

Supporting Information

Grybchuk et al. 10.1073/pnas.1717806115

SI Methods

Cultivation of Trypanosomatids. Axenic cultures of monoxenous trypanosomatids of the genera *Leptomonas*, *Cryptosporidia*, and *Phytomonas* were obtained from (i) Life Science Research Centre, University of Ostrava, Ostrava, Czech Republic; (ii) Department of Parasitology, Charles University, Prague; (iii) Institute of Parasitology, Budweis, Czech Republic; (iv) Fundação Oswaldo Cruz, Rio de Janeiro; and (v) the Zoological Institute of the Russian Academy of Sciences, St. Petersburg, Russia. Trypanosomatids were cultivated in the Brain Heart infusion medium supplemented with 10 µg/mL of hemin, 500 units/mL of penicillin, and 0.5 mg/mL of streptomycin as described previously (105) and grown to the late logarithmic stage (10^7 – 10^8 cells/mL).

Phylogenetic Analyses of Viruses. RDRP sequences of the viruses characterized in this work were aligned with the related sequences from GenBank in the online version of MAFFT 7 using the E-INS-i method (106). Ambiguous parts of the alignments were removed with the use of TrimAl v. 1.3 (107). Positions with more than 50% gaps were filtered out by Gap Strip/Squeeze V. 2.1.0 (<https://www.hiv.lanl.gov/content/sequence/GAPSTREEZE/gap.html>). The resulting alignments had the following lengths: 412 aa (the *Narnaviridae* dataset), 273 aa (the *Tombus-/Nodaviridae* dataset), and 1,415 aa (the *Bunyaviridae* dataset).

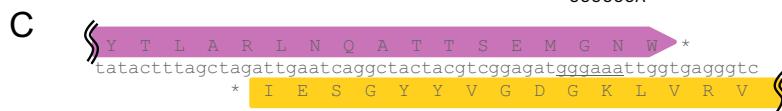
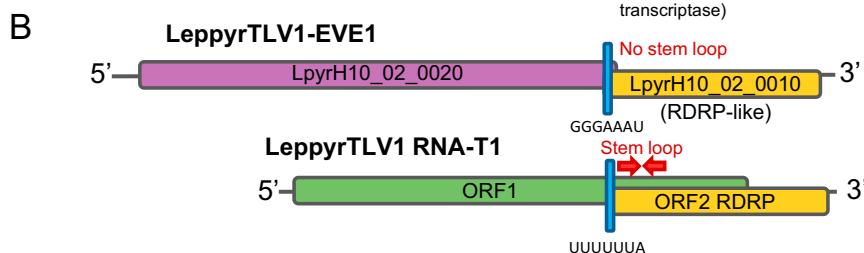
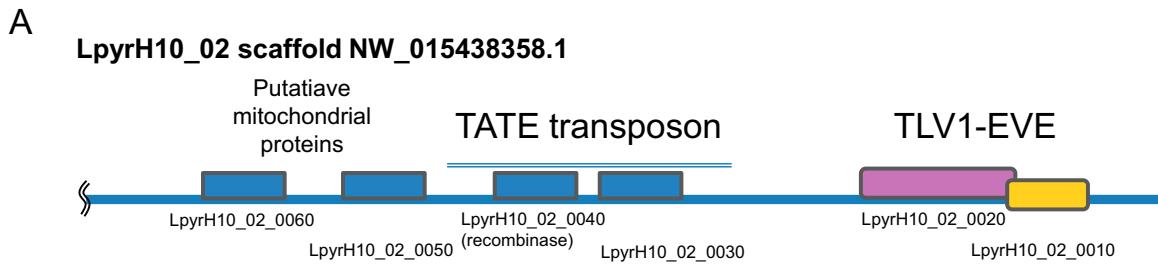
Maximum likelihood phylogenetic inference was performed in IQ-TREE v. 1.4.2 (108) with automatic selection of the best-fit amino acid substitution and site heterogeneity models (four gamma categories). The best-fitted model parameters defined by Bayesian Information Criterion were LG + F + I + G for the *Narnaviridae* and *Bunyaviridae* datasets and Blosum62 + I + G4 for the *Tombus-/Nodaviridae* dataset. Gaps were treated as missing data. Edge support was estimated with a bootstrap test (1,000 standard replicates). Bayesian inference was accomplished in MrBayes 3.2.6 (109) with the analyses ran for 1 million generations (given the observed fast convergence) and trees sampled every 100 generations. The mixed amino acid substitution model was used (resulting in 1.0 posterior probability of Blosum62 for all three datasets) with the heterogeneity over sites estimated using the G + I model. Amino acid frequencies were fixed to empirical values in the *Narnaviridae* and *Bunyaviridae* datasets and were estimated from the data matrix in the *Tombus-/Nodaviridae* dataset in accordance with the best-fit model defined by IQ-TREE v. 1.4.2 for each dataset. Other parameters were left in their default states.

Phylogeny of Trypanosomatid Hosts. The core alignments of 18S rRNA and gGAPDH genes were taken from previous work (76). Ambiguously aligned positions of 18S rRNA gene alignment were removed using Gblocks 0.91b as described previously (110). The concatenated alignments were subjected to maximum likelihood analysis in IQ-TREE (108) with partitioning by gene as well as by codon position for the gGAPDH gene. The best partitioned model of nucleotide substitutions (K3Pu + G4, TIM3 + I + G4, and TPM2u + G4 for the first, second, and third codon positions of gGAPDH gene, respectively and TNe + I + G4 for 18S rRNA gene) was selected with the use of ModelFinder (111). The statistical support of branches was estimated using 1,000 replicates of the standard bootstrap method.

Genomic and Transcriptomic Analyses. To find trypanosomatid signature sequences in the Sequence Reads Archive (SRA, <https://www.ncbi.nlm.nih.gov/sra>) and transcriptome shotgun assemblies (TSAs), TBLASTN and BLASTN searches were performed with amino acid (PFRT) and nucleotide (18S rRNA) sequences, respectively. In case of SRA-blast, the retrieved reads were assembled into contigs using the CAP3 sequence assembly program with the following parameters: minimal overlap length, 20 bp; minimum identity, 100% (112). Obtained contigs were extended by successive rounds of BLASTN searches against the original SRA. Final full-length contigs as well as hits from TSAs were subjected to blast search (megablast for nucleotide sequences and blastp for translated protein-coding sequences) against the nonredundant nucleotide collection of the NCBI database for identification. A similar approach was used to search of reads corresponding to nucleocapsid proteins in SRAs, which contained RDRPs closely related to LBVs. Obtained amino acid sequences were aligned with newly identified nucleocapsid proteins of viruses characterized in this work using BLAST pairwise alignment to confirm their identities.

Codon Usage Analysis in *Leptomonas pyrrhocoris* Viruses. Codon frequencies in the protein-coding regions of the six viral genes were analyzed using the CODONW program (codonw.sourceforge.net).

Negative-Stain Transmission Electron Microscopy. In brief, gradient-purified virus samples were applied to a carbon-coated copper grid, stained with molybdenum acetate, and examined under a Philips 201C transmission electron microscope. The morphologies and sizes of the virus particles were analyzed as described previously (113).

