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Azithromycin to prevent post-discharge morbidity and mortality in Kenyan children: A protocol for a randomized, double-blind, placebo-controlled trial

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Complete List of Authors:	Pavlinac, Patricia; University of Washington, Global Health Singa, Benson; Kenya Medical Research Institute; Childhood Acute Illness and Nutrition Network John-Stewart, G; University of Washington, Global Health, Epidemiology, Pediatrics, Allergy and Infectious Disease Richardson, BA; University of Washington, Global Health, Biostatistics Brander, Rebecca; University of Washington, Epidemiology McGrath, Christine; University of Washington, Global Health Tickell, Kirkby; University of Washington, Global Health; Childhood Acute Illness and Nutrition Network Amondi, Mary; Kenya Medical Research Institute Rwigi, Doreen; Kenya Medical Research Institute Babigumira, Joseph; University of Washington, Global Health, Pharmacy Kariuki, Sam; Kenya Medical Research Institute Nduati, Ruth; University of Nairobi Walson, Judd; University of Washington, Department of Global Health
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Title: Azithromycin to prevent post-discharge morbidity and mortality in Kenyan children: A protocol for a randomized, double-blind, placebo-controlled trial

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Authors: Pavlinac PB¹, Singa BO^{7,8}, John-Stewart GC¹⁻⁴, Richardson BA^{1,5}, Brander RL², McGrath CJ¹, Tickell KD^{1,8}, Amondi M⁷, Rwigy D⁷, Babigumira JB^{1,6}, Kariuki S⁷, Nduati R⁹, Walson JL^{1-4,8}

Affiliations: Departments of ¹Global Health, ²Epidemiology, ³Pediatrics, ⁴Allergy and Infectious Disease, ⁵Biostatistics, ⁶Pharmacy, University of Washington, Seattle, WA, USA; ⁷Kenya Medical Research Institute, Nairobi, Kenya; ⁸The Childhood Acute Illness & Nutrition Network, Nairobi, Kenya; ⁹University of Nairobi, Nairobi, Kenya

Correspondence: Patricia Pavlinac, 325 9th Ave, Box 359931, Seattle, WA 98104 (ppav@uw.edu)

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ABSTRACT

Introduction: Child mortality due to infectious diseases remains unacceptably high in much of sub-Saharan Africa. Children who are hospitalized represent a particularly vulnerable population, both during and following hospitalization. Children being discharged from hospital represent an accessible high-risk population in which targeted use of antibiotics could offer clinical benefit.

Methods and analysis: In this randomized, double-blind, placebo-controlled trial, 1400 children aged 1 to 59 months discharged from 2 hospitals in western Kenya, in Kisii and Homa Bay, will be randomized to either a 5-day course of azithromycin or placebo to determine whether a short-course of azithromycin reduces rates of re-hospitalization and/or death in the subsequent 6-month period. The primary analysis will be modified intention-to-treat and will compare the rates of re-hospitalization or death in children treated with azithromycin or placebo using Cox proportional hazard regression. The trial will also explore mechanistic questions including the effect of a short course of azithromycin on enteric and nasopharyngeal infections and cause-specific re-hospitalizations. We will also identify clinical and host risk determinants of post-discharge morbidity and mortality. The emergence of antibiotic resistance among treated individuals and in a random subset of their primary caregivers will also be assessed and cost-effectiveness analyses performed to inform policy decisions.

Ethics and dissemination: Study procedures were reviewed and approved by the institutional review boards of the Kenya Medical Research Institute, the University of Washington, and the Kenyan Pharmacy and Poisons Board. The study is being externally monitored by Westat® and a data safety and monitoring committee has been assembled to monitor patient safety and evaluate the efficacy of the intervention. The results of this trial will be published in peer-reviewed scientific journals and presented at relevant academic conferences and to key stakeholders.

Trial registration number: NCT02414399

Key words: child mortality, antibiotic prophylaxis, post-discharge interventions

STRENGTHS AND LIMITATIONS OF THIS STUDY

- Randomized, placebo-controlled, double-blinded design and intention-to-treat analysis plan will ensure unbiased treatment effect measure
- Comprehensive data are collected, including biological specimens for all child participants and a subset of adult caregivers, for analyses of mechanisms of post-discharge morbidity and mortality, as well as assessments of antibiotic resistance and cost-effectiveness
- Results will likely be generalizable due to the limited exclusion criteria, large sample size, and multiple study sites
- Causes of death and re-hospitalization may not be accurate due to limited availability of medical records and recall bias in caregiver report
- The primary endpoint of this study is a combined outcome of re-hospitalization and death, which, while improving statistical power, may present challenges for interpretation

BACKGROUND

An estimated 3.5 million deaths occur annually in children less than 5 years of age in sub-Saharan Africa (SSA), approximately 70% of which are due to infectious causes.[1] One-year mortality rates as high as 15% have been documented following hospital discharge in SSA, a rate that is 8-fold higher than similarly-aged children in the community.[2-4] Children being discharged from hospital in SSA may represent an accessible high-risk population in which to target interventions to reduce mortality.

A single dose of azithromycin halved mortality rates in among Ethiopian children living in communities randomized to receive the antibiotic as part of a mass drug administration program.[5, 6] However, concerns about the potential for the emergence of antimicrobial resistance, possible toxicity, and feasibility of delivery are barriers to community-wide distribution of antibiotics. Targeted antimicrobial interventions, including the use of cotrimoxazole among HIV-infected children and the use of amoxicillin or cefdinir among malnourished children, have been shown to reduce mortality in these specific vulnerable populations.[7-10] Children who have been recently hospitalized are a high-risk population in which targeted azithromycin distribution may optimize benefit while reducing both individual and population level risks.

Among high-risk pediatric populations with history of recent illness, azithromycin may treat residual disease not eliminated during inpatient therapy, may provide prophylaxis against infectious exposures during a time of immune vulnerability following illness, and may reduce carriage of pathogenic organisms, including those associated with mucosal surface disruption, inflammation, and immune activation.

OBJECTIVE

The primary objective of this double-blind, placebo-controlled, randomized controlled trial (RCT) is to determine whether a 5-day course of azithromycin in children age 1 to 59 months discharged from hospital in western Kenya reduces rates of re-hospitalizations and/or death in the subsequent six months. The secondary objectives are (1) to evaluate possible mechanism(s) by which azithromycin may affect morbidity and mortality, by comparing reasons for re-hospitalization and prevalence of enteric and nasopharyngeal infections between the randomization arms; (2) to determine whether empiric administration of azithromycin at hospital discharge increases risk of antimicrobial resistance in commensal *Escherichia coli* (*E. coli*) and *Streptococcus pneumoniae* (*S. pneumoniae*) isolates from treated children and their primary caregiver; (3) to identify correlates and intermediate markers of post-discharge mortality and hospital readmission; (4) to determine the cost-effectiveness of post-discharge administration of a 5-day course of azithromycin in settings of varying antibiotic use, re-hospitalization rates, and mortality rates; and (5) to create a repository of stool, nasopharyngeal, and blood specimens from highly characterized, recently discharged children to be used to address future research questions.

METHODS

Reporting of this study protocol has been verified in accordance with the SPIRIT (Standard Protocol Items for Randomized Trials) recommendations.

Eligibility

Children age 1 to 59 months old weighing at least 2 kg and have been hospitalized and subsequently discharged and who are willing to participate will be eligible for inclusion. Children will be excluded if: azithromycin is contraindicated (children taking or prescribed other macrolide antibiotics, such as erythromycin or clarithromycin, or the protease inhibitor, lopinavir); they were admitted to hospital for a trauma, injury, or a birth defect; they do not plan to remain in the study site catchment area for at least 6 months; the legal guardian does not provide consent; or if a sibling was enrolled in the Toto Bora Trial on the same day of discharge. Caregivers of potentially eligible children must be at least 18 years of age or classified as an emancipated minor and be willing to participate in the Adult Contact Cohort if randomly selected.

Recruitment

Children will be recruited from the inpatient wards of Kisii Teaching & Referral and Homa Bay County Hospitals where study staff will accompany hospital staff on ward rounds to identify children being discharged each day. All discharged children, as determined by the onsite hospital clinicians, will be screened by study staff during working hours. If the caregiver is interested in participating and indicates consent for screening, the study staff will screen the child for eligibility, and if eligible, will obtain informed consent for study participation.

Informed consent includes an explanation of the potential risks and benefits of the study and additional provision for use of participant data and samples for future studies, and will be conducted in the language of the respondent's choosing (English, Kiswahili, Kisii, or Luo). The parent or guardian (primary caregiver) must sign written informed consent (or provide a witnessed thumbprint if not literate) prior to enrollment.

Enrollment

Children will be enrolled at the time of discharge by the clinical staff. At enrollment, primary caregivers will be interviewed to assess demographic information, medical history, and detailed contact information for the child. Medical records will also be used to abstract information from the hospitalization (including presenting diagnosis, medical management, length of stay, procedures performed, relevant medical history, physical examination, and laboratory data). All enrolled participants will undergo a physical examination performed by the study clinician, including measurement of height/length, weight, and mid-upper arm circumference (MUAC), each of which will be measured three times. The height, weight, and MUAC of the caregiver will also be collected. HIV status will be obtained from medical records or from performed testing if records are not available. Detailed home location and contact information will be collected to enable patient tracing.

Specimen collection

Specimens will be collected at enrollment (prior to study medication administration, as well as at 3 and 6-month follow-up visits). All children will also be asked to provide a whole stool for enteric pathogen identification and storage. Stool samples/swabs will be divided within one hour of collection for the following purposes: 1) placed in Cary-Blair for eventual bacterial culture (FecalSwab Cary-Blair Collection and Transport System™, Copan Diagnostics), 2) immediately tested for *Giardia* and *Cryptosporidium* using the immunoassay (Quik Chek™, Alere) and 3) placed in -80°C storage for future molecular determination of pathogen or commensal flora and markers of gut function (whole stool for these purposes will be divided into two separate vials). If a child cannot produce whole stool, two flocced rectal swabs (Pedatric FLOQswab™, Copan Diagnostics) will be collected and one placed in Cary-Blair and the other stored in -80°C for future analyses.

One flocced swab (Nylon Flocced Dry Swabs, Copan Diagnostics) nasopharyngeal swab will also be collected from all enrolled children at each time point, immediately placed in skim milk, tryptone, glucose, and glycerine (STGG) media, and frozen (-80°C) within 1 hour of collection for future *S. pneumoniae* culture. [11, 12] Primary caregivers in the Contact Cohort will also be asked to provide a stool sample (or 2 rectal swabs) and nasopharyngeal sample at each visit for testing and storage as described above.

Venous blood (up to 1 teaspoon [5mL]) will be collected from all enrolled children and caregivers enrolled in the Contact Cohort at each at each time point into EDTA tube and separated for the following purposes: 1) 0.5mL for immediate HIV-testing (if indicated according to Kenyan Ministry of Health guidelines), 2) 0.4mL for a thin malaria smear which will be stored at room temperature, 3) 0.4mL for a dried blood spot whole blood -80°C storage and eventual sickle cell testing and 4) 2-4mL for plasma and buffy coat isolation and -80°C storage.

Randomization

Block randomization (1:1) in random sized blocks of no more than 10, stratified by site, will be used. Primary randomization will include allocation to the Contact Cohorts at a ratio of 1:5 (resulting in 150 per treatment arm). Each subject will be assigned a Patient Identification Number (PID), and the randomization code linking each PID to the allocated treatment will be maintained by the University of Washington Research Pharmacy. Study participants, investigators, the study staff, hospital clinicians, and persons involved in data management or analysis will remain blinded to the allocation group during all data collection phases of the study.

Intervention

Caregivers of enrolled children will be provided a 5-day course of oral suspension formulation azithromycin (Zithromax® from Pfizer, 10 mg/kg on day 1, followed by 5mg/kg/day on days 2-5) or identically appearing and tasting placebo at discharge. Dosing ranges were determined such that a given child would never be under-dosed and not over-dosed by more than 20% that the weight-specific intended dose (Table 1). The day 1 dose will be split in half and the first half administered first by study clinician (to be observed by the

caregiver) followed by the second half administered by the caregiver under careful observation of the study staff. Caregivers will be provided with visual instructions in the language of their choosing (English, Kiswahili, Luo, Kisii).

Automated daily text message reminders will be sent for the four days following discharge and caregivers asked to respond with whether or not the child took the daily dose. The response text message will be free of charge to caregivers and caregivers will be reimbursed for each response at the final study visit. Caregivers are also asked to record each administered dose on the bottle and to return bottles at the 3 month follow-up visit.

Follow-up Procedures

All enrolled children and primary caregivers will be scheduled to return to the health facility at 3 and 6 months following enrollment to collect clinical information and samples. Anthropometric measurements will be obtained from all children and caregivers at both follow up visits (height/length, weight, MUAC) and caregivers will be asked about any hospitalizations occurring since the last time the child was seen by study staff. A flowchart of follow-up and sample collection is shown in Figure 5. Caregivers will be provided with 400KSH (approximately \$4USD) to cover the cost of their round-trip transportation.

Transportation cost will be reimbursed at each follow-up visit. If the participant does not return at their scheduled time, study staff will attempt to make contact with the primary caregiver via cell phone; if no telephone number is provided, or if the participant cannot be reached, study staff will trace the child to the household within 2 weeks of the scheduled follow-up time.

