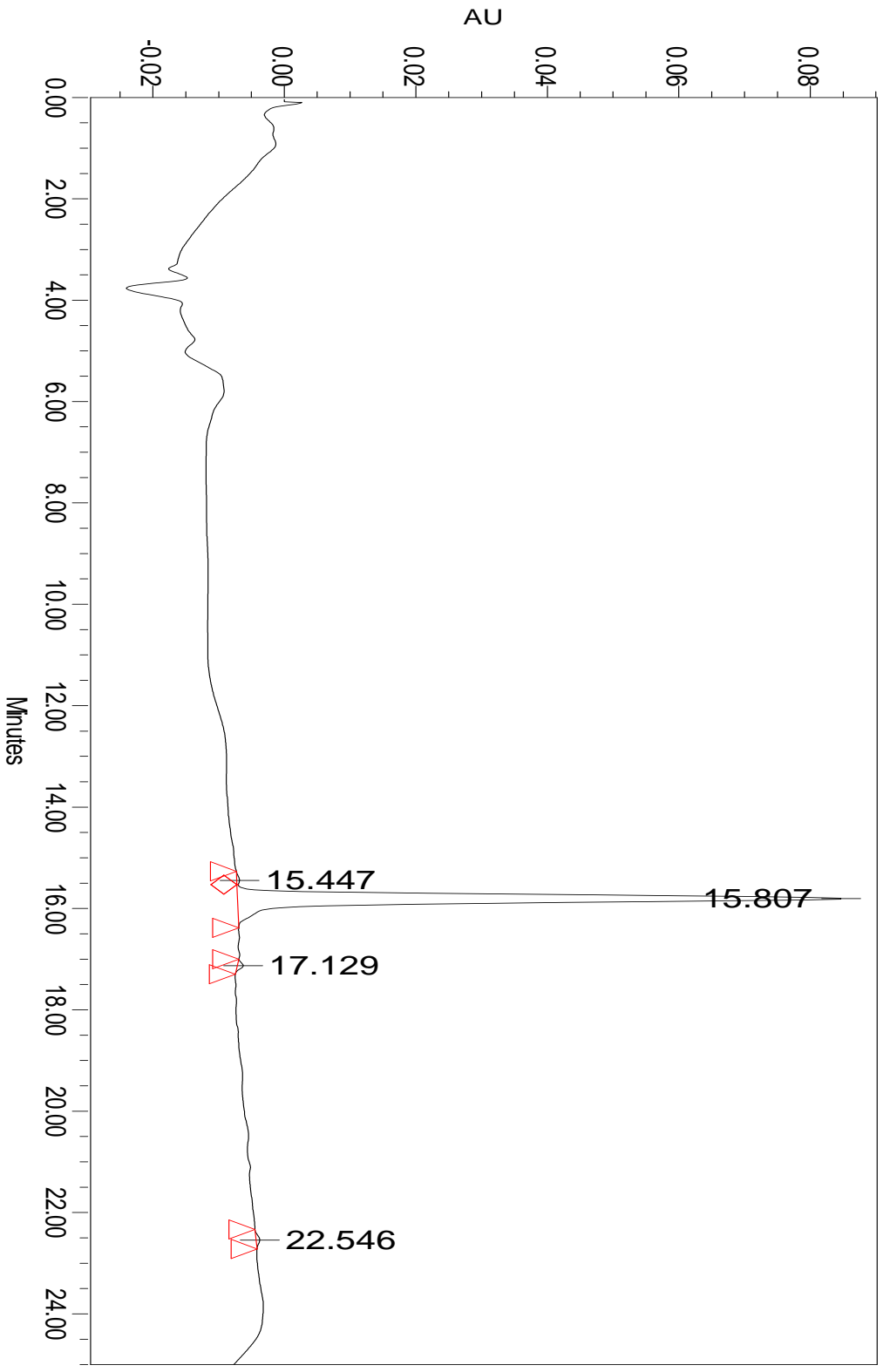


Figure S10. HPLC for peptide L1.7.

