Genomic DNA Methylation Signatures Enable Concurrent Diagnosis and Clinical Genetic Variant Classification in Neurodevelopmental Syndromes

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Pediatric developmental syndromes present with systemic, complex, and often overlapping clinical features that are not infrequently a consequence of Mendelian inheritance of mutations in genes involved in DNA methylation, establishment of histone modifications, and chromatin remodeling (the "epigenetic machinery"). The mechanistic cross-talk between histone modification and DNA methylation suggests that these syndromes might be expected to display specific DNA methylation signatures that are a reflection of those primary errors associated with chromatin dysregulation. Given the interrelated functions of these chromatin regulatory proteins, we sought to identify DNA methylation epi-signatures that could provide syndrome-specific biomarkers to complement standard clinical diagnostics. In the present study, we examined peripheral blood samples from a large cohort of individuals encompassing 14 Mendelian disorders displaying mutations in the genes encoding proteins of the epigenetic machinery. We demonstrated that specific but partially overlapping DNA methylation signatures are associated with many of these conditions. The degree of overlap among these epi-signatures is minimal, further suggesting that, consistent with the initial event, the downstream changes are unique to every syndrome. In addition, by combining these epi-signatures, we have demonstrated that a machine learning tool can be built to concurrently screen for multiple syndromes with high sensitivity and specificity, and we highlight the utility of this tool in solving ambiguous case subjects presenting with variants of unknown significance, along with its ability to generate accurate predictions for subjects presenting with the overlapping clinical and molecular features associated with the disruption of the epigenetic machinery.

Introduction

Genes encoding the epigenetic protein machinery that read, write, and erase post-translational signals on DNA and histones and remodel chromatin are implicated in a wide range of constitutional neurodevelopmental disorders.^{1–3} The pathogenesis of such disorders is likely caused by the downstream events orchestrated by the primary functional defect in these proteins of the so-called epigenetic machinery.^{1–3} Furthermore, specific mutations in these readers, writers, erasers, and chromatin remodelers are linked to the variability in clinical phenotype seen in their associated disorders. It is well established that histone modifications overlap and interact with genomic DNA methylation to affect chromatin remodeling,⁴ and thus, that mutations in the genes that are involved in histone modifications are expected to have an impact within the DNA methylome. Supporting this concept, we have previously reported DNA methylation epigenetic (epi-) signatures in the peripheral blood of subjects carrying mutations in genes involved in chromatin regulation, including the mutations in SRCAP (MIM: 611421) causing Floating-Harbor syndrome (MIM: 136140),⁵ *DNMT1* (MIM: 126375) resulting in adult-onset autosomal-dominant cerebellar ataxia, deafness, and narcolepsy (ADCA-DN [MIM: 604121]),⁶ and *ATRX* (MIM: 300032), which is responsible for the alpha thalassemia/mental retardation X-linked (ATRX) syndrome (MIM: 300448).⁷ Epi-signatures in subjects with Sotos (MIM: 117550), CHARGE (MIM: 214800), and Kabuki (MIM: 147920) syndromes have also been reported.^{8–10}

While the number of developmental and cancer-related conditions for which a DNA methylation epi-signature has been reported is increasing, the extent of overlap or distinction of these epi-signatures is not clear. Clinical overlap is a common finding in the diseases that result from such defects, and it is postulated that mechanistic overlap could be a basis for such phenotypic similarity. This is further acknowledged by noting that all of the proteins associated with these conditions regulate the epigenome through complex interactions with each other. Hence, the question has been raised as to how accurately one can use these epi-signatures as a tool in the molecular diagnosis of these conditions.^{9,10} This is particularly

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			Discovery-T	raining Coho	rt	Control Col	nort		Duchas		
Syndrome	Total	Testing Cohort	No. of Individuals	Percentage Female	Mean Age ± SD	No. of Individuals	Percentage Female	Mean Age ± SD	Probes Passing QC	Probes Found	DMRs Found
Rett	17	4	13	92%	7.6 ± 11	52	92%	$7.4~\pm~10$	448,775	N/A	no
Saethre- Chotzen	25	6	19	63%	10.6 ± 11	76	63%	$\begin{array}{c} 10.11 \ \pm \\ 10 \end{array}$	442,079	N/A	no
Weaver	7	2	5	40%	age unknown	20	40%	-	455,427	N/A	no
Coffin Siris	9	3	6	50%	5.9 ± 6	24	50%	5.8 ± 5	453,321	N/A	no
Coffin Lowry	11	3	8	12%	9.8 ± 6	32	12%	9.7 ± 6	453,285	N/A	no
ATRX	19	4	15	0%	11.4 ± 7	60	0%	11.3 ± 7	450,748	1,112	41
Floating- Harbor	17	4	13	76%	12.7	52	76%	12.2 ± 11	453,189	1,078	54
Sotos	38	10	28	57%	$8.8~\pm~4$	112	57%	8.9 ± 4	448,131	6,858	1,372
ADCA-DN	5	0	5	40%	age unknown	20	40%	_	330,788	3,562	52
Claes-Jensen	10	2	8	0%	22.5 ± 14.2	32	0%	$21.2~\pm~13$	454,978	698	14
Kabuki	44	11	33	57%	9.5 ± 6	132	57%	9.7 ± 7	401,051	919	31
CHARGE	79	40	39	41%	5.8 ± 6	156	48%	5.9 ± 6	448,876	1,320	18
GTPTS	3	0	3	33%	$2.1~\pm~5.8$	20	33%	$2.2~\pm~5.1$	454,489	707	6
SBBYSS	1	0	1	100%	6.3 ± 0	20	100%	6.5 ± 0.8	456,134	864	6

important in the context of utilizing these epi-signatures as functional evidence to classify variants of uncertain clinical significance.

In the present study, we try to address these questions by concurrently examining 14 Mendelian conditions that result from direct or indirect disruptions of the proteins involved in the regulation of the epigenome. These conditions (see Table 1) include Rett syndrome (MIM: 312750) (methyl-CpG-binding protein 2; MeCP2 [MIM: 300005]), ADCA-DN (DNA methyltransferase 1; DNMT1), Kabuki syndrome (lysine-specific methyltransferase 2D; KMT2D [MIM: 147920]), ATRX syndrome (ATRX), Sotos syndrome (nuclear receptor binding SET domain protein 1; NSD1 [MIM: 117550]), Floating-Harbor syndrome (Snf2 related CREBBP activator protein; SRCAP), Weaver syndrome (MIM: 277590) (enhancer of zeste 2 polycomb repressive complex 2 subunit; EZH2 [MIM: 601573]), CHARGE syndrome (chromodomain helicase DNA binding protein 7; CHD7 [MIM: 608892]), Claes-Jensen syndrome (MIM: 300534) (lysine-specific demethylase 5C; KDM5C [MIM: 314690]), Genitopatellar syndrome (GTPTS [MIM: 606170]), and Say-Barber-Biesecker-Young-Simpson syndrome (SBBYSS [MIM: 603736]), both caused by mutations in lysine acetyltransferase 6B (KAT6B [MIM: 605880]), and Coffin-Siris syndrome (MIM: 135900) (SWI/SNF related, matrix associated, actin dependent regulator of chromatin, subfamily B1 SMARCB1 [MIM: 601607], and AT-rich interaction domain 1B; AR1D1B [MIM: 614556]). These genes are directly involved in epigenomic regulation of the chromatin. We have also included two other conditions that result from mutations in genes that interact with the components of the epigenomic machinery. Saethre-Chotzen syndrome (MIM: 101400) is caused by mutations in the Twist family BHLH transcription factor 1 gene (*TWIST* [MIM: 601622]), encoding a transcription factor that binds to the p300 and p300/CREBBP-associated factor domains of histone acetyltransferases and regulates their activity,¹¹ and Coffin-Lowry syndrome (MIM: 303600), which results from mutations in ribosomal protein S6 kinase (*RSK2* [MIM: 300075]), a protein required for phosphorylation of histone H3 that regulates chromatin remodeling and gene expression.¹²

In this study, we have identified DNA methylation profiles in peripheral blood samples from a large cohort of individuals who carry mutations in the associated candidate genes responsible for their respective developmental syndromes, including previously unreported signatures in cohorts for GTPTS and SBBYSS (*KAT6B*) and Claes-Jensen syndrome (*KDM5C*). We have also examined the degree of positional overlap between these epi-signatures and have evaluated whether a single classification tool, built on the combined epi-signatures of these conditions, can generate accurate predictions for subjects presenting with the overlapping clinical and molecular features associated with mutations in these genes that play an essential role in the epigenetic machinery.

Material and Methods

Source of Data

This study utilized data and specimens from multiple sources. Peripheral blood DNA samples from subjects with ADCA-DN, Coffin-Siris syndrome, SBBYSS, GTPTS, and Floating-Harbor syndrome were collected from the Care4Rare Canada Consortium. Samples from subjects with clinical characteristics of Kabuki syndrome, ATRX, Saethre-Chotzen syndrome, Coffin-Lowry syndrome, Rett syndrome, Claes-Jensen syndrome, and CHARGE syndrome were collected from the Greenwood Genetic Center (Greenwood, SC, USA). The Kabuki and CHARGE cohorts were supplemented by methylation array files publicly available from GEO (GSE97362).9 Epigenomic data from subjects with Sotos and Weaver syndromes were downloaded from GEO (GSE74432).⁸ All of these subjects had clinical features of the aforementioned syndromes and were screened for mutations in the related genes. The mutation report from every subject was reviewed according to the American College of Medical Genetics Guidelines for interpretation of genomic sequence variants,¹³ and only individuals confirmed to carry a pathogenic or likely pathogenic mutation were used to identify epi-signatures (subject-level data are summarized in Table S1 where available). Control subjects were selected from our lab reference cohort, which is composed of individuals with no known aberrant epigenomic change. This reference cohort was previously preselected from a larger cohort of approximately 1,000 individuals across a broad range of age, sex, and ethnicity distribution.

Methylation Array and Quality Assessment

Genomic DNA was extracted from peripheral blood using standard techniques. Following bisulfite conversion, DNA methylation analysis of the samples was performed using the Illumina Infinium bead chip array, according to the manufacturer's protocol at the Genetic and Molecular Epidemiology Laboratory at McMaster University and the London Health Sciences Molecular Genetic Laboratory. Except for a cohort of subjects with CHARGE syndrome (n = 39), which was assayed using Illumina Infinium methylation EPIC array, all of the samples were assayed using the HumanMethylation450 bead chip. The two arrays harbor 96% overlap in CpG probes. Methylated and unmethylated intensity data were generated as idat files and imported into R 3.4.0 for analysis. Normalization was performed using Illumina normalization method with background correction using the minfi package. Probes with detection p value > 0.01 were excluded from the downstream analyses. For further quality improvement, probes located on chromosomes X and Y and probes known to contain SNPs at the CpG interrogation, or the single-nucleotide extension, were removed. As an additional quality-control step, the sex of the samples was predicted using the signal intensity of the X and Y chromosomes using the minfi package, and the files representing a discordance between the predicted and labeled sex were not used for identification of the DNA methylation profile. All of the samples were examined for genome-wide methylation density, and those deviating from a bimodal signal distribution were excluded. Files generated by the EPIC array were cast as 450k array, and the same analytical procedures used on the 450k array were applied.

Selection of Discovery/Training and Testing Cohorts and Controls

The identification of disease-specific epi-signatures was performed using a randomly selected 75% subset of the database (discovery/

training set) using caTools package. The remaining samples were only used as a testing cohort to assess the performance of the classification model developed later in the study. This procedure was not performed when ≤ 5 samples were available for a disease group (ADCA-DN, GTPTS, and SBBYSS). Given probe differences and technology variations between the two array types (450k and EPIC), it was ensured that the entire discovery/training cohort is assayed using one array (450k). Therefore, all of the CHARGEaffected subjects who were assayed using the 450k array were selected as the discovery/training cohort and the rest were included in the testing cohort. For every disease group in the discovery cohort, a sex- and age-matched control group with a sample size at least four times larger (minimum n = 20) was selected from the reference control group using MatchIt package. The methylation profile of each disease group in the discovery cohort was compared with its matched control separately to identify the disease-specific epi-signature (Table 1). Figure 1 represents the flowchart of the study.

Identification of Disease-Specific Methylation Episignatures

Analysis was performed using a modification of our previously published protocol.^{5–7,10,14} The methylation level for each probe was measured as a beta value, calculated from the ratio of the methylated signals versus the total sum of unmethylated and methylated signals, ranging between 0 (no methylation) and 1 (full methylation). This value was used for biological interpretation and visualization. For statistical analysis, wherever a normal distribution was required (linear regression modeling), beta values were logit transformed to M-values using the following equation: log2(beta/ (1-beta)). A linear regression modeling using the limma package was used to identify the differentially methylated probes. The analysis was adjusted for blood cell type compositions predicted using minfi package. The generated p values were moderated using the eBayes function in the limma package and were corrected for multiple testing using Benjamini and Hochberg method. Probes with a corrected p value < 0.01 and a methylation difference greater than 10%-20% were considered significant. The effect size cutoffs (10%-20%) were determined separately for every condition following the examination of the volcano plots generated in every comparison as previously conducted by Butcher et al.⁹ The identified probes were examined using an unsupervised hierarchical clustering to ensure their ability in separating the subjects from controls. In the case of the Coffin-Siris syndrome, which was caused by two genes in our dataset, the cohort was first split based on the causing genes and then re-analyzed regardless of the gene.

Control for Batch Effect and Robustness of the Identified Epi-signatures

Given that the study cohort was composed of data generated by multiple centers, different dates, and via various instruments, several measures were utilized to minimize possible batch effects and other sources of variability. These steps involved: (1) whenever the sample size allowed, control subjects were selected from the same batch (as with ADCA-DN); (2) if case subjects and their control subjects in a comparison were assayed in multiple batches (as with Kabuki syndrome samples), the batch variable was included as a confounding factor in the regression model; (3) where a particular sample was assayed using both 450k and EPIC arrays (CHARGE syndrome), only files from 450k were used for identification of the signature; (4) for all other disease cohorts, a



machine learning model (Support vector machine) was trained on the identified probes, and we examined how well it could distinguish the batch structure from the disease status. Failure to pass this step resulted in the findings being regarded as non-reliable.

Identification of Genomic Regions with Methylation Changes

To identify genomic regions harboring methylation changes (differentially methylated regions [DMRs]), a bump hunting approach was used by the bumphunter package.¹⁵ The analysis considered regions with greater than 10% change in the overall methylation between case and control subjects with gaps no more than 500 bp among neighboring CpGs. As suggested in the package, 1,000 bootstrapping procedure was performed to compute family-wise error rate (FWER). We selected regions containing a minimum of three consecutive probes and FWER < 0.01. The identified regions were mapped to CpG islands and coding genes. Gviz package was used for visualization of the DMRs.

Assessment of the Overlap between the Epi-signatures

Probes and regions differentially methylated in each disease group were examined to identify potential overlap. The number of probes shared in more than one disease group was visualized using a circos plot. Probes that were shared in >2and 3 disease groups were used to measure the pairwise correlations in the disease cohort. Calculated correlation coefficients were visualized using a correlation plot. Genomic regions harboring differentially methylated probes were assessed for overlap using GenomicRanges package, and regions found in more than one disease type were reported. Functional annotation clustering and gene set enrichment analysis was performed using missMethyl and ReactomePA packages for the genes harboring the shared probes.

Construction and Validation of a Multi-class Prediction Model

The identified signatures were used to build a classification model with the ability to concurrently assess a given methylation profile belonging to any of the disease groups in the study. Caret package was used for feature selection from every signature. First, a receiver operating characteristic curve analysis was performed to identify the most differentiating probes. Those probes with an area under the curve above 0.8 were retained. Next, pairwise correlations among the remaining probes were measured to identify and exclude the redundant signals with R-squared > 0.8. A multi-class support vector machine (SVM) with linear kernel was trained on

the remaining probes using e1071 package. To determine the best hyperparameters and to measure the accuracy of the model, a 10-fold cross-validation was performed. In this process, the training set was divided into ten folds. Nine folds were used for training the model and one fold for testing. After repeating this iteration for all of the ten folds, the mean accuracy was calculated and the hyperparameters with the optimal performance were selected. For every sample, the model was set to generate multiple classification scores between 0 and 1 as the probability of having a methylation profile related to every disease. To assess the sensitivity of the model, the testing cohort, which was not used for identification of the signature or construction of the SVM, was supplied to the model. To determine the specificity, we supplied all of the healthy subjects that were not used in the earlier stages of the study to the model. To understand whether this model is sensitive to other medical conditions representing developmental delay and intellectual disabilities, we tested, using the constructed model, a large number of subjects in our database with a confirmed clinical diagnosis of various diseases including autism spectrum disorders, imprinting defects, RASopathies, chromosomal aberrations, and Down syndrome. As well, we tested whether this classifier is sensitive to other diseases of epigenomic machinery for which no epi-signature was identified in this study. To further confirm that this classifier is not sensitive to the blood

cell type compositions, we downloaded normalized methylation data from isolated cell populations of healthy individuals from GEO (GSE35069)¹⁶ and supplied them to our model for prediction.

Assessment of Ambiguous Case Subjects and Variants of Unknown Significance

The approved model was used to perform a prediction on the DNA methylation profiles of individuals with variants of unknown significance in the respective genes that were not previously included in the identification of the signature or in construction and validation of the classification model. In addition, a prediction was made on subjects with predicted sex discordance, samples obtained from tissues other than blood, healthy subjects carrying pathogenic mutations, and the single subject with Kabuki syndrome resulting from the less common *KDM6A* gene mutation.

Ethics Statement

This study has been approved by the Western University Research Ethics Boards (REB ID 106302) and the Hamilton Integrated Research Ethics Board (REB ID 13-653-T). All of the samples and records were de-identified before the study.

Results

Description of the Study Cohort

Figure 1 represents the flowchart of the study. Mutation analysis of the subjects with clinical features resembling each of the 14 syndromes (Table 1) identified a total of 285 subjects with pathogenic or likely pathogenic mutations, which were classified according to the American College of Medical Genetics (ACMG) guidelines. The remaining subjects (n = 176) carried benign variants or variants of unknown significance (VUS), leaving them unsolved. Our data also included healthy female carriers with pathogenic mutations in *KDM5C* (n = 8), fibroblast samples from individuals with Sotos syndrome (n = 3), and one individual affected with Kabuki syndrome with a pathogenic truncating mutation in KDM6A. Table S1 summarizes the mutation types and demographic characteristics where available. A sample of 196 randomly selected subjects from the 285 individuals with a pathogenic mutation was used as the discovery/ training cohort for identification of epi-signatures as well as for training the classification model. The remaining 89 subjects were regarded as the testing cohort to be used for measuring the sensitivity of the classification model. For each disease group in the discovery/training cohort, a sex- and age-matched sample group four times larger (minimum n = 20) was selected from our reference healthy cohort (n = 650) for comparison. A total of 190 healthy samples never selected as a control for any of the diseases were later used to assess the specificity of the classification model. Table 1 shows the count, age, and sex distributions from every subject group in the discovery/training cohort along with the information from the matched control subjects.

Disease-Specific DNA Methylation Epi-signatures and Differentially Methylated Regions (DMRs)

The comparison of disease cohorts with their matched control subjects was performed using the subset of CpG probes that passed the quality assessment (Table 1). No reliable epi-signature was observed for five of the diseases tested here, including Saethre-Chotzen, Coffin-Siris, Coffin-Lowry, Rett, and Weaver syndromes, as the findings did not meet the criteria described in the methods. For other conditions (i.e., Floating-Harbor, ADCA-DN, Kabuki, ATRX, CHARGE, Sotos, GTPTS, SBBYSS, and Claes-Jensen syndromes), reliable epi-signatures were identified (Table 1). Of these, Sotos (n = 6,858) and ADCA-DN (n = 3,562) revealed the largest number of probes, mostly composed of hypomethylated CpGs. The identified probes from every cohort were confirmed to separate the subjects from the controls using hierarchical clustering (data not shown).