During scheduled follow-up visits, study staff will use a standardized questionnaire to ascertain history of recent illness/morbidity, post-discharge medication use including antibiotic treatment, and current condition of the child (any hospitalizations, admission and discharge date of any hospitalization, alive or dead, date of death if applicable). If caregivers report a hospitalization, causes of admission, medication administration, and length of stay will be ascertained from both caregivers and medical records, when available.

Caregivers will be encouraged, at enrollment and at each subsequent contact, to bring the child to the study health facility at any time the child is sick. Study staff will triage children to the appropriate health facility staff and will conduct a brief unscheduled visit questionnaire to ascertain adverse event information. If the unscheduled visit leads to a hospitalization, this will trigger the completion of a hospital admission form.

If at any point during follow-up a child dies, a verbal autopsy using the shortened Population Health Metrics Research Consortium questionnaire will be performed[13] If the death occurred in a hospital, data from the hospital records, including cause of death, if available, will be abstracted. If a death certificate is available, cause(s) and timing of death will be abstracted.

Final causes of re-hospitalization and death will be determined after data collection is complete by an independent adjudication committee comprised of clinicians specializing in pediatrics and infectious disease. Sources of cause of re-hospitalization (medical records and caregiver report) and causes of death (causes automatically assigned from the verbal autopsy using SmartVA-Analyze [Tariff 2.0 Method][14], hospital records, or death certificates) will be presented to the adjudication committee for final cause assignment.

Laboratory procedures and specimen collection and storage

Stool (rectal swabs), nasopharyngeal swabs and blood will be collected as described above and undergo either immediate or future laboratory testing as described in Table 2. All biological samples will be collected by staff trained in biosafety and Good Clinical Laboratory Practice (GCLP). Samples will be processed in Kenya when technology is available at one of the following laboratories: Kenya Medical Research Institute (KEMRI) (Wellcome Trust or Centre for Microbiology Research [CMR]) or at the University of Nairobi (Microbiology Department). Metagenomic analyses and/or analyses that require technology not available in

Table 1. Azithromycin dosing chart by child weight

Weight (kg)	Day 1 dose (mL)	Day 2-5 dose (mL)
2.0	0.25 x 2	0.25
2.1-2.4	0.30 x 2	0.30
2.5-2.8	0.35 x 2	0.35
2.9-3.2	0.40 x 2	0.40
3.3-3.6	0.45 x 2	0.45
3.7-4.0	0.50 x 2	0.50
4.1-4.8	0.60 x 2	0.6
4.9-5.6	0.70 x 2	0.7
5.7-6.8	0.85 x 2	0.85
6.9-8.0	1.0 x 2	1.0
8.1-9.6	1.2 x 2	1.2
9.7-11.2	1.4 x 2	1.4
11.3-13.6	1.6 x 2	1.6
13.7-16.0	2.0 x 2	2.0
16.1-19.2	2.4 x 2	2.4
19.3-23.2	2.9 x 2	2.9
23.3-25.0	3.2 x 2	3.2

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Kenya will be performed at the University of Washington. If stool culture results report *Shigella* or *Salmonella* infection, the study staff will contact the child's caregiver and encourage the caregiver to bring the child back for an evaluation and treatment if the child is symptomatic.

Table 2. Sample processing description.

Specimen Collected	Purpose	Tests Performed
Stool/ flocked rectal swabs	Bacterial ID and storage for AST	Fresh samples/rectal swabs will be cultured to identify <i>Shigella</i> , <i>Salmonella</i> , <i>Campylobacter</i> , and <i>Escherichia coli</i> using standard microbiologic methods and biochemically confirmed using bioMérieux's API [®] strips. All <i>Shigella</i> , <i>Salmonella</i> , and <i>Campylobacter</i> isolates, as well as a random subset of <i>E.coli</i> isolates will undergo antimicrobial resistance testing using disc diffusion for the following antibiotics: amoxicillin-clavulanic acid (augmentin), ampicillin, azithromycin, chloramphenicol, ciprofloxacin, ceftriaxone, ceftazidime, cefoxitin, gentamicin, imipenem, trimethoprim-sulfamethoxazole, ceftazidime/clavulanate (ESBL), cefotaxime/clavulanate (ESBL). Categorizations of susceptible, intermediate, and resistant will be determined using zone-size cut-offs outlined in CLSI interpretive standards (M100-S24 2014).
	Parasite detection	Fresh stool and rectal swabs will be tested for <i>Giardia</i> and <i>Cryptosporidium</i> using the immunoassay Giardia/ Cryptosporidium QUIK CHEK [™] .
	Storage	Stool/ flocked swabs and colonies of <i>E.coli</i> , <i>Shigella</i> spp., <i>Salmonella</i> spp., and <i>Campylobacter</i> spp. will be stored at -80°C.
Nasopharyngeal Swabs	Bacterial isolation, storage, and resistance testing	<i>Streptococcus pneumoniae</i> (<i>S. pneumoniae</i>) colonies will be isolated using standard microbiologic or molecular diagnostic protocols and susceptibility testing performed using standard microbiologic or molecular techniques. A random subset of <i>S. pneumoniae</i> isolates will undergo antimicrobial resistance testing using disc diffusion for the following antibiotics: amoxicillin-clavulanic acid (Augmentin), ampicillin, azithromycin, chloramphenicol, ciprofloxacin, ceftriaxone, ceftazidime, cefoxitin, gentamicin, imipenem, and trimethoprim-sulfamethoxazole. Categorizations of susceptible, intermediate, and resistant will be determined using zone-size cut-offs outlined in CLSI interpretive standards M100-S24 2014.
	Storage	Back-up sample and <i>S.pneumoniae</i> colonies will be will be stored at -80°C.
Blood	HIV and malaria testing	HIV testing will be performed per Kenyan National Guidelines and malaria microscopy performed using standard methods.
	Storage	Plasma and buffy coat will be stored at -80°C. Dried blood spots will be stored at room temperature.

Data Management and Confidentiality

Personal information about the participants, including medical records and data ascertained per caregiver interview, will be securely stored in files in the study offices at the study sites. Only pre-designated study staff will have access to the files. Data will be entered into an electronic database (Dacima® Electronic Data Capture) regularly by study staff. Access to the electronic database will be secured using password protected accounts for study staff. Data reports of screening, enrollment, and exclusion totals will be disseminated to the study team on a weekly basis; reports including baseline demographic characteristics, laboratory results, adherence data, and serious adverse event summaries will be distributed to study co-investigators and data monitors quarterly. Data will be regularly queried to facilitate ongoing data cleaning.

Data Analysis

Primary endpoints:

The primary study endpoint is a combined outcome of mortality and hospital readmission, as re-hospitalization is highly associated with risk of subsequent poor outcome. Re-hospitalizations that are a continuation of management from the previous hospitalization (such as elective blood transfusion) or that occur during enrollment procedures, due to a clinical deterioration post-discharge, will be excluded from the analysis. Loss to follow-up will be defined as non-attendance at both follow-up visits despite up to one month of active tracing and no clear evidence of death.

Secondary endpoints include:

1. Cause-specific re-hospitalizations assessed by questionnaire (maternal recall of diagnosis) at day 90 and day 180 follow-up visits and medical record review (discharge diagnosis). In cases when both sources are

available, information from the medical record will be considered superior. Separate analyses will be performed for each diagnosis: diarrhea, acute respiratory infection, malnutrition, or malaria.

2. Enteric pathogen carriage, operationalized as presence of a bacterial pathogen-*Shigella* species (spp.), *Campylobacter* spp., or *Salmonella* spp., or parasite- *Giardia* or *Cryptosporidium* in stool or rectal swabs assessed at day 90 and day 180 follow-up visits.
3. *Streptococcus pneumoniae* (*S. pneumoniae*) isolated from nasopharyngeal swab cultures at 90 and 180-day follow-up visits.
4. Antimicrobial resistance, specifically resistance to azithromycin, ampicillin, augmentin, trimethoprim-sulfamethoxazole, in *E.coli* and *S. pneumoniae* isolates, and presence of ESBL in *E.coli* isolates, from day 90 and day 180 samples.

To compare rates of re-hospitalization and mortality in the 6 months following hospital discharge among Kenyan children receiving 5-day azithromycin vs. placebo. Primary analyses will be modified intent-to-treat (mITT) based on randomization allocation to the 5-day course of azithromycin versus placebo. Cumulative incidence of death or first re-hospitalization will be compared between treatment groups using Cox proportional hazards regression. Participants will be censored at the date of their first re-hospitalization, or at the date of death. Median time to hospitalization-free survival will be compared between randomization groups using Kaplan-Meier (K-M) survival analysis and associated log-rank test. If the baseline assessment of randomization reveals an imbalance in characteristics between the treatment groups, we will evaluate these variables as potential confounders in a sub-analysis secondary to the mITT. Potential baseline confounders will be added stepwise in a multivariable Cox model and maintained in the model if adjustment changes the hazard ratio by more than 10%. In per protocol analyses also secondary to the mITT, we will compare treatment effects in groups defined by self-reported adherence to the 5-day course of azithromycin (5 doses vs. < 5 doses; ≥3 doses vs. <3 doses; >1 dose vs. 1 dose only). In addition, we will conduct Cox regression and K-M survival analyses for time to mortality and time to re-hospitalization as separate endpoints to understand intervention effects on these outcomes individually. The assumption of proportional hazards will be checked in all models using graphical methods including plotting a $\ln(-\ln(S(t)))$ plot for each treatment group and assessing the parallelism of the two lines and by plotting Schoenfeld residuals over time. If there is substantial missing covariate data, multiple imputation using the Markov chain Monte Carlo (MCMC) method will be used to impute covariate information. Missing outcome data (death or re-hospitalization) will not be imputed, but participants will be censored at the last follow-up visit therefore contributing some person-time to the analysis.

To evaluate possible mechanism(s) by which azithromycin may affect morbidity and mortality, by comparing reasons for re-hospitalization and change in prevalence of pathogen carriage between the randomization arms. To evaluate the association between azithromycin and the rates of cause-specific re-hospitalizations (hospitalization due to diarrhea, acute respiratory infection, malnutrition, or malaria) we will use Anderson-Gill proportional hazards modeling with previous re-hospitalizations included as time-dependent covariates in the model to capture the dependent structure of recurrence times. Because we will not have granularity in the time points other than 90-days and 6-months for assessment of pathogen carriage, we will compare the prevalence of a bacterial and parasitic pathogens (*Shigella*, *Salmonella*, *Campylobacter*, *Cryptosporidium*, *Giardia*) at 90-days and 6 months by randomization arm using generalized estimating equations (GEE) with a Poisson link, exchangeable correlation structure, and will adjust for baseline presence of a bacterial pathogen. To determine whether an observed association between the intervention and pathogen carriage wanes over time, we will test the hypothesis that the prevalence ratios comparing carriage in intervention arms are the same at the two follow-up time points using a chi-squared test.

To determine whether empiric administration of azithromycin at hospital discharge increases risk of antimicrobial resistance in commensal *E. coli* and pneumococcal isolates from treated children and their household contacts. Among children and adult household contacts in whom commensal *E. coli* and/or *S. pneumoniae* are isolated, we will compare the proportion of isolates resistant to azithromycin, ampicillin, augmentin, and trimethoprim-sulfamethoxazole, between randomization arms and Contact Cohorts for each arm, at 90-days and 6 months using GEE with a Poisson link and exchangeable correlation structure. A chi-squared test will be used to determine whether the association between intervention arm and antimicrobial resistance wanes over time. Because the likelihood of having a bacterial pathogen isolated may depend on baseline factors, including intervention arm, we will conduct secondary analyses utilizing propensity scores to

account for the potential differential likelihood of having antimicrobial susceptibility testing performed, which will allow us to make inference to the entire study population and their contacts.

To identify correlates and intermediate markers of post-discharge mortality and hospital-readmission among hospitalized children. Enrollment hospital admission diagnosis, indicators of malnutrition, age, HIV-exposure and HIV-infection status, sickle cell anemia, and randomization arm will be assessed in a multivariable Cox regression model to identify correlates of the primary endpoint of death and/or hospital-readmission independent of the treatment effect. In addition, Cox regression models will also be built for correlates of mortality and correlates of re-admission individually to understand distinct cofactors for each of these outcomes.