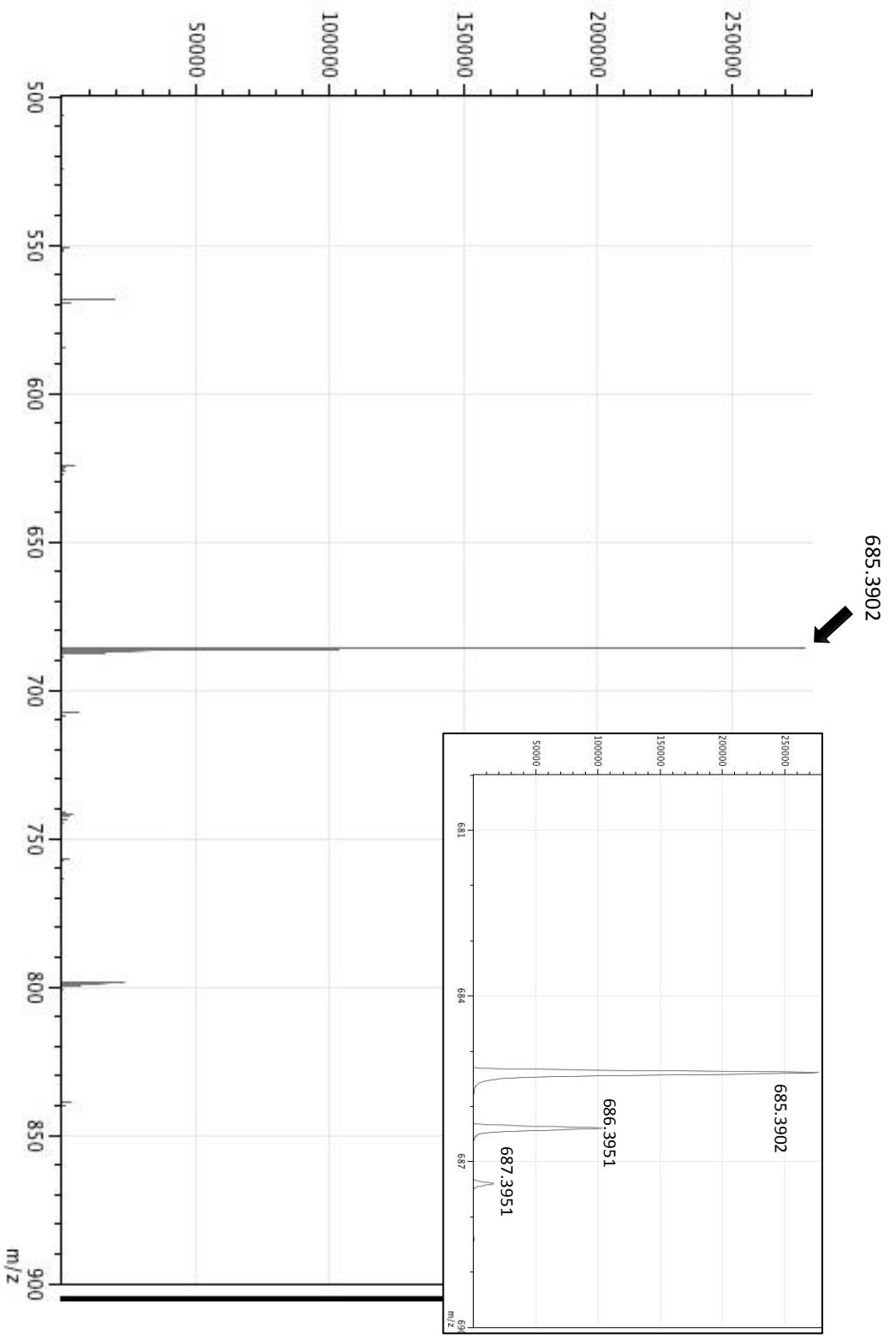


Figure S11. HPLC for peptide L1

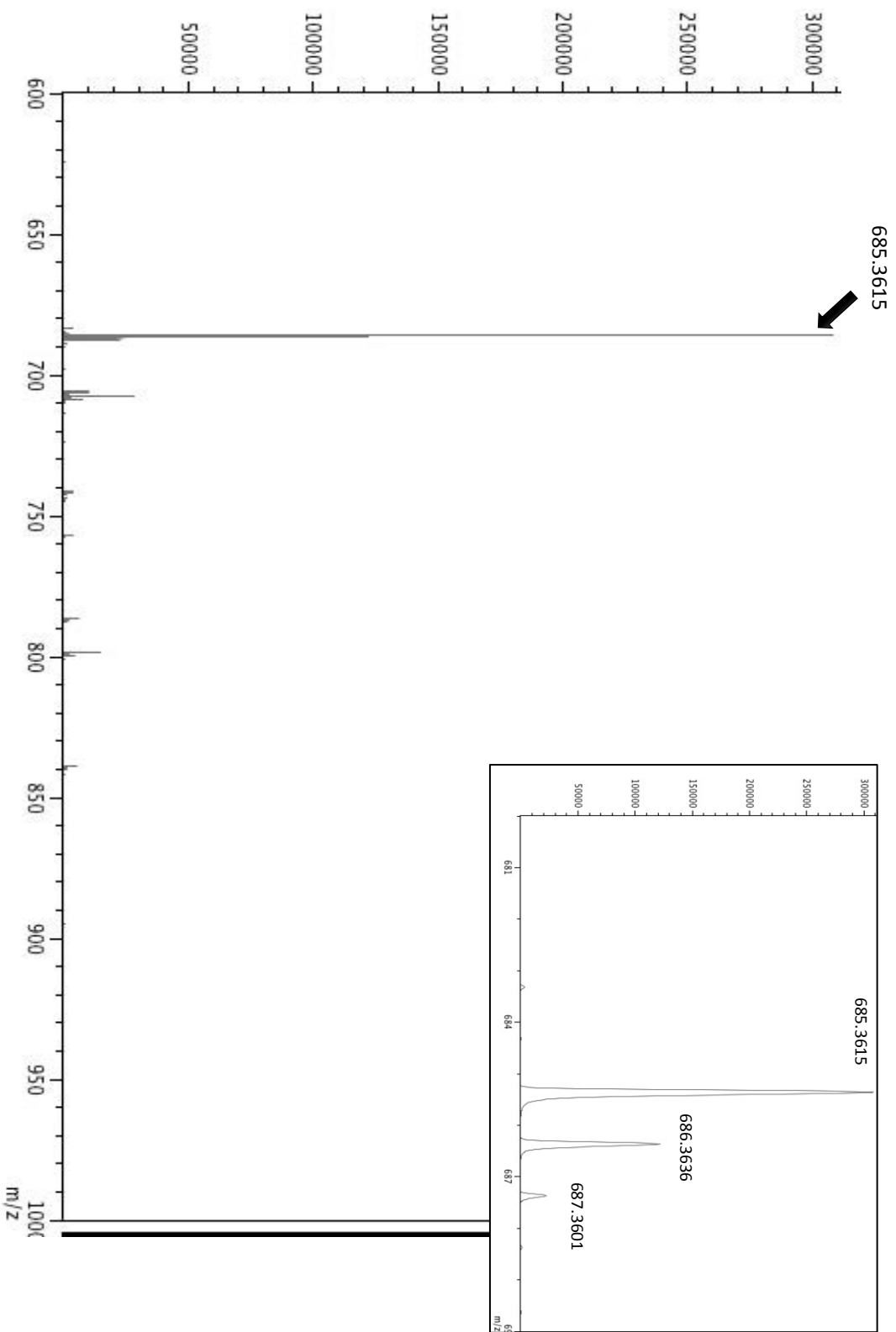


Figure S12. MALDI-TOF of peptide L1.3

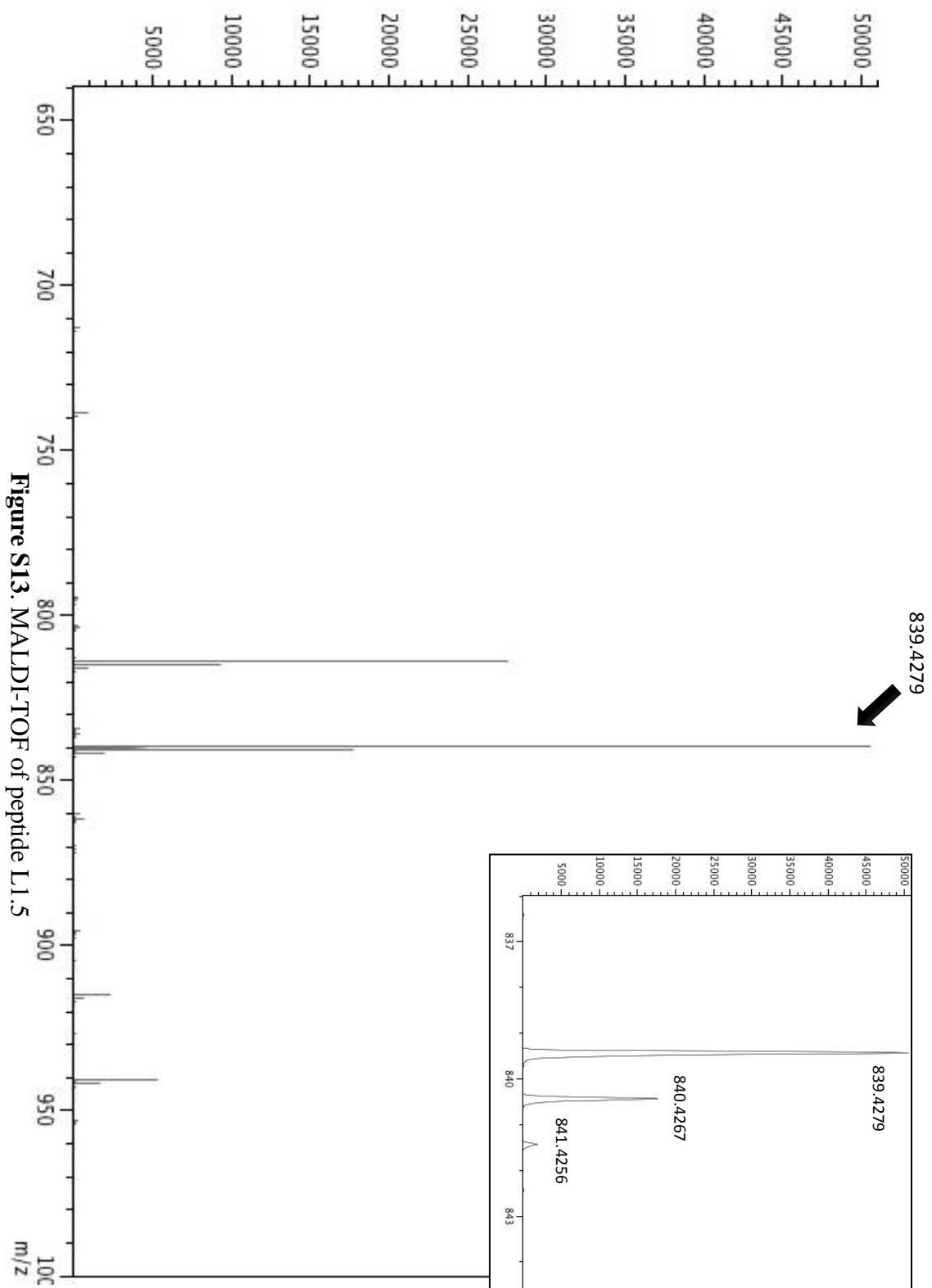


Figure S13. MALDI-TOF of peptide L1.5

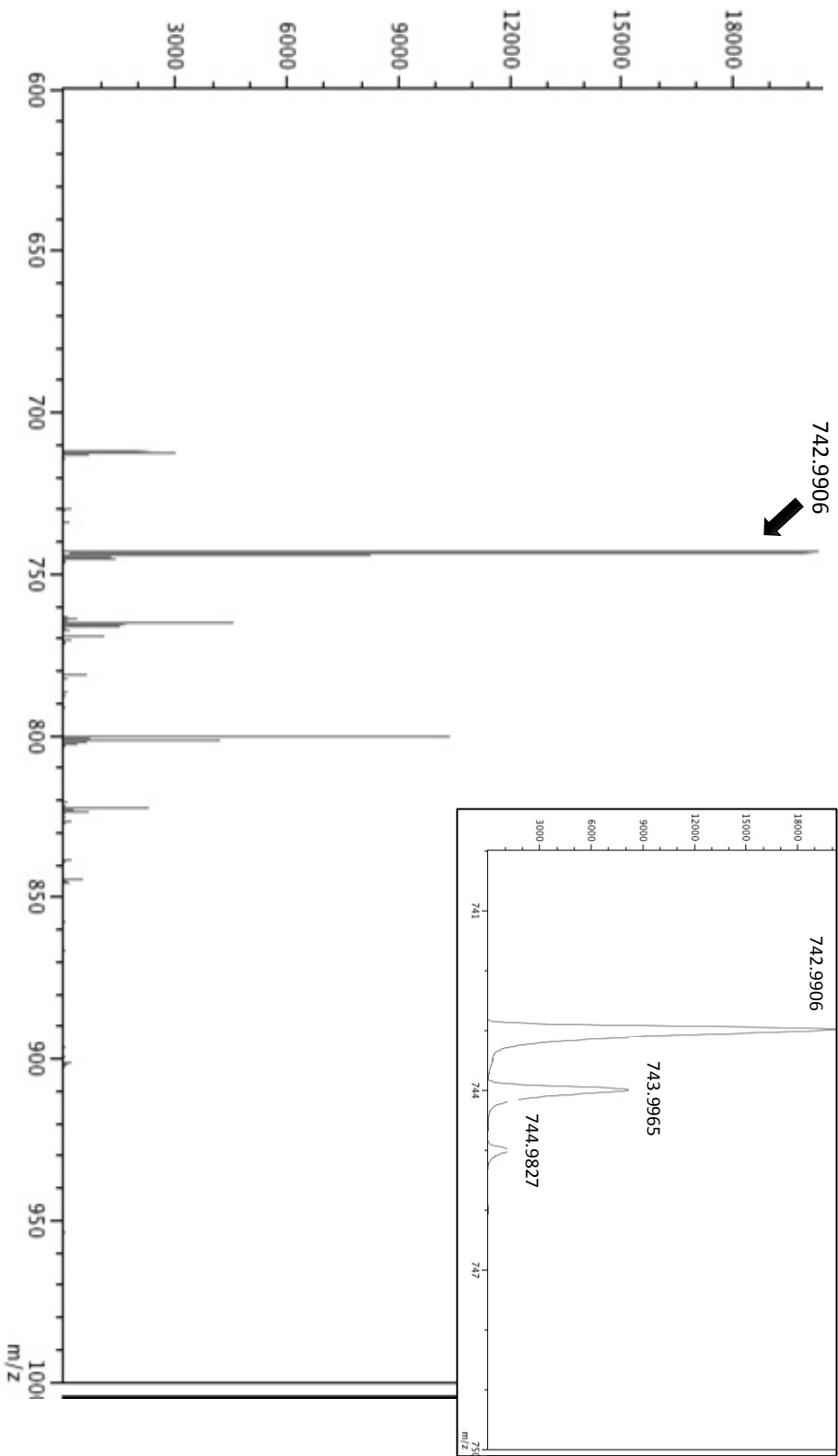


