

Supporting Information

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SI Methods

Cultivation of Trypanosomatids. Axenic cultures of monoxenous trypanosomatids of the genera *Leptomonas*, *Crithidia*, and *Phytomonas* were obtained from (i) Life Science Research Centre, University of Ostrava, Ostrava, Czech Republic; (ii) Department of Parasitology, Charles University, Prague; (iii) Institute of Parasitology, Budweis, Czech Republic; (iv) Fundação Oswaldo Cruz, Rio de Janeiro; and (v) the Zoological Institute of the Russian Academy of Sciences, St. Petersburg, Russia. Trypanosomatids were cultivated in the Brain Heart infusion medium supplemented with 10 $\mu\text{g}/\text{mL}$ of hemin, 500 units/mL of penicillin, and 0.5 mg/mL of streptomycin as described previously (105) and grown to the late logarithmic stage (10^7 – 10^8 cells/mL).

Phylogenetic Analyses of Viruses. RDRP sequences of the viruses characterized in this work were aligned with the related sequences from GenBank in the online version of MAFFT 7 using the E-INS-i method (106). Ambiguous parts of the alignments were removed with the use of TrimAl v. 1.3 (107). Positions with more than 50% gaps were filtered out by Gap Strip/Squeeze V. 2.1.0 (<https://www.hiv.lanl.gov/content/sequence/GAPSTREEZE/gap.html>). The resulting alignments had the following lengths: 412 aa (the *Narnaviridae* dataset), 273 aa (the *Tombus-/Nodaviridae* dataset), and 1,415 aa (the *Bunyaviridae* dataset).

Maximum likelihood phylogenetic inference was performed in IQ-TREE v. 1.4.2 (108) with automatic selection of the best-fit amino acid substitution and site heterogeneity models (four gamma categories). The best-fitted model parameters defined by Bayesian Information Criterion were LG + F + I + G for the *Narnaviridae* and *Bunyaviridae* datasets and Blosum62 + I + G4 for the *Tombus-/Nodaviridae* dataset. Gaps were treated as missing data. Edge support was estimated with a bootstrap test (1,000 standard replicates). Bayesian inference was accomplished in MrBayes 3.2.6 (109) with the analyses ran for 1 million generations (given the observed fast convergence) and trees sampled every 100 generations. The mixed amino acid substitution model was used (resulting in 1.0 posterior probability of Blosum62 for all three datasets) with the heterogeneity over sites estimated using the G + I model. Amino acid frequencies were fixed to empirical values in the *Narnaviridae* and *Bunyaviridae* datasets and were estimated from the data matrix in the *Tombus-/Nodaviridae* dataset in accordance with the best-fit model defined by IQ-TREE v. 1.4.2 for each dataset. Other parameters were left in their default states.

Phylogeny of Trypanosomatid Hosts. The core alignments of 18S rRNA and gGAPDH genes were taken from previous work (76). Ambiguously aligned positions of 18S rRNA gene alignment were removed using Gblocks 0.91b as described previously (110). The concatenated alignments were subjected to maximum likelihood analysis in IQ-TREE (108) with partitioning by gene as well as by codon position for the gGAPDH gene. The best partitioned model of nucleotide substitutions (K3Pu + G4, TIM3 + I + G4, and TPM2u + G4 for the first, second, and third codon positions of gGAPDH gene, respectively and TNe + I + G4 for 18S rRNA gene) was selected with the use of ModelFinder (111). The statistical support of branches was estimated using 1,000 replicates of the standard bootstrap method.