To determine the cost-effectiveness of post-discharge administration of a 5-day course of azithromycin in settings of varying antibiotic use, re-hospitalization rates, and mortality rates. Costs analysis: We will assess the costs of all supplies, services and equipment necessary to implement the intervention (direct medical costs). The perspective will be that of the healthcare provider, i.e. Kenya's Ministry of Health. Using WHO guidelines and its ingredients approach, we will quantify the resources and associated unit costs required to deliver a 5-day course of azithromycin, organized in standard expenditure categories: personnel (salaries), supplies including drugs, equipment, services, space and overhead. We will also measure the costs of severe child hospitalizations, the costs for the different types of personnel employed (e.g. nurses/doctors) and the time demanded from them for conducting the intervention.[15] When data are missing, they will be complemented by data extracted from the literature and other available sources. Full incremental costs will be derived, with estimation of the potential healthcare cost-offset realized in avoiding severe hospitalizations. Costs will be measured in local currency (Kenyan Shilling) and converted into US\$. Our main metric will be cost per child treated. Cost-effectiveness analysis (CEA): we will develop a CEA mathematical model, and estimate incremental costs and cost-effectiveness for implementation of the intervention. The model will include two components: costs (described immediately above) and health benefits. The study will provide clinical outcomes (mortality/morbidity) over a 6-month follow-up period. Subsequently, deaths averted, life-years saved and disability-adjusted life years (DALYs) averted by the intervention will be estimated. We will estimate: a) incremental costs and b) incremental cost-effectiveness of the intervention vs. status quo. *Incremental costs* are the net sum of the costs to implement the intervention compared with status quo, and the costs averted due to the decrease in severe child hospitalizations. *Incremental cost-effectiveness* ratios (ICERs) will be estimated as cost per death averted, cost per life-year saved, and cost per DALY averted. We will use recent estimates of disability weights for estimation of DALYs.[16, 17] Short-term (over study follow-up i.e. 6 months) and longer-term time horizons (extrapolated to 1, 5, and 10 years) will be used. DALYs and costs will be discounted at 3% per year, consistent with CEA guidelines (undiscounted results will also be presented). Sources of uncertainty in the results will be explored in univariate and probabilistic sensitivity analysis. [18, 19] Finally, we will compare our findings to CEA estimates for other health interventions in sub-Saharan Africa.[20, 21]

Data and Safety Monitoring

A Data Safety and Monitoring Committee (DSMC) was established at study initiation to monitor severe adverse events (SAEs) and to evaluate the statistical analysis plan and associated stopping rules. The DSMC includes expertise in clinical trials, statistics, child mortality assessment, ethics, and pediatric care in resource limited settings. Adverse events will be monitored by the DSMC. Monthly adverse event summaries will be sent to the DSMC safety officer and individual-child SAE forms, which include detailed medication history to evaluate possible drug interactions, will be sent to the safety officer per request. Each SAE will be assigned the plausibility of relatedness to study drug by study PIs. The data will not be presented by intervention group unless requested by DSMB safety officer. These reports will be descriptive (no statistical analyses). The DSMC will make recommendations regarding any imbalances in safety outcomes.

A single interim analysis for re-hospitalization-free survival will be prepared by the study statistician using O'Brien-Fleming boundaries for benefit and harm when 50% of expected person time (350 child-years) has been accrued. Assuming 157 events will be available at half of the person-time accrual, a z-score critical value of 2.797, or p -value < 0.005 , from a Kaplan Meier log-rank test will determine the cut-off of statistical significance. A symmetric boundary will be used for benefit and harm. The DSMC will review this analysis and make a determination about study continuation. Futility will not be a basis for stopping rules because of the trials' value in understanding mechanisms of post-discharge worsening and antibiotic resistance. Assuming the

DSMC decides to continue the trial after the interim analysis, an alpha of 0.045 will be used as the statistical significance boundary at the final analysis.

Statistical Power

To compare rates of re-hospitalization and mortality in the 6 months following hospital discharge among Kenyan children receiving 5-day azithromycin vs. placebo. The total sample size required was calculated for the primary endpoint of time to death or hospital re-admission within the 6 month post-discharge period, assuming an alpha level of 0.05, power of 0.80, and a ratio of treatment to placebo random assignment of 1:1. In SSA, it is estimated that 2-15% of children aged less than 5 years died within 6 months of hospital discharge and 15.5% of children who survived discharge from the district hospital were re-admitted with the same diagnosis within 6-months[2, 4, 22] Assuming that an additional 5-10% of children are re-admitted for other conditions, we expect that re-hospitalizations will occur in 20.5 to 30.5% of children enrolled in the study. Combined with our expected fatality rate (2-

15%), we expect the cumulative incidence of the combined endpoint to range from 22.5 to 45.5%.[22] Based on a previous trial of mass drug administration of a single dose of azithromycin in which a single dose of the antibiotic was associated with a 49% reduction in risk of death, we calculated sample sizes using estimates of reduction in risk ranging from 30-50% with the cumulative incidence range of 22.5 to 45.5%, and found the sample size required ranged from 90 to 550 children per treatment arm.[5] Using the most conservative estimates of a hazard ratio of 0.70 and 22.5% prevalence of re-admission arm) to achieve adequate power. We will recruit to follow-up, resulting in a total planned enrollment considering mortality alone, and estimated minimum detectable hazard ratios ≤ 0.5 for mortality rates of $\geq 8\%$ and

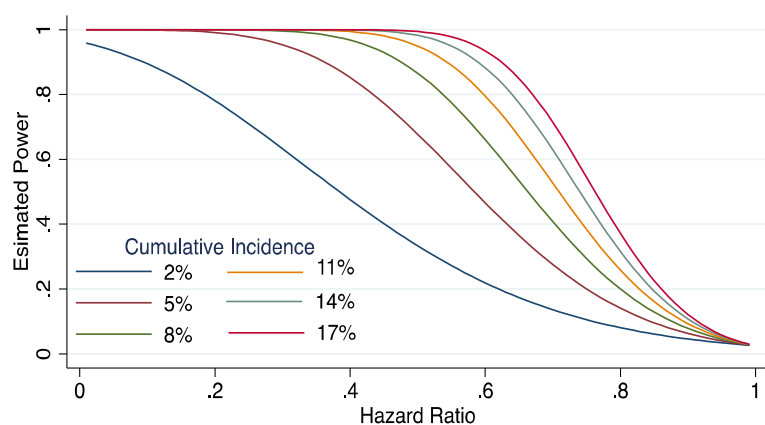
To evaluate possible mechanism(s) by

comparing reasons for re-hospitalization and prevalence of pathogen carriage between the randomization arms. We calculated the minimum detectable association between treatment arm and cause-specific re-hospitalizations (hospitalization due to diarrhea, acute respiratory infection, malnutrition, or malaria vs. any other) among enrolled children, assuming an alpha level of 0.05, power of 0.80, and a ratio of treatment to placebo of 1:1. Based on data from Kenya, re-hospitalization rates due to specific causes ranged from approximately 0.5% to 5.7% in the 6 month post-discharge period.[4] By not conditioning on the child having the same diagnosis as the initial hospitalization, we expect the cumulative incidence of cause-specific re-hospitalizations to range from 2.5% to 10%. With this range of outcome rates, we will be able to detect hazard ratios of 0.48 to 0.70 for the effect of azithromycin on specific severe morbidities.

We expect 56% of children in the placebo group to have *S. pneumoniae* isolated from nasopharyngeal swabs, providing $\geq 80\%$ power to detect a prevalence ratio of 0.85 (or 1.15) between the two treatment arms at each time point.[23-25] Based on prevalences of *Shigella*, *Salmonella*, *Campylobacter*, *Cryptosporidium*, *Giardia* among asymptomatic children in Western Kenya, we expect 10% of children in the placebo group to have a bacterial pathogen isolated at each time point, resulting in $\geq 80\%$ power to detect differences in enteric pathogen prevalences of 0.67 (1.49) at each time point.[26]

To determine whether empiric administration of azithromycin at hospital discharge increases risk of antimicrobial resistance in commensal *E. coli* and pneumococcal isolates from treated children and their household contacts. We will select a random selection of 400 *E. coli* and 400 *S. pneumoniae* isolates (200 per arm) for β -lactam and macrolide resistance testing at each timepoint. We will also store all *S. pneumoniae*, *E. coli* isolates and other isolated bacteria from stool for potential future testing in the event that resistance prevalence is lower than expected. As shown in Table 3, we will have $> 80\%$ power to detect prevalence ratios

Figure 1 Power and detectable hazard ratios given a range of mortality rates



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> 1.1, with an ability to detect the smallest effect sizes when the prevalence of resistance in the placebo group is highest.

We will enroll 300 adults in the Contact Cohort for *E. coli* and *S. pneumoniae* isolation. We expect *E. coli* to be isolated from all adults and *S. pneumoniae* isolated from between 5-55%. [23, 27, 28] Assuming an alpha of .05, a 1:1 ratio of testable isolates, and a prevalence of resistance of 50% in the placebo arm, we will have 80% power to detect a 1.4-fold higher prevalence to 1.9-fold higher resistance prevalence in the contacts of azithromycin-treated children.

To identify correlates and intermediate markers of post-discharge mortality and hospital-readmission among hospitalized Kenyan children. Conservatively estimating a 20% loss-to-follow-up rate in the RCT and a cumulative incidence of death or re-hospitalization of 22.5%, we will have >80% power to detect hazard ratios ≥ 1.3 between correlates and the outcome with exposure prevalences of $\geq 20\%$ or more and hazard ratios ≥ 1.5 for exposure prevalences $< 20\%$.

Study timeline

The trial began on June 28, 2016 and participant recruitment and follow-up will continue over a 36-month period, with anticipated final follow-up visit(s) occurring in June 2019. Primary analyses will be complete by February 2020.

Potential Challenges and Limitations

In order to ensure adequate power to detect a discernable clinically relevant difference between study groups in the primary outcome, we have combined hospital readmission with death. Preliminary studies suggest that sufficient numbers of children will reach this combined outcome. However, we have incorporated an interim analysis by the DSMB to review the accrued data and an adapted sample size could be considered if the combined event frequency is less than predicted. It is possible that since most children receive antibiotics during hospitalization, the benefit anticipated with the use of azithromycin based on previous trials of mass drug administration will not be observed. However, most hospitalized children are treated with penicillins, cephalosporins, gentamicin, or cotrimoxazole while in hospital and the broad spectrum of activity (including malaria prevention) and long half-life of azithromycin suggest that there may be additive treatment and/or prophylactic benefit. Interim analysis will allow us to determine whether children receiving specific agents during inpatient treatment are less likely to benefit and will allow us to adapt our study design, sample size, or approach if necessary. After discharge, it is difficult to ensure adherence with the full 5-day treatment course. We will measure adherence using three different measures (text message responses, bottle check boxes, and caregiver-report at follow-up visits). In addition, the mortality benefit of azithromycin observed in Ethiopia was from a single dose and in this study the first dose will be directly observed. [5] While relying on caregiver report of mortality and morbidity may lead to bias due to outcome misclassification, this misclassification should not differ between randomization arms and therefore will be non-differential. Further hospital records will be used when available to determine diagnoses. Finally, resistance prevalence may be lower than predicted, limiting power to detect clinically relevant differences in resistance prevalence between the intervention arms. We will store all isolates in the event that a greater number of isolates are needed for antimicrobial resistance testing.

Regulatory Authorities

This study has received IRB approval by the University of Washington Human Subjects Division (HSD), KEMRI Scientific and Ethics Review Unit (SERU), and the Kenya Pharmacy and Poisons Board. The clinical trial is also registered with clinicaltrials.gov (NCT02414399). Any modifications to the study protocol or consent materials will be submitted for approval all regulatory authorities before implementation. Westat® will provide external clinical, pharmacy, and laboratory monitoring.

Dissemination

Table 3. Power (%) to detect prevalence ratios of macrolide and β -lactamase resistance in 200 *E. coli* and 200 *S. pneumoniae* isolates per treatment group

Resistance Prevalence (%)	Resistance prevalence (%) in placebo group						
	10	20	30	40	50	60	70
10							
20	80						
30	>99	64					
40	>99	99	55				
50	>99	>99	98	48			
60	>99	>99	>99	>99	52		
70	>99	>99	>99	>99	98	55	
80	>99	>99	>99	>99	>99	99	64

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Results of this study will be disseminated by publication in a peer-reviewed scientific journal, presented at relevant academic conferences, and amongst participating partners and health facilities in Kenya.

Author's contributions

JLW, PBP, GJS, BAR, BOS, and RN conceived of this trial and developed of this study protocol. JLW and BOS are study co-Principal Investigators and PBP is the Project Director; BAR oversaw the statistical analyses plans; JBB developed the CEA plan; KDT developed procedures for ascertaining and reporting SAEs; CJM developed procedures related to blood specimen procedures and drug adherence measurement. GJS, CJM, RN, and PBP provide scientific expertise. RLB and MA are involved in collection and management of the data. MA and PBP coordinate and oversee implementation of all clinical study procedures and SK with assistance from DR oversees all laboratory procedures. All authors contributed to the development of this manuscript and/or study procedures, and to reading and approving the final version for publication.

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Competing interests statement

None of the authors or study co-investigators have any competing interests to declare.