Figure S14. MALDI-TOF of peptide L1.6

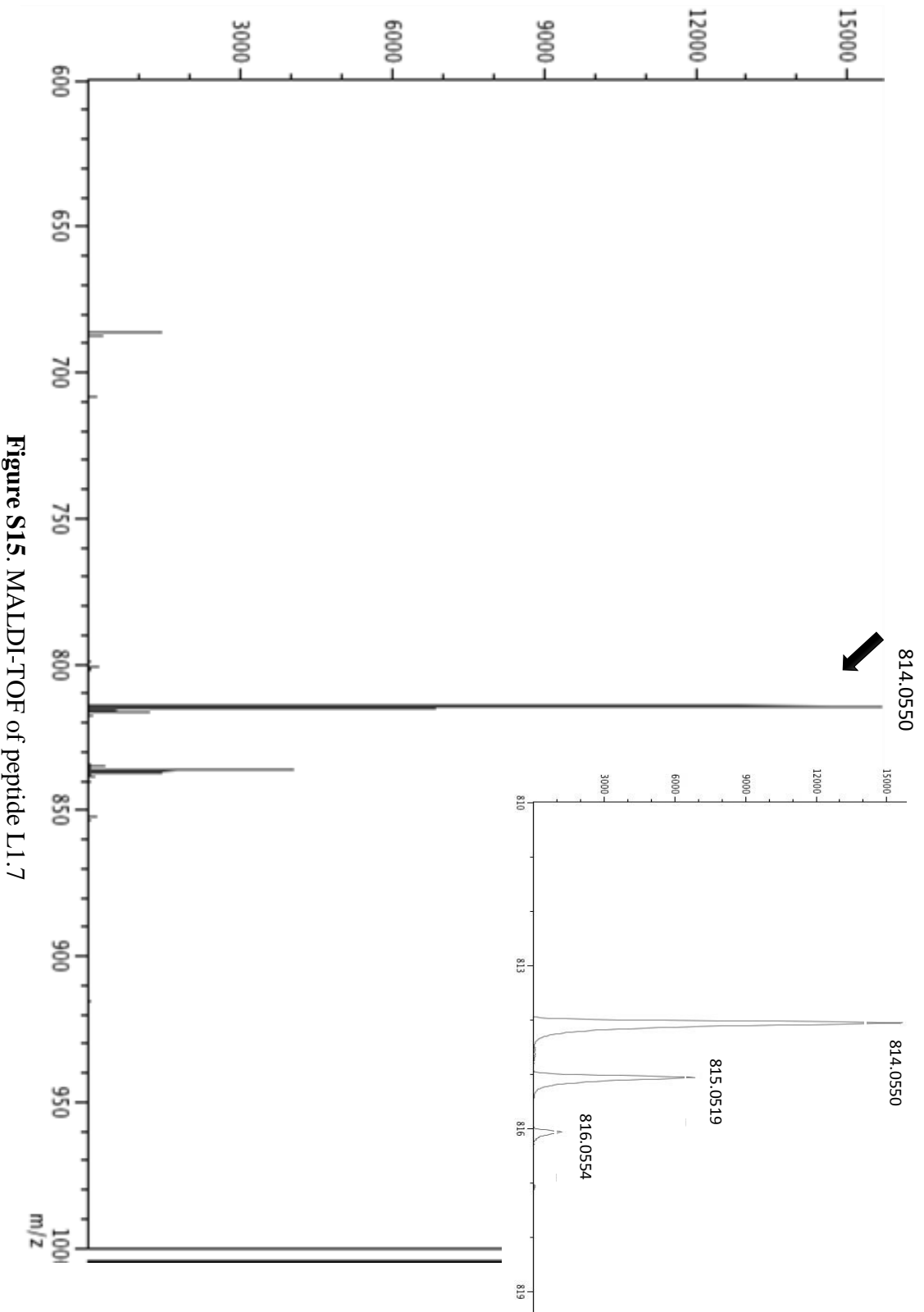


Figure S15. MALDI-TOF of peptide L1.7

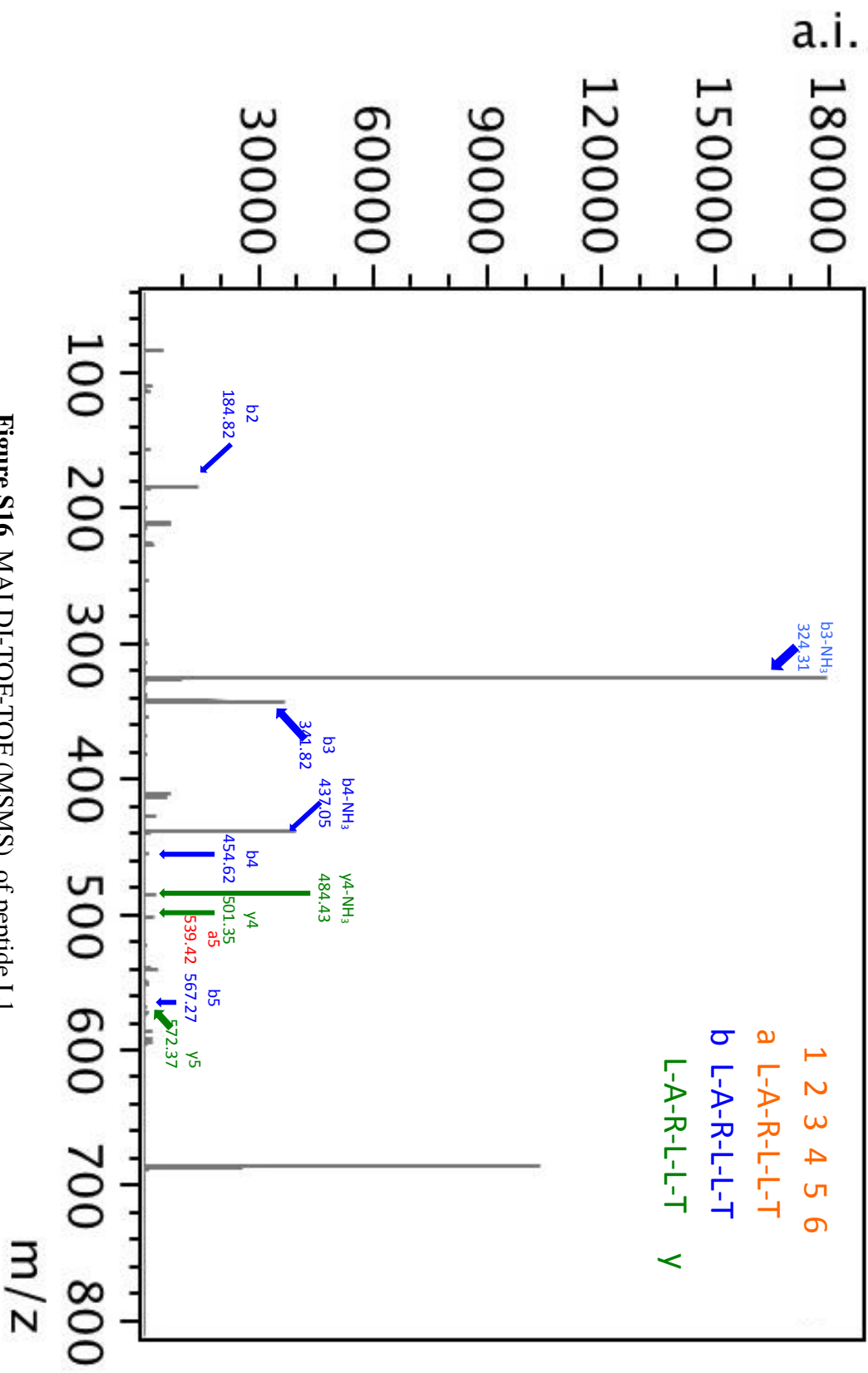


Figure S16. MALDI-TOF-TOF (MSMS) of peptide L1

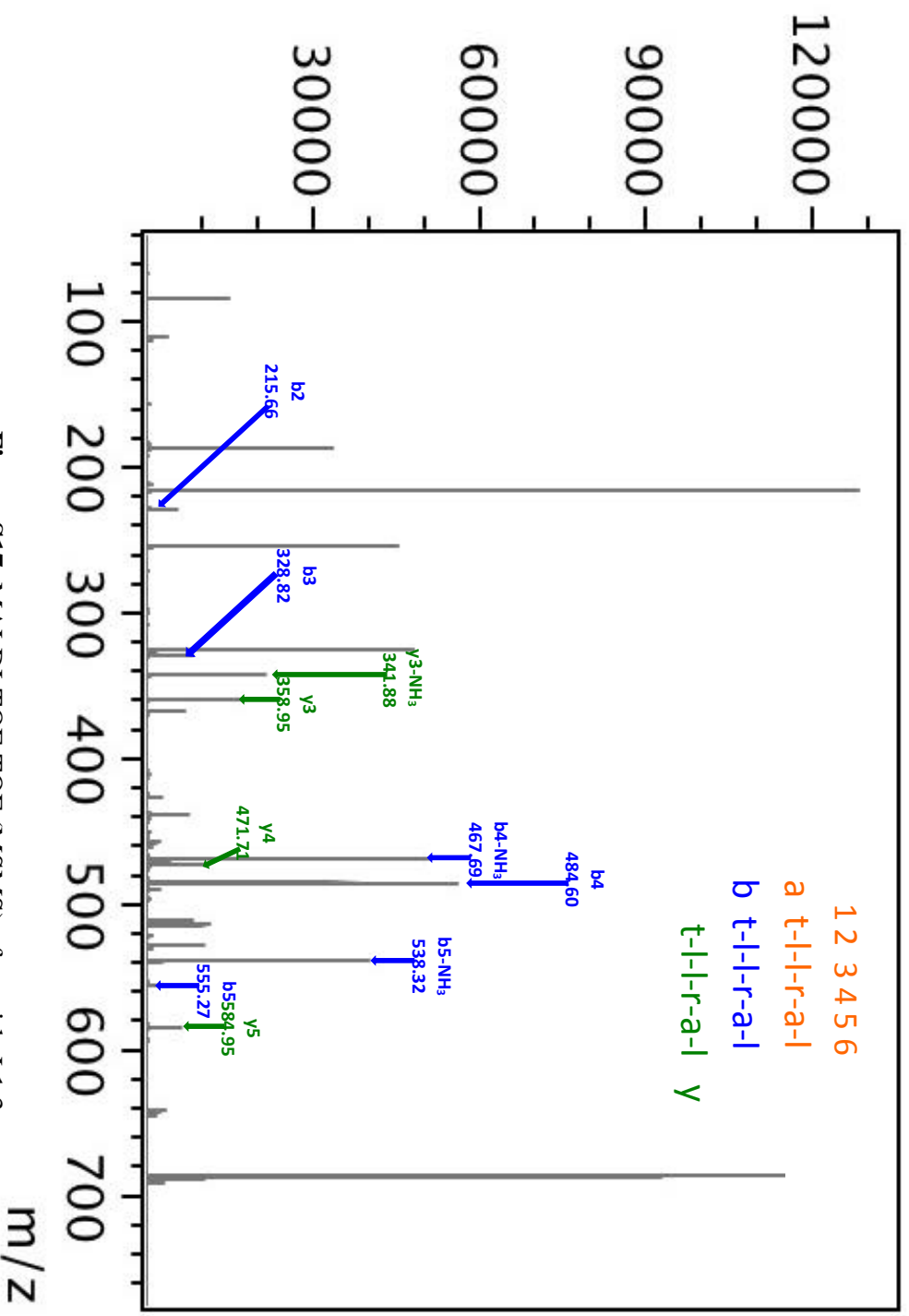