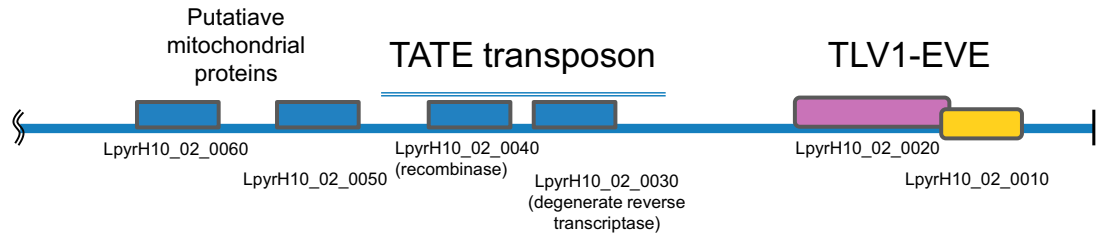
Genomic and Transcriptomic Analyses. To find trypanosomatid signature sequences in the Sequence Reads Archive (SRA, <https://www.ncbi.nlm.nih.gov/sra>) and transcriptome shotgun assemblies (TSAs), TBLASTN and BLASTN searches were performed with amino acid (PFR1) and nucleotide (18S rRNA) sequences, respectively. In case of SRA-blast, the retrieved reads were assembled into contigs using the CAP3 sequence assembly program with the following parameters: minimal overlap length, 20 bp; minimum identity, 100% (112). Obtained contigs were extended by successive rounds of BLASTN searches against the original SRA. Final full-length contigs as well as hits from TSAs were subjected to blast search (megablast for nucleotide sequences and blastp for translated protein-coding sequences) against the nonredundant nucleotide collection of the NCBI database for identification. A similar approach was used to search of reads corresponding to nucleocapsid proteins in SRAs, which contained RDRPs closely related to LBVs. Obtained amino acid sequences were aligned with newly identified nucleocapsid proteins of viruses characterized in this work using BLAST pairwise alignment to confirm their identities.

Codon Usage Analysis in *Leptomonas pyrrocoris* Viruses. Codon frequencies in the protein-coding regions of the six viral genes were analyzed using the CODONW program (codonw.sourceforge.net).

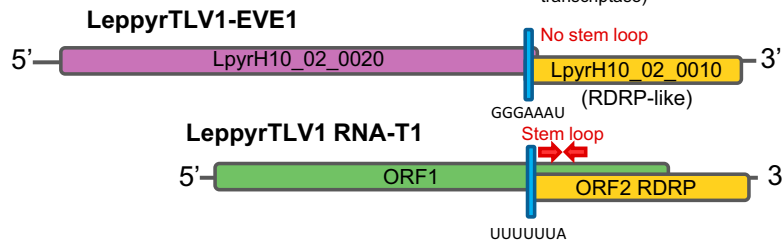
Negative-Stain Transmission Electron Microscopy. In brief, gradient-purified virus samples were applied to a carbon-coated copper grid, stained with molybdenum acetate, and examined under a Philips 201C transmission electron microscope. The morphologies and sizes of the virus particles were analyzed as described previously (113).