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SPIRIT 2013 Checklist: Recommended items to address in a clinical trial protocol and related documents*

Section/item	Item No	Description	Addressed on page number
Administrative information			
Title	1	Descriptive title identifying the study design, population, interventions, and, if applicable, trial acronym	_____
Trial registration	2a	Trial identifier and registry name. If not yet registered, name of intended registry	_____
	2b	All items from the World Health Organization Trial Registration Data Set	_____
Protocol version	3	Date and version identifier	_____
Funding	4	Sources and types of financial, material, and other support	_____
Roles and responsibilities	5a	Names, affiliations, and roles of protocol contributors	_____
	5b	Name and contact information for the trial sponsor	_____
	5c	Role of study sponsor and funders, if any, in study design; collection, management, analysis, and interpretation of data; writing of the report; and the decision to submit the report for publication, including whether they will have ultimate authority over any of these activities	_____
	5d	Composition, roles, and responsibilities of the coordinating centre, steering committee, endpoint adjudication committee, data management team, and other individuals or groups overseeing the trial, if applicable (see Item 21a for data monitoring committee)	_____

1	Introduction						
2							
3	Background and rationale	6a	Description of research question and justification for undertaking the trial, including summary of relevant studies (published and unpublished) examining benefits and harms for each intervention	_____			
4							
5							
6		6b	Explanation for choice of comparators	_____			
7							
8	Objectives	7	Specific objectives or hypotheses	_____			
9							
10	Trial design	8	Description of trial design including type of trial (eg, parallel group, crossover, factorial, single group), allocation ratio, and framework (eg, superiority, equivalence, noninferiority, exploratory)	_____			
11							
12							
13							
14	Methods: Participants, interventions, and outcomes						
15							
16	Study setting	9	Description of study settings (eg, community clinic, academic hospital) and list of countries where data will be collected. Reference to where list of study sites can be obtained	_____			
17							
18							
19	Eligibility criteria	10	Inclusion and exclusion criteria for participants. If applicable, eligibility criteria for study centres and individuals who will perform the interventions (eg, surgeons, psychotherapists)	_____			
20							
21							
22	Interventions	11a	Interventions for each group with sufficient detail to allow replication, including how and when they will be administered	_____			
23							
24							
25							
26		11b	Criteria for discontinuing or modifying allocated interventions for a given trial participant (eg, drug dose change in response to harms, participant request, or improving/worsening disease)	_____			
27							
28							
29		11c	Strategies to improve adherence to intervention protocols, and any procedures for monitoring adherence (eg, drug tablet return, laboratory tests)	_____			
30							
31							
32		11d	Relevant concomitant care and interventions that are permitted or prohibited during the trial	_____			
33							
34	Outcomes	12	Primary, secondary, and other outcomes, including the specific measurement variable (eg, systolic blood pressure), analysis metric (eg, change from baseline, final value, time to event), method of aggregation (eg, median, proportion), and time point for each outcome. Explanation of the clinical relevance of chosen efficacy and harm outcomes is strongly recommended	_____			
35							
36							
37							
38							
39	Participant timeline	13	Time schedule of enrolment, interventions (including any run-ins and washouts), assessments, and visits for participants. A schematic diagram is highly recommended (see Figure)	_____			
40							
41							
42							

1 Sample size 14 Estimated number of participants needed to achieve study objectives and how it was determined, including _____
 2 clinical and statistical assumptions supporting any sample size calculations

3
 4 Recruitment 15 Strategies for achieving adequate participant enrolment to reach target sample size _____
 5

6 **Methods: Assignment of interventions (for controlled trials)**

7 Allocation:

8
 9
 10 Sequence 16a Method of generating the allocation sequence (eg, computer-generated random numbers), and list of any _____
 11 generation factors for stratification. To reduce predictability of a random sequence, details of any planned restriction
 12 (eg, blocking) should be provided in a separate document that is unavailable to those who enrol participants
 13 or assign interventions
 14

15
 16 Allocation 16b Mechanism of implementing the allocation sequence (eg, central telephone; sequentially numbered, _____
 17 concealment opaque, sealed envelopes), describing any steps to conceal the sequence until interventions are assigned
 18 mechanism
 19

20
 21 Implementation 16c Who will generate the allocation sequence, who will enrol participants, and who will assign participants to _____
 22 interventions
 23

24 Blinding (masking) 17a Who will be blinded after assignment to interventions (eg, trial participants, care providers, outcome _____
 25 assessors, data analysts), and how
 26

27 17b If blinded, circumstances under which unblinding is permissible, and procedure for revealing a participant's _____
 28 allocated intervention during the trial
 29
 30

31 **Methods: Data collection, management, and analysis**

32
 33 Data collection 18a Plans for assessment and collection of outcome, baseline, and other trial data, including any related _____
 34 methods processes to promote data quality (eg, duplicate measurements, training of assessors) and a description of
 35 study instruments (eg, questionnaires, laboratory tests) along with their reliability and validity, if known.
 36 Reference to where data collection forms can be found, if not in the protocol
 37

38
 39 18b Plans to promote participant retention and complete follow-up, including list of any outcome data to be _____
 40 collected for participants who discontinue or deviate from intervention protocols
 41
 42

1	Data management	19	Plans for data entry, coding, security, and storage, including any related processes to promote data quality (eg, double data entry; range checks for data values). Reference to where details of data management procedures can be found, if not in the protocol	_____
2				
3				
4				
5	Statistical methods	20a	Statistical methods for analysing primary and secondary outcomes. Reference to where other details of the statistical analysis plan can be found, if not in the protocol	_____
6				
7				
8		20b	Methods for any additional analyses (eg, subgroup and adjusted analyses)	_____
9				
10		20c	Definition of analysis population relating to protocol non-adherence (eg, as randomised analysis), and any statistical methods to handle missing data (eg, multiple imputation)	_____
11				
12				
13				
14	Methods: Monitoring			
15				
16	Data monitoring	21a	Composition of data monitoring committee (DMC); summary of its role and reporting structure; statement of whether it is independent from the sponsor and competing interests; and reference to where further details about its charter can be found, if not in the protocol. Alternatively, an explanation of why a DMC is not needed	_____
17				
18				
19				
20				
21		21b	Description of any interim analyses and stopping guidelines, including who will have access to these interim results and make the final decision to terminate the trial	_____
22				
23				
24				
25	Harms	22	Plans for collecting, assessing, reporting, and managing solicited and spontaneously reported adverse events and other unintended effects of trial interventions or trial conduct	_____
26				
27				
28	Auditing	23	Frequency and procedures for auditing trial conduct, if any, and whether the process will be independent from investigators and the sponsor	_____
29				
30				
31				
32	Ethics and dissemination			
33				
34	Research ethics approval	24	Plans for seeking research ethics committee/institutional review board (REC/IRB) approval	_____
35				
36				
37	Protocol amendments	25	Plans for communicating important protocol modifications (eg, changes to eligibility criteria, outcomes, analyses) to relevant parties (eg, investigators, REC/IRBs, trial participants, trial registries, journals, regulators)	_____
38				
39				
40				
41				
42				

1	Consent or assent	26a	Who will obtain informed consent or assent from potential trial participants or authorised surrogates, and	_____
2			how (see Item 32)	
3				
4		26b	Additional consent provisions for collection and use of participant data and biological specimens in ancillary	_____
5			studies, if applicable	
6				
7	Confidentiality	27	How personal information about potential and enrolled participants will be collected, shared, and maintained	_____
8			in order to protect confidentiality before, during, and after the trial	
9				
10	Declaration of	28	Financial and other competing interests for principal investigators for the overall trial and each study site	_____
11	interests			
12				
13	Access to data	29	Statement of who will have access to the final trial dataset, and disclosure of contractual agreements that	_____
14			limit such access for investigators	
15				
16	Ancillary and post-	30	Provisions, if any, for ancillary and post-trial care, and for compensation to those who suffer harm from trial	_____
17	trial care		participation	
18				
19				
20	Dissemination policy	31a	Plans for investigators and sponsor to communicate trial results to participants, healthcare professionals,	_____
21			the public, and other relevant groups (eg, via publication, reporting in results databases, or other data	
22			sharing arrangements), including any publication restrictions	
23				
24		31b	Authorship eligibility guidelines and any intended use of professional writers	_____
25				
26		31c	Plans, if any, for granting public access to the full protocol, participant-level dataset, and statistical code	_____
27				
28				
29	Appendices			
30				
31	Informed consent	32	Model consent form and other related documentation given to participants and authorised surrogates	_____
32	materials			
33				
34	Biological	33	Plans for collection, laboratory evaluation, and storage of biological specimens for genetic or molecular	_____
35	specimens		analysis in the current trial and for future use in ancillary studies, if applicable	
36				

37
38 *It is strongly recommended that this checklist be read in conjunction with the SPIRIT 2013 Explanation & Elaboration for important clarification on the items.
39 Amendments to the protocol should be tracked and dated. The SPIRIT checklist is copyrighted by the SPIRIT Group under the Creative Commons
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41
42

BMJ Open

Azithromycin to prevent post-discharge morbidity and mortality in Kenyan children: A protocol for a randomized, double-blind, placebo-controlled trial (the Toto Bora trial)

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Complete List of Authors:	<p>Pavlinac, Patricia; University of Washington, Global Health Singa, Benson; Kenya Medical Research Institute; Childhood Acute Illness and Nutrition Network John-Stewart, G; University of Washington, Global Health, Epidemiology, Pediatrics, Allergy and Infectious Disease Richardson, BA; University of Washington, Global Health, Biostatistics Brander, Rebecca; University of Washington, Epidemiology McGrath, Christine; University of Washington, Global Health Tickell, Kirkby; University of Washington, Global Health; Childhood Acute Illness and Nutrition Network Amondi, Mary; Kenya Medical Research Institute Rwigi, Doreen; Kenya Medical Research Institute Babigumira, Joseph; University of Washington, Global Health, Pharmacy Kariuki, Sam; Kenya Medical Research Institute Nduati, Ruth; University of Nairobi Walson, Judd; University of Washington, Department of Global Health</p>
Primary Subject Heading:	Global health
Secondary Subject Heading:	Paediatrics, Infectious diseases
Keywords:	Child mortality, Post-discharge interventions, Toto Bora Trial, Targeted empiric antibiotic therapy

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Manuscripts

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Title: Azithromycin to prevent post-discharge morbidity and mortality in Kenyan children: A protocol for a randomized, double-blind, placebo-controlled trial (the Toto Bora trial)

Running head: AZM to prevent post-discharge morbidity and mortality

Authors: Pavlinac PB¹, Singa BO^{7,8}, John-Stewart GC¹⁻⁴, Richardson BA^{1,5}, Brander RL², McGrath CJ¹, Tickell KD^{1,8}, Amondi M⁷, Rwigy D⁷, Babigumira JB^{1,6}, Kariuki S⁷, Nduati R⁹, Walson JL^{1-4,8}

Affiliations: Departments of ¹Global Health, ²Epidemiology, ³Pediatrics, ⁴Allergy and Infectious Disease, ⁵Biostatistics, ⁶Pharmacy, University of Washington, Seattle, WA, USA; ⁷Kenya Medical Research Institute, Nairobi, Kenya; ⁸The Childhood Acute Illness & Nutrition Network, Nairobi, Kenya; ⁹University of Nairobi, Nairobi, Kenya

Correspondence: Patricia Pavlinac, 325 9th Ave, Box 359931, Seattle, WA 98104 (ppav@uw.edu)

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ABSTRACT

Introduction: Child mortality due to infectious diseases remains unacceptably high in much of sub-Saharan Africa. Children who are hospitalized represent an accessible population at particularly high-risk of death, both during and following hospitalization. Hospital discharge may be a critical time point at which targeted use of antibiotics could reduce morbidity and mortality in high-risk children.

Methods and analysis: In this randomized, double-blind, placebo-controlled trial (Toto Bora), 1400 children aged 1 to 59 months discharged from 2 hospitals in western Kenya, in Kisii and Homa Bay, will be randomized to either a 5-day course of azithromycin or placebo to determine whether a short-course of azithromycin reduces rates of re-hospitalization and/or death in the subsequent 6-month period. The primary analysis will be modified intention-to-treat and will compare the rates of re-hospitalization or death in children treated with azithromycin or placebo using Cox proportional hazard regression. The trial will also evaluate the effect of a short course of azithromycin on enteric and nasopharyngeal infections and cause-specific morbidities. We will also identify risk factors for post-discharge morbidity and mortality and subpopulations most likely to benefit from post-discharge antibiotic use. Antibiotic resistance in *Escherichia coli* and *Streptococcus pneumoniae* among enrolled children and their primary caregivers will also be assessed and cost-effectiveness analyses performed to inform policy decisions.

Ethics and dissemination: Study procedures were reviewed and approved by the institutional review boards of the Kenya Medical Research Institute, the University of Washington, and the Kenyan Pharmacy and Poisons Board. The study is being externally monitored and a data safety and monitoring committee has been assembled to monitor patient safety and to evaluate the efficacy of the intervention. The results of this trial will be published in peer-reviewed scientific journals and presented at relevant academic conferences and to key stakeholders.

Trial registration number: NCT02414399

Key words: child mortality, Toto Bora, targeted empiric antibiotic therapy, post-discharge interventions

STRENGTHS AND LIMITATIONS OF THIS STUDY

Strengths

- Randomized, placebo-controlled, double-blinded design and modified intention-to-treat analysis will ensure unbiased treatment effect measure
- Comprehensive data are collected, including biological specimens for all child participants and a subset of adult caregivers, for analyses of mechanisms of post-discharge morbidity and mortality, subsets of children most likely to benefit from the antibiotic, as well as assessments of antibiotic resistance and cost-effectiveness
- Results will likely be generalizable due to the limited exclusion criteria, large sample size, and multiple study sites
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Limitations

- Causes of death and re-hospitalization may be misclassified due to limited availability of medical records and recall bias in caregiver report
- The primary endpoint of this study is a combined outcome of re-hospitalization and death, which, while improving statistical power, may present challenges for interpretation
- Children in both intervention arms may receive other antibiotics over the course of follow-up

BACKGROUND

An estimated 3.5 million deaths occur annually in children less than 5 years of age in sub-Saharan Africa (SSA), approximately 70% of which are due to infectious causes. Children who were recently hospitalized have mortality rates 6 to 8-fold higher than similarly-aged children from the same community.¹⁻³ Post-discharge mortality rates as high as 15% have been documented in the 12 months following discharge, with mortality risk remaining elevated up to two years post-discharge.⁴⁻⁸ Children who are very young, malnourished, or HIV-infected are at particularly high risk of post-discharge mortality within the 3 months following discharge.^{1-4 6-8} Children being discharged from hospital in SSA may represent an accessible high-risk population in which to target interventions to reduce mortality and morbidity.