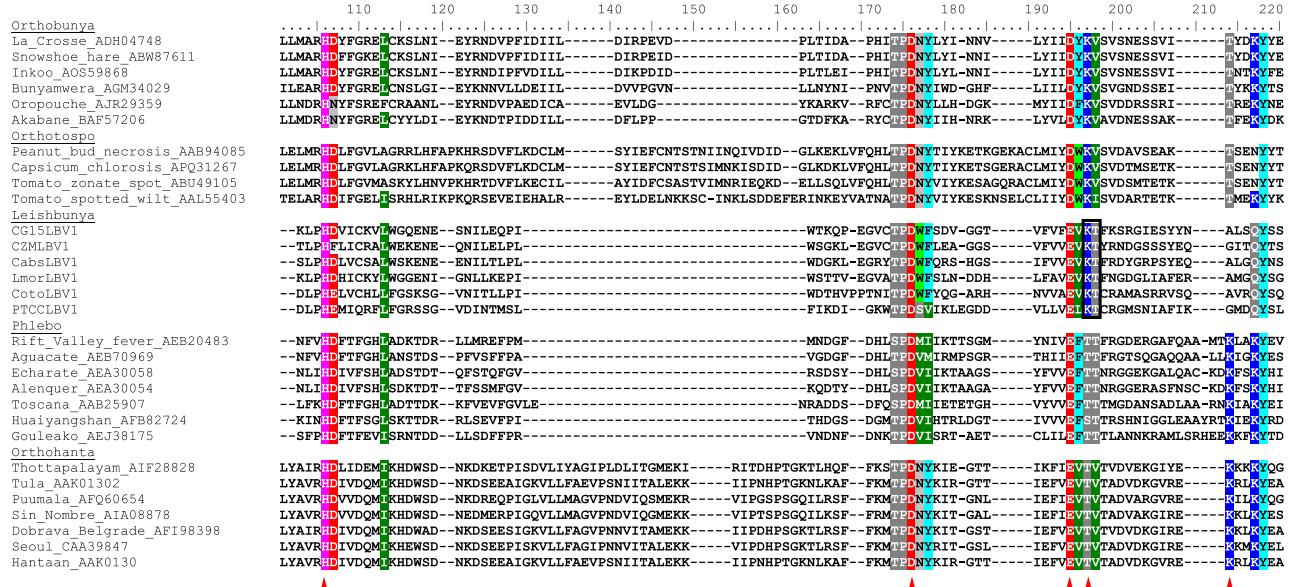


D

MOTIFS	F ₁	F ₂	A
<u>Picorna</u>			
1RDR_A_Poliovirus	(155) VTVYVKDE ELRS (164)	(169) EQGKSRLI EASSLND (183)	--KLFAD DYTG YDASLSP (243)
P03313.4_coxsackie_virus_B3	(156) VTVYVKDE LR (165)	(170) AKGKSRLI EASSLND (184)	--HLIAF DYSG YDASLSP (244)
AFK65743.1_Rhinovirus_C	(139) ITYLKDE LR (148)	(153) KVGKTRAI EASLND (167)	--NLLVF DYTN YDGSLN P (227)
P03305.1_foot-and-mouth_diseas	(160) QTFLKDE IRP (169)	(174) RAGKT RIVDVLPEVH (188)	YRNWWD DYSA FDANHCS (250)
<u>Flavi</u>			
P06935.2_west_nile_virus	(455) NMMGKRE KKP (464)	(469) KAKGSRAI WMWLGA (483)	GGKVYAD TAC WDTRITK (546)
P17763.2_dengue_virus	(452) NMMGKRE KKL (461)	(466) KAKGSRAI WMWLGA (480)	GGNMYAD TAG WDTRITE (543)
P19711.2_bovine_viral_diarrhea	(259) TAIPKNE KRD (268)	(280) VEKRP RVIQYPEAKT (294)	EPVAVSF DTKA WDTQVTS (355)
<u>Calici</u>			
Q83883.1_Norwalk_virus	(143) TAALKDE LVK (152)	(158) QKVKKRLL WGADLGT (172)	YKNHFDA DYTA WDSTQNR (233)
P27410.1_Rabbit_hemorrhagic_di	(142) ACGIKDE LRP (151)	(156) KEGKKRLL WGCDVGV (170)	ASDFLCL DYSK WDSTMSP (233)
Q6XDK8.1_Sapporo_virus	(315) KLALKDE LRP (324)	(329) AQGKR LLWGCDAGA (343)	GGVLYC LDSK WDSTQHP (404)
<u>LeppyrTLV1-EVE1</u>	(158) -SFIKDE GYD (166)	(167) EYKMP RAINSYSDST (181)	DSKVLYT DFSH FEHHRG (238)
<u>LeppyrTLV1</u>	(157) DSFIKDE SYD (166)	(167) ELKAP RSIMSYPPDV (181)	GGKVGST DFSS YECHHSG (238)
MOTIFS	B	C	D
<u>Picorna</u>			
1RDR_A_Poliovirus	(286) MPSGCSGT SIFNSMINNLII (305)	(322) KMIA Y GDDVIASYP (335)	KDYGLTM --T (355)
P03313.4_coxsackie_virus_B3	(288) MPSGCSGT SIFNSMINNIII (307)	(324) RMIAY GDDVIASYP (337)	KGYGLIM --T (357)
AFK65743.1_Rhinovirus_C	(268) MPSGISGT SIFNTIINNIII (287)	(304) KIVAY GDDVIASYP (317)	VKYGLTI --T (337)
P03305.1_foot-and-mouth_diseas	(296) MPSGCSAT SIINTILNNIYV (315)	(332) TMISY GDDIVVASD (345)	KSLCQTI --T (365)
<u>Flavi</u>			
P06935.2_west_nile_virus	(602) RGS GQVVTYALNTFTNLAVQ (621)	(662) RMAVS GDDCVVKP (674)	NAMSKVRKD IQ (697)
P17763.2_dengue_virus	(598) RGS GQVTYGLNTFTNMEAQ (617)	(656) RMAIS GDDCVVKP (668)	NDMGKVRKD IIP (691)
P19711.2_bovine_viral_diarrhea	(403) RGS QPDT SAGNSMLNVLT M (422)	(442) RIHVC GDDGFLITE (455)	HEAGKPQK-IT (480)
<u>Calici</u>			
Q83883.1_Norwalk_virus	(279) LPSGFPCT SQVNSINHWIIT (298)	(318) YFSFY GDDIEVSTD (331)	KEYGLKP --T (351)
P27410.1_Rabbit_hemorrhagic_di	(279) LPSGMPFT SVINSICHWLLW (298)	(321) PFYTY GDDGVYAMT (334)	RDYGLSP --T (356)
Q6XDK8.1_Sapporo_virus	(450) LPSGMPFT SVINSLNHMTYF (469)	(493) TVHTY GDDCLYSVC (506)	TSFGLKP --T (528)
<u>LeppyrTLV1-EVE1</u>	(284) LMSGALWT SFQNSFLNFFVM (303)	(323) KMFIE GDDGIMKH (336)	KRLGLCLK-IN (353)
<u>LeppyrTLV1</u>	(288) LMSGALWT SFQNSFLNLMMI (307)	(325) DTFIE GDDGIFRAF (338)	ARLGIKLK-IS (355)

Fig. S1. LeppyrTLV1 EVE1. (A) The endogenous viral element is located at the subtelomeric region of the *Leptomonas pyrrhocoris* chromosome (scaffold NW_015438358.1). The EVE1 element is preceded by the reverse transcriptase-coding TATE DNA transposon indicating the possible mechanism for the LeppyrTLV1 endogenization. (B) Comparison of the ORF organization of RDRP-coding RNA-T1 of 1 LeppyrTLV1 and its endogenous viral element. (C) The overlap region between the ORFs of EVE1 contains a putative slippery sequence (underlined) capable of driving a -1 ribosomal frameshift, similarly to LeppyrTLV1. (D) Multiple alignments of RDRPs of LeppyrTLV1 and LeppyrTLV1-EVE1 with RNA polymerases of Picorna-, Flavi-, and Caliciviridae. Identical residues are indicated in red; similar residues are in blue. Each block shows amino acid motifs typically found in viral RNA polymerases (59).

A



B

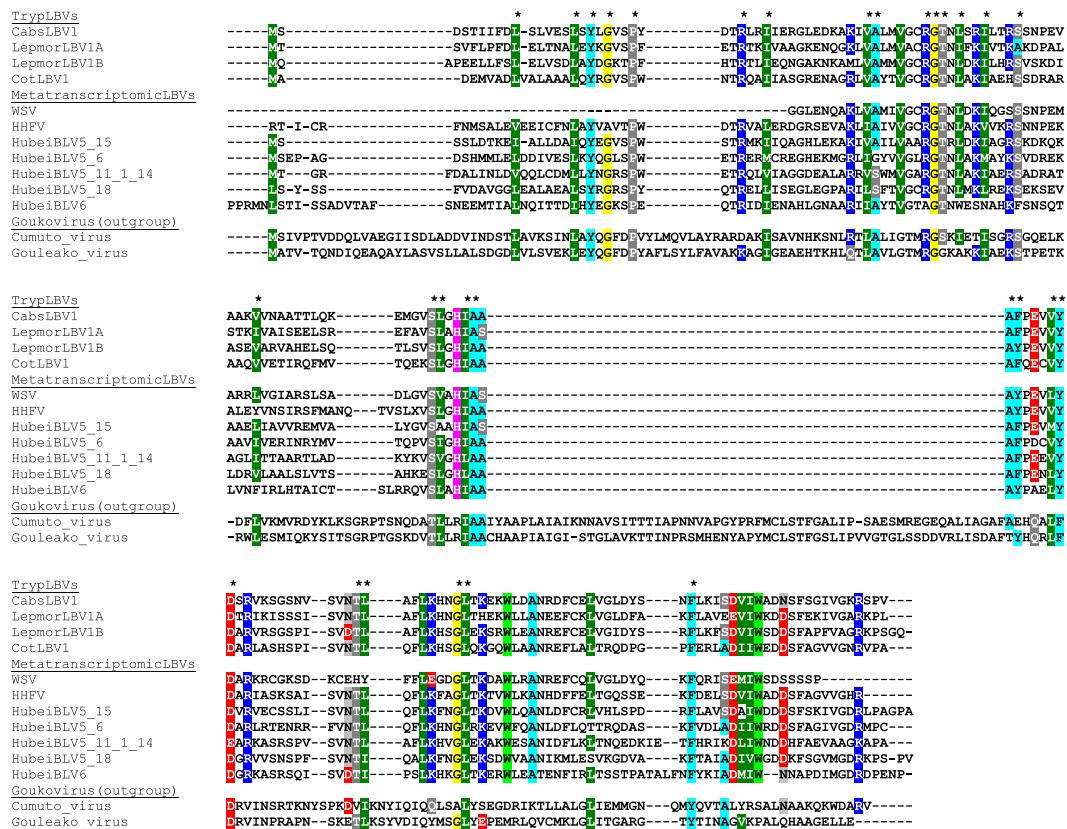


Fig. S2. Leishbunyavirus protein domains. (A) Amino acid alignment of the N-terminal endonuclease domain of the L protein of leishbunyaviruses and other bunyaviruses. Functionally important residues are marked with arrowheads. Residues implicated in nucleolytic cleavage in leishbunyaviruses are boxed. (B) Amino acid alignment of nucleocapsid proteins of leishbunyaviruses reported in ref. 34 and this work (TrypLBVs) and assembled from metatranscriptomes (metatranscriptomic BVs). Cumuto and Gouleako viruses are shown for comparison. Indels are shown as dashes. Positions with more than 82% similarity are shaded; those with 90% similarity are starred.

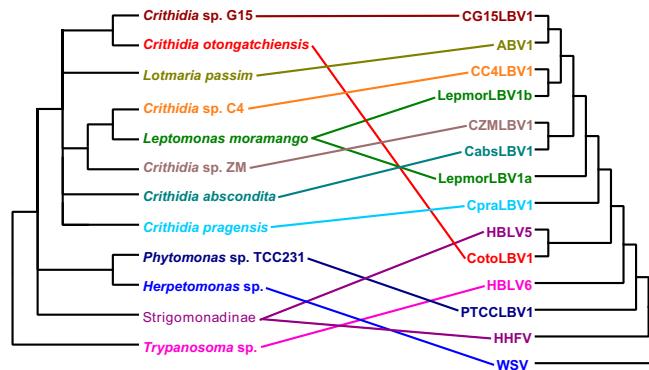


Fig. S3. Comparison of the phylogenies of leishbunyaviruses and their respective hosts. The scheme is based on the phylogenetic trees presented in Figs. 3 and 5 with simplifications: (i) no branch lengths are shown on either tree; (ii) only LBVs and LBV-containing trypanosomatids were included; and (iii) all clades with bootstrap support below 70% were collapsed and reshuffled to minimize the number of potential viral transitions between hosts.