A

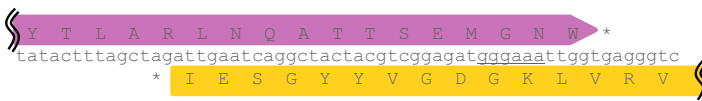
LpyrH10_02 scaffold NW_015438358.1



B



C

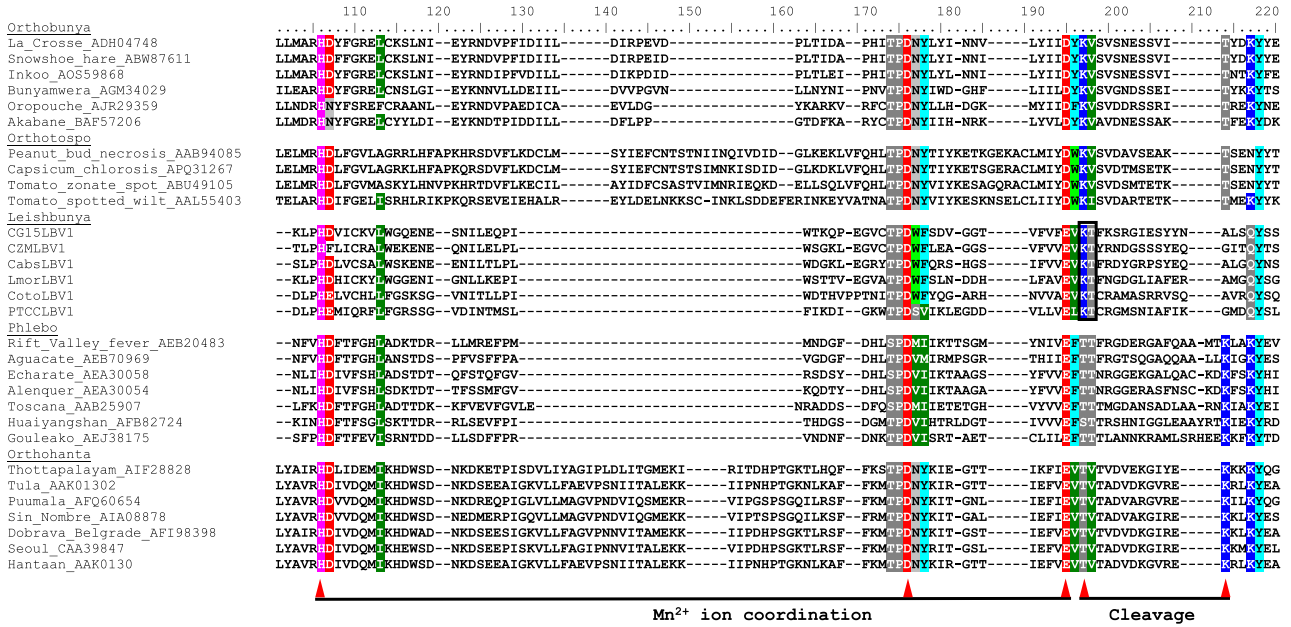


D

MOTIFS	F ₁	F ₂	A
<u>Picorna</u>			
1RDR_A_Poliiovirus	(155) VTYV KDE LRS (164) (169) EQGKS RLI EASSLND (183) (228) -- KLFAFDYTG YDASLSP (243)		
P03313.4_coxsackie_virus_B3	(156) VTYV KDE LRS (165) (170) ARGKS RLI EASSLND (184) (230) -- HLIAFDYSG YDASLSP (244)		
AFK65743.1_Rhinovirus_C	(139) ITYL KDE LRS (148) (153) KVGKT RAI EASSLND (167) (213) -- NLLVFDYTN YDGS LNP (227)		
P03305.1_foot-and-mouth_diseas	(160) QTFL KDE IRP (169) (174) RAGKT RI VDVLPVEH (188) (233) YRNVWDV DYSA FDANHCS (250)		
<u>Flavi</u>			
P06935.2_west_nile_virus	(455) NMMG KRE KKP (464) (469) KAKGS RAI WFMWLG (483) (529) GGVYAD D TAG WD TRITK (546)		
P17763.2_dengue_virus	(452) NMMG KRE KKL (461) (466) KAKGS RAI WYMWLG (480) (526) GGNMYAD D TAG WD TRITE (543)		
P19711.2_bovine_viral_diarrhea	(259) TAlP KNE KRD (268) (280) VEKRP RV IQYPEAKT (294) (338) EPVAVSF D TKA WD TQVTS (355)		
<u>Calici</u>			
Q83883.1_Norwalk_virus	(143) TAAL KDE LVK (152) (158) QKVK RLL WGADLGT (172) (216) YKNHFD A DY TAW DSTQNR (233)		
P27410.1_Rabbit_hemorrhagic_di	(142) ACGL KDE LRP (151) (156) KEGK RLL WGCDVGV (170) (216) ASDFLCL DY SK WD STMSP (233)		
Q6XDK8.1_Sapporo_virus	(315) KLAL KDE LRP (324) (329) AQGK RLL WGCDAGA (343) (387) GGVLYCL DY SK WD STQHP (404)		
<u>LeppyrTLV1-EVE1</u>	(158) - SFI KDE GYD (166) (167) EYKMP RAI NSYSDST (181) (221) DSK VLYT D FSH FE SHHRG (238)		
<u>LeppyrTLV1</u>	(157) DSFI KDE SYD (166) (167) ELKAP RSI MSYPDDV (181) (221) GKVG ST D FS S Y E CHHSG (238)		