Consistent with the methylation profiles, the bump hunting approach did not identify any genomic segment to be differentially methylated in subjects with Saethre-Chotzen, Coffin-Siris, Coffin-Lowry, Rett, and Weaver syndromes. For the other nine syndromes, however, multiple genomic coordinates containing a minimum of 3 consecutive CpG probes, an average regional methylation difference > 0.10, and a family-wise error rate (FWER) < 0.01 were identified. Subjects with Sotos syndrome showed the largest number of identified regions (n = 1,372), mostly composed of hypomethylated segments (data not shown).

To further ensure that adjustment for blood cell type compositions has not masked a potential methylation profile in the five syndromes with negative results, we repeated the analysis without inclusion of the blood cell type estimates correction. Similar to the previous analyses, a significant methylation profile was not detected. At this stage, we concluded that either no epigenomic profile existed for these five conditions, or their methylation changes were too obscure to pass the thresholds and quality assessment criteria set in this study (see Material and Methods). Thus, our subsequent analyses described below focused on the syndromes for which a differential epigenomic profile was observed: i.e., Floating-Harbor, ADCA-DN, Kabuki, ATRX, GTPTS, SBBYSS, CHARGE, Sotos, and Claes-Jensen syndromes.

Overlap between the Epi-signatures and DMRs

Of the total number of 15,408 probes that composed the epi-signatures of the 9 conditions, only 1,598 (\sim 10%) CpGs were shared across more than one disease group. Among these conditions, Floating-Harbor and Claes-Jensen syndromes were found to share the largest proportion of their epi-signatures with others (mainly with Sotos), whereas this number was minimal to negligible for other conditions, including ADCA-DN which, despite having a large number of CpGs in its epi-signature, shared less than 3% of its probes with other diseases (Figure 2). The



Figure 2. Quantity of Probes from the Epi-signatures of Every Nine Conditions that Are Shared with Each Other Thickness of the bonds represents the number of the shared probes by every two diseases as shown by digits on the circumference of the plot. This plot does not visualize the 217 probes that are shared by more than two conditions.

number of probes that were shared by more than two conditions was limited to 217 probes. Table S2 shows the 217 probes and the related conditions. A gene ontology analysis of the 217 probes using missMethyl package found multiple ontology terms related to histone modifications to be enriched in the harboring genes, including S-methyltransferase activity and histone-lysine N-methyltransferase activity (Table S3). Next, genes overlapping these probes were evaluated using the ReactomePA package to identify pathways enriched in them (Table S4). Lysine histone transferase activity was found as the only enriched pathway with a multiple testing corrected p value < 0.05, which is composed of four genes (*PRDM9, SETDB1*, *HIST1H3E*, *NSD1*). These four genes also contained the most number of probes shared by different conditions (Table S2), which included *PRDM9* with five probes found in all of the conditions except for ADCA-DN, GTPTS, and SBBYSS; *HIST1H3E*, with two probes shared by Floating-Harbor, ATRX, Claes-Jensen, GTPTS, and SBBYSS syndromes; and *NSD1* and *SETDB1* with three and two probes, respectively, shared by Sotos, Floating-Harbor, and Claes-Jensen syndromes. Table S2 also shows that 21 probes in



HOXA5 (MIM: 142952) are shared by three conditions (Kabuki, CHARGE, and GTPTS).

A pairwise correlation analysis on the methylation levels of all of these 217 probes for each affected subject in the disease group revealed that while samples from the same condition showed the strongest correlation with each other, only a weak to moderate positive correlation existed among the samples from across disease groups. The only exception was with subjects with Floating-Harbor and Sotos syndromes, which revealed a moderate negative correlation with each other, despite sharing the most number of probes of all. We further narrowed down the probes to 18 CpGs that were shared by more than three disease groups and re-evaluated the correlation of the subjects based on the methylation levels of these 18 probes. We observed a greater degree of negative correlations between different syndrome groups (Figure 3). The members of every condition, however, correlated well with each other in both of these analyses.

Table 2 shows the genomic coordinates that are shared between every two diseases. Similar to the results described at single probe analysis, Floating-Harbor and Sotos syndromes shared the most differentially methylated coordinates (n = 30), more than half of which showed an opposite direction of change in methylation levels by the two syndromes. As previously observed by the shared CpGs (Table S2), a segment in the promoter of *PRDM9* was among the regions containing at least three consecutive hypomethylated probes in both ATRX and Sotos syndromes (Figure 4). The same segment contains disperse

Figure 3. Pairwise Correlation between Samples with Different Conditions using the Methylation Values of Ten Probes that Are Shared by More than Three Conditions

Red represents positive and blue represents negative correlation. Every visible square represents the correlations of one subject on x axis with its correspondence on y axis. Abbreviations: F-H, Floating-Harbor; C-J, Cales-Jensen

probes hypomethylated in Claes-Jensen syndrome but hypermethylated in Floating-Harbor syndrome (Table S2). The only region with multiple consecutive differentially methylated probes that was shared by more than two syndromes was a segment in the promoter of *HOXA5*. This segment is significantly hypomethylated in GTPTS but hypermethylated in both CHARGE and Kabuki syndromes (Table 2).

Overall, our analyses showed limited overlap between the epi-signatures, which tend to occur only in

a few genomic coordinates. Within these regions, the methylation levels do not correlate well across the diseases, and in many cases the direction of methylation change (hypo- versus hypermethylation) is opposite. These results suggested that it might be possible to combine all of the epi-signatures for building a single classification model for concurrent classification of all of the diseases.

Development and Validation of a Classification Model for Prediction of Disease Classes

To develop a classification model, a multi-class support vector machine (SVM) with linear kernel was trained using a subset of 929 of the most differentiating and non-redundant probes selected from the epi-signature of every syndrome (Table S5). Due to small sample size for GTPTS (n = 3) and SBBYSS (n = 1), these conditions were not included in the model, and thus the training was performed only for the remaining seven syndromes. One model was trained for all of the affected subjects from these seven syndromes (n = 141) and the control subjects. Only the samples from the discovery/training cohort were used for training. The model was set to generate seven classification scores between 0 and 1 as the probability of having a methylation profile related to any of the seven syndromes. Ten-fold cross-validation of this model revealed an accuracy of 99.6%, and it correctly predicted the class of all of the 141 affected subject that were used for its training (Figures 5A-5G).

To determine the sensitivity of our model, 71 subjects from the testing cohort were supplied to the classification

Chr	Disease 1	Coordinates 1	Methylation Difference 1	Probes 1	Disease 2	Coordinates 2	Methylation Difference 2	Probes 2	Direction of Change	Overlapping Gene(s)	Distance to CpG Island (bp)
19	Floating- Harbor	8591364-8591776	-0.4	4	Kabuki	8591364-8591776	-0.28	4	same	MYO1F	0
11	Floating- Harbor	65360123-65360327	-0.27	3	Kabuki	65360123-65360327	-0.19	3	same	KCNK7	0
1	Sotos	227746111-227747468	-0.35	7	Kabuki	227746191– 227746882	0.14	4	opposite	-	0
3	Sotos	49170496-49171051	-0.22	7	Kabuki	49170599-49170794	0.16	4	opposite	LAMB2	12,032
5	Sotos	1856325-1857828	-0.2	7	Kabuki	1856713-1857477	0.16	4	opposite	_	0
12	Sotos	133179887-133180698	-0.25	5	Kabuki	133179887– 133180238	0.11	4	opposite	LRCOL1	0
5	Sotos	101119084-101119766	-0.24	5	Kabuki	101119084– 101119566	0.11	4	opposite	-	512,283
7	Sotos	27170241-27170552	0.19	6	Kabuki	27170388-27170994	-0.14	13	opposite	HOXA4	0
19	Sotos	51330265-51330469	-0.24	4	Kabuki	51330265-51330469	0.13	4	opposite	KLK15	0
15	Sotos	29968032-29968195	-0.3	3	Kabuki	29968032–29968195	0.18	3	opposite	_	410
7	Sotos	27170717-27171051	0.15	9	Kabuki	27170388-27170994	-0.14	13	opposite	_	78
2	Sotos	109746691-109747003	-0.21	5	Kabuki	109746691– 109746754	-0.14	4	same	SH3RF3	0
7	Sotos	142494148 - 142494492	-0.16	6	Kabuki	142494148– 142494492	-0.15	6	same	-	71
5	Sotos	176827082-176827697	-0.16	5	Kabuki	176827392– 176827697	-0.16	3	same	PFN3	0
10	Sotos	132099067-132100019	-0.17	3	Kabuki	132099067– 132100019	0.15	3	opposite	-	109,734
7	CHARGE	27182493-27183946	0.23	20	Kabuki	27182493-27183816	0.16	18	same	HOXA5, HOXA-AS3	0
7	CHARGE	27184316-27184521	0.17	8	Kabuki	27184369-27184441	0.14	4	same	HOXA-AS3	0
17	Floating- Harbor	7486551-7486874	-0.24	7	Claes–Jensen	7486551–7486874	-0.29	7	same	-	0
1	Sotos	247694041-247694531	-0.25	6	Claes–Jensen	247694041– 247694531	-0.24	6	same	GCSAML,GCSAML- AS1, OR2C3	0
5	Sotos	176559334-176559563	-0.36	3	Claes–Jensen	176559334– 176559563	-0.32	3	same	-	0
13	Sotos	113242878-113243141	-0.21	3	Claes–Jensen	113242878– 113243141	-0.32	3	same	TUBGCP3 (396bp away)	221

Table 2. Overlapping Genomic Coordinates with Differential Methylation in Nine Conditions Compared to Control Subjects

(Continued on next page)

Tab	e 2. Continue	ed									
Chr	Disease 1	Coordinates 1	Methylation Difference 1	Probes 1	Disease 2	Coordinates 2	Methylation Difference 2	Probes 2	Direction of Change	Overlapping Gene(s)	Distance to CpG Island (bp)
6	Sotos	32120625-32121433	-0.31	27	Floating- Harbor	32120773-32120933	-0.15	10	same	_	396
6	Sotos	31650735-31651249	-0.17	17	Floating- Harbor	31650735-31650835	-0.19	6	same	-	0
6	Sotos	31650735-31651249	-0.17	17	Floating- Harbor	31650916-31651278	-0.15	11	same	_	0
6	Sotos	32847702-32847845	-0.29	10	Floating- Harbor	32847702-32847845	0.18	10	opposite	_	0
17	Sotos	36997420-36997740	-0.38	7	Floating- Harbor	36997420-36997740	-0.23	7	same	C17orf98	0
8	Sotos	39171866–39172120	-0.31	7	Floating- Harbor	39172020-39172120	0.2	6	opposite	ADAM5 (61bp away)	206,442
10	Sotos	50649666-50650248	-0.39	5	Floating- Harbor	50649666-50650248	0.18	5	opposite	-	42,882
6	Sotos	33282867-33283055	-0.16	12	Floating- Harbor	33282867-33283184	-0.18	19	same	-	0
2	Sotos	129659018-129659946	-0.24	7	Floating- Harbor	129659316– 129659946	0.19	6	opposite	-	0
5	Sotos	83016779-83017553	-0.25	6	Floating- Harbor	83017000-83017644	0.19	6	opposite	HAPLN1	341
4	Sotos	11369349-11370872	-0.18	8	Floating- Harbor	11370314-11370565	0.25	3	opposite	MIR572	0
8	Sotos	102235927-102236831	-0.23	6	Floating- Harbor	102236283– 102236831	0.23	5	opposite	_	0
2	Sotos	165811816-165812159	-0.26	5	Floating- Harbor	165811816– 165812159	0.22	5	opposite	SLC38A11	112,922
7	Sotos	92672812-92673176	-0.25	5	Floating- Harbor	92672812-92673176	0.2	5	opposite	-	0
4	Sotos	155702409-155703138	-0.18	6	Floating- Harbor	155702409– 155703138	0.19	6	opposite	RBM46	0
1	Sotos	1003126-1003529	-0.25	4	Floating- Harbor	1003126-1003529	-0.29	4	same	-	0
3	Sotos	159557552-159558031	-0.24	4	Floating- Harbor	159557552– 159558031	0.23	4	opposite	SCHIP1, IQCJ-SCHIP1	74,528
2	Sotos	164204628-164204915	-0.24	4	Floating- Harbor	164204752– 164205343	0.31	6	opposite	-	0

(Continued on next page)

Tab	e 2. Continued	1									
Chr	Disease 1	Coordinates 1	Methylation Difference 1	Probes 1	Disease 2	Coordinates 2	Methylation Difference 2	Probes 2	Direction of Change	Overlapping Gene(s)	Distance to CpG Island (bp)
22	Sotos	50737978-50738890	-0.22	4	Floating- Harbor	50737978-50738890	-0.29	4	same	PLXNB2	0
12	Sotos	56617576-56617737	-0.28	3	Floating- Harbor	56617576-56617737	-0.23	3	same	-	258
9	Sotos	139258524-139259074	-0.26	3	Floating- Harbor	139258524– 139259074	-0.23	3	same	CARD9	0
19	Sotos	3480363–3480672	-0.21	5	Floating- Harbor	3480363-3480672	-0.2	5	same	SMIM24	1,110
19	Sotos	49222892-49223278	-0.2	5	Floating- Harbor	49222892-49223278	-0.28	5	same	MAMSTR	0
4	Sotos	165898666-165898848	-0.19	5	Floating- Harbor	165898666– 165898848	0.19	5	opposite	TRIM61	20,219
1	Sotos	45278971-45279349	-0.21	4	Floating- Harbor	45278971-45279349	-0.23	4	same	BTBD19	0
19	Sotos	2428350-2428677	-0.19	4	Floating- Harbor	2428350-2429209	-0.2	6	same	LMNB2	0
19	Sotos	1063624-1064218	-0.19	3	Floating- Harbor	1063624-1064218	-0.24	3	same	ABCA7	0
4	Sotos	46126066-46126253	-0.18	3	Floating- Harbor	46126066-46126448	0.25	7	opposite	GABRG1	265,650
12	Sotos	47219737-47219958	-0.14	9	Floating- Harbor	47219626-47219920	0.18	9	opposite	SLC38A4	4,954
5	Sotos	110062384-110062837	-0.16	6	Floating- Harbor	110062384– 110062837	0.23	6	opposite	TMEM232	11,768
11	Sotos	32449254-32449638	-0.17	3	ADCA-DN	32449163-32450692	0.24	11	opposite	WT1	0
7	CHARGE	27137922-27138712	-0.15	4	Sotos	27137922-27138396	-0.17	3	same	HOTAIRM1	1,185
7	CHARGE	53254947-53255065	0.15	3	Sotos	53254947-53255065	0.18	3	same	_	0
5	CHARGE	161178574–161178796	0.14	3	Sotos	161178574– 161178796	0.17	3	same	-	203,189
5	ATRX	23507450-23507752	-0.26	10	Sotos	23507573-23507656	-0.22	5	same	PRDM9	1,682,751
15	ATRX	39871808-39872186	-0.26	6	Sotos	39871876-39872186	-0.16	5	same	_	341
3	ATRX	109056349-109056897	0.29	4	Sotos	109056349– 109056897	-0.26	4	opposite	DPPA4	219,259
6	ATRX	34499314-34499504	-0.28	4	Sotos	34499314-34499504	-0.17	4	same	PACSIN1	0
16	ATRX	58019866-58019984	-0.37	3	Sotos	58019866-58019984	-0.27	3	same	TEPP	0

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Tab	le 2. Continued										
ਤੈ	Disease 1	Coordinates 1	Methylation Difference 1	Probes 1	Disease 2	Coordinates 2	Methylation Difference 2	Probes 2	Direction of Change	Overlapping Gene(s)	Distance to CpG Island (bp)
10	ATRX	43698834-43699070	-0.33	3	Sotos	43698834 - 43699070	-0.32	3	same	RASGEF1A	656
18	ATRX	77905119–77905751	-0.18	10	Sotos	77905663-77905751	0.16	4	opposite	PARD6G-AS1 (56bp away)	0
	ATRX	228291486–228291705	0.23	5	Sotos	228291591- 228291682	0.18	ε	same	I	301
18	ATRX	74499372-74499584	0.2	5	Sotos	74499372-74499584	-0.17	5	opposite	1	0
-	ATRX	3634867-3635100	-0.22	4	Sotos	3634867-3635100	-0.2	4	same	TP73	0
~	GTPTS	27182493–27185282	-0.36	51	Kabuki	27182493–27183816	0.15	18	opposite	HOXA5	0
	GTPTS	27182493–27185282	-0.36	51	CHARGE	27182493–27183946	0.23	20	opposite	HOXAS	0
14	GTPTS	54418728-54418851	-0.38	3	SBBYSS	54418728-54418851	-0.2	3	same	BMP4	0

model. All of these subjects received a score of close to 1 for the syndrome to which they belonged to but a low score for the other six conditions, showing that our model is 100% sensitive for detecting any of the seven conditions examined (Figures 5A–5G). To estimate the specificity of our method, we used 190 normal samples from our reference cohort that were not used for epigenomic profiling of any of the diseases in this study or training the model. All of these samples received scores close to zero by our model for all of the seven disease groups, showing a specificity of 100% (Figure 5H).

To determine whether this classification algorithm is sensitive to the composition of blood cell types, we downloaded a total of 60 methylation array data from 6 healthy individuals,¹⁶ each being assayed for whole blood, peripheral blood mononuclear cells (PBMCs), and granulocytes, as well as for seven isolated cell populations (CD4⁺ T cells, CD8⁺ T cells, CD56⁺ NK cells, CD19⁺ B cells, CD14⁺ monocytes, neutrophils, and eosinophils). The methylation data from these 60 samples were imported into our classification model. Our algorithm assigned a score of close to zero for having any of the seven conditions to all of the files, suggesting that the composition of blood cells will not influence the performance of our model. The scores obtained in this assessment are presented in Table S6.

The 7-Disease Classification Model Is Not Sensitive to Other Developmental Conditions

Next, we examined whether this classification model could distinguish between the epigenomic profile of the seven conditions and the five other previously tested syndromes (Saethre-Chotzen, Coffin-Siris, Coffin-Lowry, Rett, and Weaver syndromes) for which we did not find an epi-signature. Hence, the model was supplied with all of the 69 samples with pathogenic mutations in genes from these five syndromes, as well as to 55 subjects with clinical features similar to these conditions but with VUS variants in the related genes. All of these subjects received low scores for any of the seven disease classes in the algorithm. Next, to determine the performance of this classifier in subjects with developmental delay/intellectual disability (DD/ID) of other etiologies, the model was supplied with the methylation levels of additional cohorts composed of individuals with autism spectrum disorders (n = 146), various chromosomal abnormalities (n = 12), Down syndrome (n = 7), imprinting conditions (Angelman, Beckwith-Wiedemann, Prader-Willi syndrome, n = 50), and various forms of RASopathies (n = 97). All of these subjects had a confirmed clinical and molecular diagnosis of their respected conditions. We found that similar to our previous observations, each of these subjects received low scores for all of the seven diseases in our classification model, further demonstrating that the epi-signatures of these seven conditions are highly specific. These results are demonstrated in Figure 5H.