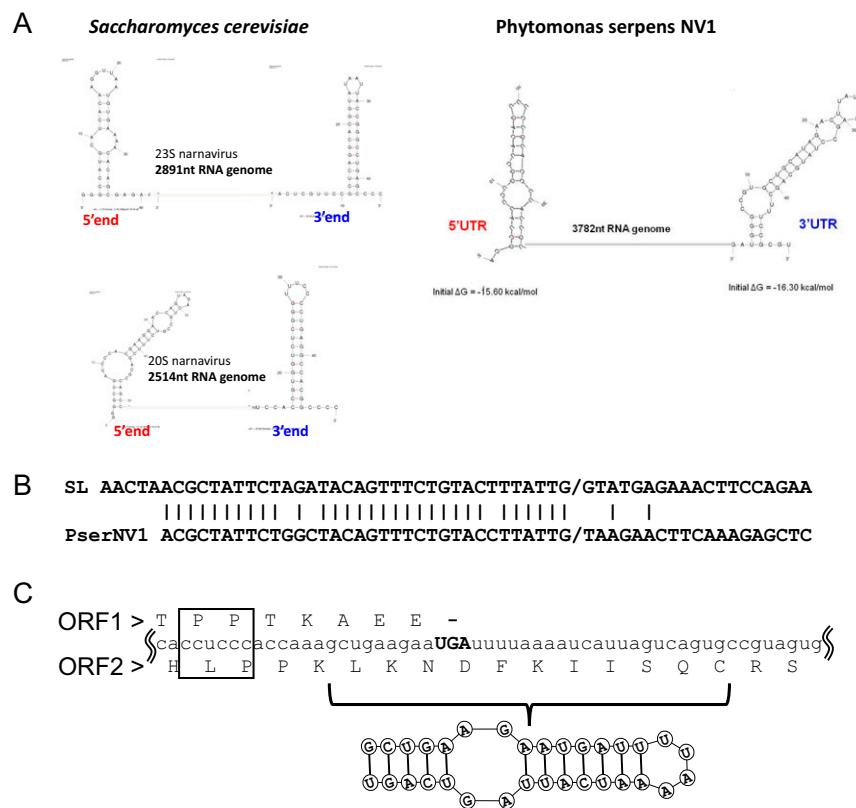


Fig. S4. Features of genomic RNAs of PserNV1 and LepseyNLV1. (A) Comparison of terminal complementary sequences and stem-loop structures with those of narnaviruses from *Saccharomyces cerevisiae*. (B) Nucleotide alignment of the 5' end of PserNV1 and the SL RNA gene from *Phytomonas serpens*. The SL/miniexon boundary is indicated by a slash symbol. (C) Features of a potential frameshift region between ORF1 and ORF2 of the LepseyNLV1 small segment. The location of a hairpin motif is marked by a bracket, which has a predicted ΔG of -6 kCal [OligoAnalyzer 3.1 (114)]. A putative pyrimidine-rich slippery sequence (CCUCCC) positioned 6 nt upstream is shown in the box.

Table S1. Complete list and properties of the studied trypanosomatid species

Species	Isolate	Reference to trypanosomatid description	Host	Geographic origin	Presence of viruses
<i>Crittidia abscondita</i>	127AL	Yurchenko et al. (2009) (1)	<i>Largus</i> sp.	Ecuador	Bunyavirales
<i>Crittidia brachyflagelli</i>	340VL	Jirků et al. (2012) (2)	<i>Prepos cf accinctus</i>	Costa Rica	Not observed
<i>Crittidia brevicula</i>	101	Kostygov et al. (2014) (3)	<i>Nabis flavomarginatus</i>	Russia	Not observed
<i>Crittidia brevicula</i>	KV1	Kostygov et al. (2014) (3)	<i>Gerris lacustris</i>	Russia	Not observed
<i>Crittidia brevicula</i>	F6	Kostygov et al. (2014) (3)	<i>Nabis flavomarginatus</i>	Russia	Not observed
<i>Crittidia fasciculata</i>	Finn-01.02	Hamilton et al. (2015) (4)	<i>Drosophila falleni</i>	USA	Not observed
<i>Crittidia fasciculata</i>	COLPROTO53	Wallace and Clark (1959) (5)	<i>Phaenicia sericata</i>	USA	Not observed
<i>Crittidia fasciculata</i>	COLPROTO48	Wallace (1943) (6)	<i>Anopheles quadrimaculatus</i>	France	Not observed
<i>Crittidia fasciculata</i>	Ecu-08	Yurchenko et al. (2014) (7)	Family Syphidae	Ecuador	Bunyavirales
<i>Crittidia fasciculata</i>	128S1	Yurchenko et al. (2009) (1)	Tribe Mirini	Ecuador	Not observed
<i>Crittidia permixta</i>	MCZ-11	Yurchenko et al. (2014) (7)	<i>Cordilura albipes</i>	Czech Republic	Bunyavirales
<i>Crittidia pragensis</i>	G15	Votyka et al. (2012b) (8)	<i>Rhynocoris rapax</i>	Ghana	Bunyavirales
<i>Crittidia sp.</i>	ZM	Podlipaev and Frolov (1987) (9)	<i>Lygocoris lucorum</i>	Russia	Bunyavirales
<i>Crittidia sp.</i>	C4	Podlipaev et al. (2004) (10)	<i>Limnoporus rufoscutellatus</i>	Russia	Bunyavirales
<i>Crittidia sp.</i>	Cfm9	Merzlyak et al. (2001) (11)	<i>Nabis flavomarginatus</i>	Russia	Not observed
<i>Crittidia sp.</i>	CL6	Podlipaev and Rokitskaya (1999) (12)	<i>Nabis limbatus</i>	Russia	Not observed
<i>Crittidia thermophila</i>	320AR	Jirků et al. (2012) (2)	<i>Largus maculatus</i>	Costa Rica	Not observed
<i>Crittidia thermophila</i>	COLPROTO18	Roitman et al. (1977) (13)	<i>Cosmoclopius</i> sp.	Brazil	Not observed
<i>Crittidia thermophila</i>	COLPROTO54	Roitman et al. (1977) (13)	<i>Zelus leucogrammus</i>	Costa Rica	Not observed
<i>Crittidia thermophila</i>	15EC	Yurchenko et al. (2006b) (14)	<i>Ricolla similisima</i>	Brazil	Not observed
<i>Crittidia thermophila</i>	34EC	Yurchenko et al. (2009) (1)	<i>Jadera obscura</i>	Costa Rica	Not observed
<i>Crittidia thermophila</i>	MMO-09	Yurchenko et al. (2014) (7)	<i>Pachycerina cf vaga</i>	Madagascar	Bunyavirales
<i>Crittidia thermophila</i>	F165	Votyka et al. (2012a) (15)	<i>Pyrhocoris apterus</i>	France	LeppyrTLV1
<i>Crittidia thermophila</i>	F19	Votyka et al. (2012a) (15)	<i>Pyrhocoris apterus</i>	France	LeppyrTLV1, LeppyrOV1
<i>Crittidia thermophila</i>	H10	Votyka et al. (2012a) (15)	<i>Pyrhocoris apterus</i>	Czech Republic	LeppyrTLV1, LeppyrOV1, LeppyrTLV1-EVE
<i>Crittidia jadraeae</i>	10VL	Votyka et al. (2012a) (15)	<i>Dysdercus obliquus</i>	Costa Rica	Not observed
<i>Crittidia jadraeae</i>	121AL	Votyka et al. (2012a) (15)	<i>Dysdercus obscuratus</i>	Ecuador	Not observed
<i>Leptomonas moramango</i>	122AL	Votyka et al. (2012a) (15)	<i>Dysdercus obscuratus</i>	Costa Rica	Not observed
<i>Leptomonas pyrrhocoris</i>	14BT	Votyka et al. (2012a) (15)	<i>Dysdercus lunulatus</i>	Costa Rica	Not observed
<i>Leptomonas pyrrhocoris</i>	25EC	Votyka et al. (2012a) (15)	<i>Dysdercus sp.</i>	Costa Rica	Not observed
<i>Leptomonas pyrrhocoris</i>	28EC	Votyka et al. (2012a) (15)	<i>Dysdercus sp.</i>	Costa Rica	Not observed
<i>Leptomonas pyrrhocoris</i>	324RV	Votyka et al. (2012a) (15)	<i>Dysdercus obscuratus</i>	Costa Rica	Not observed
<i>Leptomonas pyrrhocoris</i>	329MV	Votyka et al. (2012a) (15)	<i>Dysdercus mimulus</i>	Costa Rica	Not observed
<i>Leptomonas pyrrhocoris</i>	CH278	Votyka et al. (2012a) (15)	<i>Dysdercus poecilus</i>	China	Not observed
<i>Leptomonas pyrrhocoris</i>	G58	Votyka et al. (2012a) (15)	<i>Dysdercus fasciatus</i>	Ghana	Not observed
<i>Leptomonas pyrrhocoris</i>	K06	Votyka et al. (2012a) (15)	<i>Scantius aegyptius</i>	Cyprus	Not observed
<i>Leptomonas pyrrhocoris</i>	P59	Votyka et al. (2012a) (15)	<i>Pyrhocoris marginatus</i>	Burkina Faso	Not observed (LeppyrTLV1-EVE)
<i>Leptomonas pyrrhocoris</i>	SERG	Flegontov et al. (2016) (16)	<i>Dysdercus sp.</i>	Russia	Not observed
<i>Leptomonas pyrrhocoris</i>	PP1	Frolov et al. (2014) (17)	<i>Pyrhocoris apterus</i>	Russia	Not observed (LeppyrTLV1-EVE)
<i>Leptomonas pyrrhocoris</i>	PP2	Frolov et al. (2014) (17)	<i>Pyrhocoris apterus</i>	France	Not observed (LeppyrTLV1-EVE)
<i>Leptomonas pyrrhocoris</i>	LP	Yurchenko et al. (2006a) (18)	<i>Scantius aegyptius</i>	USA	Not observed
<i>Leptomonas scartii</i>	F221	Votyka et al. (2012a) (15)	<i>Dysdercus suturellus</i>	Costa Rica	<i>Narnaviridae</i>
<i>Leptomonas seymouri</i>	ATCC	Wallace (1977) (19)	<i>Prepopus</i> sp.	Brazil	Not observed
<i>Leptomonas taroles</i>	47VL	Yurchenko et al. (2008) (20)	<i>Lycopersicon esculentum</i>	Brazil	Not observed
<i>Phytomonas serpens</i>	9T (CB)	Jankevicius et al. (1989)	<i>Lycopersicon esculentum</i>	Brazil	Not observed
<i>Phytomonas serpens</i>	9T (UCR)	Jankevicius et al. (1989)	<i>Lycopersicon esculentum</i>	Brazil	<i>Narnaviridae</i>