MOTIFS	B	C	D
<u>Picorna</u>			
1RDR_A_Poliiovirus	(286) MP SG CSG T SIF NS MINNLI (305) (322) K MIAY GDD VIASYP (335) (348) K DYGLTM--- T (355)		
P03313.4_coxsackie_virus_B3	(288) MP SG CSG T SIF NS MINNIII (307) (324) R MIAY GDD VIASYP (337) (350) K DYGLTM--- T (357)		
AFK65743.1_Rhinovirus_C	(268) MP SG ISG T SIF NT IINNIII (287) (304) K IVAY GDD VIASYP (317) (330) V KYGLTI--- T (337)		
P03305.1_foot-and-mouth_diseas	(296) MP SG CSA T SII NT ILNNIYV (315) (332) T MSI Y GDD IVVASD (345) (358) K SLGQTI--- T (365)		
<u>Flavi</u>			
P06935.2_west_nile_virus	(602) RG SG QVV T YAL NT FTNLAVQ (621) (662) R MAVS GDD CVVKP- (674) (687) N AMSKVRKDIQ (697)		
P17763.2_dengue_virus	(598) RG SG QVG T YGL NT FTNMEAQ (617) (656) R MAIS GDD CVVKP- (668) (681) N DMGKVRKDIP (691)		
P19711.2_bovine_viral_diarrhea	(403) RG SG QPD T SAG NS MLNVLTM (422) (442) R IHV C GDD GFLLITE (455) (471) H EAGK PQK -IT (480)		
<u>Calici</u>			
Q83883.1_Norwalk_virus	(279) LP SG FP C T S QV NS INHWIIT (298) (318) Y FSFY GDD EIVSTD (331) (344) K EYGL KP --- T (351)		
P27410.1_Rabbit_hemorrhagic_di	(279) LP SG MP F T S V NS ICHWLLW (298) (321) P FYTY GDD GVYAMT (334) (349) R DYGL S P--- T (356)		
Q6XDK8.1_Sapporo_virus	(450) LP SG MP F T S V NS LNHMTYF (469) (493) T VHTY GDD CLYSVC (506) (521) T SFGL KP --- T (528)		
<u>LeppyrTLV1-EVE1</u>	(284) LM SG AL WT S FQ NS FLNFFVM (303) (323) K MFIE GDD GIMKHF (336) (344) K RLGL CL K L -IN (353)		
<u>LeppyrTLV1</u>	(288) LM SG AL WT S FQ NS FLNMMI (307) (325) D TFIE GDD GIFRAF (338) (346) A RLG L K L K L -IS (355)		