Classification of Variants of Unknown Significance, Healthy Carriers, Atypical Case Subjects, and Non-blood Methylation Profile

We next applied our classification model to a heterogeneous group of subjects (n = 176) from our cohort displaying some clinical features related to multiple of the aforementioned developmental conditions for which the variant pathogenicity was uncertain (Table S7; Figure 6), in addition to a few other samples from sources other than blood, healthy carriers, and a subject with Kabuki syndrome resulting from KDM6A mutations. Of the 36 subjects within this cohort with clinical features of Kabuki syndrome but with benign or VUS variants in KMT2D, seven were predicted to have a DNA methylation profile specific to Kabuki syndrome. The only subject with Kabuki syndrome with a KDM6A mutation also received a high classification score for Kabuki syndrome. In this cohort, there was also one individual with Floating-Harbor syndrome with a discordance between the predicted and the reported sex. This subject was predicted to belong to the Floating-Harbor class by our model. Later assessment confirmed that it was only a sample-labeling error. Our data contained healthy female carriers of pathogenic mutations in KDM5C (n = 8). All of these subjects received low scores for any of the seven conditions, including the Claes-Jensen syndrome. Two subjects with similar clinical presentations to ATRX but with no confirmed variants in the ATRX gene were predicted as not having any of the seven diseases including ATRX. Half of the 16 samples with VUS in NSD1 were predicted, based on the methylation score, as having ATRX. Our data contained four subjects suspected of having CHARGE syndrome for which sequence analysis results were not available at the time of the study, but who were all predicted to have CHARGE syndrome by our methylation prediction model. Subsequent sequence analysis reports of CHD7 for these four subjects showed that truncating exon-intron boundary mutations exist in two of them (GenBank: NG_007009.1,

Figure 4. Hypomethylation of the 5' UTR of *PRDM9* in Both Sotos-Affected and ATRX-Affected Subjects

The figure illustrates a 302-base pair region containing 10 CpG probes overlapping 5' UTR of *PRDM9*. From top to bottom: chromosome ideogram, CpG probes, gene region, and methylation level data. Blue, ATRX; green, Sotos; pink, controls; line, average methylation; shadow, 95% confidence interval; dots, methylation values from every single sample (0-1).

c.4533+1G>A [p.(=)]; GenBank: NG_007009.1, c.6937-1G>A [p.(=)]). The other two subjects shared one splice site mutation (GenBank: NG_007009.1, c.5405-17G>A [p.(=)]) which has previously been reported in CHARGE syndrome and confirmed

to induce aberrant splicing using RNA-seq analysis in another subject with CHARGE syndrome. Of the remaining 51 subjects with similar phenotypic presentations to CHARGE syndrome, for whom CHD7 mutation screening had not provided a conclusive result, 23 were predicted as CHARGE, 27 were assigned low likelihood for all of the seven conditions, and 1 was predicted as having a methylation profile similar to Kabuki syndrome. Further followup and sequencing of this subject identified a pathogenic truncating variant (p.Arg5097* [c.15289C>T]; GenBank: NG_027827.1) in KMT2D, which confirmed the diagnosis of Kabuki syndrome. The three fibroblast samples from the Sotos-affected subjects were each predicted to have a similar methylation profile to the peripheral blood DNA epi-signature of Sotos syndrome. These findings recommended the high capacity of our method to assign new molecular diagnoses to subjects for whom the sequence analysis was not available or not interpretable, or the initial clinical suspicion was not correct. The complete classification outcome for these subjects is presented in Figure 6 and Table S7.

Discussion

Here we report syndrome-specific, minimally overlapping, DNA methylation epi-signatures in peripheral blood from a large cohort of individuals who possess pathogenic mutations in genes involved in the regulation of epigenomic machinery. Fourteen syndromes were represented in this cohort (Table 1), with reliable epi-signatures being observed for nine syndromes evaluated. We also describe here the development of a single classification algorithm for seven of these syndromes which has a highly sensitive and specific performance in predicting disease class, and which confidently rejects the probability of healthy individuals or other subjects with DD/ID to be affected by any of these seven conditions. Our results support the





A 7-disease SVM classifier concurrently generates seven scores for every subject as the probability of having a DNA methylation profile similar to any of the seven diseases with a confirmed DNA methylation signature. y axis represents scores 0-1, with higher scores indicating a higher chance of carrying a methylation profile related to any of the seven conditions. x axis represents the seven classification scores generated for the same group of tested subjects. These include the probability of having a similar DNA methylation profile to Kabuki syndrome, ATRX, Sotos syndrome, CHARGE syndrome, Floating-Harbor syndrome, ADCA-DN, and Claes-Jensen, respectively. By default, the SVM classifier defines a cut-off of 0.5 for predicting the class; however, the vast majority of the tested individuals received a score close to 0 or 1. Therefore, for the purpose of better visualization, the points are jittered. Every point represents the probability score received for a single sample. This figure represents scores obtained by both the subjects in the training and testing cohorts. Shown are probability scores for belonging to any of the seven classes for: 44 subjects with Kabuki syndrome (A); 19 subjects with ATRX syndrome (B), 38 subjects with Sotos syndrome (C), 79 subjects with CHARGE syndrome (D), 17 subjects with Floating-Harbor syndrome (E), 5 subjects with ADCA-DN (F), and 10 subjects with intellectual disability due to KDM5C (G). The last panel (H) shows the probabilities of belonging to any of the seven disease groups for 436 subjects with other conditions presenting with DD/ID including diseases of epigenomic machinery for which no epi-signature was found, multiple chromosomal aberrations, Down syndrome, various forms of RASopathies, autism spectrum disorders, and imprinting defect conditions, together with 190 healthy control subjects which were not used in any previous step in the study.





A 7-disease SVM classifier concurrently generates seven scores for every subject as the probability of having a DNA methylation profile similar to any of the seven diseases with a confirmed DNA methylation signature. y axis represents scores 0-1, with higher scores indicating a higher chance of carrying a methylation profile related to any of the seven conditions. x axis represents the seven classification scores generated for the same group of tested subjects. These include the probability of having a similar DNA methylation profile to Kabuki syndrome, ATRX, Sotos syndrome, CHARGE syndrome, Floating-Harbor syndrome, ADCA-DN, and Claes-Jensen, respectively. Every point represents the probability score obtained for a single sample. Shown are probability scores for belonging to any of the seven classes for: 8 healthy female carriers with pathogenic mutations in *KDMSC* (A), 16 subjects with VUS variants in *NSD1* (B), 36 subjects with VUS and benign variants in *KMT2D* (C), and 55 subjects with similar features to CHARGE syndrome but no sequence data available or with VUS variants in *CHD7* (D).

hypothesis that the disruption of normal functions by the genes that regulate epigenetic marks, particularly those associated with histone modifications, generate unique DNA methylation epi-signatures with minimal overlaps across the conditions.

Epigenetic marks are established and maintained by an intricate network of proteins that shape the epigenome.

These proteins function as writers that establish the epigenetic marks, readers that interpret them, and erasers that remove the epigenetic marks.¹⁷ The fourth group of remodeller proteins comprises chromatin remodeling complexes that further regulate the epigenome to ensure the contextual (tissue-specific and temporal) accuracy of chromatin. Together, when properly integrated, these highly coordinated and interactive proteins act within multi-protein, multifunctional complexes on chromatin to ensure the tissue- and lineage-specific transcriptional activation and suppression that is essential for proper embryonic and fetal development.¹³ Mutations in the genes encoding these proteins can compromise the functional complexity and integration of the epigenetic machinery, lead to errors in the epigenetic signatures on chromatin, and result in a broad range of pediatric genetic disorders,^{2,3} including those investigated in the present study. Thus, while the original genetic mutation in that particular epigenetic regulator may be primarily detrimental to normal neurodevelopment (and be a significant contributor to that clinical phenotype), all cells may carry an epigenetic echo (or legacy) of that original genetic defect in the form of a DNA methylation signature that may also contribute to the clinical phenotype. This association between a primary mutation in a gene encoding a regulator of chromatin accessibility and a secondary pattern of DNA methylation is likely due to the cross-talk between those interconnected pathways responsible for the post-translational histone modifications and DNA methylation.¹⁷

In the context of the cohort addressed in this paper, ADCA-DN and Rett syndrome resulted from mutations in proteins that write (DNMT1) and read (MeCP2) methylation marks on the DNA. Other syndromes resulted from errors in writers (KMT2D, NSD1, and KAT6B) and erasers (KDM5C) of the histone marks, and chromatin remodeling proteins (CHD7, SRCAP, ATRX). Our results show that syndromes caused by defects in chromatin remodeling proteins (CHARGE, Floating-Harbor, and ATRX), DNA methylation writer (ADCA-DN), histone mark erasers (Claes-Jensen syndrome), and histone mark writers (GTPTS, SBBYSS, Sotos, and Kabuki) generate specific DNA methylation signatures. On the other hand, in case subjects where the defect occurs in reading the methylation marks, such as in Rett syndrome, our study did not observe any consistent change in DNA methylation patterns, suggesting that the mutations in the multifunctional MeCP2 protein that lead to extensive temporal and spatial errors in levels of metabolites and gene expression across the entire genome, affect the function of multiple target pathways across cell lineages, rather than related to any lineage-specific epi-signature;¹⁸ however, a possibility of a methylation signature in an alternate tissue cannot be discounted.

Overlap in the DNA Methylation Signatures Is Limited to the Initiating Event

Phenotypic overlap is a common finding in various conditions resulting from the disruption of the epigenomic machinery. Clinical overlap between CHARGE and Kabuki syndrome and the difficulty in distinguishing between the two conditions by facial features alone has been reported.¹⁹ Sotos syndrome can be challenging to differentiate from Weaver or Beckwith-Wiedemann syndromes.²⁰ Similarly, Floating-Harbor syndrome exhibits some clinical overlap with Rubinstein-Taybi syndrome (MIM: 180849) (another disease of epigenomic machinery) as a result of the molecular interaction between the genes causing the two syndromes.²¹ Bjornsson et al.² suggest that the phenotypic overlap that is seen in some imprinting disorders and a group of multiple congenital anomalies and DD/ID syndromes also result from mutations in the genes encoding components of the epigenetic machinery. Thus, overlap in these shared molecular targets and biological pathways may be the basis for phenotypic overlap in many of such conditions. The shared presence of a small number of genes known to be crucial to embryonic development within these DNA methylation profiles of the conditions we evaluated has not been thoroughly investigated. Butcher et al.⁹ showed a gain of methylation in HOXA5 in both Kabuki and CHARGE syndromes, which was also observed in this study (Table 2). Additionally, we observed the same region to be significantly hypomethylated in subjects with GTPTS. Butcher et al.⁹ also report a segment in the MYO1F (MIM: 601480) gene body to be the most hypomethylated region in Kabuki syndromeaffected individuals. Our previous study on subjects with Floating-Harbor syndrome found the same region to be significantly hypomethylated.⁵ Our current data also shows that two CpG probes from the same MYO1F region are also differentially methylated in CHARGE syndrome (Table S2), although with an opposite direction of change (hypermethylation) to those seen in Kabuki and Floating-Harbor syndromes. Hence, the primary mutational events in epigenetic regulators associated with these syndromes may initiate aberrant DNA methylation and then hardwire these methylation changes into the epigenome as abnormal repressive (or activation) signals at loci encoding developmentally relevant transcription factors, many of which are observed in our study (Table 2) including WT1 (MIM: 607102) and MAMSTR (MIM: 610349) (Sotos), DPPA4 (MIM: 614125) (ATRX), and BMP4 (MIM: 112262) (GTPTS).

The concurrent screening of all of the single probes and regions with differential methylation levels in any of the diseases reported in this study have found that the existing overlap is limited to few specific regions and sites. Examination of DMRs, except for the segment mentioned above in HOXA5, did not reveal any region with a minimum of three consecutive probes to be shared by more than two conditions, and half of the shared regions (including HOXA5), showed an opposite direction of change in methylation levels. At the single CpG probe level, we found only 217 probes that were shared by more than two disease groups, the methylation levels of which were not strongly correlated in different conditions or were negatively correlated as observed between Sotos and Floating-Harbor, despite sharing the largest number of probes. Limiting the analysis to the probes shared by more than three conditions identified further discordance between the disorders (Figure 3). In addition, ADCA-DN shared the least number of probes and DMRs (only 1

DMR with opposite methylation change; Table 2) with other conditions, possibly due to the causative gene being a DNA methyltransferase versus other conditions resulting from histone modifications errors. Also, GTPTS and SBBYSS, which are caused by defects in the same gene, did not reveal any overlap in their epi-signatures and the total number of the probes they did share was less than a few dozen.

These limited shared probes were enriched and most frequently occurred in the genes related to laminin interaction (LAMB2/LAMC1), non-integrin membrane-ECM interactions (LAMB2/LAMC1/NRXN1), and synapse function (PTPRF/DLG4/NRXN1). Also, an intriguing finding was that multiple probes were shared across syndromes and were associated with genes functioning in pathways involving histone acetylation and methylation. Multiple ontology terms related to histone modifications were found to be enriched in the genes harboring these signatures, including S-methyltransferase activity and histone-lysine N-methyltransferase activities (Tables S3 and S4). These pathways involved three genes possessing histone methyltransferase activity (PRDM9, SETDB1, NSD1) and one coding for a subunit of histone H3 (HIST1H3E).

These results indicate that the shared pathways across these developmental syndromes include the initiating events leading to the disruption of the histone modification machinery. Once the initial mutation is established, each condition encompasses differential downstream paths leading to the generation of distinct epi-signatures. This is further supported by the single subject in our database with KDM6A mutation, whom we had predicted as having Kabuki syndrome, using the same model that is built on the epigenetic signature of KMT2D mutations. The immediate targets of these two genes are not overlapping; however, they mutually regulate a large number of genes downstream²² and their disruption leads to a single medical condition with indistinguishable clinical features. This further suggests that the epigenetic profile of these conditions is additionally composed of the downstream changes unique to the syndrome rather than the primary epigenomic event. In addition to the aforementioned components related to histone acetylation and methylation, the HOXA5 promoter was found to be differentially methylated in three syndromes, although with an opposite direction of methylation change. HOXA5 encodes a transcription factor that spatiotemporally regulates the body segmentation and morphogenesis during development. It is well recognized that its expression is tightly regulated by the methylation status of its promoter.²³ Hypermethylation of HOXA5 promoter in CHARGE and Kabuki and its hypomethylation in GTPTS may partly explain the similarity in the phenotypic overlap between CHARGE and Kabuki syndromes, but not in GTPTS, supporting the concept that different epigenetic changes in a region might result in alternative phenotypic outcomes.

A Model Accounting for DNA Methylation Signatures as Legacies of Mutations in Genes of the Epigenetic Machinery

From our results, we propose a mechanistic model by which syndrome-specific DNA methylation signatures arise and are maintained. Central to our model is the concept that the initial dysfunction during early development, which is linked to a specific mutated epigenetic reader, writer, eraser, or remodeler, leaves a specific clinically identifiable and syndrome-related phenotype. In addition, a broader methylation legacy or "echo" remains throughout the genome. That epigenetic echo, reflected in (but not necessarily limited to) DNA methylation, may be a secondary contributor to the overall clinical phenotype through inappropriate activation (or repression) of gene expression in a cell-lineage-dependent manner. For example, Kabuki is among a number of syndromes (including CHARGE and Rubinstein-Taybi) that in addition to displaying clinical features of intellectual disability and distinct craniofacial features, can also present with immune dysfunction. This clinical feature can increase susceptibility to infections and an inability to generate or maintain immunological memory.²⁴ Similarly, altered KMT2D function appears to impact cardiac development and cardiomyocyte function and may lead to defects in ion transport.²⁴ Several of the target genes reported in these papers and associated with these physiological functions were detected in the epi-signatures reported here in our paper, supporting the efficacy and surrogacy of peripheral blood DNA as a source for identifying the Kabuki-specific epi-signature. Furthermore, our approach raises the potential to directly identify epigenetically silenced genes that could be the focus of targeted epigenetic or gene-editing therapies. Such therapies could be focused either on the primary mutation harbored by the epigenetic machinery gene²⁵ or at the secondary targets that reside within the epi-signatures and display altered methylation within promoter or enhancer regions.

An additional application of the epigenetic signature detection extends beyond developmental syndromes to novel, targeted cancer therapy. In infrequent cases, young subjects with developmental syndromes including Kabuki, Coffin-Siris, and Sotos syndromes have been diagnosed with rare tumors at an early age.^{26–28} This is not surprising given that these very same epigenetic regulatory genes (*KMT2D*, *NSD1*, and *SMARCB1*) are frequently mutated in many somatic cancer types.¹⁷ Hence, such epigenetic information reflected by these epi-signatures has the potential to lead to the development of new targeted therapies for various adult cancers that are similarly associated with mutated genes encoding the epigenetic machinery.

DNA Methylation Signatures in Gene Defects across Different Tissues, Mutations, and Zygosity

A clinically applicable aspect of this study relates to whether DNA methylation signatures sourced from peripheral blood samples can act as a surrogate for aberrant DNA methylation patterns that have their primary functional consequences in the developing nervous system and brain. Previous studies have generated conflicting results on the correlation of DNA methylation patterns across tissues and cell types.^{29–32} We were able to directly address this issue of tissue surrogacy, in that training our prediction algorithm on the DNA methylation signatures from peripheral blood of Sotos syndrome-affected individuals also assigned high scores for the three fibroblast DNA samples also obtained from Sotos syndrome-affected individuals. This suggests that easily accessible peripheral blood DNA can be functionally comparable to clinically relevant yet inaccessible target tissues such as the brain. Thus, from a developmental standpoint, some DNA methylation changes associated with primary defects in epigenetic machinery genes are well established before the differentiation of organs in embryonic development and are maintained in these particular cell lineages or have similar functions across tissues.

In any event, this has to be confirmed by studying multiple organs of the affected individuals in different disorders. The limited findings that we have presented in this regard comply with this observation. While ADCA-DN occurs due to one mutation present in all organs, the disease exclusively exhibits features of the nervous system involvement. Bjornsson et al.,² based on this observation, postulates that various tissues show a specific and dosagedependent response to disruption of epigenomic machinery. One observation in our study that may support this model is that the predictive algorithm that we trained using the epi-signature of the Claes-Jensen syndrome cannot detect any of the eight healthy female carriers of KDM5C pathogenic mutations. This indicates that the disruption of epigenetic machinery will be well represented in all tissues, but will lead to a functional disruption only when the dosage is beyond a critical point. This supports the clinical utility of peripheral blood as a surrogate tissue for screening for these conditions. Our study also suggests that the observed signature can be specific to the mutation type. GTPTS and SBBYSS are both caused by mutations in KAT6B. Depending on the location of the coding sequence and type of the mutation (i.e., missense versus frameshift), the gene product can be absent (loss of function) or abnormal (change of function), resulting in two distinct syndromes with overlapping features.³³ Our data show that these two conditions, despite resulting from mutations in the same gene, have distinct epi-signatures. This finding suggests that epi-signatures can potentially be useful in differentiating distinct diseases with overlapping features that are caused by a shared gene defect, where the gene sequencing alone cannot always assign a diagnosis.

The Clinical Implications of the Multi-class DNA Methylation Classification Model

The minimal overlap in the DNA methylation signatures of each of the seven conditions has allowed for training a classifier with the capacity of concurrent assessment of any given subject for all of the seven disease groups with a complete accuracy, sensitivity, and specificity. Furthermore, using datasets generated across multiple institutions, we have demonstrated generalized applicability of this approach in clinical setting that is not sensitive to the confounders such as batch effect. The model not only distinguished related disorders of the epigenetic machinery from each other, but also assigned low probability scores to subjects with various forms of DD/ID ranging from autism spectrum disorders to chromosomal aberrations and imprinting defect conditions that are not caused by mutations in these genes. This method will have a practical value in the clinical diagnosis of such genetic conditions where the disease is rare, clinical features are overlapping, and sequence mutation screening does not always produce conclusive results. In Kabuki syndrome, for instance, the mutation screening of KDM6A and KMT2D, together, identifies pathogenic mutations in only 70% of subjects.³⁴ A similar figure of 65%–70% is reported for atypical cases of CHARGE syndrome.³⁵ The vast majority of rare missense and in-frame in/del variants are currently not interpretable, and as VUSs present a significant challenge in clinical interpretation in diagnostic laboratories and genetic clinics. Some subjects also may carry pathogenic mutations in noncoding regions which are often not screened (e.g., promoter, intronic regions).