Table S1. Cont.

Species	Isolate	Reference to trypanosomatid description	Host	Geographic origin	Presence of viruses
<i>Phytonomonas serpens</i>	30T	Jankevicius et al. (1989) (21)	<i>Lycopersicon esculentum</i>	Brazil	<i>Nannaviridae</i>
<i>Phytonomonas serpens</i>	1G	Da Silva and Roitman (1990) (22)	<i>Phthia picta</i>	Brazil	Not observed
<i>Phytonomonas serpens</i>	15T, COLPROT186	Jankevicius et al. (1989) (21)	<i>Solanum lycopersicum</i>	Brazil	Not observed
<i>Phytonomonas sp.</i>	TCC231	Jankevicius et al. (1989) (21)	<i>Lycopersicon esculentum</i>	Brazil	Bunyavirales
<i>Phytonomonas sp.</i>	COLPROT079	Kastelein and Parsadi (1988) (23)	<i>Allamanda cathartica</i>	Suriname	Not observed
<i>Phytonomonas sp.</i>	COLPROT080	Conchon et al. (1989) (24)	<i>Citrus bergamia</i>	Brazil	Not observed

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Table S2. Sequences of viral RDRPs with working abbreviations of viral names used in phylogenetic inferences

Clade	Short name	Virus name	Accession no.	Host
<i>Tombus</i> -like viruses from <i>Leptomonas pyrrhocoris</i> (Fig. 1D)				
<i>Tombusviridae</i>	JINRV	<i>Japanese iris necrotic ring virus</i>	BA92792.1	Plants (<i>Iris ensata</i>)
<i>Tombusviridae</i>	TCV	<i>Turnip crinkle virus</i>	AAP78486.1	Plants
<i>Tombusviridae</i>	MNSV	<i>Melon necrotic spot virus</i>	ABG67516.1	Plants
<i>Tombusviridae</i>	CarMV	<i>Carnation mottle virus</i>	CAB38331.1	Plants
<i>Tombusviridae</i>	OMMV	<i>Olive mild mosaic virus</i>	AE50092.1	Plants (<i>Olea europaea</i>)
<i>Tombusviridae</i>	PoLV	<i>Pothos latent virus</i>	Q9IWA0.1	Plants
<i>Tombusviridae</i>	TBSV	<i>Tomato bushy stunt virus</i>	AAT67237.1	Crustacea: <i>Procambarus clarkia</i>
<i>Tombusviridae</i>	CTLV14	<i>Changjiang tombus-like virus 14</i>	APG76248.1	Myriapoda: Scutigeridae
<i>Tombusviridae</i>	HTLV13	<i>Hubei tombus-like virus 13</i>	APG76577.1	Insecta: Gerridae sp.
<i>Tombus-like viruses</i>	STLV6	<i>Sanxia tombus-like virus 6</i>	APG76428.1	Mollusca: Octopodidae sp.
<i>Tombus-like viruses</i>	BTLV9	<i>Beihai tombus-like virus 9</i>	APG76191.1	Insecta: <i>Paracerceis melanotum</i> , <i>Paracerceis calamorum</i> , <i>Ceratiagnion auranticum</i> , <i>Brachydiplos chalybea</i> , <i>Orthotrichum albistylum</i> , <i>Pseudotheremis zonata</i> , <i>Chironomus</i> sp.
<i>Tombus-like viruses</i>	HTLV22	<i>Hubei tombus-like virus 22</i>	APG76327.1	Myriapoda: <i>Diplopoda</i> sp., <i>Ototigmus scaber</i> , <i>Scolopocryptops</i> sp., <i>Ototigmus scaber</i> , <i>Myriapoda</i> sp.
<i>Tombus-like viruses</i>	HTLV36	<i>Huberi tombus-like virus 36</i>	APG76457.1	Crustacea: <i>Procambarus clarkia</i>
<i>Tombus-like viruses</i>	CTLV22	<i>Changjiang tombus-like virus 22</i>	APG76278.1	Unknown
<i>Tombus-like viruses</i>	BCU1	<i>Brandmavirus UC1</i>	AHA86931.1	Mollusca: <i>Pomacea canaliculata</i>
<i>Tombus-like viruses</i>	WTLV17	<i>Wenzhou tombus-like virus 17</i>	APG76615.1	Myriapoda: <i>Diplopoda</i> sp., <i>Ototigmus scaber</i> , <i>Scolopocryptops</i> sp., <i>Ototigmus scaber</i> , <i>Myriapoda</i> sp.
<i>Tombus-like viruses</i>	HTLV35	<i>Hubei tombus-like virus 35</i>	APG76480.1	Trypanosomatidae
<i>Tombus-like viruses</i>				Mollusca: <i>Solen strictus</i>
<i>Tombus-like viruses</i>				Mollusca: Octopodidae sp.
<i>Tombus-like viruses</i>				Unknown
<i>Tombus-like viruses</i>				Nematoda: <i>Ascaridia</i> sp.
<i>Tombus-like viruses</i>				Insecta: <i>Apis mellifera</i>
<i>Tombus-like viruses</i>				Insecta: <i>Anopheline</i> sp.
<i>Tombus-like viruses</i>				Insecta: <i>Apis mellifera</i>
<i>Tombus-like viruses</i>				Insecta: <i>Drosophila</i> sp.
<i>Tombus-like viruses</i>				Insecta (Diptera): <i>Atherigona orientalis</i> , <i>Chrysomya megacephala</i> , <i>Lucilia sericata</i> , <i>Musca domestica</i> , <i>Sarcophaga dux</i> , <i>S. peregrina</i> , <i>Sarcophaga</i> sp.
<i>Tombus-like viruses</i>	LeppyT1-EVE1	<i>LeppyT1V1 endogenous virus element</i>		Crustacea: <i>Amphibalanus rhizophorae</i>
<i>Tombus-like viruses</i>	BTLV1	<i>Leptomonas pyrrhocoris tombus-like virus 1</i>	APG76183.1	Crustacea: <i>Brachyura</i> sp., <i>Achelata</i> sp., <i>Penaeoidea</i> spp., <i>Ibacus novemdentatus</i> , <i>Anomura</i> sp., <i>Penaeidae</i> sp., <i>Charybdis bimaculata</i> , <i>Charybdis rufodactylus</i> , <i>Latreillidae</i> sp., <i>Ovalipes punctatus</i>
<i>Tombus-like viruses</i>	BTLV17	<i>Beihai tombus-like virus 17</i>	APG76207.1	
<i>Tombus-like viruses</i>	BTLV18	<i>Beihai tombus-like virus 18</i>	AOG30801.1	
<i>Tombus-like viruses</i>	Towan	<i>Towan virus</i>		
<i>Tombus-like viruses</i>	JTLV2	<i>Jingmen tombus-like virus 2</i>	APG76305.1	
<i>Tombus-like viruses</i>	LSV1	<i>Lake Sinai virus 1</i>	AEP26193.1	
<i>Tombus-like viruses</i>	AaCV	<i>Anopheline-associated C virus</i>	AGW51750.1	
<i>Tombus-like viruses</i>	CBPV	<i>Chronic bee paralysis virus</i>	ACOB2537.1	
<i>Tombus-like viruses</i>	Dansoman	<i>Dansoman virus</i>	AKHA0306.1	
<i>Tombus-like viruses</i>	HTLV42	<i>Hubei tombus-like virus 42</i>	APG76280.1	
<i>CBPV-like viruses</i>	BTLV19	<i>Beihai tombus-like virus 19</i>	APG76134.1	
<i>CBPV-like viruses</i>	WeTLV4	<i>Wenling tombus-like virus 4</i>	APG76579.1	

Table S2. Cont.