Fig. S1. LeppyrTLV1 EVE1. (A) The endogenous viral element is located at the subtelomeric region of the *Leptomonas pyrrochoris* chromosome (scaffold NW_015438358.1). The EVE1 element is preceded by the reverse transcriptase-coding TATE DNA transposon indicating the possible mechanism for the LeppyrTLV1 endogenization. (B) Comparison of the ORF organization of RDRP-coding RNA-T1 of 1 LeppyrTLV1 and its endogenous viral element. (C) The overlap region between the ORFs of EVE1 contains a putative slippery sequence (underlined) capable of driving a -1 ribosomal frameshift, similarly to LeppyrTLV1. (D) Multiple alignments of RDRPs of LeppyrTLV1 and LeppyrTLV1-EVE1 with RNA polymerases of *Picorna*-, *Flavi*-, and *Caliciviridae*. Identical residues are indicated in red; similar residues are in blue. Each block shows amino acid motifs typically found in viral RNA polymerases (59).

A



B

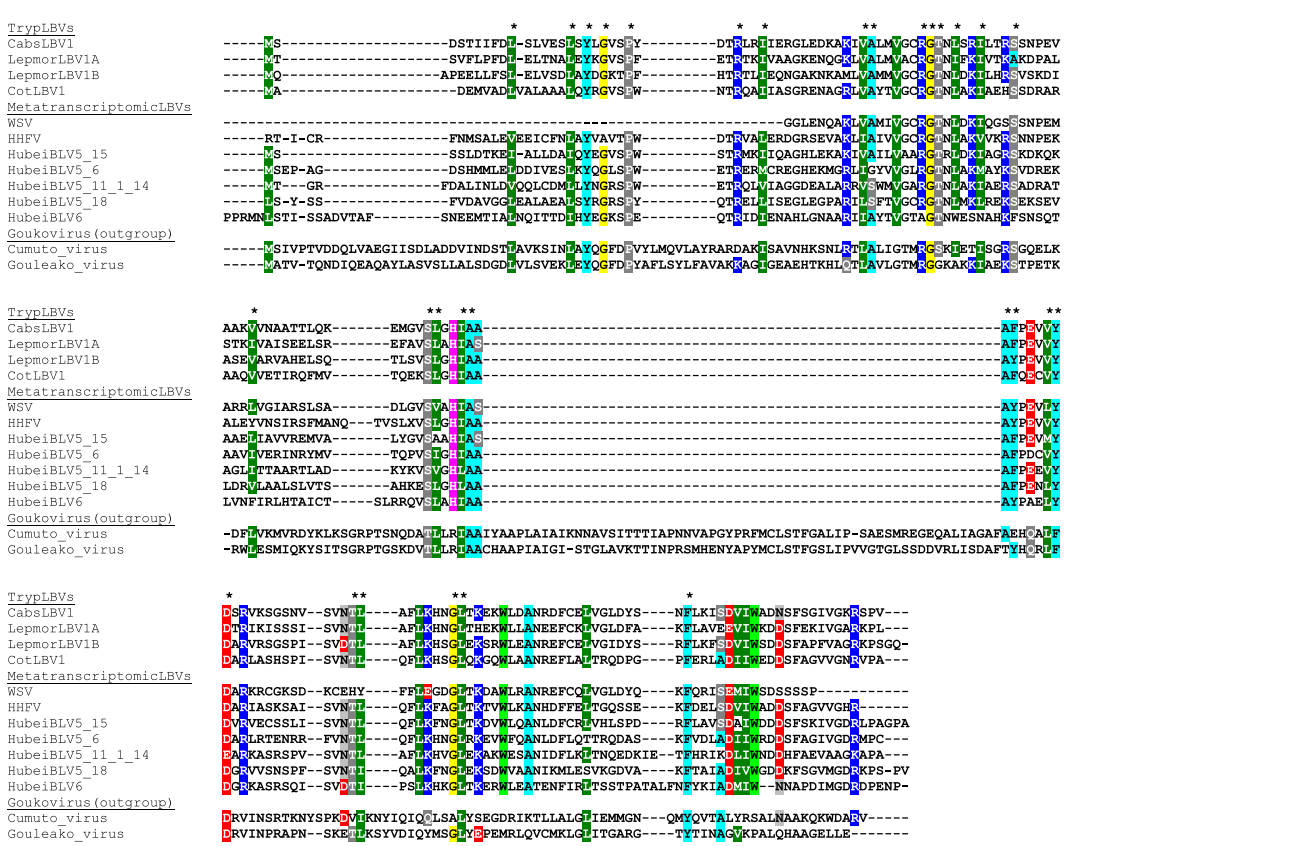


Fig. S2. Leishbunyavirus protein domains. (A) Amino acid alignment of the N-terminal endonuclease domain of the L protein of leishbunyaviruses and other bunyaviruses. Functionally important residues are marked with arrowheads. Residues implicated in nucleolytic cleavage in leishbunyaviruses are boxed. (B) Amino acid alignment of nucleocapsid proteins of leishbunyaviruses reported in ref. 34 and this work (TrypLBVs) and assembled from metatranscriptomes (metatranscriptomic BVs). *Cumuto* and *Gouleako* viruses are shown for comparison. Indels are shown as dashes. Positions with more than 82% similarity are shaded; those with 90% similarity are starred.