The method presented here provides an innovative approach to assigning pathogenicity to these variants by using the epi-signatures of each condition studied. Although the findings would have to be complemented with further clinical assessment, our model has shown its capacity to resolve a large proportion of the unsolved cases in our dataset by assigning new classifications to more than half of the subjects for whom sequence variant assessment alone had not provided a definitive answer. Of interest was a 4-year-old male with cleft palate, dysmorphic ears, microcephaly, and developmental delay, who was suspected of having CHARGE syndrome and who was screened for mutations in CHD7 where only a heterozygous missense VUS variant (GenBank: NM_017780.3, c.2185A>G; GenBank: NM_017780.3, p.Lys729Glu) was found. The classification model we have described above predicted that this subject had an epigenomic profile similar, not to CHARGE syndrome (score = 0.02), but to Kabuki syndrome (score = 0.92). This subject was later confirmed to carry a pathogenic truncating variant in KMT2D and was subsequently diagnosed with Kabuki syndrome. It is worthwhile to note that in many of these conditions, such as Kabuki syndrome, the disease is often caused by de novo variants. Identification of a de novo variant increases the chance of the mutation being pathogenic, although this is not always sufficient to assign pathogenicity to a variant. Unlike the assessment of the "de novo" status of a genetic variant, DNA methylation analysis does not require parental sample DNA to assess the impact of the variant in the proband. However, where both analyses are available, combined evidence could be used to

further strengthen the case for variant pathogenicity status.

With more samples becoming available from such conditions and more diseases being studied for the discovery of epi-signatures, the methodology that we have presented here can provide the basis of application in routine molecular diagnostics. Microarray technology has been shown to be an invaluable tool for the diagnosis of epigenetic conditions including imprinting disorders,³⁶ and as we have shown in the present study, a combination of microarray technology with machine learning will have a great, and as yet relatively unexplored, potential in resolving a large number of subjects with unsolved etiology that are frequently seen in medical genetics practice.

Supplemental Data

Supplemental Data include seven tables and can be found with this article online at https://doi.org/10.1016/j.ajhg.2017.12.008.

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Web Resources

GenBank, https://www.ncbi.nlm.nih.gov/genbank/ GEO, http://www.ncbi.nlm.nih.gov/geo/ OMIM, http://www.omim.org/

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Supplemental Data

Genomic DNA Methylation Signatures Enable

Concurrent Diagnosis and Clinical Genetic Variant

Classification in Neurodevelopmental Syndromes

Erfan Aref-Eshghi, David I. Rodenhiser, Laila C. Schenkel, Hanxin Lin, Cindy Skinner, Peter Ainsworth, Guillaume Paré, Rebecca L. Hood, Dennis E. Bulman, Kristin D. Kernohan, Care4Rare Canada Consortium, Kym M. Boycott, Philippe M. Campeau, Charles Schwartz, and Bekim Sadikovic Table S1 - Characteristics of the patient cohort used in the study

Disease	Id	Sex	Age	Gene	Mutation
ATRX	MS0564	m	6.3	ATRX	c.6254G>A; p.R2085H
ATRX	MS0565	m	18	ATRX	c.736T>C; p.R246C
ATRX	MS0566	m	1.4	ATRX	c.6593A>G; p.H2198R
ATRX	MS0568	m	18.5	ATRX	c.758T>C; p.L253S
ATRX	MS0569	m		ATRX	c.109C>T; p.R37X
ATRX	MS0570	m	21	ATRX	c.4817G>A; p.S1606N
ATRX	MS0571	m		ATRX	c.865T>C; p.C220R
ATRX	MS0572	m	0.7	ATRX	c.5786A>G; p.K1929R
ATRX	MS0573	m	18	ATRX	c.952G>T; p.G249C
ATRX	MS0574	m	14	ATRX	c.730A>C; p.I244L
ATRX	MS0589	m	4.6	ATRX	c.7156C>T; p.R2386X
ATRX	MS0591	m	4.6	ATRX	c.536A>G, p.N179S
ATRX	MS0593	m	4.4	ATRX	deletion of exon 207
ATRX	MS0594	m		ATRX	c.109C>T; p.R37X
ATRX	MS0595	m	27	ATRX	c.7366_7367 InsA; p.M2456N fs X42
ATRX	MS0596	m	14.5	ATRX	c.109C>T; p.R37X
ATRX	MS0597	m	2.5	ATRX	c.736C>T; P.R246C
ATRX	MS0598	m	17.5	ATRX	c.109C>T; p.R37X
ATRX	MS0599	m	14	ATRX	c.109C>T; p.R37X
CHARGE	CHD7-1	f	4	CHD7	c.7282C>T (p.Arg2428*)
CHARGE	CHD7-10	f	0.02	CHD7	c.2905_2906del (p.Arg969Glyfs*25)
CHARGE	CHD7-11	m	12	CHD7	c.7636G>T (p.Glu2546*)
CHARGE	CHD7-12	m	19	CHD7	c.361delC (p.Gly121Valfs*90)
CHARGE	CHD7-13	m	19	CHD7	c.2504_2508delATCTT (p.Arg835Serfs*14)
CHARGE	CHD7-14	m	4	CHD7	c.7717-7720del (p.Gln2537*)
CHARGE	CHD7-15	m	11	CHD7	c.5458C>T (p.Arg1820*)
CHARGE	CHD7-16	m	7	CHD7	exon 1 deletion

CHARGE	CHD7-17	m	15	CHD7	c.5405-17G>A
CHARGE	CHD7-18	f	10	CHD7	c.5405-7G>A
CHARGE	CHD7-19	f	4	CHD7	c.2097-1G>A
CHARGE	CHD7-2	m	6	CHD7	c.3526C>T (p.Gln1176*)
CHARGE	CHD7-3	f	14	CHD7	c.934C>T (p.Arg312*)
CHARGE	CHD7-34	m		CHD7	c.5097dupA (p.Ala1700Serfs*37)
CHARGE	CHD7-36	m		CHD7	c.799G>T (p.Glu267*)
CHARGE	CHD7-4	m	13	CHD7	c562C>T (p.Gly188*)
CHARGE	CHD7-40	m		CHD7	c.1312C>T (p.Gln438*)
CHARGE	CHD7-41	m		CHD7	c.6322G>A (p.Gly2108Arg)
CHARGE	CHD7-46	f		CHD7	c.5050G>A (p.Gly1684Ser)
CHARGE	CHD7-47	f		CHD7	c.5210+3A>G
CHARGE	CHD7-5	m	18	CHD7	c.1327delATGGG (p.Met443Asnfs*130)
CHARGE	CHD7-51	f		CHD7	c.4087delC (p.Leu1363Serfs*9)
CHARGE	CHD7-52	m		CHD7	c.2498+1G>T
CHARGE	CHD7-53	f		CHD7	c.1918delG (p.Gly640Lysfs*71)
CHARGE	CHD7-55	f		CHD7	c.604 C>T (p.Gln202*)
CHARGE	CHD7-56	m		CHD7	c.4393C>T (p.Arg1465*)
CHARGE	CHD7-58	f		CHD7	c.5029C>T (p.Arg1677*)
CHARGE	CHD7-6	m	24	CHD7	c.2504_2508delATCTT (p.Tyr835Serfs*14)
CHARGE	CHD7-60	m		CHD7	c.3177T>G (p.Tyr1059*)
CHARGE	CHD7-61	m		CHD7	c.2839C>T (p.Arg947*)
CHARGE	CHD7-63	m		CHD7	c.4361_4362delAG (p.Gln1454Profs*21)
CHARGE	CHD7-64	f		CHD7	c.2362C>T (p.Gln788*)
CHARGE	CHD7-68	f		CHD7	c.8507delC (p.Pro2836Argfs*53)
CHARGE	CHD7-69	f		CHD7	c.3655C>T (p.Arg1219*)
CHARGE	CHD7-7	m	3	CHD7	c.1990G>T (p.Glu664*)
CHARGE	CHD7-71	f		CHD7	c.1141_1142delAT (p.Met381Alafs*23)
CHARGE	CHD7-72	m		CHD7	c.3082A>G (p.Ile1028Val)
CHARGE	CHD7-8	f	6	CHD7	c.3377dupT (p.Leu1126Phefs*46)
			•	1	

CHARGE	CHD7-9	m	9	CHD7	c.2585delA (p.Leu862Serfs*26)
CHARGE	MS0979	f	0.5	CHD7	c.7252C>T, p.R2418X
CHARGE	MS0982	m	0.8	CHD7	c.3655C>T, p.R1219X
CHARGE	MS0983	f	0.1	CHD7	c.7282C>T, p.R2428X
CHARGE	MS0984	f	0.005	CHD7	c.959_960delAG
CHARGE	MS0985	m	0.2	CHD7	c.5607+1G>A
CHARGE	MS0987	m	0.001	CHD7	c.4517dupG
CHARGE	MS0989	f	1.1	CHD7	c.6716delA
CHARGE	MS0991	f	15.4	CHD7	c.3654C>G, p.Y1218X
CHARGE	MS0992	m	2.1	CHD7	c.3640C>T, p.Q1214X
CHARGE	MS0995	m	0.1	CHD7	c.1972G>T, p.E658X
CHARGE	MS0996	f	3.2	CHD7	c.3778+1G>A
CHARGE	MS1002	m	13.5	CHD7	c.2464G>T, p.E822X
CHARGE	MS1003	f	0.8	CHD7	c.7957C>T, p.R2653X
CHARGE	MS1004	m	24.7	CHD7	c.4393C>T, p.R1465X
CHARGE	MS1005	f	0.01	CHD7	c.1480C>T, p.R494X
CHARGE	MS1007	m	0.003	CHD7	c.559C>T, p.Q187X
CHARGE	MS1011	f	1.5	CHD7	c.4015C>T, p.R1339X
CHARGE	MS1012	m	0.01	CHD7	c.2049delG
CHARGE	MS1013	f	3.5	CHD7	c.6217C>T, p.Q2073X
CHARGE	MS1014	m	0.01	CHD7	c.5050+1G>A
CHARGE	MS1015	m	0.7	CHD7	c.5405-17G>A, abnormal splicing confirmed by RNA-Seq
CHARGE	MS1016	m	17.5	CHD7	c.7047C>A, p.Y2349X
CHARGE	MS1019	f	0.01	CHD7	c.7252C>T, p.R2418X
CHARGE	MS1021	f	8.7	CHD7	c.5627C>G, p.S1876X
CHARGE	MS1022	m	0.01	CHD7	c.1924_1931del8
CHARGE	MS1023	m	0.5	CHD7	c.7879C>T, p.R2627X
CHARGE	MS1024	m	0.001	CHD7	c.6044delT
CHARGE	MS1027	f	0.3	CHD7	c.3339delG
CHARGE	MS1029	m	0.1	CHD7	c.4402delG

CHARGE	MS1030	m	0.2	CHD7	c.5074G>T, p.G1692X
CHARGE	MS1032	m	1.5	CHD7	c.3548 delA
CHARGE	MS1035	m	0.2	CHD7	c.6243C>A, p.C2081X).
CHARGE	MS1036	f	0.9	CHD7	c.5768dupG
CHARGE	MS1037	f	1	CHD7	c.7282C>T, p.R2428X
CHARGE	MS1038	m	2.1	CHD7	c.1152delA
CHARGE	MS1039	f	0.1	CHD7	c.2614-2_2614-1dupAG
CHARGE	MS1044	m	0.1	CHD7	c.5574delA
CHARGE	MS1045	m	0.2	CHD7	c.4862G>A, p.W1621X
CHARGE	MS1046	f	0.1	CHD7	c.3359delG
CHARGE	MS1049	m	10.1	CHD7	7219delA
Coffin-Siris Syndrome	MS0674	m		ARID1B	
Coffin-Siris Syndrome	MS0676	f		ARID1B	
Coffin-Siris Syndrome	MS0678	f		ARID1B	
Coffin-Siris Syndrome	MS0679	f		SMARCB1	
Coffin-Siris Syndrome	MS0680	m		ARID1B	
Coffin-Siris Syndrome	MS0681	f		SMARCB1	
Coffin-Siris Syndrome	MS0683	m		SMARCB1	
Coffin-Siris Syndrome	MS0684	m		ARID1B	
Coffin-Siris Syndrome	MS0685	f		ARID1B	
ADCA-DN	MS0494	f		DNMT1	DNMT1 - NM_001130823.1:c.1709C>T [p.Ala570Val]
ADCA-DN	MS0495	f		DNMT1	DNMT1 - NM_001130823.1:c.1709C>T [p.Ala570Val]
ADCA-DN	MS0496	m		DNMT1	DNMT1 - NM_001130823.1:c.1709C>T [p.Ala570Val]
ADCA-DN	MS0497	m		DNMT1	DNMT1 - NM_001130823.1:c.1709C>T [p.Ala570Val]
ADCA-DN	MS0498	m		DNMT1	DNMT1 - NM_001130823.1:c.1709C>T [p.Ala570Val]
Kabuki	KMT2D-1	f	15	KMT2D	c.15061C>T (p.Arg5021*)
Kabuki	KMT2D-10	m	6	KMT2D	c.4135_4136delA (p.Met1379Valfs*52)
Kabuki	KMT2D-11	m	16	KMT2D	c.11710C>T (p.Gln3904*)
Kabuki	KMT2D-2	m	1	KMT2D	c.16318delG (p.Glu5440Argfs*16)

Kabuki	KMT2D-22	f		KMT2D	c 11158C T (p Glp 3720*)
Kabuki	KW12D-22	1		KM12D	e.11196e>1 (p.6m3720)
Kabuki	KMT2D-25	f		KMT2D	c.16521+1G>T
Kabuki	KMT2D-26	f		KMT2D	c.1940dupC (p.Pro648Thrfs*2)
Kabuki	KMT2D-29	m		KMT2D	c.15088C>T (p.Arg5030Cys)
Kabuki	KMT2D-3	m	18	KMT2D	c15030dupA (p.Glu5011Argfs*13)
Kabuki	KMT2D-30	m		KMT2D	c.5135delA (p.Lys1712Argfs*10)
Kabuki	KMT2D-32	m		KMT2D	c.16052G>A (p.Arg5351Gln)
Kabuki	KMT2D-33	f		KMT2D	c.11203C>T (p.Gln3735*)
Kabuki	KMT2D-36	f		KMT2D	c.15536G>A (p.Arg5179His)
Kabuki	KMT2D-4	f	16	KMT2D	c.8172_8173delC (p.Phe2724Glnfs*5)
Kabuki	KMT2D-5	m	15	KMT2D	c.6595delT (p.Tyr2199Ilefs*65)
Kabuki	KMT2D-6	m	11	KMT2D	c.14055-14056delCA (p.His4685Glnfs*4)
Kabuki	KMT2D-7	m	14	KMT2D	c.6295C>T (p.Arg2099*)
Kabuki	KMT2D-8	m	20	KMT2D	c.4135_4136delA (p.Met1379Valfs*52)
Kabuki	KMT2D-9	m	18	KMT2D	c.12592C>T (p.Arg4198*)
Kabuki	MS0608	m	7	KMT2D	MLL2 c.1801_1822dup 22
Kabuki	MS0711	f	6.7	KMT2D	c.13059delG
Kabuki	MS0713	m	1.9	KMT2D	c.839+1delG
Kabuki	MS0719	f	3.9	KMT2D	c.15844C>T (p.R5282X)
Kabuki	MS0720	m	21.6	KMT2D	c.16294C>T (p.R5432W)
Kabuki	MS0721	f	0	KMT2D	c.8488C>T (p.R2830X)
Kabuki	MS0738	f	3.8	KMT2D	c.4168dupG
Kabuki	MS0740	m	4.3	KMT2D	c.15289C>T (p.R5097X)
Kabuki	MS0744	m	2.6	KMT2D	c.4419-2A>G
Kabuki	MS0774	f	16.6	KMT2D	c.548delC
Kabuki	MS0776	f	3.3	KMT2D	c.7411C>T (p.R2471X)
Kabuki	MS0778	f	24.1	KMT2D	c.1966dupC
Kabuki	MS0782	f	9.5	KMT2D	c.6200delA
Kabuki	MS0783	f	9.3	KMT2D	c.7933C>T (p.R2645X)
Kabuki	MS0784	f	5.8	KMT2D	c.13450C>T (p.R4484X)

Kabuki	MS0786	f	19.1	KMT2D	c.16048A>T (p.K5350X)
Kabuki	MS0787	m	7.1	KMT2D	c.10201C>T (p.Q3401X)
Kabuki	MS0788	m	3.4	KMT2D	c.16360C>T (p.R5454X)
Kabuki	MS0790	m	3.1	KMT2D	c.8692C>T (p.Q2898X)
Kabuki	MS0792	f	4.1	KMT2D	c.14878C>T (p.R4960X)
Kabuki	MS0793	f	23.1	KMT2D	c.6265A>T (p.K2089X)
Kabuki	MS0794	f	6.9	KMT2D	c.10740+1G>A
Kabuki	MS0795	m	2.2	KMT2D	c.13652T>A (p.L4551X)
Kabuki	MS0796	f	1	KMT2D	c.11596C>T (p.Q3866X)
Kabuki	MS0842	f	4	KMT2D	c. 4135_4136delAT
Claes-Jensen	MS0690	m	30	KDM5C	c.1510G>A; p.V504M
Claes-Jensen	MS0691	m	6	KDM5C	c.1439C>T; p.P480L
Claes-Jensen	MS0692	m	26	KDM5C	c.4439_4440delAG; p.fsR1481fxX9
Claes-Jensen	MS0693	m	42	KDM5C	IVS11+5G>A; p.E468GfsX2
Claes-Jensen	MS0695	m	8	KDM5C	c.1510G>A; p.V504M
Claes-Jensen	MS0697	m	2	KDM5C	c.1439C>T; p.P480L
Claes-Jensen	MS0723	m	37	KDM5C	c.229G>A; p.A77T
Claes-Jensen	MS0726	m	28	KDM5C	c.4439_4440delAG; p.fsR1481fxX9
Claes-Jensen	MS0727	m	13	KDM5C	c.229G>A; p.A77T
Claes-Jensen	MS0728	m	26	KDM5C	c.1510G>A; p.V504M
Rett	MS0516	f	1	MECP2	R106W
Rett	MS0517	f	25	MECP2	R168X
Rett	MS0518	f	34	MECP2	P302R
Rett	MS0519	f	2	MECP2	c1085del1113
Rett	MS0520	f	1	MECP2	T158M
Rett	MS0521	f	3	MECP2	917_939del123
Rett	MS0522	f	3	MECP2	deletion in exon 4
Rett	MS0523	f	1	MECP2	T158M
Rett	MS0524	f	4	MECP2	P225R
Rett	MS0525	f	6	MECP2	1157_1197 del 41c (c terminal deletion)