Clade	Short name	Virus name	Accession no.	Host
CBPV-like viruses	WCV4	Wenzhou crab virus 4	APG76640.1	Crustacea: <i>Charybdis hellerii</i> , <i>Charybdis japonica</i> , <i>Charybdis lucifera</i>
CBPV-like viruses	WTLV18	Wenzhou tombs-like virus 18	APG76097.1	Mollusca: <i>Barbatia virescens</i> , <i>Sinonovacula constricta</i> , <i>Tegillarca granosa</i> , <i>Crassostrea arizakensis</i> , <i>Mytilus coruscus</i>
CBPV-like viruses	PHVA	<i>Plasmopara halstedii</i> virus A	ADK55578.1	Oomycetes: <i>Plasmopara halstedii</i>
CBPV-like viruses	SmVA	<i>Sclerotophthora macrospora</i> virus A	BAC11954.1	Oomycetes: <i>Sclerotophthora macrospora</i>
Nodaviridae	FHV	<i>Flock house virus</i>	CAA54399.1	Insecta (Coleoptera): <i>Costelytra zealandica</i>
Nodaviridae	NoV	<i>Nodamura virus</i>	AAP97860.1	Insecta (Diptera): <i>Culex tritaeniorhynchus</i>
Nodaviridae	PaV	<i>Pariacoto virus</i>	AAF71691	Insecta (Lepidoptera): <i>Spodoptera eridania</i>
Nodaviridae	SJNNV	<i>Striped Jack nervous necrosis virus</i>	BAB64329.1	Chordata: <i>Cocinero</i> sp.
Leishbunyaviruses (Fig. 3D)				
<i>Phlebovirus</i>	CDUV	<i>Chandiru virus</i>	AEA30057.1	Arthropod vectors (Diptera), vertebrate hosts
<i>Phlebovirus</i>	PTV	<i>Punta Toro phlebovirus</i>	ALL45372.1	Arthropod vectors (Diptera), vertebrate hosts
<i>Phlebovirus</i>	BUJV	<i>Bujaru virus</i>	APG68880.1	Arthropod vectors (Diptera), vertebrate hosts
<i>Phlebovirus</i>	Adana	<i>Adana virus</i>	AJk91618.1	Arthropod vectors (Diptera), vertebrate hosts
<i>Phlebovirus</i>	SALV	<i>Salehabad phlebovirus</i>	AGA82741.1	Arthropod vectors (Diptera), vertebrate hosts
<i>Phlebovirus</i>	SFNV	<i>Sandfly fever Naples virus</i>	CAA8478.1	Arthropod vectors (Diptera), vertebrate hosts
<i>Phlebovirus</i>	RVFV	<i>Rift Valley fever virus</i>	ABP1507.1	Arthropod vectors (Diptera), vertebrate hosts
<i>Phlebovirus</i>	SFTSV	<i>Severe fever with thrombocytopenia virus</i>	ADP04470.1	Arthropod vectors (Diptera or ticks), vertebrate hosts
<i>Phlebovirus</i>	BHAV	<i>Bhanja virus</i>	AFG066272.1	Arthropod vectors (Diptera or ticks), vertebrate hosts
<i>Phlebovirus</i>	LSV	<i>Lone Star virus</i>	AGL50921.1	Arthropod vectors (Diptera or ticks), vertebrate hosts
<i>Phlebovirus</i>	EgAN	<i>EgAN 1825-61 virus</i>	AEL29654.1	Arthropod vectors (ticks), vertebrate hosts
<i>Phlebovirus</i>	UU.K.V	<i>Uukuniemi phlebovirus</i>	BAA01590.1	Arthropod vectors (ticks), vertebrate hosts
<i>Phlebovirus</i>	Khasan	<i>Khasan virus</i>	All79370.1	Arthropod vectors (ticks), vertebrate hosts
<i>Phlebovirus</i>	Badu	<i>Badu phasivirus</i>	AMA19446.1	Insects (mosquitoes)
<i>Phasivirus</i>	WMPV	<i>Wutai mosquito phasivirus</i>	AGC39270.1	Insects (mosquitoes)
<i>Phasivirus</i>	PCLPV	<i>Phasi Charioen-like phasivirus</i>	AKP18602.1	Insects (mosquitoes)
<i>Phasivirus</i>	WFV	<i>Wuhan fly phasivirus</i>	AGG39259.1	Insects (mosquitoes)
<i>Tenuivirus</i>	RaSV	<i>Ramu stunt virus</i>	ALB83282.1	Arthropod vectors (Hemiptera), plant hosts
<i>Tenuivirus</i>	RGSV	<i>Rice grassy stunt tenuivirus</i>	BAA89602.1	Arthropod vectors (Hemiptera), plant hosts
<i>Tenuivirus</i>	RiSV	<i>Rice stripe tenuivirus</i>	AFM93792.1	Arthropod vectors (Hemiptera), plant hosts
<i>Tenuivirus</i>	Cumuto	<i>Cumuto virus</i>	AHH60917.1	Insects (mosquitoes)
<i>Goukovirus</i>	Gouleako	<i>Gouleako virus</i>	AE38175.1	Insects (mosquitoes)
<i>Goukovirus</i>	YIV	<i>Yichang insect virus</i>	AGG39273.1	Insects (mosquitoes)
<i>Goukovirus</i>	ABV1	<i>Apis bunyavirus 1</i>	ARO50045.1	Honey bee (<i>Apis mellifera</i>) + Trypanosomatidae (<i>Lotmaria passim</i>)
<i>Leishbunyaviridae</i>	CG15LBV1	<i>Critthidia</i> sp. G.5 Leishbunyavirus 1		Trypanosomatidae
<i>Leishbunyaviridae</i>	LmoriLBV1b	<i>Leptomonas moramango</i> Leishbunyavirus 1b		Trypanosomatidae
<i>Leishbunyaviridae</i>	CC4LBV1	<i>Critthidia</i> sp. C.4 Leishbunyavirus 1		Trypanosomatidae
<i>Leishbunyaviridae</i>	CZMLBV1	<i>Critthidia</i> sp. Z.M Leishbunyavirus 1		Trypanosomatidae
<i>Leishbunyaviridae</i>	CabstLBV1	<i>Critthidia abscondita</i> Leishbunyavirus 1		Trypanosomatidae
<i>Leishbunyaviridae</i>	LmorLBV1a	<i>Leptomonas moramango</i> Leishbunyavirus 1a		Honey bee (<i>Apis mellifera</i>) + ?
<i>Leishbunyaviridae</i>	DubV	<i>Duke bunyavirus</i>	ARE30258.1	Trypanosomatidae
<i>Leishbunyaviridae</i>	CpralLBV1	<i>Critthidia pragensis</i> Leishbunyavirus 1		

Table S2. Cont.

Clade	Short name	Virus name	Accession no.	Host
Leishbunyaviridae	HBLV5	Hubei bunya-like virus 5	APG79301.1	Insects (Diptera) + Trypanosomatidae (Strigomonadinae)
Leishbunyaviridae	CotolBV1	<i>Critchidia</i> otongatchiensis Leishbunyavirus 1	APG79326.1	Trypanosomatidae
Leishbunyaviridae	HBLV6	Hubei bunya-like virus 6		Horse leech (<i>Whitmania pigra</i>) + Trypanosomatidae (Trypanosoma sp)
Leishbunyaviridae	PTCCLBV1	Phytomonas sp. TCC231 Leishbunyavirus 1	AJG39239.1	Humpbacked Fly (<i>Megaselia scalaris</i>) + Trypanosomatidae (Strigomonadinae)
Leishbunyaviridae	HHFV	Huangshi Humpbacked Fly virus		Arthropods (Araneae) + Diptera, Heteroptera + Trypanosomatidae (Herpetomonas sp)
Leishbunyaviridae	WSV	Wuhan Spider virus	AJG39269.1	Arthropod vectors (ticks), vertebrate hosts
Nairoviridae, Orthonairovirus	DONV	Dugbe orthonairovirus	AMT75392.1	Arthropod vectors (ticks), vertebrate hosts
Nairoviridae, Orthonairovirus	CCHFONV	Crimean-Congo hemorrhagic fever orthonairovirus	ARRB51463.1	Arthropod vectors (ticks), vertebrate hosts
Arenaviridae	LCMAV	Lymphocytic choriomeningitis mammarenavirus	AMZ00419.1	Vertebrates
Arenaviridae	MaMAV	Machupo mammarenavirus	AF993553.1	Vertebrates
Arenaviridae	GGV	Alethinophid 1 reparenavirus (Golden Gate virus)	AGH06042.1	Vertebrates
Arenaviridae	Boa	Alethinophid 3 reparenavirus (Boa arenavirus)	ALAF04559.1	Vertebrates
Phasmaviridae, Orthophasmavirus	KPOFV	Kiglualik phantom orthophasmavirus	AJG39258.1	Insects
Phasmaviridae, Orthophasmavirus	WCOPV	Wuchang cockroach orthophasmavirus 1	AKN56913.1	Insects
Fantiviridae, Orthohantavirus	FERV	Ferak orthoflavivirus	AKN56884.1	Insects
Fantiviridae, Orthohantavirus	JONV	Jonchet orthojonavirus	APH07644.1	Vertebrates
Fantiviridae, Orthohantavirus	HOHV	Hantaan orthohantavirus	AI25337.1	Vertebrates
Fantiviridae, Orthohantavirus	KhaOHV	Khabarovsk orthohantavirus	AA573287.2	Arthropod vectors (Eriophyidae), Plant hosts
Finoviridae, Emaravirus	EMARAV	European mountain ash ringspot-associated virus	ADZ54688.1	Arthropod vectors (Eriophyidae), Plant hosts
Finoviridae, Emaravirus	RREV	Rose rosette emaravirus	AIY28466.1	Arthropod vectors (Thripidae), Plant hosts
Tospoviridae, Orthotospovirus	TSWV	Tomato spotted wilt orthotospovirus	ACM189280.1	Arthropod vectors (Thripidae), Plant hosts
Tospoviridae, Orthotospovirus	IYSV	Iris yellow spot orthotospovirus	AGX32061.1	Insects
Tospoviridae, Orthotospovirus	HeHV	Herbert herbevirus	AGX32057.1	Insects
Peribunyaviridae	TaHV	Tai herbevirus	AKX73309.1	Arthropod vectors (ticks), vertebrate hosts
Peribunyaviridae	BUNV	Bunyamwera virus		Arthropod vectors (ticks), vertebrate hosts
Peribunyaviridae	OROV	Oropouche orthobunyavirus	ALB07205.1	Arthropod vectors (ticks), vertebrate hosts
Narnaviruses of trypanosomatids (Fig. 4D)	SAOV1	Soybean-associated ourmiaivirus 1	ALM62238	Plants
Ournia-like viruses	SAOV2	Soybean-associated ourmiaivirus 2	ALM62250	Plants
Ournia-like viruses	SsOLV1	Sclerotinia sclerotiorum ourmia-like virus 1	ALD89138	Fungi
Ournia-like viruses	RsOLV1	Rhizoctonia solani ourmia-like virus 1	ALD89131	Fungi
Ournia-like viruses	BOV	Botryotis ourmiaivirus	CZ26310	Fungi
Ournia-like viruses	SsOLV2	Sclerotinia sclerotiorum ourmia-like virus 2	ALD89139	Fungi
Ourniaivirus	CsVC	Cassava virus C	ACI03053	Plants
Ourniaivirus	EpCV	Epirus cherry virus	ACF16357	Plants
Ourniaivirus	OuMV	Ournia melon virus	ACF16360	Plants
Narnavirus	ScNV-20S	Saccharomyces cerevisiae 20S RNA narnavirus	AAC98925	Fungi
Narnavirus	ScNV-23S	Saccharomyces cerevisiae 23S RNA narnavirus	AAC98708	Fungi
Narnavirus	NarEnv	Narnaviridae environmental sample	ALT35956	Unknown
Narnavirus	PserNV1	Phytomonas serpens narnavirus 1		Trypanosomatidae
Narnavirus	LepseyNLV1	Leptomonas seymouri narna-like virus 1		Trypanosomatidae