Targeted antibiotic interventions, including the use of cotrimoxazole among HIV-infected children and the use of amoxicillin or cefdinir among children with severe acute malnutrition (SAM), have been shown to reduce morbidity and mortality in these specific vulnerable populations.⁹⁻¹² Other trials of targeted antibiotic use in vulnerable populations, including cotrimoxazole in HIV-exposed uninfected (HEU) children and in children with SAM, have failed to demonstrate a mortality benefit.^{13 14} In contrast, non-targeted mass drug administration of a single dose of azithromycin halved mortality rates among Ethiopian children living in communities randomized to receive the antibiotic.^{15 16} Concerns about the potential emergence of antibiotic resistance, possible toxicity, and feasibility of delivery are barriers to the non-targeted antibiotic distribution strategies.

A short-course of azithromycin given to children with recent severe illness being discharged from hospital may optimize benefit while reducing both individual and population level risks. Azithromycin may reduce post-discharge morbidity and mortality through infection related mechanisms such as treating undiagnosed, incompletely treated or nosocomial infections or by protecting against new or recrudescing infections that occur during recovery. Azithromycin may also act through non antimicrobial pathways such as by anti-inflammatory and/or immunomodulatory effects.

OBJECTIVE

The primary objective of this double-blind, placebo-controlled, randomized controlled trial (RCT) is to determine whether a 5-day course of azithromycin in children age 1 to 59 months discharged from hospital in western Kenya reduces rates of re-hospitalizations and/or death in the subsequent six months. The secondary objectives are (1) to evaluate possible mechanism(s) by which azithromycin may affect morbidity and mortality, by comparing reasons for re-hospitalization and prevalence of enteric and nasopharyngeal infections between the randomization arms; (2) to determine whether empiric administration of azithromycin at hospital discharge increases risk of antibiotic resistance in commensal *Escherichia coli* (*E. coli*) and *Streptococcus pneumoniae* (*S. pneumoniae*) isolates from treated children and their primary caregiver; (3) to identify correlates and intermediate markers of post-discharge mortality and hospital readmission; (4) to determine the cost-effectiveness of post-discharge administration of a 5-day course of azithromycin in settings of varying antibiotic use, re-hospitalization rates, and mortality rates; and (5) to create a repository of stool, nasopharyngeal, and blood specimens from highly characterized, recently discharged children, half of whom are treated with azithromycin, to be used to address future research questions.

METHODS

Reporting of this study protocol has been verified in accordance with the SPIRIT (Standard Protocol Items for Randomized Trials) recommendations.

Eligibility

Children age 1 to 59 months old weighing at least 2 kg who have been hospitalized, and subsequently discharged, will be eligible for inclusion. Caregivers of potentially eligible children must be at least 18 years of age or classified as an emancipated minor and be willing to participate in the Contact Cohort if randomly selected. Children will be excluded if: azithromycin is contraindicated (children taking or prescribed other macrolide antibiotics, such as erythromycin or clarithromycin, or the protease inhibitor, lopinavir); they were admitted to hospital for a trauma, injury, or a birth defect; they do not plan to remain in the study site catchment area for at least 6 months; the legal guardian does not provide consent; or if a sibling was enrolled in the trial on the same day of discharge.

Recruitment

Children will be recruited from the inpatient wards of Kisii Teaching & Referral and Homa Bay County Hospitals where study staff will accompany hospital staff on ward rounds to identify children being discharged each day. All discharged children, as determined by the onsite hospital clinicians, will be screened by study staff during working hours. If the caregiver is interested in participating and indicates consent for screening, the study staff will screen the child for eligibility, and if eligible, will obtain informed consent for study participation. Informed consent includes an explanation of the potential risks and benefits of the study and additional provision for use of participant data and samples for future studies, and will be conducted in the language of the respondent's choosing (English, Kiswahili, Kisii, or Luo). The parent or guardian (primary caregiver) must sign written informed consent (or provide a witnessed thumbprint if not literate) prior to enrollment.

Enrollment

Children will be enrolled at the time of discharge by the clinical staff. At enrollment, primary caregivers will be interviewed to assess demographic information, medical history, and detailed contact information for the child (Table 1). Medical records will also be used to abstract information from the hospitalization (including presenting diagnosis, medical management, length of stay, procedures performed, relevant medical history, physical examination, and laboratory data). All enrolled participants will undergo a physical examination performed by the study clinician, including measurement of height (in children ≥ 24 months), length (in children <24 months), weight, and mid-upper arm circumference (MUAC), each of which will be measured three times. HIV status will be obtained from medical records or from performed testing if records are not available. Detailed home location and contact information will be collected to enable patient tracing.

Table 1. Summary of data collected among enrolled children at each study visit

Enrollment visit (hospital discharge)	3 month follow up visit	6 month follow up visit	Unscheduled visits
<ul style="list-style-type: none"> Questionnaire of sociodemographic, clinical history, treatments prescribed in hospital and at discharge, hospitalization costs, dietary factors, household factors, and environmental exposures Physical exam Anthropometry Abstraction of medical records (if re-hospitalized) Verbal autopsy (or abstracted medical records) Heel/finger prick (HIV and malaria) Stool collection (<i>Shigella</i>, <i>Salmonella</i>, <i>Campylobacter</i>, <i>Escherichia coli</i>, <i>Cryptosporidium</i>, and <i>Giardia</i>) Nasopharyngeal swab collection (<i>Streptococcus pneumoniae</i>) 	<ul style="list-style-type: none"> Questionnaire of study drug administration, and reported illnesses, hospitalization costs, change in clinical history, and treatments since last visit Physical exam Anthropometry Abstraction of medical records (if re-hospitalized) Verbal autopsy (or abstracted medical records) Heel/finger prick (HIV and malaria) Stool collection (<i>Shigella</i>, <i>Salmonella</i>, <i>Campylobacter</i>, <i>Escherichia coli</i>, <i>Cryptosporidium</i>, and <i>Giardia</i>) Nasopharyngeal swab collection (<i>Streptococcus pneumoniae</i>) 	<ul style="list-style-type: none"> Questionnaire of reported illnesses, hospitalization costs, change in clinical history, and treatments since last visit Physical exam Anthropometry Abstraction of medical records (if re-hospitalized) Verbal autopsy (or abstracted medical records) Heel/finger prick (HIV and malaria, sickle-cell) Stool collection (<i>Shigella</i>, <i>Salmonella</i>, <i>Campylobacter</i>, <i>Escherichia coli</i>, <i>Cryptosporidium</i>, and <i>Giardia</i>) Nasopharyngeal swab collection (<i>Streptococcus pneumoniae</i>) 	<ul style="list-style-type: none"> Questionnaire of reported illnesses since last scheduled visit, change in clinical history, and treatments since last visit Abstraction of medical records (if re-hospitalized) Verbal autopsy (or abstracted medical records)

Specimen collection

Specimens will be collected at enrollment (prior to study medication administration, as well as at 3 and 6-month follow-up visits). All children will also be asked to provide a whole stool for enteric pathogen identification and storage. Stool samples will be divided within one hour of collection for the following purposes: 1) placed in Cary-Blair for eventual bacterial culture (FecalSwab Cary-Blair Collection and Transport System™, Copan Diagnostics), 2) immediately tested for *Giardia* and *Cryptosporidium* using the immunoassay (Quik Chek™, Alere) and 3) placed in -80°C storage for future molecular determination of pathogen or commensal flora and markers of gut function (whole stool for these purposes will be divided into two separate vials). If a child cannot produce whole stool, two flocced rectal swabs (Pediatric FLOQswab™,

Copan Diagnostics) will be collected and one placed in Cary-Blair and the other stored in -80°C for future analyses.

One flocced dry nasopharyngeal swab (Copan Diagnostics) will also be collected from all enrolled children at each time point, immediately placed in skim milk, tryptone, glucose, and glycerine (STGG) media, and frozen (-80°C) within 1 hour of collection for future *S.pneumoniae* culture.^{17 18} Primary caregivers in the Contact Cohort will also be asked to provide a stool sample (or 2 rectal swabs) and nasopharyngeal sample at each visit for testing and storage as described above.

Venous blood (up to 1 teaspoon [5mL]) will be collected from all enrolled children and caregivers enrolled in the Contact Cohort at each at each time point into EDTA tube and separated for the following purposes: 1) 0.5mL for immediate HIV-testing (if indicated according to Kenyan Ministry of Health guidelines), 2) 0.4mL for a thin malaria smear which will be stored at room temperature, 3) 0.4mL for a dried blood spot, and 4) 2-4mL for plasma and buffy coat isolation and -80°C storage.

Randomization

Block randomization (1:1) in random sized blocks of no more than 10, stratified by site, will be used. Primary randomization will include allocation to the Contact Cohorts at a ratio of 1:5 (resulting in 150 per treatment arm). Each subject will be assigned a Patient Identification Number (PID), and the randomization code linking each PID to the allocated treatment will be generated by a designated statistician and maintained by the University of Washington Research Pharmacy. Study participants, investigators (other than the statistician), the study staff, hospital clinicians, and persons involved in data management or analysis will remain blinded to the allocation group during all data collection phases of the study.

Intervention

Caregivers of enrolled children will be provided a 5-day course of oral suspension formulation azithromycin (Zithromax® from Pfizer, 10 mg/kg on day 1, followed by 5mg/kg/day on days 2-5) or identically appearing and tasting placebo at discharge. Identically appearing bottles will be pre-labelled with the PID. Dosing ranges were determined such that a given child would never be under-dosed or over-dosed by more than 20% of the weight-specific intended dose (Table 2). The day 1 dose will be split in half and the first half administered by the study clinician (to be observed by the caregiver) followed by the second half administered by the caregiver under careful observation of the study staff. Caregivers will be provided with visual instructions in the language of their choosing (English, Kiswahili, Luo, Kisii).

Automated daily text message drug administration reminders will be sent for the four days following discharge and caregivers asked to respond with whether or not the child took the daily dose. The response text message will be free of charge to caregivers and caregivers will be reimbursed for each response at the final study visit. Caregivers are also asked to record each administered dose on the bottle and to return bottles at the 3 month follow-up visit.

Follow-up Procedures

All enrolled children and primary caregivers will be scheduled to return to the health facility at 3 and 6 months following enrollment to collect clinical information and samples. Anthropometric measurements will be obtained from all children and caregivers at both follow up visits (height/length, weight, MUAC) and caregivers will be asked about any hospitalizations occurring since the last time the child was seen by study staff. Caregivers will be provided with 400KSH (approximately \$4USD) to cover the cost of their round-trip transportation.

Transportation cost will be reimbursed at each follow-up visit. If the participant does not return at their scheduled time,

Table 2. Azithromycin dosing chart by child weight

Weight (kg)	Day 1 dose (mL)	Day 2-5 dose (mL)
2.0	0.25 x 2	0.25
2.1-2.4	0.30 x 2	0.30
2.5-2.8	0.35 x 2	0.35
2.9-3.2	0.40 x 2	0.40
3.3-3.6	0.45 x 2	0.45
3.7-4.0	0.50 x 2	0.50
4.1-4.8	0.60 x 2	0.6
4.9-5.6	0.70 x 2	0.7
5.7-6.8	0.85 x 2	0.85
6.9-8.0	1.0 x 2	1.0
8.1-9.6	1.2 x 2	1.2
9.7-11.2	1.4 x 2	1.4
11.3-13.6	1.6 x 2	1.6
13.7-16.0	2.0 x 2	2.0
16.1-19.2	2.4 x 2	2.4
19.3-23.2	2.9 x 2	2.9
23.3-25.0	3.2 x 2	3.2

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study staff will attempt to make contact with the primary caregiver via cell phone; if no telephone number is provided, or if the participant cannot be reached, study staff will trace the child to the household within 2 weeks of the scheduled follow-up time.

During scheduled follow-up visits, study staff will use a standardized questionnaire to ascertain history of recent illness/morbidity, post-discharge medication use including antibiotic treatment, and current condition of the child (any hospitalizations, admission and discharge date of any hospitalization, vital status, date of death if applicable). If caregivers report a hospitalization, causes of admission, medication administration, cost of admission, and length of stay will be ascertained from both caregivers and medical records, when available.

Caregivers will be encouraged, at enrollment and at each subsequent contact, to bring the child to the study health facility at any time the child is sick. Study staff will triage children to the appropriate health facility staff and will conduct a brief unscheduled visit questionnaire to ascertain adverse event information. If the unscheduled visit leads to a hospitalization, this will trigger the completion of a hospital admission form.

If at any point during follow-up a child dies, a verbal autopsy using the Population Health Metrics Research Consortium Shortened Verbal Autopsy Questionnaire.¹⁹ If the death occurred in a hospital, data from the hospital records, including cause of death, if available, will be abstracted. If a death certificate is available, cause(s) and timing of death will be abstracted.

Final causes of re-hospitalization and death will be determined after data collection is complete by an independent adjudication committee comprised of clinicians specializing in pediatrics and infectious disease. Sources of cause of re-hospitalization (medical records and caregiver report) and causes of death (causes automatically assigned from the verbal autopsy using SmartVA-Analyze [Tariff 2.0 Method]²⁰, hospital records, or death certificates) will be presented to the adjudication committee for final cause assignment.

Laboratory Procedures

Stool (rectal swabs), nasopharyngeal swabs and blood will be collected as described above and undergo either immediate or future laboratory testing as described in Table 3. All biological samples will be collected by staff trained in biosafety and Good Clinical Laboratory Practice (GCLP). Samples will be processed in Kenya when technology is available at the Kenya Medical Research Institute (KEMRI) Wellcome Trust or Centre for Microbiology Research [CMR]. Metagenomic analyses and/or analyses that require technology not available in Kenya will be performed at the University of Washington. If stool culture results report *Shigella* or *Salmonella* infection, the study staff will contact the child's caregiver and encourage the caregiver to bring the child back for an evaluation and potential treatment if the child is symptomatic.