Rett	MS0526	f	1.5	MECP2	R255X
Rett	MS0527	m	0.7	MECP2	507 insAG
Rett	MS0555	f	6	MECP2	deletion in exon 3 and 4
Rett	MS0557	f	29	MECP2	R106W
Rett	MS0559	f	3	MECP2	T158M
Rett	MS0561	f	11	MECP2	R255X
Rett	MS0563	f	4	MECP2	partial deletion of exon 4
Coffin-Lowry	MS0513	f	6	RSK2	c.1520insA; p. fs507X
Coffin-Lowry	MS0514	m		RSK2	c.1894C>T; p. R632X
Coffin-Lowry	MS0515	m	11.5	RSK2	c.371G>S; p.fs372X
Coffin-Lowry	MS0528	m	11.5	RSK2	c.2065C>T; p.Q688X
Coffin-Lowry	MS0530	m	4	RSK2	c.2186G>A; p. R729Q
Coffin-Lowry	MS0532	m	1.3	RSK2	c.340C>T; p.W114R
Coffin-Lowry	MS0533	m	13	RSK2	c.727C>T; p. X243
Coffin-Lowry	MS0534	m	22.8	RSK2	IVS14+1G>A
Coffin-Lowry	MS0535	m	12	RSK2	c.62bp deletion spanning 35bp at 3'end of intron 7 and 29 bp at 5' of exon 8
Coffin-Lowry	MS0536	m	2	RSK2	c.386insCTTT; p.fs 130 X141
Coffin-Lowry	MS0538	m	8	RSK2	c.1155delT; p. fs385-X40 codons downstream
Sotos	11D/6637	m	10	NSD1	ex15-19 del
Sotos	11D/6718	f	10	NSD1	c.1716delC (p.Cys573Valfs*26)
Sotos	11D_0326	f	9	NSD1	chr5:175,366,008-177,470,488 (hg19)
Sotos	11D_0328	f	7	NSD1	chr5:175,764,262-177,059,256 (hg19)
Sotos	A1208	f	3.5	NSD1	c.6454C>T (p.Arg2152*)
Sotos	DL117330	f	13.2	NSD1	c.5445C>G (p.Tyr1815*)
Sotos	DL122057	m	3	NSD1	c.4843delT (p.Tyr1615Thrfs*27)
Sotos	DL151889	m	2.2	NSD1	microdeletion of distal 5q35.2
Sotos	DL159249	f	12	NSD1	c.6349C>T (p.Arg2117*)
Sotos	DL168744	m	2.2	NSD1	c.1492C>T (p.Arg498*)
Sotos	DL179067	m	18	NSD1	c.6454C>T (p.Arg2152*)
Sotos	DL38402	m	19.7	NSD1	c.1583delA (p.Lys528Argfs*8)

Sotos	DL50448	m	8	NSD1	c.2014-2018delACAGA (p.Thr672Glufs*9)
Sotos	DL50450	m	41	NSD1	c.2014-2018delACAGA (p.Thr672Glufs*9)
Sotos	DL50452	f	2	NSD1	c.2014-2018delACAGA (p.Thr672Glufs*9)
Sotos	DL76010	f	1.6	NSD1	c.1810C>T (p.Arg604*)
Sotos	DL87406	m	0.5	NSD1	5q35.2-35.3 (RP11-67P18 to RP11-423H2), FISH BAC RP11-99N22 deleted
Sotos	DL89813	m	10.6	NSD1	c.1801A>T (p.Lys601*)
Sotos	DL94609	m	20	NSD1	4977_4978insG (p.Arg1660Alafs*13)
Sotos	HK-10067	m		NSD1	c.6013C>T (p.Arg2005*)
Sotos	HK-11302	f		NSD1	c.5397_c.5398insT (p.Gly1800Trpfs*3)
Sotos	HK-11807	f		NSD1	c.5887A>T (p.Lys1963*)
Sotos	HK-13565	f		NSD1	c.2607_c.2608insA (p.Arg870Lysfs*5)
Sotos	HK-15164	f		NSD1	WGD-Whole gene deletion
Sotos	HK-1540	m		NSD1	WGD-Whole gene deletion
Sotos	HK-15438	f		NSD1	WGD-Whole gene deletion
Sotos	HK-2129	f		NSD1	c.4417C>T (p.Arg1473*)
Sotos	HK-4470	f		NSD1	c.1754C>G (p.Ser585*)
Sotos	HK-4913	f		NSD1	c.2362C>T (p.Arg788*)
Sotos	HK-5107	f		NSD1	WGD-Whole gene deletion
Sotos	HK-5439	f		NSD1	c.3728delG (p.Ser1243Thrfs*14)
Sotos	HK-5530	m		NSD1	c.2334_c.2335insA (p.His779Thrfs*30)
Sotos	HK-5533	m		NSD1	c.2314delG (p.Ala772Glnfs*19)
Sotos	HK-5589	f		NSD1	c.1310C>G (p.Ser437*)
Sotos	HK-5977	f		NSD1	c.6349C>T (p.Arg2117*)
Sotos	HK-6779	m		NSD1	c.3067C>T (p.Arg1023*)
Sotos	HK-6966	f		NSD1	c.5296C>A (R1766X p.= (p.Arg1766Arg))
Sotos	HK-7783	m		NSD1	c.4217delGAAA (p.Arg1406Asnfs*12)
Floating Harbor	MS0433	m		SRCAP	
Floating Harbor	MS0434	f		SRCAP	
Floating Harbor	MS0435	m		SRCAP	
Floating Harbor	MS0437	f		SRCAP	

Floating Harbor	MS0439	f		SRCAP	
Floating Harbor	MS0440	f		SRCAP	
Floating Harbor	MS0441	f		SRCAP	
Floating Harbor	MS0442	f		SRCAP	
Floating Harbor	MS0443	m		SRCAP	
Floating Harbor	MS0444	m		SRCAP	
Floating Harbor	MS0445	m		SRCAP	
Floating Harbor	MS0446	f		SRCAP	
Floating Harbor	MS0447	f		SRCAP	
Floating Harbor	MS0449	f		SRCAP	
Floating Harbor	MS0450	f		SRCAP	
Floating Harbor	MS0451	m		SRCAP	
Floating Harbor	MS0452	f		SRCAP	
Saethre-Chotzen Syndrome	MS0611	f	0.003	TWIST	c.385_405dup21
Saethre-Chotzen Syndrome	MS0612	m	5.7	TWIST	c.261del13
Saethre-Chotzen Syndrome	MS0613	f	0.9	TWIST	c.156delC
Saethre-Chotzen Syndrome	MS0614	m	38	TWIST	A G>T transversion at nucleotide 376 was detected (E126X)
Saethre-Chotzen Syndrome	MS0615	m	30	TWIST	A 21 basepair insertion beginning at nucleotide 406 was detected.
Saethre-Chotzen Syndrome	MS0617	f	33.5	TWIST	The deletion of a cytosine at nucleotide 156 was detected.
Saethre-Chotzen Syndrome	MS0618	m	17.7	TWIST	Insertion of 21 base pairs beginning at nucleotide 418
Saethre-Chotzen Syndrome	MS0624	f	20.7	TWIST	c.211C>T, p.Q71X
Saethre-Chotzen Syndrome	MS0625	m	0.7	TWIST	c.325C>T, p.Q109X
Saethre-Chotzen Syndrome	MS0626	m	0.1	TWIST	c.396_416dup21
Saethre-Chotzen Syndrome	MS0632	f	0.01	TWIST	c.193G>T, p.E65X
Saethre-Chotzen Syndrome	MS0633	f	23.3	TWIST	c.472T>C, p.F158L
Saethre-Chotzen Syndrome	MS0638	f	0.35	TWIST	entire TWIST gene deleted
Saethre-Chotzen Syndrome	MS0648	f	0.003	TWIST	MLPA analysis detected a full gene deletion involving the TWIST gene.
Saethre-Chotzen Syndrome	MS0653	f	0.7	TWIST	A G>T transition at nucleotide 160 (stop codon for a glycine at amino acid 54)
Saethre-Chotzen Syndrome	MS0654	f	20.5	TWIST	c.397_417dup21
Saethre-Chotzen Syndrome	MS0656	m	0.6	TWIST	c.120_145del26
1				1	

Saethre-Chotzen Syndrome	MS0657	f	23.5	TWIST	c.149delC
Saethre-Chotzen Syndrome	MS0660	f	12.3	TWIST	A 21 base pair deletion beginning at nucleotide 394 was detected.
Saethre-Chotzen Syndrome	MS0665	f	21.5	TWIST	c.352C>G, p.R118G
Saethre-Chotzen Syndrome	MS0668	f	0.8	TWIST	c.376G>T, p.Gln126X
Saethre-Chotzen Syndrome	MS0669	f	28.7	TWIST	c.490C>T, p.Q164X
Saethre-Chotzen Syndrome	MS0688	m	0.02	TWIST	c.149delC
Saethre-Chotzen Syndrome	MS0689	f	0.1	TWIST	c.149delC
Saethre-Chotzen Syndrome	MS0844	f	23.5	TWIST	full gene deletion of TWIST
Weaver	A117	f		EZH2	c.394C>T (p.Pro132Ser)
Weaver	A123	m		EZH2	c.457_459del (p.Tyr153del)
Weaver	A134	f		EZH2	c.2080C>T (p.His694Tyr)
Weaver	A1359	m		EZH2	c.2050C>T (p.Arg684Cys)
Weaver	A212	f		EZH2	c.398A>G (p.Tyr133Cys)
Weaver	DL129940	m		EZH2	c.1876G>A (p.Val626Met)
Weaver	F0123	m		EZH2	c.553G>C (p.Asp185His)
GTPTS	MS0673	f		KAT6B	
GTPTS	MS0675	m		KAT6B	
GTPTS	MS0677	m		KAT6B	
SBBYSS	MS0682	f		KAT6B	

Age and mutation information from some of the samples have been masked to protect the identities. Reference sequence for all mutations is hg19, except for NSD1 mutations (hg18, unless specified otherwise [11D-0326 and 11D-0328])

Table	S2 –	Probes	shared	by the	epi-sig	gnatures	of r	nore	than	two	condit	ions

Probe	Chr	Position	Genes	Enhancer	CpG island	ADCA- DN	Floating Harbor	ATRX	Kabuki	CHARGE	Claes- Jensen	Sotos	GTPTS	SBBYSS
cg20768358	chr1	3574999	TP73		OpenSea			Yes	Yes			Yes		
cg04015962	chr1	10949192			OpenSea				Yes		Yes	Yes		
cg06790069	chr1	40203513	PPIE		N_Shore		Yes			Yes				Yes
cg02283735	chr1	44011587	PTPRF	TRUE	OpenSea				Yes		Yes	Yes		
cg06766016	chr1	45278971	BTBD19		Island		Yes				Yes	Yes		
cg19530728	chr1	45279132	BTBD19		Island		Yes				Yes	Yes		
cg04605287	chr1	54953486			N_Shore				Yes		Yes	Yes		
cg08128444	chr1	68966466			S_Shelf			Yes		Yes		Yes		
cg20457732	chr1	71172486		TRUE	OpenSea		Yes				Yes	Yes		
cg03847896	chr1	112154295		TRUE	OpenSea		Yes	Yes		Yes				
cg25066665	chr1	150335507	RPRD2		N_Shore		Yes				Yes	Yes		
cg15448220	chr1	150897856	SETDB1		N_Shore		Yes				Yes	Yes		
cg10589385	chr1	150898437	SETDB1		N_Shore		Yes				Yes	Yes		
cg13069100	chr1	155035008	ADAM15;EFNA4		Island		Yes			Yes				Yes
cg09938479	chr1	178455912			Island		Yes		Yes		Yes	Yes		
cg26004771	chr1	178456093			Island		Yes				Yes	Yes		
cg22790377	chr1	228291668	Clorf35		S_Shore		Yes	Yes	Yes					
cg20462561	chr1	228890820			Island		Yes		Yes			Yes		
cg03982998	chr1	238024671	LOC100130331		OpenSea			Yes		Yes		Yes		
cg11166453	chr1	247681781			Island		Yes				Yes	Yes		Yes
cg06362358	chr1	248855545			S_Shore		Yes	Yes	Yes					
cg03701930	chr10	1981436			OpenSea				Yes	Yes		Yes		
cg08584759	chr10	11912126	C10orf47		Island				Yes	Yes		Yes		
cg20137746	chr10	27235598			OpenSea		Yes	Yes				Yes		
cg01001533	chr10	44184868			N_Shore				Yes			Yes	Yes	
cg24530234	chr10	44224014			Island		Yes	Yes				Yes		
cg12845268	chr10	63657363		TRUE	OpenSea				Yes		Yes	Yes		

cg09411922	chr10	75121857			S_Shelf			Yes		Yes		Yes		
cg12667595	chr10	81743362			Island		Yes		Yes			Yes		
cg16844333	chr10	89167457			OpenSea	Yes	Yes					Yes		
cg23615741	chr10	101297642		TRUE	Island		Yes		Yes				Yes	Yes
cg27245056	chr10	122356491			OpenSea		Yes	Yes		Yes				
cg07040013	chr10	132099553			OpenSea	Yes			Yes			Yes		
cg22079043	chr11	1785631	CTSD;HCCA2		S_Shore		Yes				Yes	Yes		
cg13924715	chr11	10750890			OpenSea			Yes	Yes			Yes		
cg05488217	chr11	58730897		TRUE	N_Shore		Yes			Yes			Yes	
cg17379860	chr11	61159602	TMEM216		N_Shore			Yes			Yes	Yes		
cg04638150	chr11	62273418	AHNAK	TRUE	OpenSea			Yes	Yes			Yes		
cg01178624	chr11	65360327	KCNK7		Island		Yes		Yes					Yes
cg22203628	chr11	69241075			OpenSea		Yes			Yes		Yes		
cg04006327	chr11	72533487	ATG16L2	TRUE	Island		Yes			Yes				Yes
cg03274739	chr11	98437942			OpenSea		Yes			Yes		Yes		
cg06137123	chr11	129444480			OpenSea			Yes	Yes	Yes		Yes		
cg05237469	chr11	133446415			OpenSea			Yes	Yes		Yes			
cg10635895	chr12	2339439	CACNA1C		Island		Yes				Yes	Yes		
cg01837362	chr12	34492938			N_Shore			Yes	Yes			Yes		
cg16615454	chr12	50475167	ACCN2		Island			Yes				Yes	Yes	
cg03146625	chr12	54448729	HOXC4		S_Shore				Yes	Yes		Yes		
cg17970299	chr12	54772804	ZNF385A	TRUE	Island		Yes	Yes	Yes					
cg08425810	chr12	58132558	AGAP2		Island				Yes	Yes		Yes		
cg00047843	chr12	83321412	TMTC2	TRUE	OpenSea			Yes		Yes		Yes		
cg10082647	chr12	107348855	C12orf23		N_Shore						Yes	Yes	Yes	
cg11367159	chr12	110044531		TRUE	OpenSea		Yes				Yes	Yes		
cg10504392	chr12	110044639		TRUE	OpenSea		Yes				Yes	Yes		
cg13810723	chr12	119592343	SRRM4		S_Shore	Yes		Yes				Yes		
cg08083251	chr13	23309930			OpenSea		Yes	1	Yes			Yes		
cg13614409	chr13	31506752	C13orf26		OpenSea		Yes		Yes			Yes		

cg26168643	chr13	88328009	SLITRK5		N_Shore			Yes	Yes	Yes				
cg24626752	chr13	88328274	SLITRK5		N_Shore				Yes	Yes				Yes
cg05757365	chr13	88328471	SLITRK5		N_Shore				Yes	Yes				Yes
cg08602346	chr13	113242997	TUBGCP3		S_Shore						Yes	Yes	Yes	
cg14009504	chr14	71555837	PCNX	TRUE	OpenSea			Yes		Yes		Yes		
cg09214243	chr15	29968124			S_Shore				Yes		Yes	Yes		
cg12414681	chr15	29968195			S_Shore				Yes		Yes	Yes		
cg09456065	chr15	56759655			S_Shelf			Yes		Yes		Yes		
cg02703787	chr15	93256789			N_Shore		Yes	Yes					Yes	
cg27009812	chr16	3062597	CLDN9	TRUE	N_Shore					Yes	Yes	Yes		
cg04715503	chr16	3062795	CLDN9	TRUE	N_Shore					Yes	Yes	Yes		
cg00524708	chr16	31159558	PRSS36		N_Shore			Yes	Yes			Yes		
cg10424751	chr16	32289757			Island			Yes	Yes			Yes		
cg00130223	chr16	33070551			Island				Yes	Yes		Yes		
cg03203197	chr16	34587120			Island				Yes	Yes		Yes		
cg01517680	chr16	49499006		TRUE	Island		Yes	Yes				Yes		
cg12499872	chr16	58019893	TEPP		Island	Yes		Yes				Yes		
cg09482050	chr16	67686832	RLTPR		N_Shore		Yes	Yes				Yes		
cg06911613	chr16	85846184			S_Shore				Yes			Yes		Yes
cg04387835	chr17	4649076	ZMYND15		OpenSea			Yes	Yes			Yes		
cg17900689	chr17	4649262	ZMYND15		OpenSea			Yes	Yes			Yes		
cg03613822	chr17	7115140	DLG4		N_Shelf		Yes	Yes	Yes					
cg19786602	chr17	7966326			OpenSea			Yes	Yes			Yes		
cg06663305	chr17	8095813			S_Shelf		Yes	Yes	Yes					
cg16006004	chr17	41278241	BRCA1;NBR2		Island		Yes		Yes					Yes
cg07075026	chr17	47091521	IGF2BP1		Island		Yes			Yes		Yes		
cg10950924	chr17	47092072	IGF2BP1	TRUE	Island		Yes			Yes		Yes		
cg18040354	chr17	53800484	TMEM100		OpenSea		Yes			Yes	Yes			
cg12131208	chr17	58499700	C17orf64	TRUE	S_Shore				Yes		Yes	Yes		
cg21122199	chr17	58499720	C17orf64	TRUE	S_Shore	Yes					Yes	Yes		

cg08122232	chr17	74100132	EXOC7		S_Shore		Yes				Yes	Yes	Yes	
cg12082025	chr19	1064218	ABCA7		Island		Yes	Yes				Yes		
cg14576825	chr19	1676136			Island	Yes							Yes	Yes
cg18795569	chr19	3480508	C19orf77		N_Shore		Yes	Yes				Yes		
cg02085507	chr19	6739192	TRIP10		N_Shore		Yes		Yes			Yes		
cg22987448	chr19	8591364	MYO1F		Island		Yes		Yes	Yes				
cg08283130	chr19	8591776	MYO1F		Island		Yes		Yes	Yes				
cg14266237	chr19	12995624	KLF1		N_Shore		Yes				Yes			Yes
cg19492423	chr19	18700933	C19orf60		S_Shore		Yes	Yes	Yes		Yes			Yes
cg21977234	chr19	18701320	C190rf60		S_Shore		Yes	Yes			Yes			Yes
cg26931208	chr19	49244592	IZUMO1		S_Shore		Yes	Yes				Yes		
cg26750002	chr19	49244674	IZUMO1;RASIP1		S_Shore		Yes	Yes				Yes		
cg17434634	chr19	49522954		TRUE	Island		Yes						Yes	Yes
cg05934682	chr19	50394298	IL411	TRUE	S_Shore			Yes	Yes			Yes		
cg11383165	chr19	56002283	SSC5D		Island				Yes			Yes		Yes
cg16151651	chr2	1609119			OpenSea		Yes				Yes	Yes		
cg18396357	chr2	50201413	NRXN1		OpenSea		Yes	Yes		Yes				
cg09692695	chr2	74668286	RTKN		Island			Yes			Yes	Yes		
cg25334934	chr2	121269348		TRUE	OpenSea				Yes		Yes	Yes		
cg18478731	chr2	129659682			Island		Yes		Yes			Yes		
cg11942181	chr2	129659946			Island		Yes		Yes			Yes		
cg09015973	chr2	131673771	ARHGEF4		Island				Yes			Yes		Yes
cg03727500	chr2	232348334		TRUE	Island				Yes		Yes	Yes		
cg11559198	chr2	232348794			Island				Yes		Yes	Yes		
cg05857996	chr20	2675418	EBF4	TRUE	S_Shore				Yes		Yes	Yes		
cg24263062	chr20	2730191	EBF4		Island				Yes		Yes	Yes		
cg12176783	chr20	62694000	TCEA2		Island		Yes		Yes					Yes
cg05437132	chr21	10990903	TPTE		Island	1		Yes		Yes		Yes		
cg04767174	chr21	15069190			Island			Yes	Yes			Yes		
cg08597025	chr21	15077096			Island			Yes		Yes		Yes		