Table S2. Cont.

Clade		Short name	Virus name	Accession no.	Host
<i>Narnavirus</i>		PIRV4	<i>Phytophthora infestans RNA virus 4</i>	AEM89291	Oomycetes
<i>Narnavirus</i>	TSA: TdalM		<i>Teleopsis dalmani transcribed RNA</i>	GBBP01132666.1	<i>Trypanosomatidae (Naemimonas sp.)</i>
<i>Mitovirus</i>	CdMV1a	<i>Cryphonectria cubensis mitovirus 1a</i>		AAR01970	Fungi (mitochondria)
<i>Mitovirus</i>	SsMV9	<i>Sclerotinia sclerotiorum mitovirus 9</i>		AHF48625	Fungi (mitochondria)
<i>Mitovirus</i>	BcMV3	<i>Botrytis cinerea mitovirus 3</i>		CEZ26302	Fungi (mitochondria)
<i>Mitovirus</i>	OMV3a	<i>Ophiostoma mitovirus 3a</i>		CAA06228	Fungi (mitochondria)
<i>Mitovirus</i>	SsMV3	<i>Sclerotinia sclerotiorum mitovirus 3</i>		AGC24232	Fungi (mitochondria)
<i>Levivirus</i>	GA	<i>Enterobacteria phage GA</i>		CAA27499	Bacteria
<i>Levivirus</i>	MS2	<i>Enterobacteria phage MS2</i>		P00585	Bacteria

CBPV, Chronic bee paralysis virus.

Table S3. Accession numbers and/or sequences reported in this work

Species	Isolate	Viral name	Accession nos.
<i>Critchidia otongatchiensis</i>	Ecu-08	CotoLBV1	KX373292, KX451144, KX451145, KX683300
<i>Critchidia</i> sp.	G15	CG15LBV1	KX373291
<i>Critchidia</i> sp.	ZM	CZMLBV1	KX373293
<i>Critchidia</i> sp.	C4	CC4LBV1	KY322668
<i>Critchidia abscondita</i>	127AL	CabsLBV1	KX507299, KX507300, KX507301
<i>Critchidia pragensis</i>	MCZ-11	CpraLBV1	KY322669
<i>Leptomonas moramango</i>	MMO-09	LepmorLBV1	KX280012-KX280017
<i>Phytomonas</i> sp.	TCC231	PTCCLBV1	KY322667
<i>Leptomonas pyrrhocoris</i>	F165	LeppyrTLV1	KX373302, KX373303
<i>Leptomonas pyrrhocoris</i>	F19	LeppyrTLV1	MG008317
<i>Leptomonas pyrrhocoris</i>	H10	LeppyrTLV1	KX373300, KX373301
<i>Phytomonas serpens</i>	9T (UCR)	PserNV1	KU882057, KY322666
<i>Phytomonas serpens</i>	30T	PserNV1	—
<i>Leptomonas seymouri</i>	ATCC	LseyNLV1	KU935604, KU935605, KX373304, KX373305
<i>Leptomonas pyrrhocoris</i>	F19	LeppyrOV1	—
<i>Leptomonas pyrrhocoris</i>	H10	LeppyrOV1	KX373294-KX373299
<i>Leptomonas pyrrhocoris</i>	P59	LeppyrTLV1-EVE1	KY364836, KY364842
<i>Leptomonas pyrrhocoris</i>	PP1	LeppyrTLV1-EVE1	KY364837, KY364843
<i>Leptomonas pyrrhocoris</i>	PP2	LeppyrTLV1-EVE1	KY364835, KY364841
<i>Leptomonas pyrrhocoris</i>	LP	LeppyrTLV1-EVE1	KY364839