Table S4. Metatranscriptomic contigs assembled from SRA depositions

Name of contig	Sequence	Identity	SRA accession	Name of virus found in the SRA	SRA name	Biomaterial included in the sample	Ref. to the SRA
SRX1712638_HBLV6_PFR	GATTGGAAAGCTGGAGAAAGATTGAGGATGAGCTGGCGGCTCAGCTAGACGCGG- ACGGAATGGCAGACAGCGCCAGTGAATGTGCTCAAGAACCTCGAG CCATGGCAGATCAGGTAATCTGCGCAAAATTTGGGGTTTCGCAACATTT- GGATAAATTGGCGAAACCCAAAGTAAATACATGAACCAACCAAGCGTTCTCCG- CCAGGGGCTGGGCAACCGTACGTCTAGTGAACGCTTGGAA TGAATGA- CAITAAAACCAATCCCTTACCTGGCAGTAAACACCCAGAAAGTGTGACTCAAT- CAITCCGTGGAAAGCCGGATTTCCGGGCTTTTGGAGAACAACTGCCCTAT- CAGCTAGTGTAGTGTAGTGTAGTGTAGTGTAGTGTAGTGTAGTGTAGTGTAG- TTAGGTTTCGATTCCGGAGAGGAGGAGGAGGAGGAGGAGGAGGAGGAGGAGG- GGG	<i>Trypanosoma parafagellar</i> rod protein 2 <i>Herpetomonas</i> sp. 185 ribosomal RNA	SRX1712638	Hubei bunya-like virus 6 (HBLV6)	Leech mix Hubei	<i>Whitmania pigra</i> (horse leech)	Shi (2016) (1)
SRX833697_WSV_185	CCATGGCAGATCAGGTAATCTGCGCAAAATTTGGGGTTTCGCAACATTT- GGATAAATTGGCGAAACCCAAAGTAAATACATGAACCAACCAAGCGTTCTCCG- CCAGGGGCTGGGCAACCGTACGTCTAGTGAACGCTTGGAA TGAATGA- CAITAAAACCAATCCCTTACCTGGCAGTAAACACCCAGAAAGTGTGACTCAAT- CAITCCGTGGAAAGCCGGATTTCCGGGCTTTTGGAGAACAACTGCCCTAT- CAGCTAGTGTAGTGTAGTGTAGTGTAGTGTAGTGTAGTGTAGTGTAGTGTAG- TTAGGTTTCGATTCCGGAGAGGAGGAGGAGGAGGAGGAGGAGGAGGAGGAGG- GGG	<i>Trypanosoma parafagellar</i> rod protein 2 <i>Herpetomonas</i> sp. 185 ribosomal RNA	SRX833697	<i>Wuhan spader</i> virus (WSV)	Spiders	<i>Neoscona, Parasteatoda,</i> <i>Plexippus, Pirata, Araneae</i> spp.	Li (2015) (2)
SRX1711976_HBLV5_185	ATCAGCTCGTGTGGCGTGTAGTGTAGTGTAGTGTAGTGTAGTGTAGTGTAG- TTAGGTTTCGATTCCGGAGAGGAGGAGGAGGAGGAGGAGGAGGAGGAGGAG- GGG	Strigomonadinae 185 ribosomal RNA	SRX1711976	Hubei bunya-like virus 5 (HBLV5)	Diptera mix Hubei	<i>Drosophila, Episyphus, Sarcophaga,</i> <i>Muscina, Plecticus</i>	Shi (2016) (1)
SRX833692_HHFV_185	CCGGCTCTTTTTCAGCAACAACTGCCCTATCAGCTAGTGTAGTGTAGTGTAG- CTGCCATGGCTTTCAGCGGAGCGGGGATTAAGGTTCCGATTCGGGAGGGAG- CCTGAGAAATAGCTACACTTCTACGGAGGCGCAGCAGCGGGCGAAA TTTGCCA- ATGTCAAAAGAAAACGATGAGGCAAGCAAGAAAATAGGTTTGT	Strigomonadinae 185 ribosomal RNA	SRX833692	<i>Huangshi humpbacked</i> fly virus (HHFV)	Insects mix 4 (insect in the mountain)	<i>Psychoda, Velarifictorus, Crocothemis,</i> Phoridae spp., Lampyridae spp., <i>Aphelinus, Hyalopterus, Aulacorthum</i>	Li (2015) (2)