Figure S17. MALDI-TOF-TOF (MS/MS) of peptide L1.3

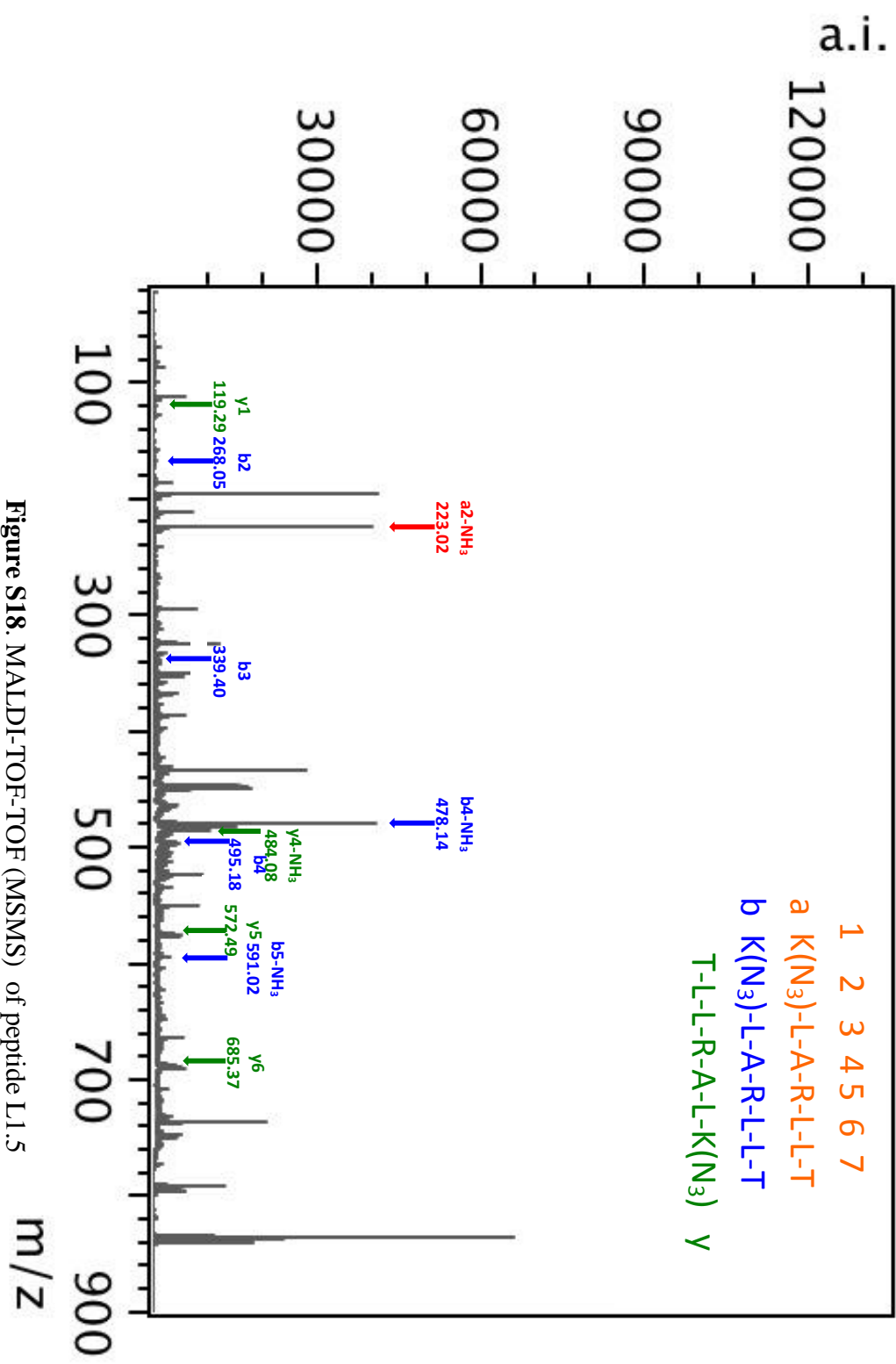


Figure S18. MALDI-TOF-TOF (MS/MS) of peptide L1.5

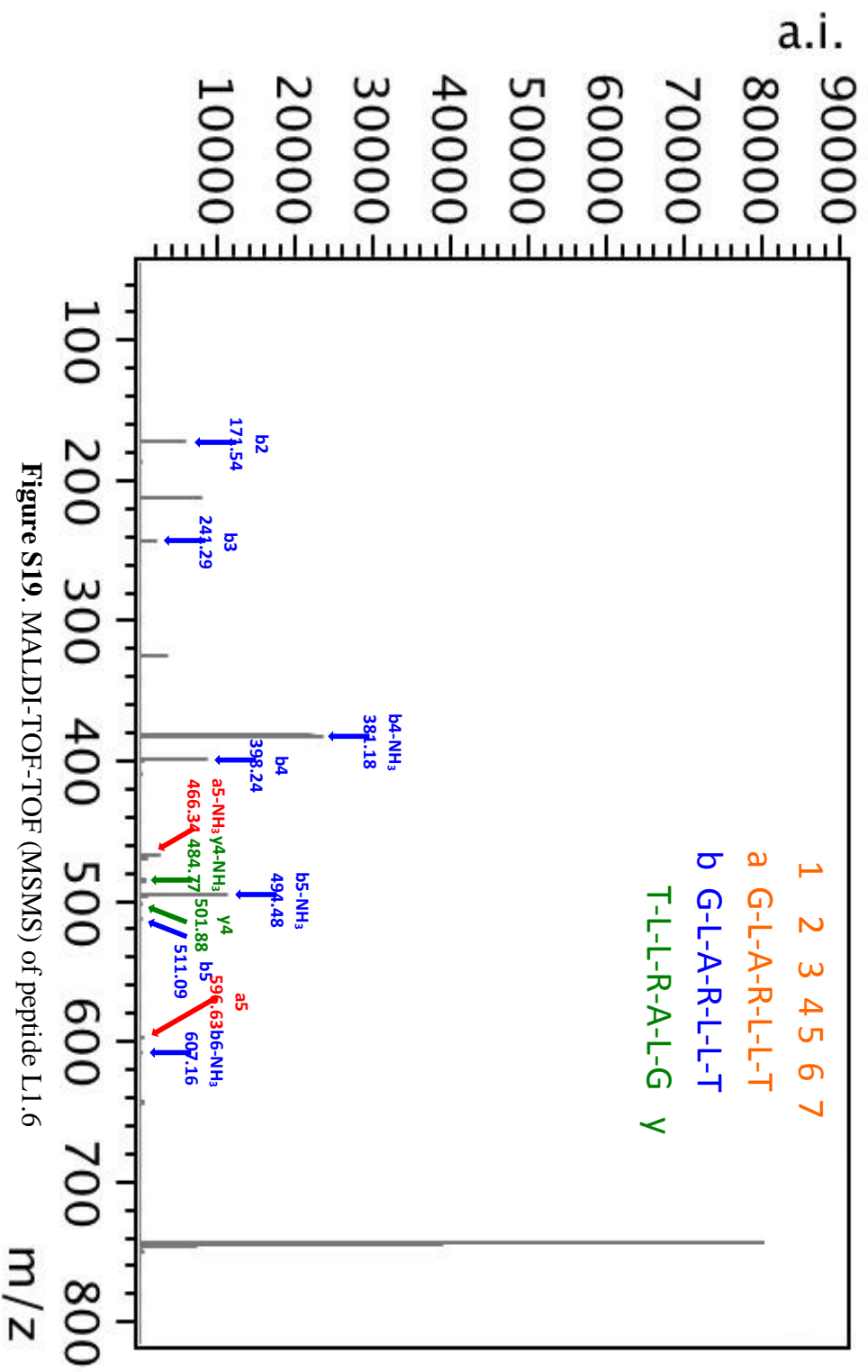


Figure S19. MALDI-TOF-TOF (MS/MS) of peptide L1.6