Table S4. Metatranscriptomic contigs assembled from SRA depositions

Name of contig	Sequence	Identity	SRA accession	Name of virus found in the SRA	Source of the contig
SRX712638_HBLV6_PFR	GATTGCGAAGCTGGAGAAAGATGGGATGAGCTGGCGCTCTCAGTACAGCGG- ACGGAAATGGCACAGCCAGTCAGTGCTCAAGAACCTCGAG CCATGCGAGATAGACCTAAATCTGCCGAAAATTTGGGTTCCGGAAACATT- GGATAACTTGCACAAAGCAAGCTAAATCATGAACCAAGGGTTCCTCG- CCACGGGGTTCGGGCAACCTACGCTCTAGTGTAGCCCTGGCAAATGATA- CATTAAACAAATGCTTCACTGGCAGTAACCCAGAAAGTGTGACTCAATT- CATTCGGTGGAAAAGCGGATTCCGGCTCTTGAACAAACTCCCCAT- CAGCTAGTGTGGCACTGTAAGTGGCATGGCTAGTCAGCGGAGGGGA- GGC ATAGGGTTGATGGCCGTGTAAGTGGACTGCATGGGTTGACGGGGGGAA- GGGCACAGGGCGCAAAATTCCCCTAATGTCAGAAAAAAAGATGAGCAGCCA- AAAGAAAT	Trypanosome parafagellar rod protein 2 Herpetomonas sp. 18S ribosomal RNA	SRX1712638 SRX833697	Leech mix Hubei virus 6 (HBLV6) Wuhan spader virus (WSV)	Whitmania pigra (horse leech) Neoscona, Parasteatoda, Plexippus, Pirata, Araneae spp. Li (2015) (2)
SRX833697_WSV_18S	CCATGCGAGATAGACCTAAATCTGCCGAAAATTTGGGTTCCGGAAACATT- GGATAACTTGCACAAAGCAAGCTAAATCATGAACCAAGGGTTCCTCG- CCACGGGGTTCGGGCAACCTACGCTCTAGTGTAGCCCTGGCAAATGATA- CATTAAACAAATGCTTCACTGGCAGTAACCCAGAAAGTGTGACTCAATT- CATTCGGTGGAAAAGCGGATTCCGGCTCTTGAACAAACTCCCCAT- CAGCTAGTGTGGCACTGTAAGTGGCATGGCTAGTCAGCGGAGGGGA- GGC ATAGGGTTGATGGCCGTGTAAGTGGACTGCATGGGTTGACGGGGGGAA- GGGCACAGGGCGCAAAATTCCCCTAATGTCAGAAAAAAAGATGAGCAGCCA- AAAGAAAT	Trypanosome parafagellar rod protein 2 Herpetomonas sp. 18S ribosomal RNA	SRX1712638 SRX833697	Leech mix Hubei virus 6 (HBLV6) Wuhan spader virus (WSV)	Whitmania pigra (horse leech) Neoscona, Parasteatoda, Plexippus, Pirata, Araneae spp. Li (2015) (2)
SRX711976_HBLV5_18S	CCGGCGCTTTGACGAAACACTGCCCCTATCAGTAGTGTAGTGGCGCTGTAGTGGA- CTGCCATGGGTTGACGGGAGGGGATTAAGGTTGATCTCCGAGGGAG- CCTGAGAAAATAGCTTACCAACTCTACGGAGGCCAGGGGGCAAATTGCCCA- ATGTCAGAAAACAAATGCTTCAATGGCAGTAAACCCAGAGCTTGTACTCAAT- TGACAGTAAAACAAATGCTTCAATGGCAGTAAACCCAGAGCTTGTACTCAAT- TCAATTGGGTTGTTGGCTCTTGTGACTTGTGAAACAACGCCCC- TATGAGCTGTGATGGCGTGTGAGTGGCATGGCGTGGAGGAGCTACAT- GGAGGGCAGAGGGGGCAATTGGCCAATGTCAAACAAACAGATAGGCAG- CGAAAAAGAAATAGAGTTGTTGAGTCCATTGGATTGTCTCATTTCAATAGGGATA- TTAAACCCATCGAAAATCTAGTAAACATTGGAGGAAAGAATGTTGGTCCAGCA- CCCGCGTAAATTCAGCTCAGCTCAGCTCAAAGGCTTAT	Strigomonadinae 18S ribosomal RNA	SRX1711976	Hubei bunya-like virus 5 (HBLV5)	Drosophila, Episyrrhus, Sarcophaga, Muscina, Precticus Shi (2016) (1)
SRX833692_HHFV_18S	CCGGCGCTTTGACGAAACACTGCCCCTATCAGTAGTGTAGTGGCGCTGTAGTGGA- CTGCCATGGGTTGACGGGAGGGGATTAAGGTTGATCTCCGAGGGAG- CCTGAGAAAATAGCTTACCAACTCTACGGAGGCCAGGGGGCAAATTGCCCA- ATGTCAGAAAACAAATGCTTCAATGGCAGTAAACCCAGAGCTTGTACTCAAT- TGACAGTAAAACAAATGCTTCAATGGCAGTAAACCCAGAGCTTGTACTCAAT- TCAATTGGGTTGTTGGCTCTTGTGACTTGTGAAACAACGCCCC- TATGAGCTGTGATGGCGTGTGAGTGGCATGGCGTGGAGGAGCTACAT- GGAGGGCAGAGGGGGCAATTGGCCAATGTCAAACAAACAGATAGGCAG- CGAAAAAGAAATAGAGTTGTTGAGTCCATTGGATTGTCTCATTTCAATAGGGATA- TTAAACCCATCGAAAATCTAGTAAACATTGGAGGAAAGAATGTTGGTCCAGCA- CCCGCGTAAATTCAGCTCAGCTCAGCTCAAAGGCTTAT	Strigomonadinae 18S ribosomal RNA	SRX833692	Huangshi humpbacked fly virus (HHFV)	Psychoda, Velarifictorus, Crocothemis, Phoridae spp., Lampyridae spp., Aphelinus, Hyalopterus, Aulacorthum Li (2015) (2)
SRX2422212_ABV1_colonyA_18S	TGACAGTAAAACAAATGCTTCAATGGCAGTAAACCCAGAGCTTGTACTCAAT- TCAATTGGGTTGTTGGCTCTTGTGACTTGTGAAACAACGCCCC- TATGAGCTGTGATGGCGTGTGAGTGGCATGGCGTGGAGGAGCTACAT- GGAGGGCAGAGGGGGCAATTGGCCAATGTCAAACAAACAGATAGGCAG- CGAAAAAGAAATAGAGTTGTTGAGTCCATTGGATTGTCTCATTTCAATAGGGATA- TTAAACCCATCGAAAATCTAGTAAACATTGGAGGAAAGAATGTTGGTCCAGCA- CCCGCGTAAATTCAGCTCAGCTCAGCTCAAAGGCTTAT	Leishmaninae 18S ribosomal RNA	SRX2422225	Apis bunyavirus 1	RNA-seq of Apis mellifera: South Africa colony 11 Remnant (2017) (3)
SRX833692_HHFV_nuc	GGAGGACCATCTGCAAGTCAATAGTGTGCTTAGAGGTTGAAGAGATTGTTT- TAACCTAGCCATGTTGGCTGAGCTGGGACACAGCTGGCCCTTAGAGA- GAGATGGAAGGTCGGAGGTTGCGAAAGTTAATAGCCATCTGTTGGGTGCGAGG- GGTACCAACCTCGCCAAAGGTGGTAAAGGAGATAAATCTGAAAGGCCCT- CGAATAGCTGATAGATCCGCAAGGTGGTAAAGGAGATAAATCTGAAAGGCCCT- GNNCTGCTCTCTGGACATATTGGCGCTTACCCGGCTTACGGTACTCTGAT- GCAAGAAATTSGCTCCAAGAGSTGCTATCTGCTCAACACCCTCCAGTTCCTGAA- ATTTCGAGGTGCTGACCAAGAGGGTGGCTTAAGGGCTAAATAGCAGCTTCTG- AACTCACAGGCCAAAGTTCAAGAAAGTTGAGTGGCTGGCGATGTGATATGG- GCAGAGCTACCTCCGGTAGCCGAAACTCTGGGTTAGGTACAGG CCAATCTGATAAGATCCAAAGTTCAGCAAGCTGGGATGATGCTGGGTGCAAGGA- CTTGTGTCGATTGGCGTAGCTGCTGTCAGCAACCTGGGCTCTCACTGGCTCA- CATTGGTTCTCTTAACCAAGGTTGCTTATAGCCAGGAGATGGCTACGCTCA- AGTCAGATAATGTGAAACATTTCTCCTGCTAACTAAGTGGCTTACCAAG- GATGCTGAGGTGAGGCAAACAGGGAGTCTGGCTGGCTGCTGAGTGGCTAC- CCAGAAGGTTCAACGCAATCTGAAATAGTGTGATAGCTGGCTGATAGCAGCTCATGGC- AAAGCCTTAAGTCAAGGTTGAGGATGGGAGCTGGCTATGGCTGAGATAATGTTG- AGAGAAGGTTATGAGAGATGGGAGCTGGCTATGGCTGAGATAAGACTTGAAC- GGGAAACGAACTCTGGTAAATGGCTTATAGCTGATAGCTGGCTACGCTCA- CAGTGATCGGGAGAAATACTGGTAACTAGTGTGATAGCTGGCTGATAGCTGGC- GGACATATCGCTGCAAGCTTCTGATGTTAGGCTGAGACTTGAAC- TGAAAAGCAGGAGATTGTTGAGATACTGCTACAGTCTTAAGGAGACAATGGCTC- GGAAAAGGTTGGGTTCAACCTAATTGAGTTCTACAAACCAAGGCAA- GATGCTGAGGTGAGGATAAGTTGAGATATTAGTGGAGATATTAGTGGAGATGATGTT- TGCCTGGGATCTGGGAGATAAGGATGCTTGTAGCAGTGTGCTTAAACCTA- TGGAAATTCAAGTTAAGGCC	LBV nucleocapsid	SRX833692	Huangshi humpbacked fly virus (HHFV)	Psychoda, Velarifictorus, Crocothemis, Phoridae spp., Lampyridae spp., Aphelinus, Hyalopterus, Aulacorthum Li (2015) (2)
SRX833697_WSV_nuc	GGGGGATGGGAAATCAGGCAAACACTGGGATGATGCTGGGTGCAAGGA- CTTGTGTCGATTGGCGTAGCTGCTGTCAGCAACCTGGGCTCTCACTGGCTCA- CATTGGTTCTCTTAACCAAGGTTGCTTATAGCCAGGAGATGGCTACGCTCA- AGTCAGATAATGTGAAACATTTCTCCTGCTAACTAAGTGGCTTACCAAG- GATGCTGAGGTGAGGCAAACAGGGAGTCTGGCTGGCTGCTGAGTGGCTAC- CCAGAAGGTTCAACGCAATCTGAAATAGTGTGATAGCTGGCTGATAGCAGCTCATGGC- AAAGCCTTAAGTCAAGGTTGAGGATGGGAGCTGGCTATGGCTGAGATAATGTTG- AGAGAAGGTTATGAGAGATGGGAGCTGGCTATGGCTGAGATAAGACTTGAAC- GGGAAACGAACTCTGGTAAATGGCTTATAGCTGATAGCTGGCTACGCTCA- CAGTGATCGGGAGAAATACTGGTAACTAGTGTGATAGCTGGCTGATAGCTGGC- GGACATATCGCTGCAAGCTTCTGATGTTAGGCTGAGACTTGAAC- TGAAAAGCAGGAGATTGTTGAGATACTGCTACAGTCTTAAGGAGACAATGGCTC- GGAAAAGGTTGGGTTCAACCTAATTGAGTTCTACAAACCAAGGCAA- GATGCTGAGGTGAGGATAAGTTGAGATATTAGTGGAGATATTAGTGGAGATGATGTT- TGCCTGGGATCTGGGAGATAAGGATGCTTGTAGCAGTGTGCTTAAACCTA- TGGAAATTCAAGTTAAGGCC	LBV nucleocapsid	SRX833697	Wuhan spader virus (WSV)	Spiders Neoscona, Parasteatoda, Plexippus, Pirata, Araneae spp. Li (2015) (2)
SRX711976_HBLV5_nuc_contig6	AATGCTCTGAGCCAGTGGGATAGCAGCTACAGTGTGATGCTGGCTGAGATAATGTTG- AAAGCCTTAAGTCAAGGTTGAGGATGGGAGCTGGCTATGGCTGAGATAAGACTTGAAC- AGAGAAGGTTATGAGAGATGGGAGCTGGCTATGGCTGAGATAAGACTTGAAC- GGGAAACGAACTCTGGTAAATGGCTTATAGCTGATAGCTGGCTACGCTCA- CAGTGATCGGGAGAAATACTGGTAACTAGTGTGATAGCTGGCTGATAGCTGGC- GGACATATCGCTGCAAGCTTCTGATGTTAGGCTGAGACTTGAAC- TGAAAAGCAGGAGATTGTTGAGATACTGCTACAGTCTTAAGGAGACAATGGCTC- GGAAAAGGTTGGGTTCAACCTAATTGAGTTCTACAAACCAAGGCAA- GATGCTGAGGTGAGGATAAGTTGAGATATTAGTGGAGATATTAGTGGAGATGATGTT- TGCCTGGGATCTGGGAGATAAGGATGCTTGTAGCAGTGTGCTTAAACCTA- TGGAAATTCAAGTTAAGGCC	LBV nucleocapsid	SRX1711973	Hubei bunya-like virus 5 (HBLV5)	Drosophila, Episyrrhus, Sarcophaga, Muscina, Precticus Shi (2016) (1)

Table S4. Cont.