cg21916358	chr21	15077562			Island			Yes	Yes	Yes		Yes		
cg10044179	chr21	15352983	C21orf81		S_Shore			Yes	Yes	Yes		Yes		
cg23720384	chr21	23747055			OpenSea			Yes	Yes				Yes	
cg14172108	chr21	34405553			N_Shore				Yes		Yes	Yes		
cg00274965	chr21	34405681			Island				Yes		Yes	Yes		
cg03389890	chr22	16868045			Island				Yes			Yes	Yes	
cg16054907	chr22	50706065	MAPK11		Island		Yes	Yes		Yes				
cg07225641	chr3	46618325	LRRC2;TDGF1		Island	Yes						Yes	Yes	
cg08234664	chr3	49170668	LAMB2		OpenSea		Yes		Yes		Yes	Yes		
cg14099457	chr3	49170794	LAMB2		OpenSea				Yes		Yes	Yes		
cg10313047	chr3	52351355	DNAH1		OpenSea		Yes		Yes			Yes		
cg00487142	chr3	109056357	DPPA4		OpenSea		Yes	Yes				Yes		
cg14836960	chr3	109056618	DPPA4		OpenSea		Yes	Yes				Yes		
cg21750709	chr3	109056897	DPPA4		OpenSea		Yes	Yes				Yes		
cg01571001	chr3	128215433			Island		Yes		Yes					Yes
cg01757168	chr3	128215565			Island		Yes		Yes	Yes				Yes
cg01757168 cg09406615	chr3 chr3	128215565 165555103	ВСНЕ	TRUE	Island OpenSea		Yes Yes		Yes	Yes		Yes	Yes	Yes
cg01757168 cg09406615 cg10663765	chr3 chr3 chr3	128215565 165555103 194014592	ВСНЕ	TRUE	Island OpenSea Island		Yes Yes		Yes Yes	Yes	Yes	Yes Yes	Yes	Yes
cg01757168 cg09406615 cg10663765 cg23855319	chr3 chr3 chr3 chr3 chr4	128215565 165555103 194014592 2062392	BCHE NAT8L	TRUE	Island OpenSea Island Island		Yes Yes Yes		Yes	Yes	Yes Yes	Yes Yes Yes	Yes	Yes
cg01757168 cg09406615 cg10663765 cg23855319 cg16091553	chr3 chr3 chr3 chr4 chr4	128215565 165555103 194014592 2062392 46126245	BCHE NAT8L GABRG1	TRUE	Island OpenSea Island Island OpenSea		Yes Yes Yes Yes		Yes Yes	Yes	Yes Yes Yes	Yes Yes Yes	Yes	Yes
cg01757168 cg09406615 cg10663765 cg23855319 cg16091553 cg19676181	chr3 chr3 chr3 chr4 chr4 chr4	128215565 165555103 194014592 2062392 46126245 56660398	BCHE NAT8L GABRG1	TRUE TRUE	Island OpenSea Island Island OpenSea S_Shore		Yes Yes Yes Yes Yes		Yes Yes	Yes	Yes Yes Yes	Yes Yes Yes Yes	Yes	Yes
cg01757168 cg09406615 cg10663765 cg23855319 cg16091553 cg19676181 cg06013872	chr3 chr3 chr3 chr4 chr4 chr4 chr4	128215565 165555103 194014592 2062392 46126245 56660398 72203823	BCHE NAT8L GABRG1 SLC4A4	TRUE	Island OpenSea Island Island OpenSea S_Shore OpenSea		Yes Yes Yes Yes Yes Yes	Yes	Yes Yes	Yes Yes Yes	Yes Yes Yes	Yes Yes Yes Yes	Yes	Yes
cg01757168 cg09406615 cg10663765 cg23855319 cg16091553 cg19676181 cg06013872 cg03670393	chr3 chr3 chr3 chr4 chr4 chr4 chr4 chr4 chr4	128215565 165555103 194014592 2062392 46126245 56660398 72203823 81128690	BCHE NAT8L GABRG1 SLC4A4	TRUE TRUE TRUE TRUE	Island OpenSea Island Island OpenSea S_Shore OpenSea Island	Yes	Yes Yes Yes Yes Yes Yes Yes	Yes	Yes Yes	Yes Yes Yes	Yes Yes Yes	Yes Yes Yes Yes Yes	Yes Yes	Yes
cg01757168 cg09406615 cg10663765 cg23855319 cg16091553 cg19676181 cg06013872 cg03670393 cg04511182	chr3 chr3 chr3 chr4 chr4 chr4 chr4 chr4 chr4	128215565 165555103 194014592 2062392 46126245 56660398 72203823 81128690 103269584	BCHE NAT8L GABRG1 SLC4A4	TRUE TRUE TRUE	Island OpenSea Island Island OpenSea S_Shore OpenSea Island S_Shelf	Yes	Yes Yes Yes Yes Yes Yes Yes	Yes	Yes Yes	Yes Yes Yes	Yes Yes Yes	Yes Yes Yes Yes Yes Yes	Yes	Yes
cg01757168 cg09406615 cg10663765 cg23855319 cg16091553 cg19676181 cg06013872 cg03670393 cg04511182 cg13023646	chr3 chr3 chr3 chr4 chr4 chr4 chr4 chr4 chr4 chr4 chr4	128215565 165555103 194014592 2062392 46126245 56660398 72203823 81128690 103269584 165898835	BCHE NAT8L GABRG1 SLC4A4 TRIM61	TRUE TRUE TRUE TRUE	Island OpenSea Island Island OpenSea S_Shore OpenSea Island S_Shelf OpenSea	Yes	Yes Yes Yes Yes Yes Yes Yes Yes	Yes Yes	Yes Yes	Yes Yes Yes Yes	Yes Yes Yes	Yes Yes Yes Yes Yes Yes Yes	Yes Yes Yes	Yes
cg01757168 cg09406615 cg10663765 cg23855319 cg16091553 cg19676181 cg06013872 cg03670393 cg04511182 cg13023646 cg11685843	chr3 chr3 chr3 chr4 chr4 chr4 chr4 chr4 chr4 chr4 chr4	128215565 165555103 194014592 2062392 46126245 56660398 72203823 81128690 103269584 165898835 176349171	BCHE NAT8L GABRG1 SLC4A4 TRIM61	TRUE TRUE TRUE	Island OpenSea Island Island OpenSea S_Shore OpenSea Island S_Shelf OpenSea OpenSea	Yes	Yes Yes Yes Yes Yes Yes Yes Yes	Yes	Yes Yes Yes Yes	Yes Yes Yes Yes Yes	Yes Yes Yes	Yes Yes Yes Yes Yes Yes Yes	Yes Yes Yes	Yes
cg01757168 cg09406615 cg10663765 cg23855319 cg16091553 cg19676181 cg06013872 cg03670393 cg04511182 cg13023646 cg11685843 cg09166973	chr3 chr3 chr3 chr4 chr4 chr4 chr4 chr4 chr4 chr4 chr4	128215565 165555103 194014592 2062392 46126245 56660398 72203823 81128690 103269584 165898835 176349171 23507573	BCHE NAT8L GABRG1 SLC4A4 TRIM61 PRDM9	TRUE TRUE TRUE TRUE	Island OpenSea Island Island OpenSea S_Shore OpenSea Island S_Shelf OpenSea OpenSea OpenSea	Yes	Yes Yes Yes Yes Yes Yes Yes Yes Yes	Yes Yes Yes Yes	Yes Yes Yes Yes	Yes Yes Yes Yes Yes Yes	Yes Yes Yes	Yes Yes Yes Yes Yes Yes Yes Yes	Yes Yes Yes	Yes
cg01757168 cg09406615 cg10663765 cg23855319 cg16091553 cg19676181 cg06013872 cg03670393 cg04511182 cg13023646 cg11685843 cg09166973 cg07011961	chr3 chr3 chr3 chr4 chr4 chr4 chr4 chr4 chr4 chr4 chr4	128215565 165555103 194014592 2062392 46126245 56660398 72203823 81128690 103269584 1658988355 176349171 23507573 23507594	BCHE NAT8L GABRG1 SLC4A4 TRIM61 PRDM9 PRDM9	TRUE TRUE TRUE	Island OpenSea Island Island OpenSea S_Shore OpenSea Island S_Shelf OpenSea OpenSea OpenSea	Yes	Yes Yes Yes Yes Yes Yes Yes Yes Yes	Yes Yes Yes Yes Yes	Yes Yes Yes	Yes Yes Yes Yes Yes Yes	Yes Yes Yes Yes Yes Yes	Yes Yes Yes Yes Yes Yes Yes Yes Yes	Yes Yes Yes	Yes
cg01757168 cg09406615 cg10663765 cg23855319 cg16091553 cg19676181 cg06013872 cg03670393 cg04511182 cg13023646 cg11685843 cg09166973 cg07011961 cg25472530	chr3 chr3 chr3 chr4 chr4 chr4 chr4 chr4 chr4 chr4 chr4	128215565 165555103 194014592 2062392 46126245 56660398 72203823 81128690 103269584 165898835 176349171 23507573 23507594 23507617	BCHE NAT8L GABRG1 SLC4A4 TRIM61 PRDM9 PRDM9 PRDM9	TRUE TRUE TRUE TRUE	IslandOpenSeaIslandOpenSeaS_ShoreOpenSeaIslandS_ShelfOpenSeaOpenSeaOpenSeaOpenSeaOpenSeaOpenSeaOpenSeaOpenSeaOpenSeaOpenSeaOpenSeaOpenSeaOpenSeaOpenSeaOpenSeaOpenSeaOpenSeaOpenSea	Yes	Yes Yes Yes Yes Yes Yes Yes Yes Yes Yes	Yes Yes Yes Yes Yes Yes Yes Yes	Yes Yes Yes Yes	Yes Yes Yes Yes Yes Yes Yes	Yes Yes Yes Yes Yes Yes Yes	Yes Yes Yes Yes Yes Yes Yes Yes Yes	Yes Yes Yes	Yes

cg01667892	chr5	23507656	PRDM9		OpenSea			Yes			Yes	Yes		
cg23688479	chr5	42756851	CCDC152		OpenSea		Yes	Yes				Yes		
cg16026035	chr5	70742893			OpenSea			Yes			Yes	Yes		
cg06501366	chr5	78365687	BHMT2	TRUE	Island		Yes	Yes				Yes		
cg08328513	chr5	78365691	BHMT2;DMGDH	TRUE	Island		Yes	Yes				Yes		
cg04117076	chr5	88991074		TRUE	OpenSea			Yes		Yes		Yes		
cg14575983	chr5	99922152	FAM174A		OpenSea			Yes		Yes		Yes		
cg18665594	chr5	101119420			OpenSea		Yes		Yes			Yes		
cg19526166	chr5	110062729	TMEM232		OpenSea		Yes	Yes			Yes	Yes		
cg26817546	chr5	135538704		TRUE	OpenSea		Yes				Yes	Yes		
cg08951638	chr5	141706213		TRUE	S_Shore	Yes	Yes					Yes		
cg19731612	chr5	176559334	NSD1	TRUE	Island		Yes				Yes	Yes		
cg17493885	chr5	176559558	NSD1		Island		Yes				Yes	Yes		
cg18121224	chr5	176559563	NSD1		Island		Yes				Yes	Yes		
cg26092675	chr6	26225258	HIST1H3E		N_Shore		Yes	Yes			Yes			Yes
cg13836098	chr6	26225268	HIST1H3E		N_Shore		Yes	Yes			Yes			Yes
cg22040809	chr6	26522578	HCG11		Island	Yes					Yes	Yes		
cg17762073	chr6	34024220	GRM4	TRUE	Island			Yes			Yes	Yes		
cg20646500	chr6	42536105	UBR2		S_Shelf		Yes	Yes		Yes				
cg17097119	chr6	49917872	DEFB133		OpenSea			Yes		Yes		Yes		
cg00389785	chr6	55035173			N_Shelf		Yes	Yes		Yes		Yes		
cg18090145	chr6	67741714			OpenSea			Yes	Yes	Yes				
cg14079463	chr6	127796989	C6orf174		Island		Yes	Yes			Yes			
cg04580344	chr6	127797022	C6orf174		Island	Yes		Yes			Yes			
cg08883853	chr7	1026168	CYP2W1		Island					Yes		Yes		Yes
cg12995372	chr7	11066024	PHF14		OpenSea			Yes		Yes		Yes		
cg24231558	chr7	16626326			Island			Yes		Yes		Yes		
cg18081818	chr7	23246105			Island						Yes	Yes	Yes	
cg05407003	chr7	23246146			Island						Yes	Yes	Yes	
cg11015251	chr7	27170554	HOXA4	TRUE	Island				Yes	Yes			Yes	

cg20974609	chr7	27181671	HOXA5		N_Shore				Yes	Yes			Yes	
cg11724970	chr7	27182493	HOXA5		N_Shore				Yes	Yes			Yes	
cg05076221	chr7	27182637	HOXA5		Island				Yes	Yes			Yes	
cg23936031	chr7	27183133	HOXA5		Island				Yes	Yes			Yes	
cg04863892	chr7	27183375	HOXA5		Island				Yes	Yes			Yes	
cg19759481	chr7	27183401	HOXA5		Island				Yes	Yes			Yes	
cg02916332	chr7	27183591	HOXA5		Island				Yes	Yes			Yes	
cg17569124	chr7	27183643	HOXA5		Island				Yes	Yes			Yes	
cg02005600	chr7	27183686	HOXA5		Island				Yes	Yes			Yes	
cg25307665	chr7	27183694	HOXA5		Island				Yes	Yes			Yes	
cg14014955	chr7	27183701	HOXA5		Island				Yes	Yes			Yes	
cg02646423	chr7	27183794	HOXA5		Island				Yes	Yes			Yes	
cg20517050	chr7	27183806	HOXA5		Island				Yes	Yes			Yes	
cg23204968	chr7	27183816	HOXA5		Island				Yes	Yes			Yes	
cg20817131	chr7	27184167	HOXA5		Island				Yes	Yes			Yes	
cg25390165	chr7	27184188	HOXA5		Island				Yes	Yes			Yes	
cg14882265	chr7	27184375	HOXA5		Island				Yes	Yes			Yes	
cg17432857	chr7	27184438	HOXA5		Island	Yes			Yes	Yes			Yes	
cg00969405	chr7	27184441	HOXA5		Island				Yes	Yes			Yes	
cg03368099	chr7	27184521	HOXA5		Island				Yes	Yes			Yes	
cg27151303	chr7	27184821			Island				Yes	Yes			Yes	
cg24040595	chr7	27185512	HOXA6		Island				Yes	Yes			Yes	
cg23129930	chr7	27186993	HOXA6		Island				Yes	Yes			Yes	
cg05928186	chr7	27187102	HOXA6		Island				Yes	Yes			Yes	
cg02943305	chr7	31685368	CCDC129	TRUE	OpenSea		Yes	Yes		Yes				
cg27385590	chr7	49619841		TRUE	OpenSea		Yes					Yes	Yes	
cg26766885	chr7	96220185		TRUE	OpenSea					Yes		Yes		Yes
cg05715492	chr7	98991138	ARPC1B	TRUE	S_Shore		Yes		Yes					Yes
cg08234689	chr7	127910927			Island			Yes			Yes	Yes		
cg26209990	chr7	127911258			Island			Yes	1		Yes	Yes		

cg11688949	chr8	27115956	STMN4		OpenSea	Yes					Yes	Yes		
cg20893180	chr8	36957379		TRUE	OpenSea						Yes	Yes	Yes	
cg08620329	chr8	67454546		TRUE	Island		Yes	Yes				Yes		
cg12109823	chr8	67454595		TRUE	Island		Yes	Yes				Yes		
cg02173067	chr8	111214984			OpenSea		Yes					Yes	Yes	
cg12873476	chr8	142402728		TRUE	S_Shore		Yes	Yes			Yes			
cg23171972	chr8	143203477			S_Shore					Yes		Yes		Yes
cg13746854	chr9	34370894	KIAA1161	TRUE	Island				Yes		Yes			Yes
cg11510586	chr9	72027409		TRUE	Island		Yes		Yes			Yes		
cg14035368	chr9	139258938	CARD9;DNLZ		Island		Yes			Yes		Yes		

Gene Ontology	Gene Ontology Term	Ν	DE	P-value
GO:0007625	grooming behavior	17	3	3.80E-05
GO:2000821	regulation of grooming behavior	4	2	0.000149
GO:0097109	neuroligin family protein binding	5	2	0.000504
GO:0033130	acetylcholine receptor binding	8	2	0.0007
GO:0060484	lung-associated mesenchyme development	10	2	0.001066
GO:0060044	negative regulation of cardiac muscle cell proliferation	10	2	0.001096
GO:0018024	histone-lysine N-methyltransferase activity	43	3	0.001101
GO:0016279	protein-lysine N-methyltransferase activity	55	3	0.001883
GO:0016278	lysine N-methyltransferase activity	56	3	0.001955
GO:0042054	histone methyltransferase activity	56	3	0.00215
GO:0055022	negative regulation of cardiac muscle tissue growth	14	2	0.002192
GO:0061117	negative regulation of heart growth	14	2	0.002192
GO:0006978	DNA damage response, signal transduction by p53 class mediator resulting in transcription of p21 class mediator	16	2	0.002608
GO:0001891	phagocytic cup	19	2	0.002654
GO:0047865	dimethylglycine dehydrogenase activity	1	1	0.00278
GO:0042772	DNA damage response, signal transduction resulting in transcription	17	2	0.003039
GO:0061627	S-methylmethionine-homocysteine S-methyltransferase activity	1	1	0.003144
GO:0033477	S-methylmethionine metabolic process	1	1	0.003144
GO:0071625	vocalization behavior	15	2	0.003453
GO:0072274	metanephric glomerular basement membrane development	1	1	0.003898

Table S3 - Top gene ontology terms found to be enriched in the genes harboring probes shared by the epi-signatures of more than two conditions

N: Number of genes in the GO term; DE: number of genes that are differentially methylated

TD		D D C	,	11		ID	<u> </u>
ID	Description	BgRatio	pvalue	p.adjust	qvalue	geneID	Count
R-HSA-3214841	PKMTs methylate histone lysines*	73/10281	0.000116	0.025528	0.022841	PRDM9/SETDB1/HIST1H3E/NSD1	4
R-HSA-6804759	Regulation of TP53 Activity through Association with Co- factors	14/10281	0.001057	0.104046	0.093094	ZNF385A/TP73	2
R-HSA-3000171	Non-integrin membrane-ECM interactions	64/10281	0.001419	0.104046	0.093094	LAMB2/LAMC1/NRXN1	3
R-HSA-6794362	Protein-protein interactions at synapses	71/10281	0.001914	0.105267	0.094186	PTPRF/DLG4/NRXN1	3
R-HSA-8849932	SALM protein interactions at the synapses	21/10281	0.002401	0.105641	0.094521	PTPRF/DLG4	2
R-HSA-3000157	Laminin interactions	33/10281	0.00588	0.184787	0.165336	LAMB2/LAMC1	2
R-HSA-8874081	MET activates PTK2 signaling	33/10281	0.00588	0.184787	0.165336	LAMB2/LAMC1	2
R-HSA-168638	NOD1/2 Signaling Pathway	36/10281	0.006969	0.191659	0.171484	CARD9/MAPK11	2