SRX2422212_ABV1_colonyA_185	TGACAGTAAACCAATGCTTCTACTGGCAGTAAACCCAGACGTTGACTCAAT- TCATTCGTCGAAAGCGGCTTTTCGCGGCTTTTTCAGCAACAACTGCC- TATCAGCTGTGTGGCGTGTAGTGTAGTGTAGTGTAGTGTAGTGTAGTGTAG- GGATTAGGTTTCGATTCCGGAGAGGAGGAGGAGGAGGAGGAGGAGGAGGAG- GGAGGCGAGCGCGCAAAATGCCCAATGTCAAAACCAAAACCAACGATGAGGCGAG- CGAAAAGAAATAGAGTTGTAGTGTAGTGTAGTGTAGTGTAGTGTAGTGTAG- TTTAAAACCAATGCAAAATCTAGTAAACAAATGGAGCACAAAGTCTGTGCCAGCA- CCCGGGTAAATTCAGCTCCAAAAGCGTATAT	Leishmaniinae 185 ribosomal RNA	SRX2422225	<i>Apis bunyavirus 1</i>	RNA-seq of <i>Apis mellifera</i> South Africa colony 11	<i>Apis mellifera</i>	Remnant (2017) (3)
SRX833692_HHFV_nuc	GGAGGAGTCTCAGATCAATATGCTGCCCTTAGAGTTGAAGATTGTTT- TAACCTAGCTTATGCTGTGCGCTTGGCACACACGCTGTGGCTTTAGAGA- GAGATGGAAGGTCGAGGTTCCGAAAGTTAAGCACTGTTGGGCTGCAGG- GGTACCAACCTGCGCAAGCTGTTAAAAGAGTAAATATCTGAAAAGGCCCT- CGAATAGTGAATAGCATCCGCTCTTTCAATGGCCAAACAGACTGTCTCCCTGG- GNNCTGCTCCTTGGACATATTCGCGCGCTTACCGGAGGATGCTACGAT- GCAAGAAATCGTCCAAAGAGTGTCTCTCAACACCCCTCCAGTCTCTGAA- AATTCAGAGGCTCAGCAAGGTTGCTGTGAAGGTTAATCAGGACTTCTTCG- AACTCAGAGGCTCAGCAAGGTTGAGAAAGTTGATGAGCTGCTCCGATGTGATATGG- GCAGACATAGCTTCGCTGGGCTGGTAGTCCAGG	LBV nucleocapsid	SRX833692	<i>Huangshi humpbacked</i> fly virus (HHFV)	Insects mix 4 (insect in the mountain)	<i>Psychoda, Velarifictorus, Crocothemis,</i> Phoridae spp., Lampyridae spp., <i>Aphelinus, Hyalopterus, Aulacorthum</i>	Li (2015) (2)
SRX833697_WSV_nuc	GGGGATTGGAAATCAGGCAAACTGGTTGCGATGATCGTGGGTCAGGGGAA- CCAAATCTTGAATCAAGGTTCTTCGAGCAATCCGGAGATGGCAGCTCGC- CTTGTGGCATGGCGTGGCTGTCCAGCAGACCTGGGCGTCTCAGTGGCTCA- CAITGCTTCTGTTATCCAGAGGTTTATGATGATCCAGGAAAGGTTCCGGCA- AGTCAGATAGTGAACACTATTTCTTCTTGAAGGTCAGGCTCACCAG- GATGCTTGGCTGAGGCAACAGGGAGTTCGCCAGCTGGTGGGCTTGACTA- CCAGAAATTCAGCCAAATCTGAAATGATCTGGTGTGATGAGCTCATCGCC	LBV nucleocapsid	SRX833697	<i>Wuhan spader virus</i> (WSV)	Spiders	<i>Neoscona, Parasteatoda, Plexippus,</i> <i>Pirata, Araneae</i> spp.	Li (2015) (2)
SRX1711976_HBLV5_nuc_contig6	ATGCTGAGCCAGCTGGAGATAGCCATGATGCTTGGCTTGCAGGATAGTTG- AAAGCCTTAAAGTATCAGGGTTGATCCATGGGAAACGAGAGAGGATGTGC- AGAGAAAGTCAAGAGAGAGAGGAGGCTTATGGCTATGTAGTGGATTGAG- GGAAACGAACTTCTAAATGCGTATTAAGAGTGTGGATCGGAGAGAGGCTG- CAGTATCGTGGAGAGAAATCAATAGGATATGTTAATCACTCAGCCTGTATCTAAT- TGAAAACAGAAATTTGTGAATACGCTACAGTTCTTAAAGCAATGGTCTGC- GGAAAGAGGTTTGGTTCAAGCTAATTTAGATTTCTTACAAAACCCAGAGGAA- GATCTAGCAAGTTTGGATCTGGCAGATATTTGGAGAGATGATGATTT- TGCTGGGATTTGGGAGATAGGATGCCTTGTAGTGCAGTTGTTAGAAACCTA- TGAATTCAGTTATGGCG	LBV nucleocapsid	SRX1711973	Hubei bunya-like virus 5 (HBLV5)	Diptera mix Hubei	<i>Drosophila, Episyphus, Sarcophaga,</i> <i>Muscina, Plecticus</i>	Shi (2016) (1)