Table 3. Sample storage and processing descriptions

Specimen Collected	Purpose	Tests Performed
Stool/ flocked rectal swabs	Bacterial ID and storage for AST	Fresh samples/rectal swabs will be cultured to identify <i>Shigella</i> , <i>Salmonella</i> , <i>Campylobacter</i> , and <i>Escherichia coli</i> (<i>E. coli</i>) using standard microbiologic methods and biochemically confirmed using bioMérieux's API [®] strips. All <i>Shigella</i> , <i>Salmonella</i> , and <i>Campylobacter</i> isolates, as well as a random subset of <i>E. coli</i> isolates will undergo antibiotic resistance testing using disc diffusion for the following antibiotics: amoxicillin-clavulanic acid (augmentin), ampicillin, azithromycin, chloramphenicol, ciprofloxacin, ceftriaxone, ceftazidime, cefoxitin, gentamicin, imipenem, trimethoprim-sulfamethoxazole, ceftazidime/clavulanate (ESBL), cefotaxime/clavulanate (ESBL). Categorizations of susceptible, intermediate, and resistant will be determined using zone-size cut-offs outlined in Clinical and Laboratory Standards Institute (CLSI) interpretive standards.
	Parasite detection	Fresh stool and rectal swabs will be tested for <i>Giardia</i> and <i>Cryptosporidium</i> using the immunoassay Giardia/ Cryptosporidium QUIK CHEK [™] .
	Storage	Stool/ flocked swabs and colonies of <i>E. coli</i> , <i>Shigella</i> spp., <i>Salmonella</i> spp., and <i>Campylobacter</i> spp. will be stored at -80°C.
Nasopharyngeal Swabs	Bacterial isolation, storage, and resistance testing	<i>Streptococcus pneumoniae</i> (<i>S. pneumoniae</i>) colonies will be isolated using standard microbiologic or molecular diagnostic protocols and susceptibility testing performed using standard microbiologic or molecular techniques. A random subset of <i>S. pneumoniae</i> isolates will undergo antimicrobial resistance testing using disc diffusion for the following antibiotics: amoxicillin-clavulanic acid (Augmentin), ampicillin, azithromycin, chloramphenicol, ciprofloxacin, ceftriaxone, ceftazidime, cefoxitin, gentamicin, imipenem, and trimethoprim-sulfamethoxazole. Categorizations of susceptible, intermediate, and resistant will be determined using zone-size cut-offs outlined in CLSI interpretive standards.
	Storage	Back-up sample and <i>S. pneumoniae</i> colonies will be stored at -80°C.

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Blood	HIV and malaria testing	HIV testing will be performed per Kenyan National Guidelines and malaria microscopy performed using standard methods.
	Storage	Plasma and buffy coat will be stored at -80°C. Dried blood spots will be stored at room temperature.

Data Management and Confidentiality

Personal information about the participants, including medical records and data ascertained per caregiver interview, will be securely stored in files in the study offices at the study sites. Only pre-designated study staff will have access to the files. Data will be entered into an electronic database (Dacima® Electronic Data Capture) regularly by study staff. Access to the electronic database will be secured using password protected accounts for study staff. Data reports of screening, enrollment, and exclusion totals will be disseminated to the study team on a weekly basis; reports including baseline demographic characteristics, laboratory results, adherence data, and serious adverse event summaries will be distributed to study co-investigators and data monitors quarterly. Data will be regularly queried to facilitate ongoing data cleaning.

Data Analysis

Primary endpoints:

The primary study endpoint is a combined outcome of mortality and hospital readmission, as re-hospitalization is highly associated with risk of subsequent poor outcome. Re-hospitalizations that are a continuation of management from the previous hospitalization (such as elective blood transfusion) or that occur during enrollment procedures, due to a clinical deterioration post-discharge, will be excluded from the analysis. Loss to follow-up will be defined as non-attendance at both follow-up visits despite one month of active tracing and no clear evidence of death.

Secondary endpoints include:

1. Cause-specific re-hospitalizations assessed by questionnaire (maternal recall of diagnosis) at month 3 and month 6 follow-up visits and medical record review (discharge diagnosis). In cases when both sources are available, information from the medical record will be considered as the primary source. Separate analyses will be performed for each diagnosis: diarrhea, acute respiratory infection, malnutrition, or malaria.
2. Mild, moderate, and severe events that did not result in re-hospitalization, including diarrhea, vomiting, skin rash, lip swelling, difficulty breathing/wheeze, and seizure will be ascertained by caregivers identified by the study clinicians during clinical exams at scheduled follow-up visits or during unscheduled visits. Severity (grade 1-3) will be defined according to 2014 Division of AIDS (DAIDS) Table for Grading the Severity of Adult and Pediatric Adverse Events.
3. Enteric pathogen carriage, operationalized as presence of a bacterial pathogen-*Shigella* species (spp.), *Campylobacter* spp., or *Salmonella* spp., or parasite- *Giardia* or *Cryptosporidium* in stool or rectal swabs assessed at month 3 and month 6 follow-up visits.
4. *Streptococcus pneumoniae* (*S. pneumoniae*) isolated from nasopharyngeal swab cultures at month 3 and month 6 follow-up visits.
5. Antibiotic resistance, specifically resistance to azithromycin, ampicillin, augmentin, ciprofloxacin, trimethoprim-sulfamethoxazole, in *E.coli* and *S. pneumoniae* isolates, and presence of ESBL in *E.coli* isolates, from month 3 and month 6 follow-up visits.

To compare rates of re-hospitalization and mortality in the 6 months following hospital discharge among Kenyan children receiving 5-day azithromycin vs. placebo. Primary analyses will be modified intent-to-treat (mITT) based on randomization allocation to the 5-day course of azithromycin versus placebo. Cumulative incidence of death or first re-hospitalization will be compared between treatment groups using Cox proportional hazards regression. Participants will be censored at the date of their first re-hospitalization, or at the date of death. Median time to hospitalization-free survival will be compared between randomization groups using Kaplan-Meier (K-M) survival analysis and associated log-rank test. If the baseline assessment of randomization reveals an imbalance in characteristics between the treatment groups, we will evaluate these variables as potential confounders in a sub-analysis secondary to the mITT. Potential baseline confounders will be added stepwise in a multivariable Cox model and maintained in the model if adjustment changes the hazard

ratio by more than 10%. In per protocol analyses also secondary to the mITT, we will compare treatment effects in groups defined by self-reported adherence to the 5-day course of azithromycin (5 doses vs. < 5 doses; ≥ 3 doses vs. < 3 doses; > 1 dose vs. 1 dose only). In addition, we will conduct Cox regression and K-M survival analyses for time to mortality and time to re-hospitalization as separate endpoints to understand intervention effects on these outcomes individually. The assumption of proportional hazards will be checked in all models using graphical methods including plotting a $\ln(-\ln(S(t)))$ plot for each treatment group and assessing the parallelism of the two lines and by plotting Schoenfeld residuals over time. If there is substantial missing covariate data, multiple imputation using the Markov chain Monte Carlo (MCMC) method will be used to impute covariate information. Missing outcome data (death or re-hospitalization) will not be imputed, but participants will be censored at the last follow-up visit therefore contributing some person-time to the analysis. In sensitivity analyses, we will compare treatment effects in children whose caregivers report no additional antibiotic use over follow-up and separately, who report no additional azithromycin use specifically, and in subsets of children defined by age, site, and discharge diagnosis.

To evaluate possible mechanism(s) by which azithromycin may affect morbidity and mortality, by comparing reasons for re-hospitalization and change in prevalence of pathogen carriage between the randomization arms. To evaluate the association between azithromycin and the rates of cause-specific re-hospitalizations (hospitalization due to diarrhea, acute respiratory infection, malnutrition, or malaria) we will use Anderson-Gill proportional hazards modeling with previous re-hospitalizations included as time-dependent covariates in the model to capture the dependent structure of recurrence times. Because we will not have granularity in the time points other than 3 months and 6 months for assessment of pathogen carriage, we will compare the prevalence of a bacterial and parasitic pathogens (*Shigella*, *Salmonella*, *Campylobacter*, *Cryptosporidium*, *Giardia*) at 3 and 6 months by randomization arm using generalized estimating equations (GEE) with a Poisson link, exchangeable correlation structure, and will adjust for baseline presence of a bacterial pathogen. To determine whether an observed association between the intervention and pathogen carriage wanes over time, we will test the hypothesis that the prevalence ratios comparing carriage in intervention arms are the same at the two follow-up time points using a chi-squared test.

To determine whether empiric administration of azithromycin at hospital discharge increases risk of antibiotic resistance in commensal *E. coli* and pneumococcal isolates from treated children and their household contacts. Among children and adult household contacts in whom commensal *E. coli* and/or *S. pneumoniae* are isolated, we will compare the proportion of isolates resistant to azithromycin, ampicillin, augmentin, ciprofloxacin, and trimethoprim-sulfamethoxazole, between randomization arms and Contact Cohorts for each arm, at 3 and 6 months using GEE with a Poisson link and exchangeable correlation structure. A chi-squared test will be used to determine whether the association between intervention arm and antibiotic resistance wanes over time. Because the likelihood of having a bacterial pathogen isolated may depend on baseline factors, including intervention arm, we will conduct secondary analyses utilizing propensity scores to account for the potential differential likelihood of having antibiotic susceptibility testing performed, which will allow us to make inference to the entire study population and their contacts. Also we will compare resistance proportions among children (as opposed to among isolates) where absence of an isolated bacteria is considered not resistant.

To identify correlates and intermediate markers of post-discharge mortality and hospital-readmission among hospitalized children. Enrollment hospital admission diagnosis, indicators of malnutrition, age, HIV-exposure and HIV-infection status, sickle cell anemia, and randomization arm will be assessed in a multivariable Cox regression model to identify correlates of the primary endpoint of death and/or hospital-readmission independent of the treatment effect. In addition, Cox regression models will also be built for correlates of mortality and correlates of re-admission individually to understand distinct cofactors for each of these outcomes.

To determine the cost-effectiveness of post-discharge administration of a 5-day course of azithromycin in settings of varying antibiotic use, re-hospitalization rates, and mortality rates. Costs analysis: We will assess the costs of all supplies, services and equipment necessary to implement the intervention (direct medical costs). The perspective will be that of the healthcare provider, i.e. Kenya's Ministry of Health. Using WHO guidelines and its ingredients approach, we will quantify the resources and associated unit costs required to deliver a 5-day course of azithromycin, organized in standard expenditure categories: personnel (salaries), supplies including drugs, equipment, services, space and overhead. We will also measure the costs of severe child hospitalizations, the costs for the different types of personnel employed (e.g. nurses/doctors) and the time

1
2 demanded from them for conducting the intervention.²¹ When data are missing, they will be complemented by
3 data extracted from the literature and other available sources. Full incremental costs will be derived, with
4 estimation of the potential healthcare cost-offset realized in avoiding severe hospitalizations. Costs will be
5 measured in local currency (Kenyan Shilling) and converted into US\$. Our main metric will be cost per child
6 treated. Cost-effectiveness analysis (CEA): we will develop a CEA mathematical model, and estimate
7 incremental costs and cost-effectiveness for implementation of the intervention. The model will include two
8 components: costs (described immediately above) and health benefits. The study will provide clinical outcomes
9 (mortality/morbidity) over a 6-month follow-up period. Subsequently, deaths averted, life-years saved and
10 disability-adjusted life years (DALYs) averted by the intervention will be estimated. We will estimate: a)
11 incremental costs and b) incremental cost-effectiveness of the intervention vs. status quo. *Incremental costs*
12 are the net sum of the costs to implement the intervention compared with status quo, and the costs averted
13 due to the decrease in severe child hospitalizations. *Incremental cost-effectiveness* ratios (ICERs) will be
14 estimated as cost per death averted, cost per life-year saved, and cost per DALY averted. We will use recent
15 estimates of disability weights for estimation of DALYs.^{22 23} Short-term (over study follow-up i.e. 6 months) and
16 longer-term time horizons (extrapolated to 1, 5, and 10 years) will be used. DALYs and costs will be discounted
17 at 3% per year, consistent with CEA guidelines (undiscounted results will also be presented). Sources of
18 uncertainty in the results will be explored in univariate and probabilistic sensitivity analysis.^{24 25} Finally, we will
19 compare our findings to CEA estimates for other health interventions in sub-Saharan Africa.^{26 27}

22 Data and Safety Monitoring

23 A Data Safety and Monitoring Committee (DSMC) will be established at study initiation to monitor
24 severe adverse events (SAEs) and to evaluate the statistical analysis plan and associated stopping rules. The
25 DSMC includes expertise in clinical trials, statistics, child mortality assessment, ethics, and pediatric care in
26 resource limited settings. Adverse events will be monitored by the DSMC. Monthly adverse event summaries
27 will be sent to the DSMC safety officer and individual-child SAE forms, which include detailed medication
28 history to evaluate possible drug interactions, will be sent to the safety officer per request. Each SAE will be
29 assigned the plausibility of relatedness to study drug by study PIs. The data will not be presented by
30 intervention group unless requested by DSMB safety officer. These reports will be descriptive (no statistical
31 analyses). The DSMC will make recommendations regarding any imbalances in safety outcomes.