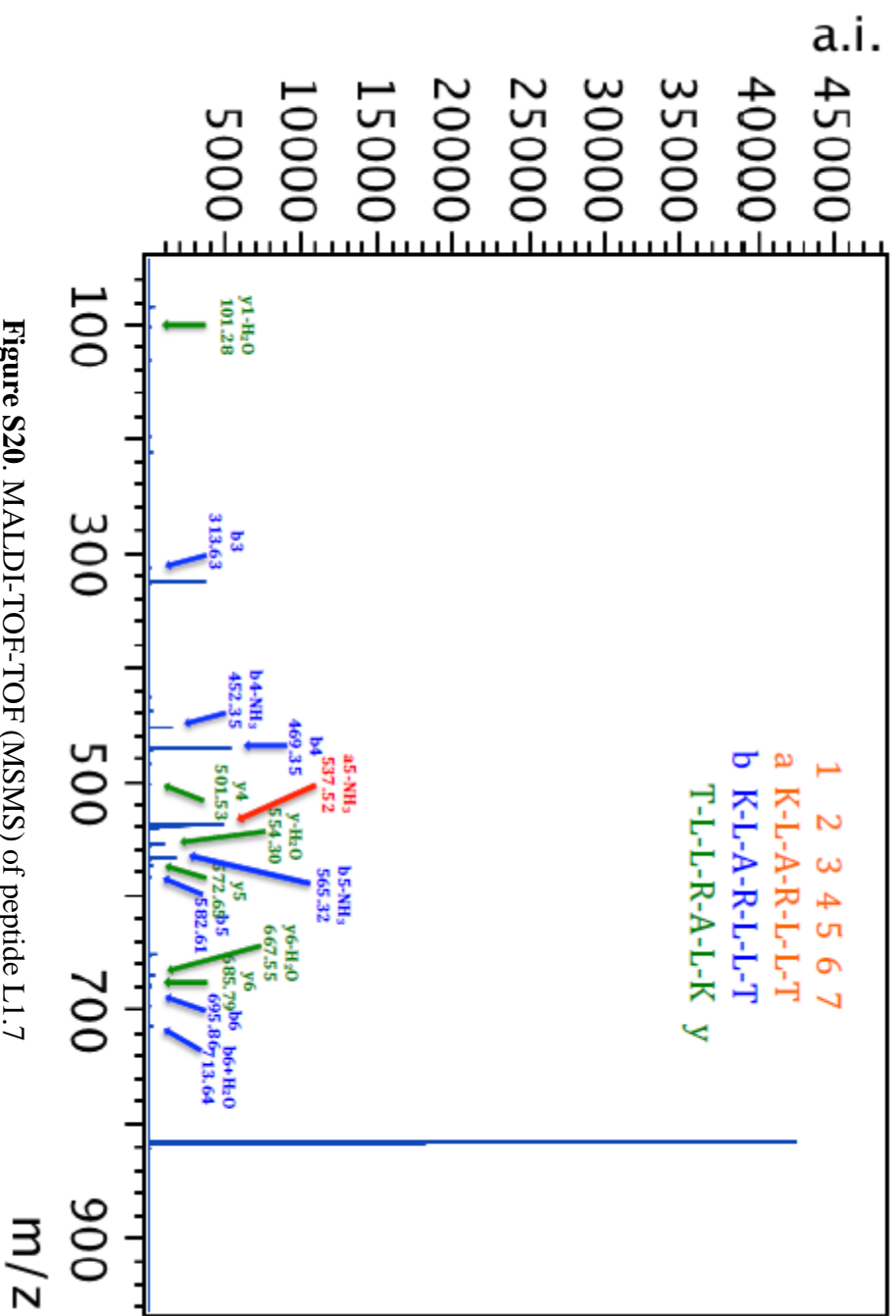


Figure S20. MALDI-TOF-TOF (MSMS) of peptide L1.7

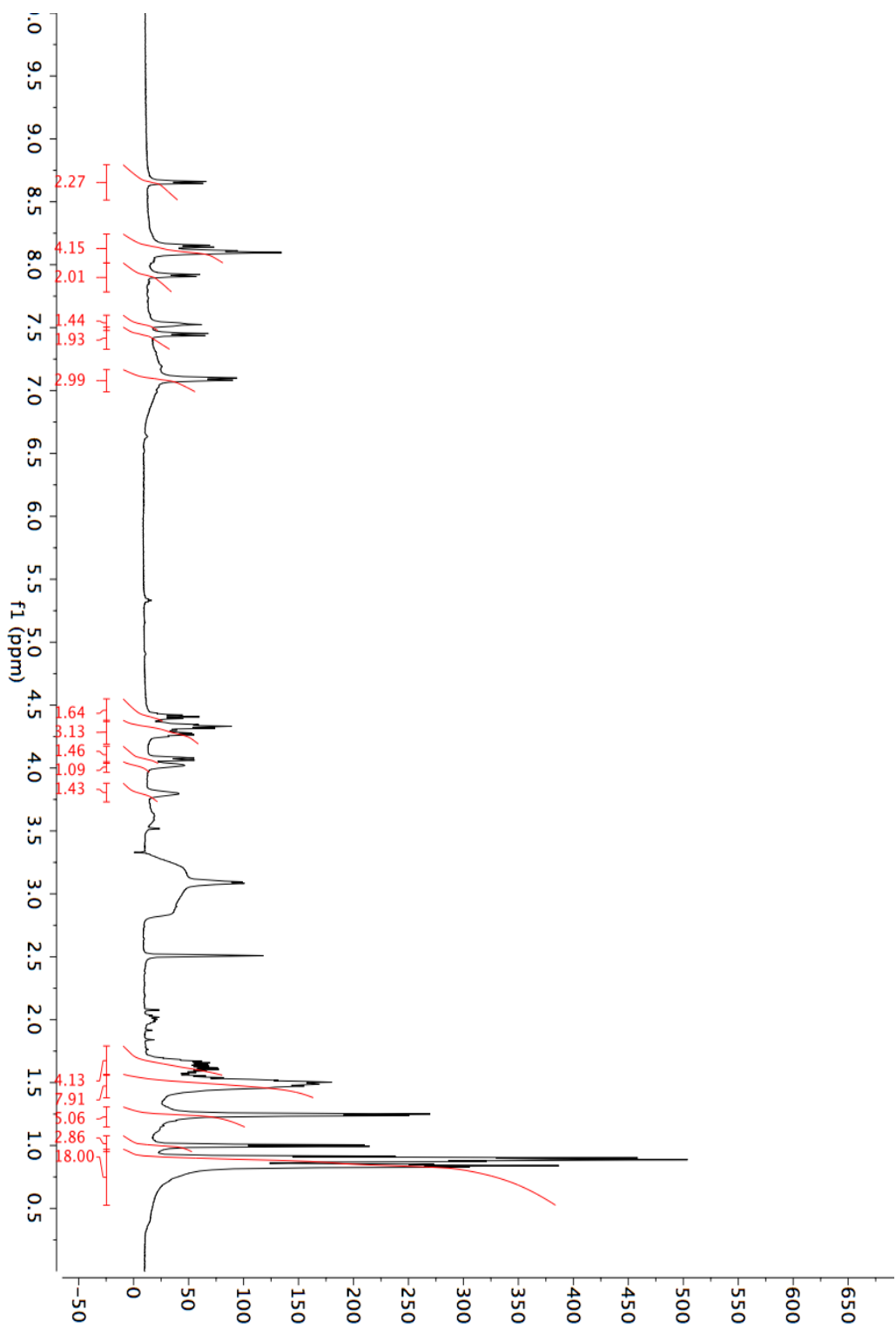


Figure S21. ¹H NMR spectrum for peptide L1.

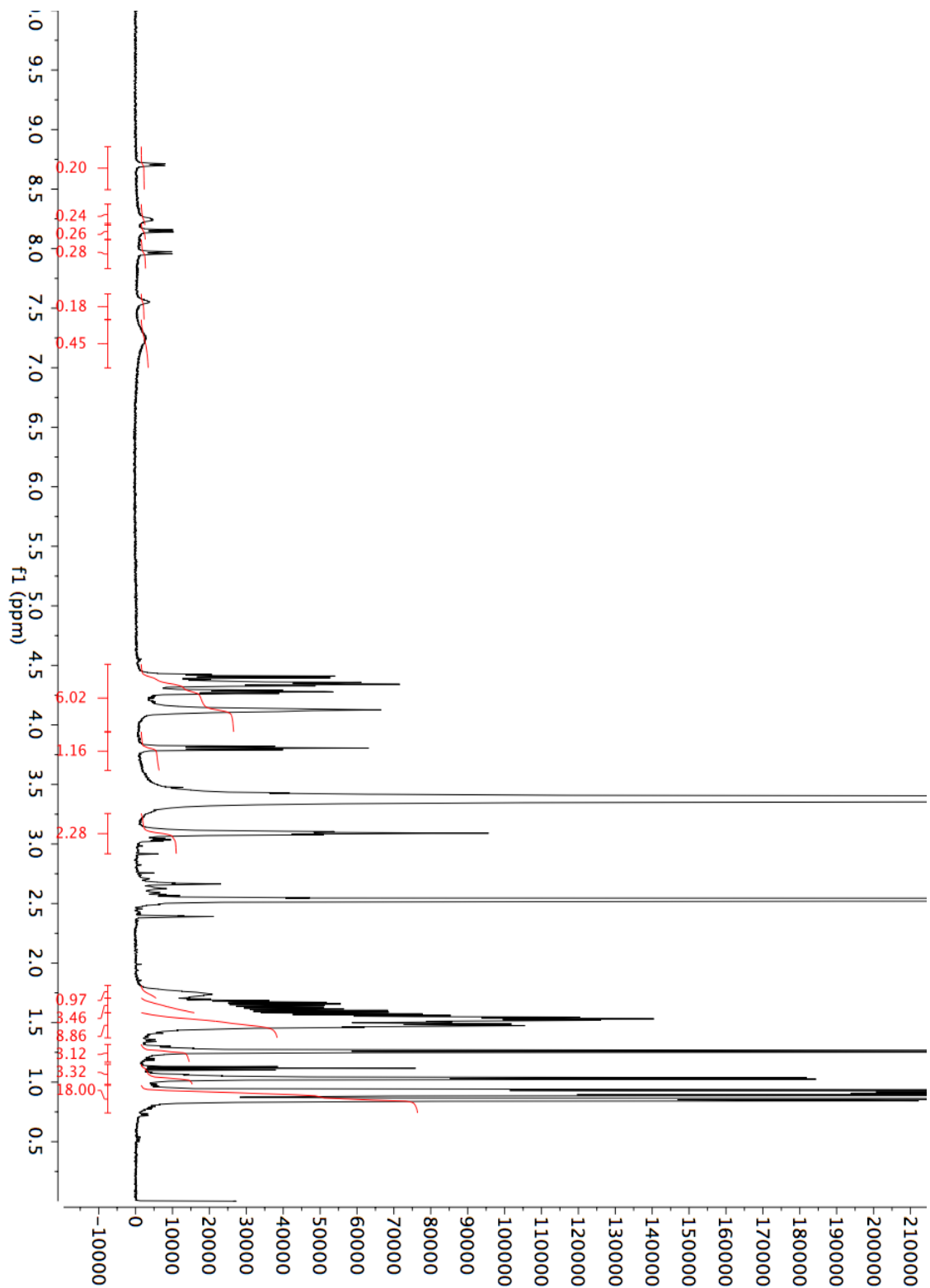


Figure S22. ¹H NMR spectrum for peptide L1.1.