Table S5. Codon usage in LeppyrOV1 and LeppyrTLV1 ORFs

Sequence	T3	C3	A3	G3	GC
LeppyrOV1					
ORF1	0.40	0.30	0.18	0.31	0.51
ORF2	0.35	0.32	0.19	0.33	0.53
ORF3	0.39	0.28	0.20	0.34	0.52
ORF4	0.33	0.34	0.17	0.34	0.55
ORF5	0.39	0.27	0.15	0.33	0.57
ORF6	0.45	0.27	0.16	0.32	0.48
Average	0.38	0.30	0.18	0.33	0.53
LeppyrTLV1					
ORF1	0.34	0.26	0.33	0.32	0.45
ORF2	0.41	0.29	0.28	0.23	0.47
ORF3	0.34	0.24	0.20	0.22	0.47
Average	0.36	0.26	0.27	0.26	0.46
Leppyr genome					
	0.14	0.38	0.1	0.39	0.77

A3, C3, G3, and T3 are frequencies of the corresponding nucleotides in the third position; GC is the overall GC content.

Table S6. Complementary terminal sequences (panhandles) of LBV1s and other Bunyavirales

Virus	5'	3'
LBV1s		
L segment		
CG15LBV1	ACACAAAGAGAA	TGTTCTTTGTGT
LepmorLBV1b	ACACAAAGACAA	TATTCTTTGTGT
LepmorLBV1a	ACACAAAGATAA	ND
CZMLBV1	ACACAAAGAGAA	TGTTCTTTGTGT
CotoLBV1	ACACAAAGACGA	TATTCTTTGTGT
PTCCLBV1	ACACAAAGAAGA	TATTCTTTGTGT
Consensus	ACACAAAGANAA	TRTTCTTTGTGT
M segment		
CabsLBV1-M	ACACAAAGAGAA	TGTTCTTTGTGT
CotoLBV1-M	ACTCAAAGACGA	ND
LepmorLBV1a-M	ACACAAAGATAA	TATTCTTTGTG-
LepmorLBV1b-M	ACACAAAGACAA	ND
consensus	ACWCAAAGABRA	TRTTCTTTGTGT
S segment		
CabsLBV1-S	ACACACGGAAAA	TATTCTTTGTGT
CotoLBV1-S	CACAACGACGA	ND
LepmorLBV1a-S	ACACATAGACAA	ND
LepmorLBV1b-S	ACACACAGATAA	ND
Consensus	ACACAHVGAHRA	TRTTCTTTGTGT
Phenuiviridae		
<i>Gouleako virus</i>		
L segment	ACACAAAGACAC	TGGACTTTGTGT
M segment	ACACAGTGACCC	GGGACTTTGTGT
S segment	ACACAGTGACCT	GGGACTTTGTGT
<i>Rift Valley fever virus</i>		
L segment	ACACAAAGGCGC	CGGTCTTTGTGT
M segment	ACACAAAGACGG	CGGTCTTTGTGT
S segment	ACACAAAGACCC	GGAGCTTTGTGT
<i>Rice grassy stunt virus</i>		
L segment	ACACAAAGTCCT	CAGACTTTGTGT
M segment	ACACAAAGTCCT	CAGACTTTGTGT
S segment	ACACAAAGTCCT	CAGACTTTGTGT
Other Bunyavirales		
<i>Hantaan orthohantavirus</i>		
L segment	TAGTAGTAGACT	AGCATACTACTA
M segment	TAGTAGTAGACT	AGCATACTACTA
S segment	TAGTAGTAGACT	AGCATACTACTA
<i>Crimean-Congo hemorrhagic fever orthonairovirus</i>		
L segment	TCTCAAAGATAT	ATTTCTTTGAGA
M segment	TCTCAAAGAAAT	ATTTCTTTGAGA
S segment	TCTCAAAGAAAC	ATTTCTTTGAGA

Alignments of 5' and 3' terminal sequences L, M, and S segments of LBV1s and selected other Bunyavirales are shown. Nucleotides involved in bulge formation are underlined. ND, not determined.