32 A single interim analysis for re-hospitalization-free survival will be prepared by the study statistician
33 using O'Brien-Fleming boundaries for benefit and harm when 50% of expected person time (350 child-years)
34 has been accrued. Assuming 157 events will be available at half of the person-time accrual, a z-score critical
35 value of 2.797, or p -value < 0.005, from a Kaplan Meier log-rank test will determine the cut-off of statistical
36 significance. A symmetric boundary will be used for benefit and harm. The DSMC will review this analysis and
37 make a determination about study continuation. Futility will not be a basis for stopping rules because of the
38 trials' value in understanding mechanisms of post-discharge worsening and antibiotic resistance. Assuming the
39 DSMC decides to continue the trial after the interim analysis, an alpha of 0.045 will be used as the statistical
40 significance boundary at the final analysis.

44 Statistical Power

45 To compare rates of re-hospitalization and mortality in the 6 months following hospital discharge among
46 Kenyan children receiving 5-day azithromycin vs. placebo. The total sample size required was calculated for
47 the primary endpoint of time to death or hospital re-admission within the 6 month post-discharge period,
48 assuming an alpha level of 0.05, power of 0.80, and a ratio of treatment to placebo random assignment of 1:1.
49 In SSA, it is estimated that 2-15% of children aged less than 5 years died within 6 months of hospital discharge
50 and 15.5% of children who survived discharge from the district hospital were re-admitted with the same
51 diagnosis within 6-months.^{1 3 8} Assuming that an additional 5-10% of children are re-admitted for other
52 conditions, we expect that re-hospitalizations will occur in 20.5 to 30.5% of children enrolled in the study.
53 Combined with our expected fatality rate (2-15%), we expect the cumulative incidence of the combined
54 endpoint to range from 22.5 to 45.5%.⁸ Based on a previous trial of mass drug administration of a single dose
55 of azithromycin in which a single dose of the antibiotic was associated with a 49% reduction in risk of death, we
56 calculated sample sizes using estimates of reduction in risk ranging from 30-50% with the cumulative incidence
57 range of 22.5 to 45.5% in the placebo-treated group, and found the sample size required ranged from 90 to
58 550 children per treatment arm.¹⁵ Using the most conservative estimates of a hazard ratio of 0.70 and 22.5%

prevalence of re-admission/death, we need to enroll 1100 children in the study (550 per arm) to achieve adequate power. We will recruit an additional 300 children ($\approx 20\%$) to account for possible loss to follow-up, resulting in a total planned enrollment of 1400 children, or 700 per treatment group. When considering mortality alone, and estimated mortality ranges of 2-17% among placebo-treated children, we will have $>80\%$ power to detect hazard ratios ≤ 0.5 for mortality rates of $\geq 8\%$ and hazard ratios ≤ 0.6 for mortality rates $\geq 11\%$ (Figure 1).

To evaluate possible mechanism(s) by which azithromycin may affect morbidity and mortality, by comparing reasons for re-hospitalization and prevalence of pathogen carriage between the randomization arms. We calculated the minimum detectable association between treatment arm and cause-specific re-hospitalizations (hospitalization due to diarrhea, acute respiratory infection, malnutrition, or malaria vs. any other) among enrolled children, assuming an alpha level of 0.05, power of 0.80, and a ratio of treatment to placebo of 1:1. Based on data from Kenya, re-hospitalization rates due to specific causes ranged from approximately 0.5% to 5.7% in the 6 month post-discharge period.³ By not conditioning on the child having the same diagnosis as the initial hospitalization, we expect the cumulative incidence of cause-specific re-hospitalizations to range from 2.5% to 10%. With this range of outcome rates, we will be able to detect hazard ratios of 0.48 to 0.70 for the effect of azithromycin on specific severe morbidities.

We expect 56% of children in the placebo group to have *S. pneumoniae* isolated from nasopharyngeal swabs, providing $\geq 80\%$ power to detect a prevalence ratio of 0.85 (or 1.15) between the two treatment arms at each time point.²⁸⁻³⁰ Based on prevalences of *Shigella*, *Salmonella*, *Campylobacter*, *Cryptosporidium*, *Giardia* among asymptomatic children in Western Kenya, we expect 10% of children in the placebo group to have a bacterial pathogen isolated at each time point, resulting in $\geq 80\%$ power to detect differences in enteric pathogen prevalences of 0.67 (1.49) at each time point.³¹

To determine whether empiric administration of azithromycin at hospital discharge increases risk of antibiotic resistance in commensal *E. coli* and *S. pneumoniae* isolates from treated children and their household contacts.

We will select a random selection of 400 *E. coli* and 400 *S. pneumoniae* isolates (200 per arm) for β -lactam and macrolide resistance testing at each timepoint. We will also store all *S. pneumoniae*, *E. coli* isolates and other isolated bacteria from stool for potential future testing in the event that resistance prevalence is lower than expected. As shown in Table 4, we will have $> 80\%$ power to detect prevalence ratios > 1.1 , with an ability to detect the smallest effect sizes when the prevalence of resistance in the placebo group is highest. We will enroll 300 adults in the Contact Cohort for *E. coli* and *S. pneumoniae* isolation. We expect *E. coli* to be isolated from all adults and *S. pneumoniae* isolated from between 5-55%.^{28 32 33} Assuming an alpha of .05, a 1:1 ratio of testable isolates, and a prevalence of

Table 4. Power (%) to detect prevalence ratios of macrolide and β -lactamase resistance in 200 *E. coli* and 200 *S. pneumoniae* isolates per treatment group

Resistance prevalence (%) in azithromycin group	Resistance prevalence (%) in placebo group							
	10	20	30	40	50	60	70	80
10								
20	80							
30	>99	64						
40	>99	99	55					
50	>99	>99	98	48				
60	>99	>99	>99	>99	52			
70	>99	>99	>99	>99	98	55		
80	>99	>99	>99	>99	>99	99	64	

of resistance of 50% in the placebo arm, we will have 80% power to detect a 1.4-fold higher prevalence to 1.9-fold higher resistance prevalence in the contacts of azithromycin-treated children.

To identify correlates and intermediate markers of post-discharge mortality and hospital-readmission among hospitalized Kenyan children. Conservatively estimating a 20% loss-to-follow-up rate in the RCT and a cumulative incidence of death or re-hospitalization of 22.5%, we will have $>80\%$ power to detect hazard ratios ≥ 1.3 between correlates and the outcome with exposure prevalences of $\geq 20\%$ or more and hazard ratios ≥ 1.5 for exposure prevalences $< 20\%$.

Study timeline

The trial began on June 28, 2016 and participant recruitment and follow-up will continue over a 36-month period, with anticipated final follow-up visit(s) occurring in June 2019. Primary analyses will be complete by February 2020.

Potential Challenges and Limitations

In order to ensure adequate power to detect a discernable clinically relevant difference between study groups in the primary outcome, we have combined hospital readmission with death. Preliminary studies

1
2 suggest that sufficient numbers of children will reach this combined outcome. However, we have incorporated
3 an interim analysis by the DSMB to review the accrued data and an adapted sample size could be considered
4 if the combined event frequency is less than predicted. It is possible that since most children receive antibiotics
5 during hospitalization, the benefit anticipated with the use of azithromycin based on previous trials of mass
6 drug administration will not be observed. However, most hospitalized children are treated with penicillins,
7 cephalosporins, gentamicin, or cotrimoxazole while in hospital and the broad spectrum of activity (including
8 malaria prevention) and long half-life of azithromycin suggest that there may be additive treatment and/or
9 prophylactic benefit. Similarly, children may receive azithromycin during follow up - either as treatment for an
10 illness or because the caregiver sought out azithromycin upon learning of the hypothesis – and this
11 azithromycin use may lead to contamination in the placebo-arm. After discharge, it is difficult to ensure
12 adherence with the full 5-day treatment course. We will measure adherence using three different measures
13 (text message responses, bottle check boxes, and caregiver-report at follow-up visits) although all are limited
14 by caregiver-report. In addition, the mortality benefit of azithromycin observed in Ethiopia was from a single
15 dose and in this study the first dose will be directly observed.¹⁵ While relying on caregiver report of mortality
16 and morbidity may lead to bias due to outcome misclassification, this misclassification should not differ
17 between randomization arms and therefore will be non-differential. Further hospital records will be used when
18 available to determine diagnoses. Finally, resistance prevalence may be lower than predicted, limiting power to
19 detect clinically relevant differences in resistance prevalence between the intervention arms. We will store all
20 isolates in the event that a greater number of isolates are needed for antibiotic resistance testing.
21
22

23 **Regulatory Authorities**

24 This study has received IRB approval by the University of Washington Human Subjects Division (HSD), KEMRI
25 Scientific and Ethics Review Unit (SERU), and the Kenya Pharmacy and Poisons Board. The clinical trial is
26 also registered with clinicaltrials.gov (NCT02414399). Any modifications to the study protocol or consent
27 materials will be submitted for approval all regulatory authorities before implementation. Westat® will provide
28 external clinical, pharmacy, and laboratory monitoring.
29
30

31 **Dissemination**

32 Results of this study will be disseminated by publication in a peer-reviewed scientific journal, presented at
33 relevant academic conferences, and amongst participating partners and health facilities in Kenya.
34
35

36 **Author's contributions**

37 JLW, PBP, GJS, BAR, BOS, and RN conceived of this trial and developed of this study protocol. JLW and BOS
38 are study co-Principal Investigators and PBP is the Project Director; BAR oversaw the statistical analyses
39 plans; JBB developed the CEA plan; KDT developed procedures for ascertaining and reporting SAEs; CJM
40 developed procedures related to blood specimen procedures and drug adherence measurement. GJS, CJM,
41 RN, and PBP provide scientific expertise. RLB and MA are involved in collection and management of the data.
42 MA and PBP coordinate and oversee implementation of all clinical study procedures and SK, with assistance
43 from DR, oversees all laboratory procedures. All authors contributed to the development of this manuscript
44 and/or study procedures, and to reading and approving the final version for publication.
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46

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50

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54 Waichungo, Angela Karani, Donald Akech, and Horace Gumba provided microbiology expertise and training in
55 nasopharyngeal swab collection, STGG media preparation, and laboratory quality assurance and control. Liru
56 Meshack Wekesa and George Bogonko provide clinical expertise and facilitate the study's integration into
57 pediatric wards at the two hospitals. Alex Awuor and Caleb Okonji, with the support of Richard Omore,
58 provided training in anthropometric measurement. We are extremely thankful to Dr. Philip Walson who
59 developed azithromycin dosing regimens. Hannah Atlas and Stephanie Belanger contributed to the standard
60

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operating procedure and case report form development and implementation. Gillian Levine played an invaluable role in the proposal development.

Competing interests statement

None of the authors or study co-investigators have any competing interests to declare.

Figure Legend

Figure 1. Power and detectable hazard ratios assuming a range of mortality rates from 2-17%

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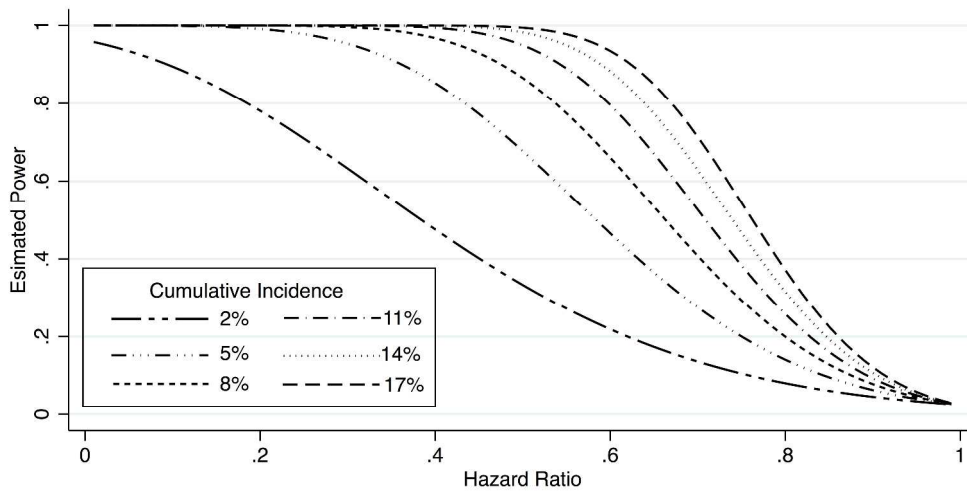


Figure 1. Power and detectable hazard ratios assuming a range of mortality rates from 2-17%

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STANDARD PROTOCOL ITEMS: RECOMMENDATIONS FOR INTERVENTIONAL TRIALS

SPIRIT 2013 Checklist: Recommended items to address in a clinical trial protocol and related documents*

Section/item	Item No	Description	Addressed on page number
Administrative information			
Title	1	Descriptive title identifying the study design, population, interventions, and, if applicable, trial acronym	_____
Trial registration	2a	Trial identifier and registry name. If not yet registered, name of intended registry	_____
	2b	All items from the World Health Organization Trial Registration Data Set	_____
Protocol version	3	Date and version identifier	_____
Funding	4	Sources and types of financial, material, and other support	_____
Roles and responsibilities	5a	Names, affiliations, and roles of protocol contributors	_____
	5b	Name and contact information for the trial sponsor	_____
	5c	Role of study sponsor and funders, if any, in study design; collection, management, analysis, and interpretation of data; writing of the report; and the decision to submit the report for publication, including whether they will have ultimate authority over any of these activities	_____
	5d	Composition, roles, and responsibilities of the coordinating centre, steering committee, endpoint adjudication committee, data management team, and other individuals or groups overseeing the trial, if applicable (see Item 21a for data monitoring committee)	_____