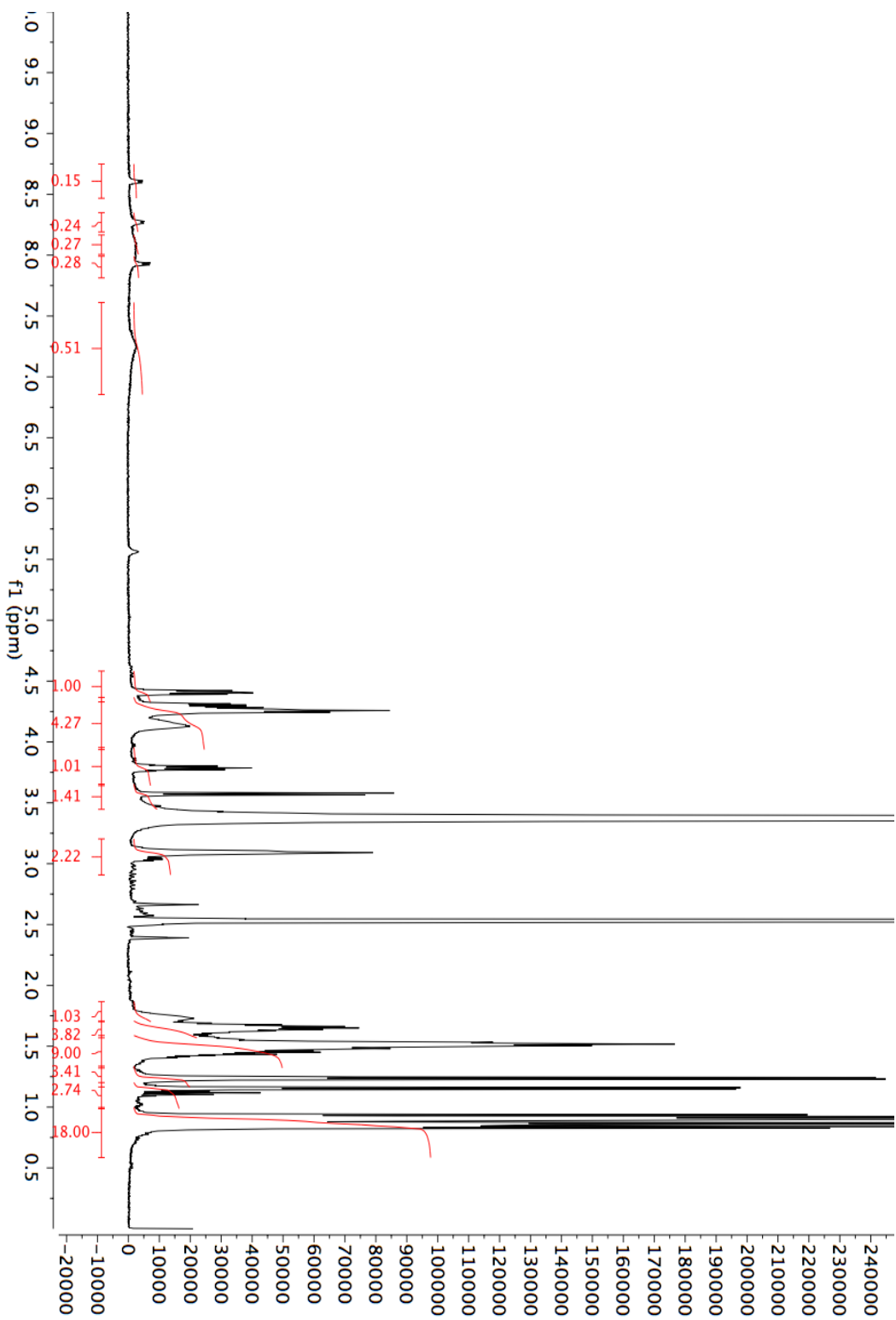


Figure S23. ¹H NMR spectrum for peptide L1.2.

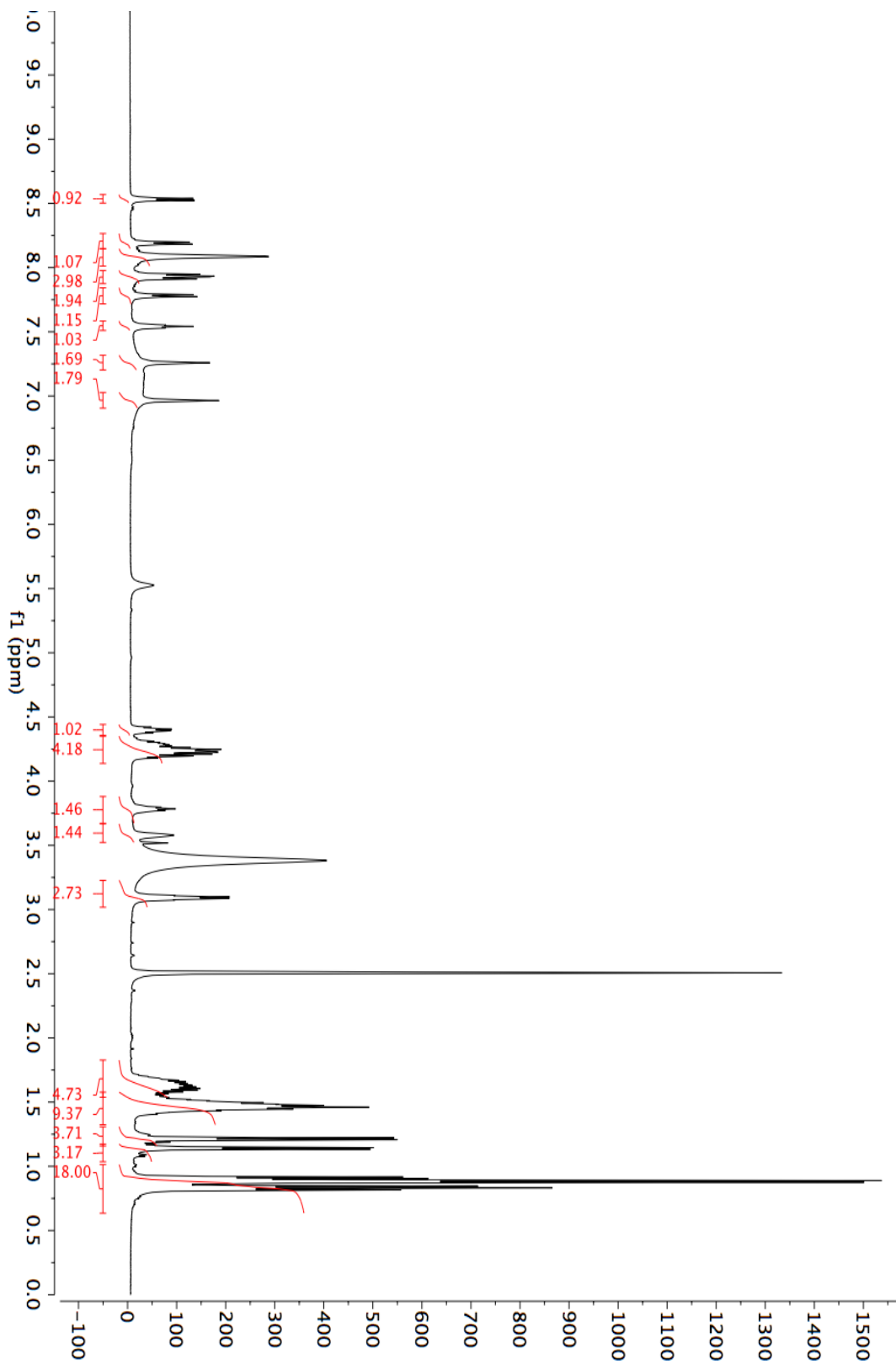


Figure S24. ^1H NMR spectrum for peptide L1.3.

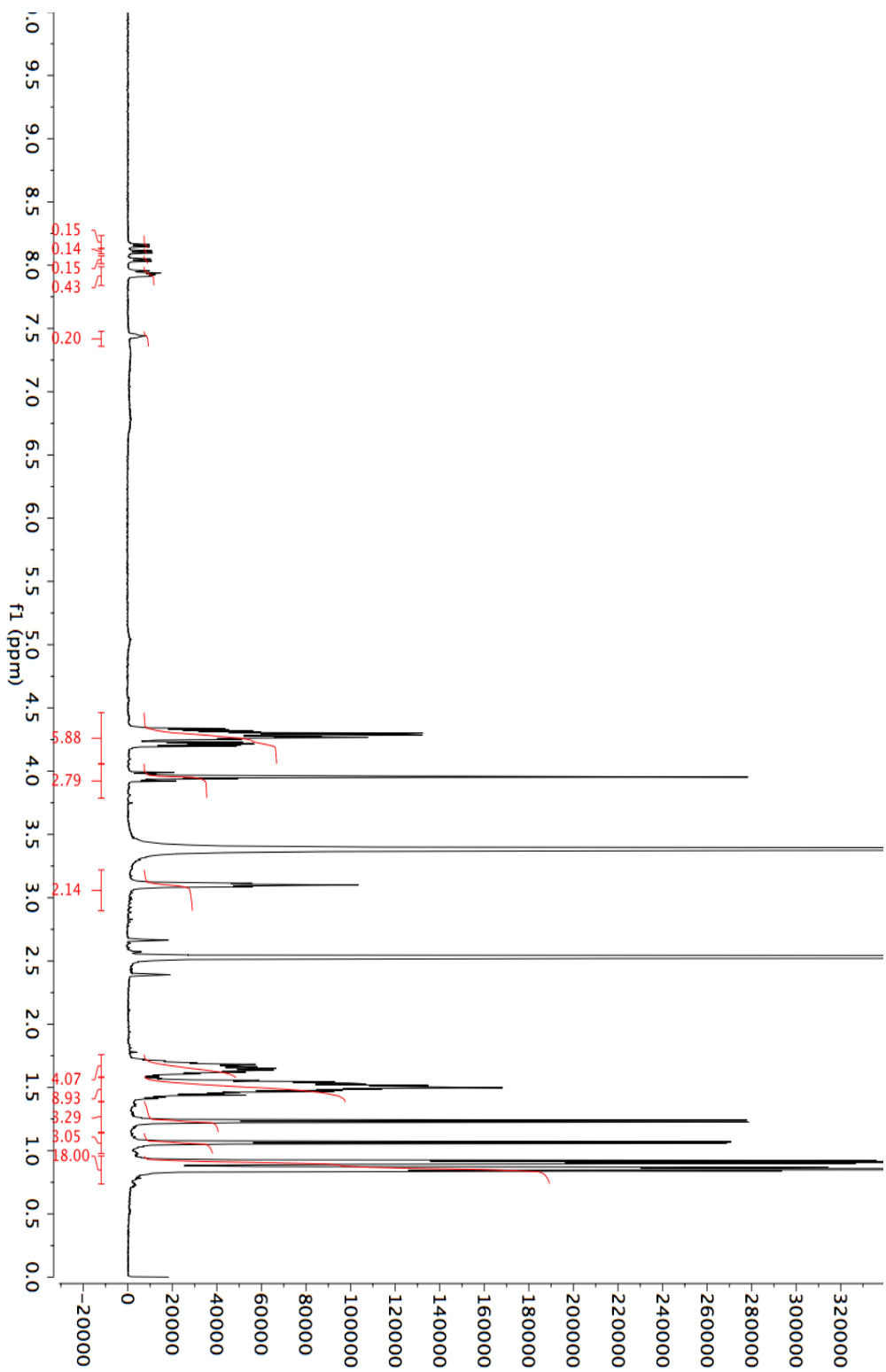


Figure S25. ¹H NMR spectrum for peptide L1.4.