Table S4 - Pathways enriched in the genes harboring probes shared by the epi-signatures of more than two conditions

*Adjusted p-value<0.05

Table S5 - Probes used	for development	of a multiclass SVM	classification model
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Probe	Chr	Position	Gene	Islands_Name	Relation_to_Island
cg27619353	chr22	38071651	LGALS1	chr22:38073037-38073412	N_Shore
cg19386379	chr11	844284	TSPAN4	chr11:842293-843396	S_Shore
cg04234016	chr12	7062109	PTPN6		OpenSea
cg12474798	chr10	64565772	ADO	chr10:64564435-64565818	Island
cg02583484	chr12	54677008	HNRNPA1	chr12:54673322-54673550	S_Shelf
cg03010887	chr16	85600235			OpenSea
cg13759905	chr2	233741920		chr2:233740668-233741879	S_Shore
cg06768599	chr14	24785488	LTB4R	chr14:24785147-24785869	Island
cg04731926	chr19	35758185	LSR	chr19:35758201-35759670	N_Shore
cg13298859	chr7	2551996	LFNG		OpenSea
cg21886367	chr14	24780825	LTB4R	chr14:24779874-24780932	Island
cg12464638	chr11	844400	TSPAN4	chr11:842293-843396	S_Shore
cg07397616	chr22	38035109	SH3BP1	chr22:38035350-38035928	N_Shore
cg18182216	chr1	150978385	FAM63A	chr1:150980735-150981092	N_Shelf
cg15095327	chr3	9944512	IL17RE		OpenSea
cg10582687	chr8	48091424			OpenSea
cg22091609	chr22	20780508	SCARF2	chr22:20779386-20780560	Island
cg21721340	chr20	62682309	SOX18	chr20:62679424-62680883	S_Shore
cg11545720	chr19	4558377	SEMA6B	chr19:4558363-4558579	Island
cg17054691	chr17	79813439	P4HB		OpenSea
cg06247837	chr17	37820135	TCAP	chr17:37823693-37824989	N_Shelf
cg05551825	chr7	5735129	RNF216		OpenSea
cg03043406	chr1	45242356	RPS8	chr1:45241013-45241900	S_Shore
cg08255475	chr16	88871329	CDT1	chr16:88871850-88872056	N_Shore
cg20978937	chr14	105399321	PLD4	chr14:105398076-105400079	Island
cg01178624	chr11	65360327	KCNK7	chr11:65359292-65360328	Island
cg00551910	chr14	70037973	C14orf162	chr14:70038108-70040302	N_Shore
cg11225330	chr1	1196747	UBE2J2		OpenSea
cg08818610	chr6	24910720	FAM65B	chr6:24910626-24911285	Island
cg13710969	chr17	79393463	BAHCC1	chr17:79393341-79393742	Island
cg24695071	chr7	2419958	EIF3B	chr7:2419808-2420340	Island
cg03173827	chr20	62688717	TCEA2	chr20:62688551-62688878	Island
cg16823042	chr12	58119992	AGAP2	chr12:58119909-58121551	Island
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cg06286618	chr16	86987702			OpenSea
cg13451000	chr11	60383256	C11orf64		OpenSea

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og02168442	chi7	23991034		chr9:102226152 102226040	Jaland
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cg1/181033		14441/53/	55555	ciii0:144410219-144410854	
cg24168538	chr4	35527016		1 10 511 550 10 511 5555	OpenSea
cg25594486	chr19	51165441	SHANKI	chr19:51165249-51165775	Island
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eg20139982 chr4 17234027 NEURL4 chr4:117847087-117847322 Island eg21399822 chr4 117847107 chr4:117847087-117847322 Island eg1716447 chr16 67233409 ELMO3 chr16:6723302-67233822 Island eg12082025 chr3 1064218 ABCA7 chr3:0162669342-62695422 Island eg12082025 chr3 113160437 WDR22 chr1:12034896-125036426 Island eg1921736 chr3 113160437 WDR52 chr3:11316029-113160641 Island eg2142221 chr6 32382997 ZBTP22 chr6:32302855-3238196 OpenSea eg23478105 chr3 19476870 chr1:19470839-9540616 S_shore eg23378105 chr17 9590256 USP43 chr1:9463829-9540616 S_shore eg2400217 chr3 23713441 chr3:1023653-10223644 S_shore eg2400217 chr3 23713441 chr3:1023654 S_shore eg2400217 chr3 23713441 chr3:1023654 S_shore <td>cg13069100</td> <td>chr1</td> <td>155035008</td> <td>ADAM15</td> <td>chr1:155034925-155036578</td> <td>Island</td>	cg13069100	chr1	155035008	ADAM15	chr1:155034925-155036578	Island
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cg01512482		1/84511/6	ZNF0/9	chi3:178430630-178431216	Island
cg02932314		42733698	C1701f104	CHT17:42753480-42734020	
cg03890037	chr5	115297989	LVRN	chr5:115297607-115298093	Island
cg04293259	chr6	30434208	G L D D G A	chr6:30434029-30434422	Island
cg09257796	chr15	27215971	GABRG3	chr15:27215951-27216856	Island
cg21429394	chr12	100750899	SLC1/A8		OpenSea
cg08513253	chr6	107955362	SOBP	chr6:10/955163-10/956693	Island
cg12277666	chrl	179560870	TDRD5	chr1:179560751-179561596	Island
cg12076931	chr9	139427171	NOTCHI	chr9:139428318-139428527	N_Shore
cg01289544	chr2	27887409	SUPT7L	chr2:27886259-27887410	Island
cg03896611	chr8	99986344		chr8:99985733-99986983	Island
cg16621833	chr6	117587588	VGLL2	chr6:117586482-117587156	S_Shore
cg20185461	chr19	18899082	COMP	chr19:18899037-18902284	Island
cg20692569	chr7	72848481	FZD9	chr7:72847933-72850032	Island
cg04626491	chr6	28584053		chr6:28583934-28584289	Island
cg23193759	chr10	71389896	C10orf35	chr10:71389862-71390355	Island
cg11807105	chr6	28784852		chr6:28781857-28782302	S_Shelf
cg01889893	chr1	8085193	ERRF11	chr1:8085554-8086854	N_Shore
cg07906855	chr3	9642468		chr3:9642387-9642695	Island
cg03308652	chr3	62354991		chr3:62354291-62355012	Island
cg07465864	chr5	112849162	YTHDC2	chr5:112849265-112849982	N_Shore
cg18457731	chr8	139734484	COL22A1		OpenSea
cg23630442	chr8	18245225		chr8:18244455-18245226	Island
cg00986580	chr14	22951241			OpenSea
cg12190994	chr7	4305061	SDK1	chr7:4303079-4305062	Island
cg09862733	chr19	49631106	PPFIA3	chr19:49631105-49631494	Island
cg13932794	chr14	100126196	HHIPL1	chr14:100125671-100126694	Island
cg16178091	chr5	37844401		chr5:37836747-37840726	S_Shelf
cg13718539	chr5	140801354	PCDHGA4	chr5:140802399-140802832	N_Shore
cg07025583	chr5	157078881	SOX30	chr5:157078327-157079564	Island
cg24411302	chr14	74893183	TMEM90A	chr14:74892466-74893184	Island
cg08602008	chr19	48076841		chr19:48076461-48076877	Island
cg14064148	chr19	10527576		chr19:10526982-10527755	Island
cg21177456	chr19	519611	C19orf20	chr19:518847-519905	Island
cg07852825	chr3	172166182	GHSR	chr3:172165372-172166738	Island
cg15962375	chr1	213563639		chr1:213562739-213562958	S_Shore
cg20810675	chr4	171604188			OpenSea
cg06358300	chr9	84302344	TLE1	chr9:84302139-84305001	Island
0	I	5.552511	l		l

cg24085707	chr17	79615652	TSPAN10	chr17:79614850-79615559	S_Shore
cg19137818	chr3	45837556	SLC6A20	chr3:45837480-45838256	Island
cg10304824	chr2	177028804	HOXD3	chr2:177029413-177029941	N_Shore
cg12050497	chr2	14773274	FAM84A	chr2:14772377-14775809	Island
cg08623383	chr1	28205926	Clorf38		OpenSea
cg15009294	chr1	214813712	CENPF		OpenSea
cg04096767	chr11	32449450	WT1	chr11:32448261-32449744	Island
cg04290586	chr12	108168987	ASCL4	chr12:108168986-108169570	Island
cg25782229	chr11	32450692	WT1	chr11:32452144-32452708	N_Shore
cg23553442	chr9	74062096		chr9:74061513-74062097	Island
cg19027255	chr10	78943401	KCNMA1		OpenSea
cg15726426	chr1	111216839	KCNA3	chr1:111216244-111217937	Island
cg10640072	chr8	99985888		chr8:99985733-99986983	Island
cg19374779	chr3	138341122	FAIM		OpenSea
cg15704225	chr2	81427138			OpenSea
cg22835435	chr19	51111185		chr19:51111184-51111519	Island
cg23713520	chr16	57318087	PLLP	chr16:57317967-57318964	Island
cg05251389	chr22	43525330	BIK		OpenSea
cg12289045	chr19	51141361	SYT3	chr19:51142284-51142986	N_Shore
cg07472704	chr13	113763758	F7	chr13:113761566-113765534	Island
cg20992319	chr2	225813194	DOCK10		OpenSea
cg14829814	chr12	108294894		chr12:108297426-108297743	N_Shelf
cg18105725	chr6	160554003	SLC22A1	chr6:160554661-160555504	N_Shore
cg07508429	chr7	44448689	NUDCD3		OpenSea
cg10424892	chr12	9602245	DDX12	chr12:9600415-9601259	S_Shore
cg01353448	chr7	31726912	C7orf16		OpenSea
cg14905403	chr17	39317349	KRTAP4-4		OpenSea
cg16119505	chr1	97186464	PTBP2	chr1:97187102-97187606	N_Shore
cg14299369	chr15	43560165	TGM5		OpenSea
cg16006141	chr1	25167475	CLIC4		OpenSea
cg10221391	chr6	29945726	HCG9	chr6:29944402-29945169	S_Shore
cg15115365	chr5	88384656			OpenSea
cg10207609	chr7	80267619	CD36		OpenSea
cg22288927	chr4	144482688		chr4:144480541-144480947	S_Shore
cg27391117	chr11	89442829	TRIM77		OpenSea
cg26825569	chr6	46704077	PLA2G7	chr6:46702736-46703316	S_Shore
cg21424209	chr6	148792600	SASH1		OpenSea
cg05753589	chr1	2064765	PRKCZ	chr1:2064628-2064855	Island
cg15136953	chr3	110793466	PVRL3	chr3:110790149-110791401	S_Shelf
cg18222192	chr11	79104769	ODZ4		OpenSea
cg00919055	chr5	149546946	CDX1	chr5:149546027-149546988	Island
cg14625770	chr9	104295276	RNF20	chr9:104295917-104296232	N_Shore
cg22109694	chr15	26360852			OpenSea
cg13629358	chr5	160112456	ATP10B		OpenSea
cg10992558					
cg10))2550	chr1	95369272	CNN3		OpenSea

Blood cell	Kabuki	ATRX	Sotos	CHARGE	Floating	ADCA-DN	Claes-	Non 7-
WB_1	0.002	0.002	0.002	0.002	0.003	0.001	0.002	0.987
WB_2	0.002	0.003	0.002	0.006	0.002	0.001	0.003	0.980
WB_3	0.003	0.004	0.002	0.002	0.002	0.001	0.003	0.983
WB_4	0.002	0.002	0.002	0.001	0.002	0.001	0.002	0.990
WB_5	0.002	0.002	0.002	0.005	0.003	0.001	0.004	0.982
WB_6	0.003	0.003	0.003	0.019	0.002	0.001	0.003	0.967
PBMC_1	0.002	0.002	0.001	0.002	0.004	0.001	0.002	0.987
PBMC_2	0.001	0.002	0.002	0.002	0.002	0.001	0.003	0.987
PBMC_3	0.002	0.001	0.001	0.001	0.002	0.000	0.001	0.991
PBMC_4	0.001	0.001	0.001	0.001	0.002	0.001	0.001	0.991
PBMC_5	0.001	0.001	0.001	0.002	0.002	0.001	0.002	0.990
PBMC_6	0.002	0.001	0.001	0.005	0.002	0.001	0.001	0.986
Gran_1	0.001	0.003	0.003	0.005	0.001	0.001	0.003	0.983
Gran_2	0.003	0.003	0.004	0.030	0.003	0.001	0.007	0.950
Gran_3	0.003	0.004	0.003	0.007	0.002	0.001	0.006	0.974
Gran_4	0.002	0.003	0.003	0.002	0.002	0.001	0.004	0.982
Gran_5	0.002	0.002	0.002	0.017	0.002	0.001	0.005	0.968
Gran_6	0.006	0.006	0.004	0.078	0.003	0.002	0.006	0.895
CD4+_1	0.001	0.001	0.001	0.001	0.002	0.001	0.001	0.992
CD4+_2	0.001	0.001	0.001	0.001	0.002	0.000	0.001	0.994
CD4+_3	0.001	0.001	0.001	0.001	0.001	0.000	0.001	0.995
CD4+_4	0.001	0.001	0.001	0.001	0.002	0.000	0.001	0.994
CD4+_5	0.001	0.001	0.000	0.001	0.003	0.001	0.001	0.993
CD4+_6	0.002	0.001	0.001	0.001	0.002	0.001	0.001	0.992
CD8+_1	0.010	0.002	0.001	0.003	0.018	0.002	0.002	0.962
CD8+_2	0.003	0.001	0.001	0.001	0.005	0.001	0.001	0.988
CD8+_3	0.003	0.001	0.001	0.001	0.004	0.001	0.001	0.988
CD8+_4	0.002	0.001	0.001	0.001	0.004	0.001	0.001	0.990
CD8+_5	0.003	0.001	0.000	0.001	0.005	0.001	0.001	0.989
CD8+_6	0.006	0.001	0.001	0.002	0.004	0.001	0.001	0.984
CD14+_1	0.001	0.004	0.003	0.007	0.002	0.001	0.003	0.979
CD14+_2	0.003	0.005	0.006	0.053	0.003	0.001	0.008	0.921
CD14+_3	0.002	0.004	0.003	0.008	0.002	0.001	0.004	0.976
CD14+_4	0.002	0.002	0.003	0.002	0.001	0.001	0.003	0.987
CD14+_5	0.002	0.002	0.002	0.015	0.002	0.001	0.005	0.971
CD14+_6	0.005	0.007	0.006	0.116	0.003	0.002	0.007	0.855
CD19+_1	0.006	0.008	0.006	0.012	0.010	0.002	0.004	0.951
CD19+_2	0.004	0.006	0.008	0.025	0.006	0.001	0.004	0.944
CD19+_3	0.027	0.009	0.009	0.015	0.016	0.002	0.007	0.916

Table S6- Prediction scores generated for methylation data from 60 samples with different blood cell types

CD19+_4	0.002	0.004	0.004	0.004	0.003	0.001	0.001	0.980
CD19+_5	0.005	0.004	0.005	0.026	0.009	0.001	0.003	0.947
CD19+_6	0.013	0.013	0.016	0.251	0.009	0.003	0.006	0.688
CD56+_1	0.004	0.002	0.002	0.002	0.010	0.001	0.002	0.978
CD56+_2	0.003	0.001	0.001	0.002	0.005	0.001	0.003	0.984
CD56+_3	0.004	0.001	0.001	0.002	0.006	0.001	0.002	0.983
CD56+_4	0.001	0.001	0.001	0.001	0.003	0.001	0.001	0.992
CD56+_5	0.001	0.001	0.001	0.001	0.003	0.001	0.001	0.992
CD56+_6	0.001	0.001	0.001	0.001	0.001	0.000	0.001	0.993
Neu_1	0.002	0.004	0.003	0.006	0.003	0.001	0.005	0.977
Neu_2	0.002	0.003	0.003	0.007	0.002	0.001	0.006	0.976
Neu_3	0.003	0.004	0.003	0.005	0.002	0.001	0.007	0.976
Neu_4	0.002	0.003	0.002	0.004	0.001	0.001	0.005	0.981
Neu_5	0.002	0.002	0.002	0.008	0.002	0.001	0.004	0.981
Neu_6	0.004	0.003	0.003	0.024	0.002	0.001	0.004	0.959
Eos_1	0.001	0.004	0.003	0.004	0.002	0.001	0.004	0.981
Eos_2	0.003	0.004	0.005	0.023	0.003	0.001	0.010	0.951
Eos_3	0.002	0.004	0.004	0.006	0.002	0.001	0.008	0.974
Eos_4	0.002	0.002	0.003	0.002	0.002	0.001	0.005	0.984
Eos_5	0.002	0.002	0.002	0.007	0.002	0.001	0.006	0.978
Eos_6	0.004	0.005	0.005	0.058	0.003	0.001	0.008	0.917