Name of contig	Sequence	Identity	SRA accession	Name of virus found in the SRA	Biomaterial included in the sample	Source of the contig
SRX1711976_HBLV5_nuc_contig15	CATTTTGATTTAACGGCGGTGGCACATCCCTACGGCCAATATTAAAG-CTGACCAGCTGGGCTATACCAACAAATTGGAAAAGAATCATCACC-CAAATTGCTGAACGGCAAGAACCGTCCGGACTAGATGCCACTAAACG-AACAAAAATCAGTTCGCTGTAGCCACACGCTTGGTAAGTCATTAATT-TCAAGAACTGAAGTGTTCAGTCACTGGAAAGCTGAAGCAAAACTSCATTGACCCACA-AAGACCCACCAATTCTCAACACGCAATAAGCTCAGGCCTCTGTTGT-CCITGAGGAGCCAGCAACGCAATCTTATAGGCCCTGAGCTGAGCCAG-ATAGAACAAATTTGGCTTCTCCAAATGCTCTGCTTGATAAATCTCATCCT-AGTGAACCATGTTGATAACCCCTCTGATGGATTGCAATCAAGTAGCAGAATCT-CTTGTGCTAGGGAGGACTCATTTCTCTCATAGGGAAAATAGTGTCTT-TCTG	LBV nucleocapsid	SRX1711974	Hebei bunya-like virus 5 (HBLV5)	Diptera mix Hubei	Shi (2016) (1)
SRX1711976_HBLV5_nuc_contig18	GCTTTCTTACTCTTCCTCGCATGCAAGTCGGCTCGAGCTGGCTCTGAGGCTCTGGTA-GAGCTCAGCTATAGGGTGAAGTCCTTACCAAGCTCGAGCTGGCTGTAAT-CCGAGGACTAGAAGGACGGCCGAATACTCTTCTTCTGCTGAGGTGTGG-GGCACCAATCTTATGAAGCTCCGAGAAAAGTGAAGTCAAGAGTACTGGA-TAGAFTGCTTGTGCTTGTGCTTGTGACCTAGTGTCAACAAAGACTCTTAG-GTCACTGGCTGGCTGCTGCTTCCCTAGAACCTGTTGAGTGGTAGTGGTAGC-AATTGCCCCCTTCGGTCAACACTATCCAGGCTCTGAGTTCAATGGCTTGA-GAAAGATGACTGGTAGAGCCAAATCAAAATGCTGGTAGCTGAAAGGGTG-ATGTTGGAAGTTCACGGCATCGTGTATAFTGTTGGGAGAGCAAGTT-TCAGAAGTGGAGATGGAGATGGCCCTGGTGTATTAGCAGTGAATCTACCAATTAGGGAGTGTAGTGTAGTGTAGT	LBV nucleocapsid	SRX1711975	Hebei bunya-like virus 5 (HBLV5)	Diptera mix Hubei	Shi (2016) (1)
SRX1711976_HBLV5_nuc_contig8	CTTCAACCAACTCTGCAAGGGCTGTCCTCCCAAGTCAAGTCGGCAATGGACT-AAAATGGGATCTCCCAGCTGGCAATGGCTTCAAGGGTAAACATCTCACAGTGG-CTTGTGAGCCAATCATCCCTCTCAAGCCAGGTGCTGAGGAAGCGAGCTG-TTGTGAGACTAACGGGGASCGGCCCAACTCTCCCATPATAGACCTCTCGGG-GAAGGCGCCGGAGGTCCCTAAAGACACCCGTACCTGCCAACAGAGCCC-TGGCATAGCTGTGTCAGGCCCCGGGAGCTAGGACCTTACTCGG	LBV nucleocapsid	SRX1711976	Hebei bunya-like virus 5 (HBLV5)	Diptera mix Hubei	Shi (2016) (1)
SRX1712638_HBLV6_nuc	CGGTGGCTGAGAACCTGAGAAATTTATCCGATTAACCTGATGAACTCTGAC-CAACATTGCAATGCTCTGAGAAACTAGGATAAGCATAGAAATGCTACTAGG-TAATGCTGCTAGAATCATAGCTTACACTGTTGGGACTCTGGAACCATTTGG-AAAGCAATGCAATAAGTCTCTTAATAGTCAACATAGTGAACCTCATGCA-CTTCACTACTGCAATATGCTACTAGTGTGCTGAGCAAGCTCATGGCACACAT-AGCAGCAGCAATACCCAGGAGTGTGTATGAGTGTGAAAGGCCAICTCGCAGTC-AAATCTCTGTGTCAGTACGATCCCTAGTGAAGCACAAGGGCTAACAAAAGAA-CGCTGAGAACCTGAGAAATTTATCCGATTAACCTGATGAACTCTGAC-	LBV nucleocapsid	SRX1712638	Hebei bunya-like virus 6 (HBLV6)	Leech mix Hubei	Shi (2016) (1)

Partial sequences of trunansomatid-specific and RV-like nuclear capsid protein genes assembled from viral metatranscriptomes. Because they have not been experimentally verified they cannot be deposited in GenBank

- Shi M, et al. (2016) Redefining the invertebrate RNA virosphere. *Nature* 540:539–543.
 - Shi M, et al. (2015) Unprecedented genomic diversity of negative-sense RNA viruses in arthropods reveals the ancestry of RNA viruses. *Nature* 526:471–475.
 - Liu CX, et al. (2015) A diverse group of novel RNA viruses from arthropods. *Virology* 481:118–127.
 - Shi M, et al. (2014) A diverse group of novel RNA viruses from arthropods. *Virology* 461:e118–e127.

Table S5. Codon usage in LeppyrOV1 and LeppyrTLV1 ORFs

Sequence	T3	C3	A3	G3	GC
LeppyrOV1					
ORF1	0.40	0.30	0.18	0.31	0.51
ORF2	0.35	0.32	0.19	0.33	0.53
ORF3	0.39	0.28	0.20	0.34	0.52
ORF4	0.33	0.34	0.17	0.34	0.55
ORF5	0.39	0.27	0.15	0.33	0.57
ORF6	0.45	0.27	0.16	0.32	0.48
Average	0.38	0.30	0.18	0.33	0.53
LeppyrTLV1					
ORF1	0.34	0.26	0.33	0.32	0.45
ORF2	0.41	0.29	0.28	0.23	0.47
ORF3	0.34	0.24	0.20	0.22	0.47
Average	0.36	0.26	0.27	0.26	0.46
Leppyr genome					
	0.14	0.38	0.1	0.39	0.77

A3, C3, G3, and T3 are frequencies of the corresponding nucleotides in the third position; GC is the overall GC content.

Table S6. Complementary terminal sequences (panhandles) of LBV1s and other Bunyavirales

Virus	5'	3'
LBV1s		
L segment		
CG15LBV1	ACACAA <u>AGAGAA</u>	TGTTCTTGTGT
LepmorLBV1b	ACACAA <u>AGACAA</u>	TATTCTTGTGT
LepmorLBV1a	ACACAA <u>AGATAA</u>	ND
CZMLBV1	ACACAA <u>AGAGAA</u>	TGTTCTTGTGT
CotoLBV1	ACACAA <u>AGACGA</u>	TATTCTTGTGT
PTCCLBV1	ACACAA <u>AGAGAA</u>	TATTCTTGTGT
Consensus	ACACAA <u>AGANAA</u>	TRTTCTTGTGT
M segment		
CabsLBV1-M	ACACAA <u>AGAGAA</u>	TGTTCTTGTGT
CotoLBV1-M	ACTCAA <u>AGACGA</u>	ND
LepmorLBV1a-M	ACACAA <u>AGATAA</u>	TATTCTTGTG-
LepmorLBV1b-M	ACACAA <u>AGACAA</u>	ND
consensus	ACWCAA <u>AGABRA</u>	TRTTCTTGTGT
S segment		
CabsLBV1-S	ACACACGGAAAA	TATTCTTGTGT
CotoLBV1-S	CACAACGACGA	ND
LepmorLBV1a-S	ACACATAGACAA	ND
LepmorLBV1b-S	ACACACAGATAA	ND
Consensus	ACACAHVGRAHRA	TRTTCTTGTGT
Phenuiviridae		
<i>Gouleako virus</i>		
L segment	ACACAA <u>AGACAC</u>	TGGACTTGTGT
M segment	ACACAGTGACCC	GGGACTTGTGT
S segment	ACACAGTGACCT	GGGACTTGTGT
<i>Rift Valley fever virus</i>		
L segment	ACACAA <u>AGGC</u> GC	CGGTCTTGTGT
M segment	ACACAA <u>AGAC</u> GG	CGGTCTTGTGT
S segment	ACACAA <u>AGACCC</u>	GGAGCTTGTGT
<i>Rice grassy stunt virus</i>		
L segment	ACACAA <u>AGTC</u> CT	CAGACTTGTGT
M segment	ACACAA <u>AGTC</u> CT	CAGACTTGTGT
S segment	ACACAA <u>AGTC</u> CT	CAGACTTGTGT
Other Bunyavirales		
<i>Hantaan orthohantavirus</i>		
L segment	TAGTAGTAGACT	AGCATA <u>CTACTA</u>
M segment	TAGTAGTAGACT	AGCATA <u>CTACTA</u>
S segment	TAGTAGTAGACT	AGCATA <u>CTACTA</u>
<i>Crimean-Congo hemorrhagic fever orthonairovirus</i>		
L segment	TCTCAA <u>AGAT</u> AT	ATTCTTTGAGA
M segment	TCTCAA <u>AGAA</u> AT	ATTCTTTGAGA
S segment	TCTCAA <u>AGAA</u> AC	ATTCTTTGAGA

Alignments of 5' and 3' terminal sequences L, M, and S segments of LBV1s and selected other Bunyavirales are shown. Nucleotides involved in bulge formation are underlined. ND, not determined.