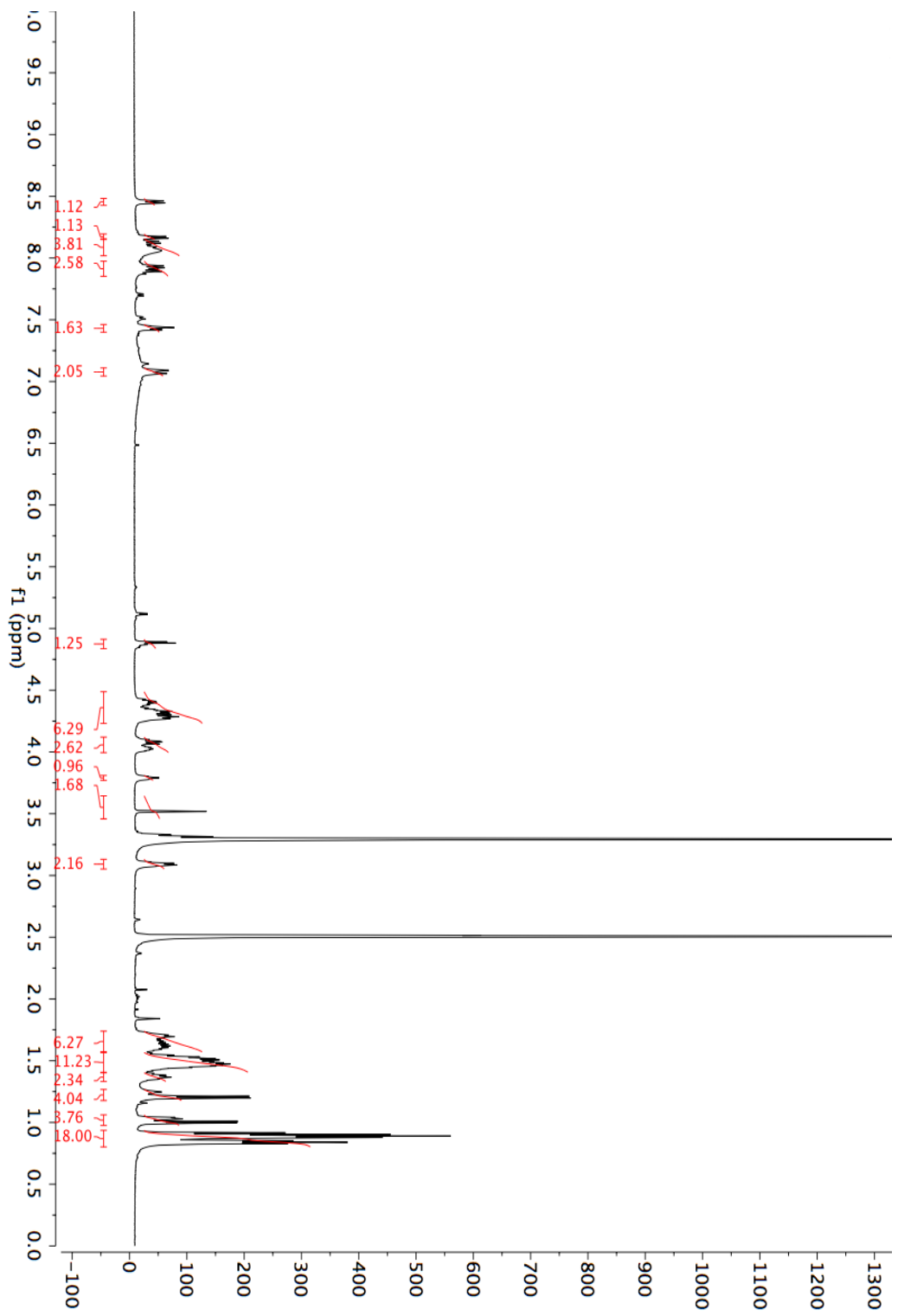


Figure S26. ¹H NMR spectrum for peptide L1.5.

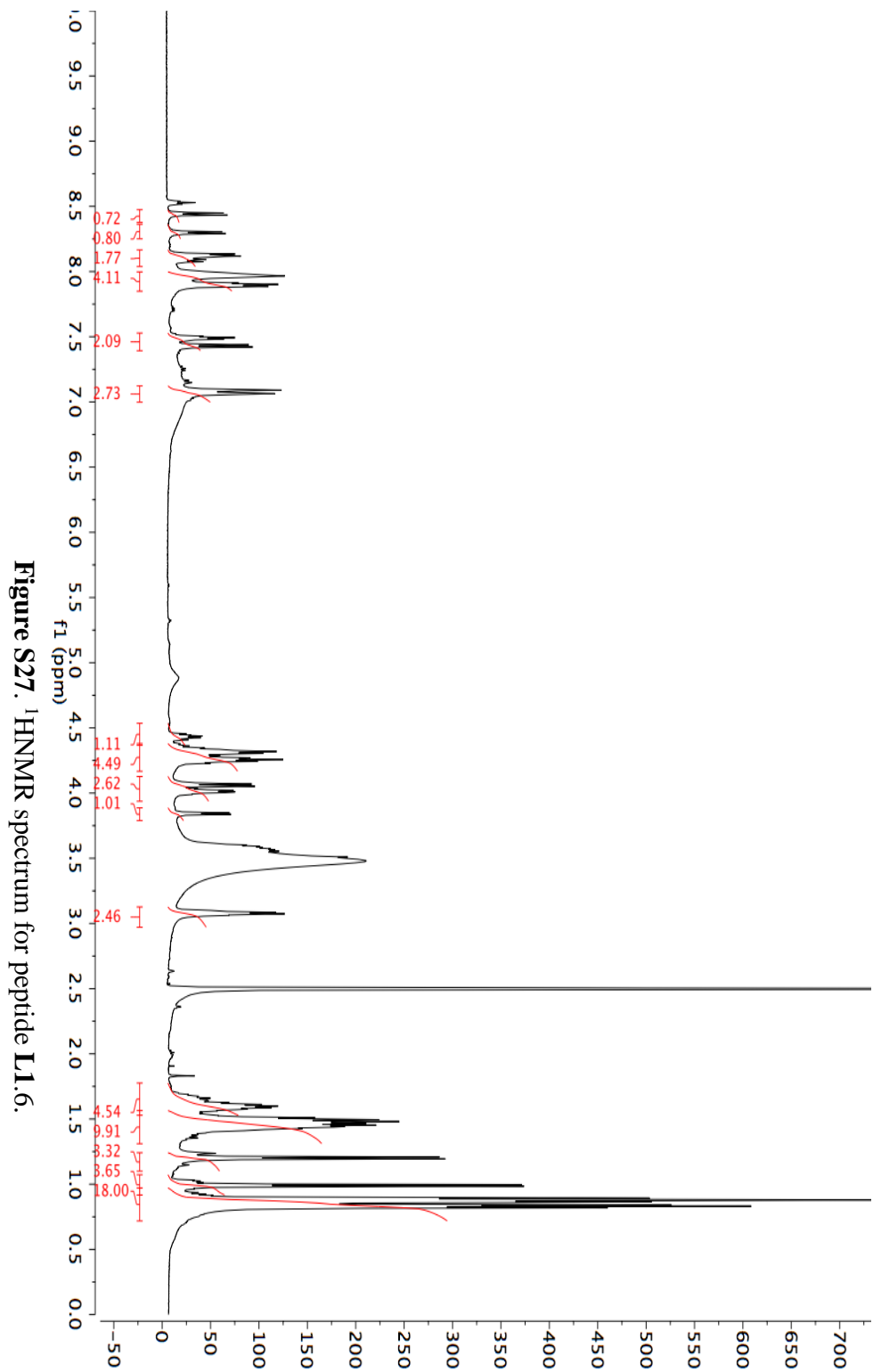


Figure S27. ¹H NMR spectrum for peptide L1.6.

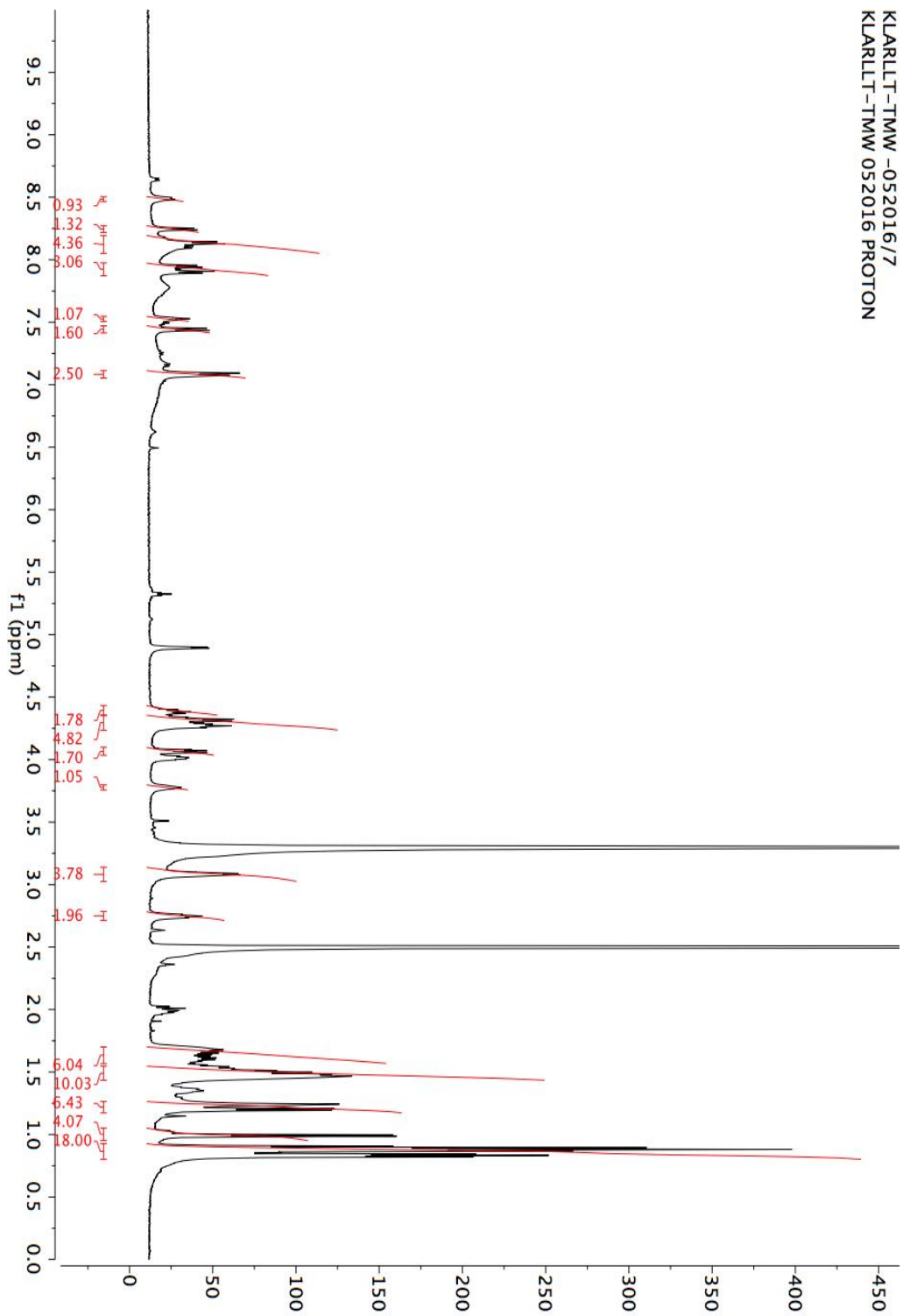


Figure S28. 1H NMR spectrum for peptide L1.7.