WB: Whole blood, PBMC	: Peripheral blood	l mononuclear cells,	Gran: Granulocytes,	Neu: Neutrophil, Eos	: Eosinophil
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Id	sex	age	Suspected	Screened	Mutation	ACMG Variant	DNA mothylation	Kabuki	ATRX	Sotos	CHARGE	Floating	ADCA-	Claes-	Non 7- disorder
			pnenotype	Gene		classification	prediction					narbor	DN	Jensen	uisoruer
MS1050	m	4.3	ATRX	ATRX	c.2254A>G; p.T752A	VUS		0	0	0	0	0	0	0	0.99
MS0567	m	11.5	ATRX	ATRX	c.5579A>G; p.N1860S	Benign		0.01	0.01	0	0	0	0	0	0.97
MS1020	f	2.5	CHARGE	CHD7	c.5041+5G>A	VUS	CHARGE	0.03	0.01	0.01	0.82	0.02	0.01	0.01	0.1
MS0978	f	0.8	CHARGE	CHD7	c.2788G>A, p.E930K	VUS		0	0	0	0	0.01	0	0	0.98
MS0980	m	16	CHARGE	CHD7	c.6775+1G>A	VUS	CHARGE	0.01	0.01	0.01	0.94	0.01	0.01	0.01	0
MS0981	m	3.5	CHARGE	CHD7	c.2058insGCAAAA	VUS		0.02	0	0	0.01	0	0	0	0.97
MS1028	m	4.3	CHARGE	CHD7	c.2185A>G, p.K729E	VUS	Kabuki	0.92	0.01	0.01	0.01	0.02	0.01	0.01	0.02
MS1031	f	7.5	CHARGE	CHD7	c.5405-17G>A (de novo)	Pathogenic	CHARGE	0.04	0.02	0.01	0.65	0.01	0.01	0.01	0.25
MS1034	m	0.01	CHARGE	CHD7	c.5405-17G>A	Pathogenic	CHARGE	0.02	0.02	0.01	0.89	0.01	0.01	0.01	0.02
MS0986	m	0.01	CHARGE	CHD7	c.5405-17G>A	VUS		0.02	0.04	0.01	0.2	0.01	0.01	0	0.71
MS0988	f	2.5	CHARGE	CHD7	c.3990-58insA, c.5210+58insA, c.5535- 68T>C	VUS		0.01	0	0	0.07	0	0	0	0.91
MS0990	m	10.1	CHARGE	CHD7	c.6104-22_6104-4del19	VUS	CHARGE	0.03	0.03	0.02	0.82	0.03	0.01	0.03	0.03
MS0993	f	1.5	CHARGE	CHD7	c.2957+5delG	VUS		0.02	0.01	0	0.17	0.01	0.01	0	0.79
MS1042	f	0.9	CHARGE	CHD7	c.2124T>C, p.S708S	VUS		0.01	0.01	0	0.15	0.01	0	0	0.81
MS1043	m	0.2	CHARGE	CHD7	c.1565G>T, pG522V	VUS		0	0	0	0.07	0	0	0	0.92
MS1047	m	3	CHARGE	CHD7	No mutation was found in CHD7	N/A	CHARGE	0.02	0.01	0.01	0.91	0.02	0.01	0.01	0.02
MS0994	f	13.1	CHARGE	CHD7	c.712G>A, p.V238M	VUS		0.04	0.02	0	0.01	0.01	0.01	0	0.91
MS0997	f	0.4	CHARGE	CHD7	c.5534+16T>C	VUS		0	0.01	0	0	0	0	0	0.97
MS0998	f	4.4	CHARGE	CHD7	c.2915A>G, Q972R	VUS	CHARGE	0.04	0.02	0.01	0.82	0.03	0.01	0.02	0.05
MS0999	f	0.3	CHARGE	CHD7	c.2053_2058dup6, c.5517dupT	VUS	CHARGE	0.02	0.01	0.01	0.92	0.01	0.01	0.02	0.01
MS1000	f	0.01	CHARGE	CHD7	c.2490C>A	VUS	CHARGE	0.02	0.02	0.01	0.92	0.01	0.01	0.01	0.01
MS1001	m	0.2	CHARGE	CHD7	c.4533+1G>A	Pathogenic	CHARGE	0.03	0.02	0.01	0.8	0.01	0.01	0.03	0.09
MS1006	m	0.001	CHARGE	CHD7	c.6321C>T, p.H2107H	VUS		0.01	0.01	0.01	0.18	0.01	0.01	0	0.78
MS1008	f	0.9	CHARGE	CHD7	c.6937-1G>A	Pathogenic	CHARGE	0.04	0.04	0.03	0.82	0.02	0.02	0.02	0.02
CHD7-20	f	5	CHARGE	CHD7	c.6322G>T (p.Gly2108Trp)	VUS		0.02	0.01	0	0.15	0.01	0	0	0.75

Table S7 - Probability scores and final predictions for undiagnosed cases and VUS variants

CHD7-21	f	3	CHARGE	CHD7	c.3746G>A (p.Arg1249Gln)	VUS	CHARGE	0.02	0.01	0.01	0.73	0.02	0	0.01	0.2
CHD7-22	f	0.01	CHARGE	CHD7	c2751G>A (p.Thr917Thr)	VUS		0	0	0	0.08	0	0	0	0.9
CHD7-23	m	0.5	CHARGE	CHD7	c15G>A	VUS	CHARGE	0.02	0.01	0.01	0.69	0.01	0.01	0.01	0.25
CHD7-24	m	2	CHARGE	CHD7	c.4225G>A (p.Val1409Met)	VUS	CHARGE	0.02	0.01	0.01	0.74	0.01	0	0.01	0.2
CHD7-25	m	13	CHARGE	CHD7	c.5436C>G (p.Asp1812Glu)	VUS	CHARGE	0.02	0.02	0.02	0.83	0.01	0.01	0.02	0.07
CHD7-26	m	4	CHARGE	CHD7	c.5633A>G (p.Asp1878Gly)	VUS		0	0	0	0	0	0	0	0.99
CHD7-27	m	4	CHARGE	CHD7	c.5848G>A (p.Ala1950Thr)	VUS		0.01	0	0	0.02	0	0	0	0.97
CHD7-28	m	13	CHARGE	CHD7	c.6304G>T (p.Val2102Phe)	VUS		0	0	0	0.01	0	0	0	0.99
CHD7-29	m	6	CHARGE	CHD7	c.3566G>A (p.Arg1189His)	VUS		0	0	0	0.03	0	0	0	0.95
CHD7-30	m	0.08	CHARGE	CHD7	intron4:c2238+1del	VUS	CHARGE	0.01	0.01	0.01	0.95	0	0	0.01	0
CHD7-31	m	5	CHARGE	CHD7	c.2049_2050insAAAGCA (p.Ala685_Lys686dup)	VUS		0	0	0	0	0	0	0	0.99
CHD7-32	m	9	CHARGE	CHD7	c.6377A>T (p.Asp2126Val)	VUS		0	0	0	0.04	0	0	0	0.95
CHD7-33	m		CHARGE	CHD7	c.4851T>G (p.=(p.Gly1617Gly))	VUS		0	0	0	0.02	0	0	0	0.97
CHD7-35	f		CHARGE	CHD7	c.8791 G>A (p.Val2931Met)	VUS	CHARGE	0.02	0.01	0.01	0.86	0.01	0	0	0.09
CHD7-37	m		CHARGE	CHD7	c.8802C>G (p.Ser2934Arg)	VUS		0	0	0	0.02	0	0	0	0.98
CHD7-38	m		CHARGE	CHD7	c.2516_2518delAGT (p.Gln839_Trp840delinsArg)	VUS	CHARGE	0.03	0.02	0.01	0.77	0.02	0.01	0.01	0.13
CHD7-39	f		CHARGE	CHD7	c.8759G>C (p.Gly2920Ala)	VUS		0	0	0	0	0	0	0	0.99
CHD7-42	f		CHARGE	CHD7	c.127A>G (p.Ile43Val)	VUS	CHARGE	0.01	0.01	0	0.96	0.01	0	0	0
CHD7-43	m		CHARGE	CHD7	c.1405A>G (p.Arg469Gly)	VUS		0	0	0	0	0	0	0	0.99
CHD7-44	f		CHARGE	CHD7	c.7763A>G (p.Asn2588Ser)	VUS	CHARGE	0.01	0.01	0	0.97	0	0	0.01	0
CHD7-45	m		CHARGE	CHD7	c.3871A>C (p.Lys1291Gln)	VUS	CHARGE	0.01	0.01	0.01	0.95	0.01	0	0.01	0
CHD7-48	m		CHARGE	CHD7	c.6193C>T (p.Arg2065Cys)	VUS	CHARGE	0.02	0.02	0.01	0.89	0.01	0.01	0.01	0.04
CHD7-49	f		CHARGE	CHD7	c.3762T>A (p.His1254Gln)	VUS	CHARGE	0.02	0.01	0.01	0.91	0.02	0	0.01	0.01
CHD7-50	m		CHARGE	CHD7	c.583C>T (p.Arg195Cys)	VUS		0	0	0	0.01	0	0	0	0.99
CHD7-54	f		CHARGE	CHD7	c.1562C>T (p.Pro521Leu)	VUS		0	0	0	0.03	0	0	0	0.97
CHD7-57	f		CHARGE	CHD7	c.5666-9C>G	VUS	CHARGE	0.01	0.01	0.01	0.96	0.01	0	0.01	0
CHD7-59	m		CHARGE	CHD7	c.1797_1799delGAA (p.Lys602del)	VUS		0	0	0	0	0	0	0	0.99
CHD7-62	f		CHARGE	CHD7	c.5429G>C (p.Arg1810Pro)	VUS	CHARGE	0.03	0.01	0	0.77	0.01	0.01	0.01	0.16
CHD7-65	m		CHARGE	CHD7	c.5827C>T (p.Arg1943Trp)	VUS		0.01	0	0	0.08	0	0	0.01	0.89

CHD7-66	m		CHARGE	CHD7	c.317A>G (p.His106Arg)	VUS		0.01	0	0	0.11	0	0	0	0.86
CHD7-67	m		CHARGE	CHD7	c.6529G>A (p.Gln2177Lys)	VUS		0.01	0.02	0.02	0.15	0.01	0	0.01	0.79
CHD7-70	f		CHARGE	CHD7	c.6356A>G (p.Asp2119Gly)	VUS	CHARGE	0.03	0.01	0.01	0.81	0.02	0.01	0.01	0.1
MS0694	f	54	Claes-Jensen - carrier	KDM5C	c.1510G>A; p.V504M	Pathogenic		0.03	0.01	0.01	0.03	0.02	0.01	0.1	0.8
MS0696	f	31	Claes-Jensen - carrier	KDM5C	c.1510G>A; p.V504M	Pathogenic		0.01	0.01	0	0.01	0.01	0	0.04	0.92
MS0725	f	66	Claes-Jensen - carrier	KDM5C	c.229G>A; p.A77T	Pathogenic		0.03	0.01	0.01	0.03	0.01	0.01	0.11	0.79
MS0729	f	17	Claes-Jensen - carrier	KDM5C	c.229G>A; p.A77T	Pathogenic		0.02	0.01	0.01	0.03	0.02	0	0.1	0.82
MS0730	f	51	Claes-Jensen - carrier	KDM5C	c.4439_4440DdelAG; p.fsR1481fxX9	Pathogenic		0.02	0.01	0	0.02	0.01	0	0.05	0.89
MS0731	f	39	Claes-Jensen - carrier	KDM5C	c.229G>A; p.A77T	Pathogenic		0.06	0.03	0.02	0.1	0.04	0.02	0.27	0.66
MS0732	f	55	Claes-Jensen - carrier	KDM5C	c.229G>A; p.A77T	Pathogenic		0.02	0.01	0.01	0.03	0.01	0	0.08	0.85
MS0733	f	54	Claes-Jensen - carrier	KDM5C	c.1510G>A; p.V504M	Pathogenic		0.05	0.02	0.01	0.06	0.03	0.01	0.14	0.68
KDM6A	f		Kabuki	KDM6A	c.2668_2669dupTA (p.Pro891Thrfs*8)	Pathogenic	Kabuki	0.65	0.01	0.01	0.07	0.02	0.01	0.02	0.21
MS0710	f	9.1	Kabuki	KMT2D	c.8382C>A (p.G2794G)	Likely_Benign		0	0	0	0	0	0	0	0.99
MS0712	m	10.8	Kabuki	KMT2D	c.2438C>T (p.P813L)	Benign		0	0	0	0	0	0	0	0.99
MS0714	f	14.4	Kabuki	KMT2D	c.15540G>C (p.V5180V)	Likely_Benign		0	0	0	0	0	0	0	0.99
MS0715	f	9.3	Kabuki	KMT2D	c.5868-25G>A	Likely_Benign		0.24	0.01	0	0.02	0.01	0	0.01	0.71
MS0716	f	1.3	Kabuki	KMT2D	c.16229G>A (p.G5410E)	VUS	Kabuki	0.9	0.01	0.01	0.02	0.01	0.01	0.01	0.04
MS0717	f	9.2	Kabuki	KMT2D	c.1938C>G (p.P646P)	Likely_Benign		0.07	0	0	0.01	0	0	0	0.91
MS0718	m	2.8	Kabuki	KMT2D	c.4265G>C (p.W1422S)	VUS	Kabuki	0.62	0.02	0.01	0.03	0.02	0.01	0.01	0.3
MS0734	f	3.6	Kabuki	KMT2D	c.15641G>T (p.R5214L)	VUS	Kabuki	0.9	0.01	0.01	0.01	0.02	0.01	0.01	0.04
MS0736	f	1.4	Kabuki	KMT2D	c.3907-15C>G	Likely_Benign		0.06	0.01	0	0.01	0.01	0	0.01	0.9
MS0742	m	1.2	Kabuki	KMT2D	c.11750_11758del9	VUS		0	0	0	0	0	0	0	0.98
MS0791	f	2.8	Kabuki	KMT2D	c.10741-42G>A	Likely_Benign		0	0	0	0	0	0	0	0.99
MS0797	m	13.8	Kabuki	KMT2D	c.12889T>C (p.S4297P)	VUS		0	0	0	0	0	0	0	0.99
MS0775	m	1.8	Kabuki	KMT2D	c.4143G>A (p.V1381V)	VUS		0	0	0	0	0	0	0	0.99
MS0777	m	13.4	Kabuki	KMT2D	c.401-11delC	Likely_Benign		0.02	0	0	0.01	0	0	0	0.97
MS0779	m	6.3	Kabuki	KMT2D	c.12913G>A (p.V4305I)	Likely_Benign		0	0	0	0	0	0	0	0.99
MS0780	m	1.3	Kabuki	KMT2D	c.6752C>T (p.S2251L)	Likely_Benign		0.01	0	0	0	0	0	0	0.99

MS0781	f	4.3	Kabuki	KMT2D	c.15108_15110del3 (p.H5036 E5037delinsQ)	VUS	Kabuki	0.94	0.01	0	0.01	0.01	0.01	0.01	0.01
MS0785	m	7.7	Kabuki	KMT2D	c.13644C>T (p.S4548S)	Benign		0	0	0	0.01	0	0	0	0.98
KMT2D-12	f	4.5	Kabuki	KMT2D	c.15143G>A (p.Arg5048His)	VUS	Kabuki	0.9	0.01	0.01	0.02	0.01	0.01	0.01	0.04
KMT2D-13	m	5	Kabuki	KMT2D	c.12028 T>C (p.Ser4010Pro)	VUS		0	0	0	0.01	0	0	0	0.98
KMT2D-14	f	15	Kabuki	KMT2D	c.16522-5_16522-4delTT	VUS	Kabuki	0.69	0.01	0.01	0.06	0.01	0.01	0.02	0.19
KMT2D-15	m	14	Kabuki	KMT2D	c.15910A>G (p.Ile5304Val)	VUS		0.01	0	0	0.17	0.01	0	0.01	0.79
KMT2D-16	m	17	Kabuki	KMT2D	c.15659G>A (p.Arg5220His)	VUS		0	0	0	0.02	0	0	0	0.96
KMT2D-17	m	9	Kabuki	KMT2D	c.10256A>G (p.Asp3419Gly)	VUS		0	0	0	0.01	0	0	0	0.98
KMT2D-18	f	16	Kabuki	KMT2D	c.8974G>A (p.Glu2992Lys)	VUS		0	0	0	0	0	0	0	0.99
KMT2D-19	f	8	Kabuki	KMT2D	c.8831A>G (p.Asn2944Ser)	VUS		0	0	0	0.01	0	0	0	0.98
KMT2D-20	f	10	Kabuki	KMT2D	c.832G>A (p.Ala278Thr)	VUS		0	0	0	0.01	0	0	0	0.99
KMT2D-21	m	6	Kabuki	KMT2D	c.682C>G (p.Arg228Gly)	VUS		0	0	0	0.01	0	0	0	0.98
KMT2D-23	m		Kabuki	KMT2D	c.15587T>A (p.Met5196Lys)	VUS		0	0	0.01	0.03	0	0	0	0.95
KMT2D-24	f		Kabuki	KMT2D	c.11150A>C (p.Gln3717Pro)	VUS		0	0	0	0	0	0	0	0.98
KMT2D-27	m		Kabuki	KMT2D	c.11578_11580dupCAG (p.Gln3863dup)	VUS		0	0	0	0.06	0	0	0	0.93
KMT2D-28	m		Kabuki	KMT2D	c.10909C>A (p.Pro3637Thr)	VUS		0.01	0.01	0	0.22	0	0	0	0.75
KMT2D-31	m		Kabuki	KMT2D	c.12662_12664dupAGC (p.Gln4221dup)	VUS		0	0	0	0.03	0	0	0	0.96
KMT2D-34	f		Kabuki	KMT2D	c.2334C>G (p.Cys778Trp)	VUS		0	0	0	0.1	0	0	0	0.88
KMT2D-35	f		Kabuki	KMT2D	c.14731_14733delCCT (p.Pro4911del)	VUS		0	0	0	0	0	0	0	0.99
KMT2D-37	f		Kabuki	KMT2D	c.15626G>T (p.Gly5209Val)	VUS	Kabuki	0.79	0.01	0.01	0.04	0.01	0.01	0.01	0.11
DL141677	m	18	Sotos_fibroblast	NSD1	ex20-21 del (1860-2086 aa)	Pathogenic	Sotos	0.03	0.1	0.65	0.03	0.05	0.01	0.03	0.1
51537F	f	1.6	Sotos_fibroblast	NSD1	c.1810C>T (p.Arg604*)	Pathogenic	Sotos	0.05	0.1	0.58	0.04	0.05	0.01	0.03	0.13
SF155	f		Sotos_fibroblast	NSD1	c.2362C>T (p.Arg788*)	Pathogenic	Sotos	0.06	0.1	0.62	0.04	0.04	0.01	0.03	0.1
DL208122	m	2.3	Sotos	NSD1	c.6437G>C (p.Cys2146Ser)	VUS	Sotos	0.03	0.03	0.68	0.06	0.01	0.01	0.02	0.17
DL136303	f	5	Sotos	NSD1	c.5779G>C (p.Ala1927Pro)	VUS	Sotos	0.01	0.02	0.86	0.04	0.01	0.01	0.01	0.04
DL199861	m	7	Sotos	NSD1	c.6412T>C (p.Cys2138Arg)	VUS	Sotos	0.03	0.03	0.65	0.06	0.02	0.01	0.02	0.19
DL181344	m	13	Sotos	NSD1	c.4949A>G (p.Asn1650Ser)	VUS		0	0	0	0	0	0	0	0.99
DL73286	m	10	Sotos	NSD1	c.3722G>C (p.Ser1241Thr)	VUS		0	0	0	0.01	0	0	0	0.99

DL159025	m	2.5	Sotos	NSD1	c.6013C>G (p.Arg2005Gly)	VUS	Sotos	0.01	0.02	0.9	0.02	0.01	0.01	0.01	0.03
HK-9776	f		Sotos	NSD1	c.1070A>G (p.Asn357Ser)	VUS		0.01	0	0	0.01	0	0	0	0.98
HK-12366	f		Sotos	NSD1	c.3446A>G (p.Asn1149Ser)	VUS		0	0	0	0.02	0	0	0	0.97
HK-5474	f		Sotos	NSD1	c.4817G>A (p.Cys1606Tyr)	VUS	Sotos	0.01	0.02	0.84	0.06	0.01	0.01	0.01	0.03
HK-11693	f		Sotos	NSD1	c.5177C>G (p.Pro1726Arg)	VUS	Sotos	0.01	0.02	0.9	0.02	0.01	0	0.01	0.04
HK-11767	f		Sotos	NSD1	c.5357A>G (p.Lys1786Arg)	VUS		0	0	0	0	0	0	0	0.99
HK-5581	m		Sotos	NSD1	c.5903T>C (p.Val1968Ala)	VUS		0.03	0.02	0.1	0.17	0.01	0.01	0.02	0.59
HK-3326	f		Sotos	NSD1	c.5990A>G (p.Tyr1997Cys)	VUS	Sotos	0.02	0.02	0.81	0.04	0.01	0.01	0.01	0.08
HK-435	m		Sotos	NSD1	c.6049C>T (p.Arg2017Trp)	VUS	Sotos	0.01	0.02	0.89	0.03	0.01	0.01	0.01	0.03
HK-6943	m		Sotos	NSD1	c.6674C>A (p.Pro2225Gln)	VUS		0	0	0	0.01	0	0	0	0.99
HK-14867	m		Sotos	NSD1	c.7421A>G (p.Gln2474Arg)	VUS		0	0	0	0	0	0	0	0.99
MS0436	f	40	Floating Harbor (Sex mismatch)	SRCAP	p.Arg2748*	Pathogenic	Floating Harbor	0.02	0.02	0.01	0.02	0.87	0.01	0.02	0.03

Reference sequence for all mutations is hg19, except for NSD1 mutations (hg18)