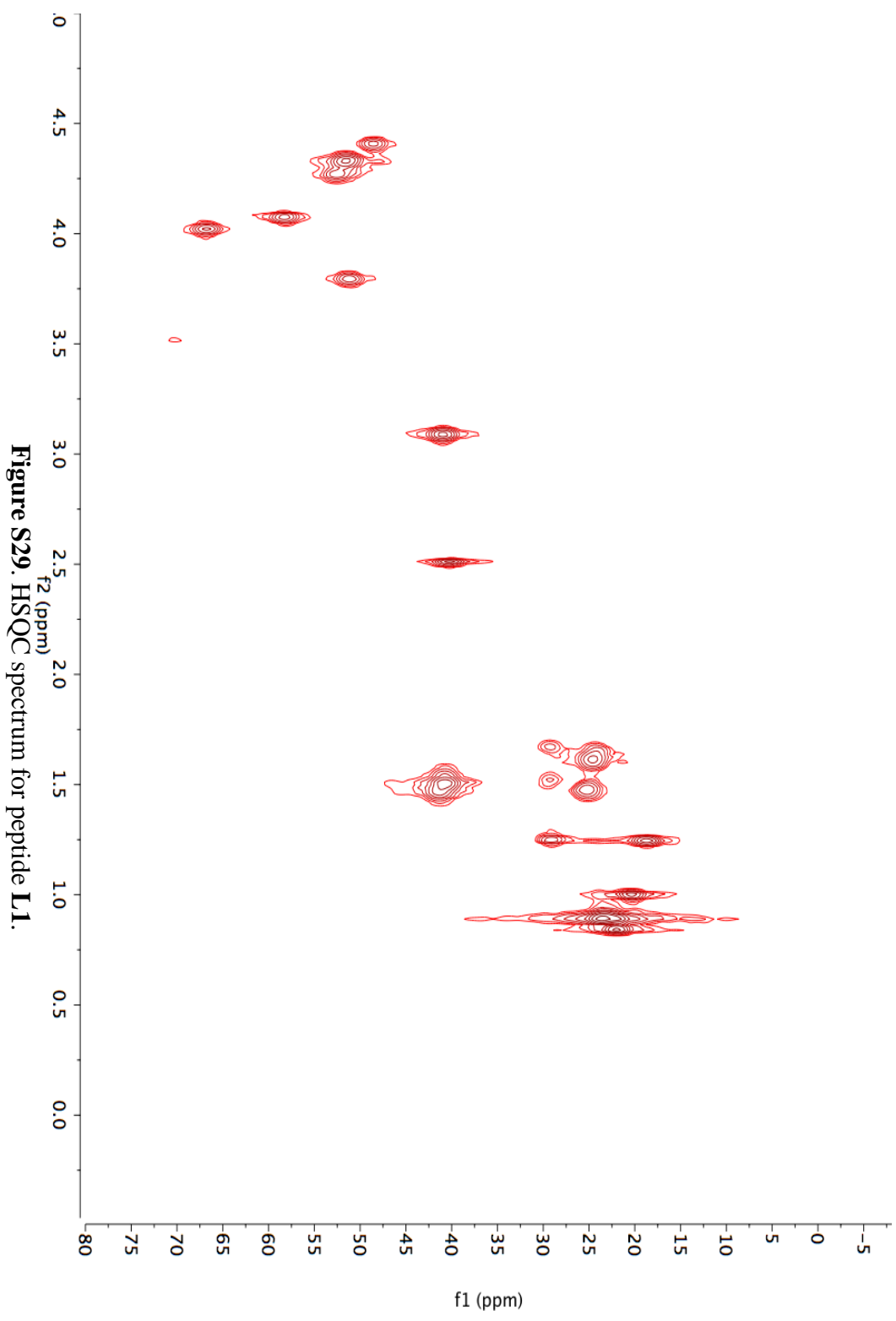


Figure S29. HSQC spectrum for peptide L1.

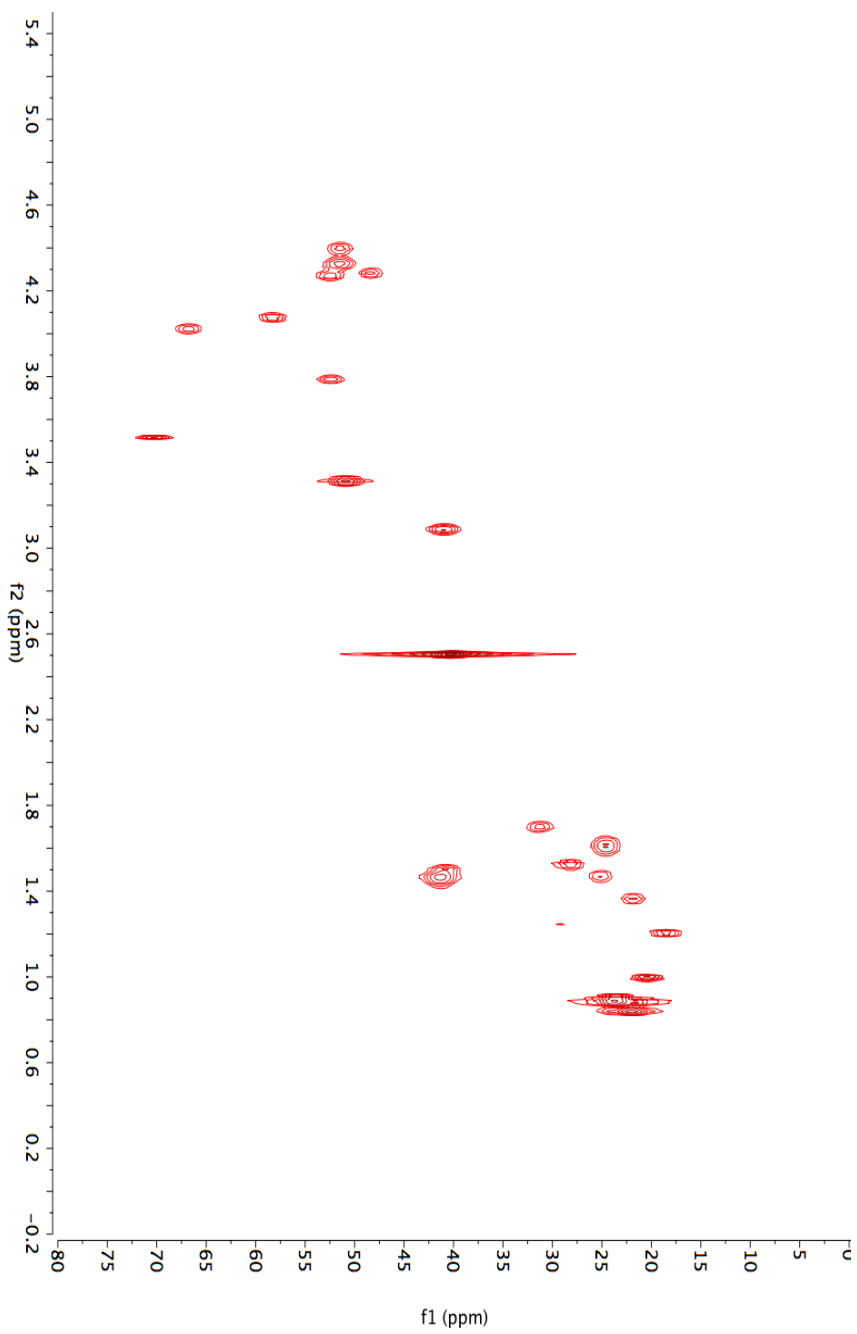


Figure S30. HSQC spectrum for peptide L1.5.

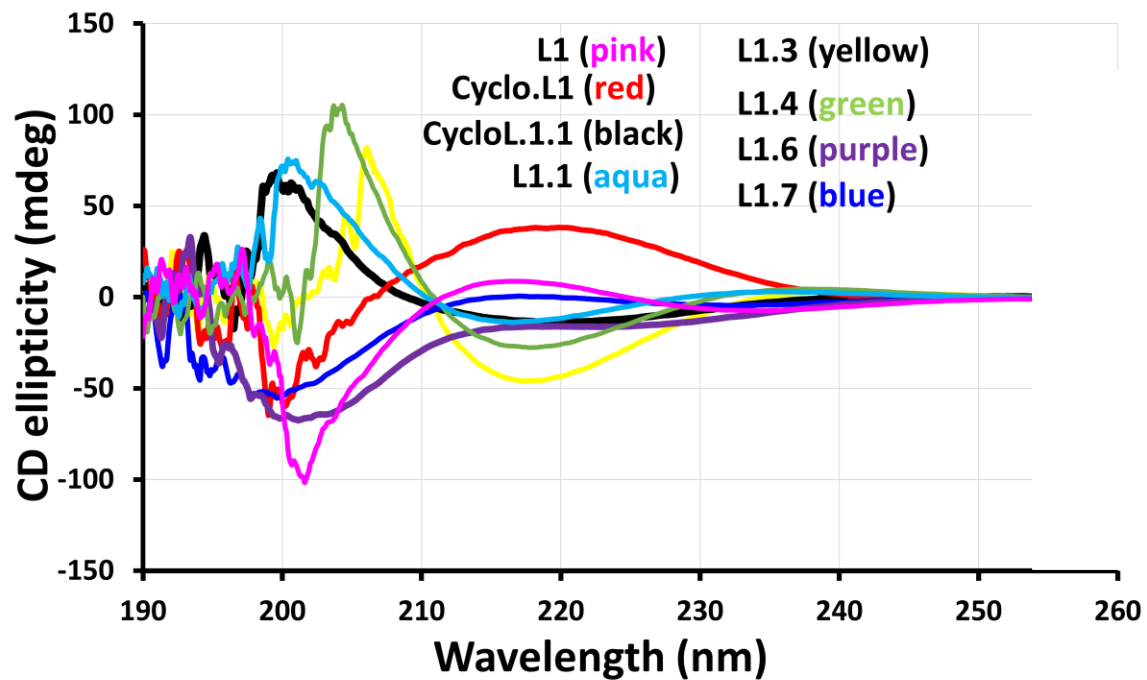


Figure S31. CD spectra of linear and cyclic peptides in methanol. 1 mg/mL of peptide was dissolved in methanol and CD spectra was acquired at 25 °C, and the spectra were average of 4 scans.