1 **Introduction**

2

3 Background and 6a Description of research question and justification for undertaking the trial, including summary of relevant
4 rationale studies (published and unpublished) examining benefits and harms for each intervention _____

5

6 6b Explanation for choice of comparators _____

7

8 Objectives 7 Specific objectives or hypotheses _____

9

10 Trial design 8 Description of trial design including type of trial (eg, parallel group, crossover, factorial, single group),
11 allocation ratio, and framework (eg, superiority, equivalence, noninferiority, exploratory) _____

12

13

14

15 **Methods: Participants, interventions, and outcomes**

16

17 Study setting 9 Description of study settings (eg, community clinic, academic hospital) and list of countries where data will
18 be collected. Reference to where list of study sites can be obtained _____

19

20 Eligibility criteria 10 Inclusion and exclusion criteria for participants. If applicable, eligibility criteria for study centres and
21 individuals who will perform the interventions (eg, surgeons, psychotherapists) _____

22

23 Interventions 11a Interventions for each group with sufficient detail to allow replication, including how and when they will be
24 administered _____

25

26 11b Criteria for discontinuing or modifying allocated interventions for a given trial participant (eg, drug dose
27 change in response to harms, participant request, or improving/worsening disease) _____

28

29 11c Strategies to improve adherence to intervention protocols, and any procedures for monitoring adherence
30 (eg, drug tablet return, laboratory tests) _____

31

32 11d Relevant concomitant care and interventions that are permitted or prohibited during the trial _____

33

34 Outcomes 12 Primary, secondary, and other outcomes, including the specific measurement variable (eg, systolic blood
35 pressure), analysis metric (eg, change from baseline, final value, time to event), method of aggregation (eg,
36 median, proportion), and time point for each outcome. Explanation of the clinical relevance of chosen
37 efficacy and harm outcomes is strongly recommended _____

38

39

40

41 Participant timeline 13 Time schedule of enrolment, interventions (including any run-ins and washouts), assessments, and visits for
42 participants. A schematic diagram is highly recommended (see Figure) _____

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1 Sample size 14 Estimated number of participants needed to achieve study objectives and how it was determined, including _____
 2 clinical and statistical assumptions supporting any sample size calculations

4 Recruitment 15 Strategies for achieving adequate participant enrolment to reach target sample size _____
 5

7 **Methods: Assignment of interventions (for controlled trials)**

9 Allocation:

11 Sequence 16a Method of generating the allocation sequence (eg, computer-generated random numbers), and list of any _____
 12 generation factors for stratification. To reduce predictability of a random sequence, details of any planned restriction
 13 (eg, blocking) should be provided in a separate document that is unavailable to those who enrol participants
 14 or assign interventions

17 Allocation 16b Mechanism of implementing the allocation sequence (eg, central telephone; sequentially numbered, _____
 18 concealment opaque, sealed envelopes), describing any steps to conceal the sequence until interventions are assigned
 19 mechanism

21 Implementation 16c Who will generate the allocation sequence, who will enrol participants, and who will assign participants to _____
 22 interventions

25 Blinding (masking) 17a Who will be blinded after assignment to interventions (eg, trial participants, care providers, outcome _____
 26 assessors, data analysts), and how

28 17b If blinded, circumstances under which unblinding is permissible, and procedure for revealing a participant's _____
 29 allocated intervention during the trial

32 **Methods: Data collection, management, and analysis**

34 Data collection 18a Plans for assessment and collection of outcome, baseline, and other trial data, including any related _____
 35 methods processes to promote data quality (eg, duplicate measurements, training of assessors) and a description of
 36 study instruments (eg, questionnaires, laboratory tests) along with their reliability and validity, if known.
 37 Reference to where data collection forms can be found, if not in the protocol

40 18b Plans to promote participant retention and complete follow-up, including list of any outcome data to be _____
 41 collected for participants who discontinue or deviate from intervention protocols

1	Data management	19	Plans for data entry, coding, security, and storage, including any related processes to promote data quality	_____
2			(eg, double data entry; range checks for data values). Reference to where details of data management	
3			procedures can be found, if not in the protocol	
4				
5	Statistical methods	20a	Statistical methods for analysing primary and secondary outcomes. Reference to where other details of the	_____
6			statistical analysis plan can be found, if not in the protocol	
7				
8		20b	Methods for any additional analyses (eg, subgroup and adjusted analyses)	_____
9				
10		20c	Definition of analysis population relating to protocol non-adherence (eg, as randomised analysis), and any	_____
11			statistical methods to handle missing data (eg, multiple imputation)	
12				
13				
14				
15	Methods: Monitoring			
16				
17	Data monitoring	21a	Composition of data monitoring committee (DMC); summary of its role and reporting structure; statement of	_____
18			whether it is independent from the sponsor and competing interests; and reference to where further details	
19			about its charter can be found, if not in the protocol. Alternatively, an explanation of why a DMC is not	
20			needed	
21				
22		21b	Description of any interim analyses and stopping guidelines, including who will have access to these interim	_____
23			results and make the final decision to terminate the trial	
24				
25				
26	Harms	22	Plans for collecting, assessing, reporting, and managing solicited and spontaneously reported adverse	_____
27			events and other unintended effects of trial interventions or trial conduct	
28				
29	Auditing	23	Frequency and procedures for auditing trial conduct, if any, and whether the process will be independent	_____
30			from investigators and the sponsor	
31				
32				
33	Ethics and dissemination			
34				
35	Research ethics	24	Plans for seeking research ethics committee/institutional review board (REC/IRB) approval	_____
36	approval			
37				
38	Protocol	25	Plans for communicating important protocol modifications (eg, changes to eligibility criteria, outcomes,	_____
39	amendments		analyses) to relevant parties (eg, investigators, REC/IRBs, trial participants, trial registries, journals,	
40			regulators)	
41				
42				
43				
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45				
46				
47				

1	Consent or assent	26a	Who will obtain informed consent or assent from potential trial participants or authorised surrogates, and	_____
2			how (see Item 32)	
3				
4		26b	Additional consent provisions for collection and use of participant data and biological specimens in ancillary	_____
5			studies, if applicable	
6				
7	Confidentiality	27	How personal information about potential and enrolled participants will be collected, shared, and maintained	_____
8			in order to protect confidentiality before, during, and after the trial	
9				
10	Declaration of	28	Financial and other competing interests for principal investigators for the overall trial and each study site	_____
11	interests			
12				
13				
14	Access to data	29	Statement of who will have access to the final trial dataset, and disclosure of contractual agreements that	_____
15			limit such access for investigators	
16				
17	Ancillary and post-	30	Provisions, if any, for ancillary and post-trial care, and for compensation to those who suffer harm from trial	_____
18	trial care		participation	
19				
20	Dissemination policy	31a	Plans for investigators and sponsor to communicate trial results to participants, healthcare professionals,	_____
21			the public, and other relevant groups (eg, via publication, reporting in results databases, or other data	
22			sharing arrangements), including any publication restrictions	
23				
24				
25		31b	Authorship eligibility guidelines and any intended use of professional writers	_____
26				
27		31c	Plans, if any, for granting public access to the full protocol, participant-level dataset, and statistical code	_____
28				
29				
30	Appendices			
31				
32	Informed consent	32	Model consent form and other related documentation given to participants and authorised surrogates	_____
33	materials			
34				
35	Biological	33	Plans for collection, laboratory evaluation, and storage of biological specimens for genetic or molecular	_____
36	specimens		analysis in the current trial and for future use in ancillary studies, if applicable	
37				

38
39 *It is strongly recommended that this checklist be read in conjunction with the SPIRIT 2013 Explanation & Elaboration for important clarification on the items.
40 Amendments to the protocol should be tracked and dated. The SPIRIT checklist is copyrighted by the SPIRIT Group under the Creative Commons
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BMJ Open

Azithromycin to prevent post-discharge morbidity and mortality in Kenyan children: A protocol for a randomized, double-blind, placebo-controlled trial (the Toto Bora trial)

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Title: Azithromycin to prevent post-discharge morbidity and mortality in Kenyan children: A protocol for a randomized, double-blind, placebo-controlled trial (the Toto Bora trial)

Running head: AZM to prevent post-discharge morbidity and mortality

Authors: Pavlinac PB¹, Singa BO^{7,8}, John-Stewart GC¹⁻⁴, Richardson BA^{1,5}, Brander RL², McGrath CJ¹, Tickell KD^{1,8}, Amondi M⁷, Rwigy D⁷, Babigumira JB^{1,6}, Kariuki S⁷, Nduati R⁹, Walson JL^{1-4,8}

Affiliations: Departments of ¹Global Health, ²Epidemiology, ³Pediatrics, ⁴Allergy and Infectious Disease, ⁵Biostatistics, ⁶Pharmacy, University of Washington, Seattle, WA, USA; ⁷Kenya Medical Research Institute, Nairobi, Kenya; ⁸The Childhood Acute Illness & Nutrition Network, Nairobi, Kenya; ⁹University of Nairobi, Nairobi, Kenya

Correspondence: Patricia Pavlinac, 325 9th Ave, Box 359931, Seattle, WA 98104 (ppav@uw.edu)

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ABSTRACT

Introduction: Child mortality due to infectious diseases remains unacceptably high in much of sub-Saharan Africa. Children who are hospitalized represent an accessible population at particularly high-risk of death, both during and following hospitalization. Hospital discharge may be a critical time point at which targeted use of antibiotics could reduce morbidity and mortality in high-risk children.

Methods and analysis: In this randomized, double-blind, placebo-controlled trial (Toto Bora Trial), 1400 children aged 1 to 59 months discharged from hospitals in western Kenya, in Kisii and Homa Bay, will be randomized to either a 5-day course of azithromycin or placebo to determine whether a short-course of azithromycin reduces rates of re-hospitalization and/or death in the subsequent 6-month period. The primary analysis will be modified intention-to-treat and will compare the rates of re-hospitalization or death in children treated with azithromycin or placebo using Cox proportional hazard regression. The trial will also evaluate the effect of a short course of azithromycin on enteric and nasopharyngeal infections and cause-specific morbidities. We will also identify risk factors for post-discharge morbidity and mortality and subpopulations most likely to benefit from post-discharge antibiotic use. Antibiotic resistance in *Escherichia coli* and *Streptococcus pneumoniae* among enrolled children and their primary caregivers will also be assessed and cost-effectiveness analyses performed to inform policy decisions.

Ethics and dissemination: Study procedures were reviewed and approved by the institutional review boards of the Kenya Medical Research Institute, the University of Washington, and the Kenyan Pharmacy and Poisons Board. The study is being externally monitored and a data safety and monitoring committee has been assembled to monitor patient safety and to evaluate the efficacy of the intervention. The results of this trial will be published in peer-reviewed scientific journals and presented at relevant academic conferences and to key stakeholders.

Trial registration number: NCT02414399

Key words: child mortality, Toto Bora, targeted empiric antibiotic therapy, post-discharge interventions

STRENGTHS AND LIMITATIONS OF THIS STUDY

Strengths

- Randomized, placebo-controlled, double-blinded design and modified intention-to-treat analysis will ensure unbiased treatment effect measure
- Comprehensive data are collected, including biological specimens for all child participants and a subset of adult caregivers, for analyses of mechanisms of post-discharge morbidity and mortality, subsets of children most likely to benefit from the antibiotic, as well as assessments of antibiotic resistance and cost-effectiveness
- Results will likely be generalizable due to the limited exclusion criteria, large sample size, and multiple study sites

Limitations

- Causes of death and re-hospitalization may be misclassified due to limited availability of medical records and recall bias in caregiver report
- The primary endpoint of this study is a combined outcome of re-hospitalization and death, which, while improving statistical power, may present challenges for interpretation
- Children in both intervention arms may receive other antibiotics over the course of follow-up

BACKGROUND

Close to 3 million deaths occur annually in children less than 5 years of age in sub-Saharan Africa (SSA), over half of which are attributed to infectious causes.¹ Children who were recently hospitalized have mortality rates 6 to 8-fold higher than similarly-aged children from the same community.²⁻⁴ Post-discharge mortality rates as high as 15% have been documented in the 12 months following discharge, with mortality risk remaining elevated up to two years post-discharge.⁵⁻⁹ Children who are very young, malnourished, or HIV-infected are at particularly high risk of post-discharge mortality within the 3 months following discharge.^{2-5 7-9} Children being discharged from hospital in SSA may represent an accessible high-risk population in which to target interventions to reduce mortality and morbidity.

Targeted antibiotic interventions, including the use of cotrimoxazole among HIV-infected children and the use of amoxicillin or cefdinir among children with severe acute malnutrition (SAM), have been shown to reduce morbidity and mortality in these specific vulnerable populations.¹⁰⁻¹³ Other trials of targeted antibiotic use in vulnerable populations, including cotrimoxazole in HIV-exposed uninfected (HEU) children and in children with SAM, have failed to demonstrate a mortality benefit.^{14 15} In contrast, non-targeted mass drug administration of a single dose of azithromycin halved mortality rates among Ethiopian children living in communities randomized to receive the antibiotic.^{16 17} Concerns about the potential emergence of antibiotic resistance, possible toxicity, and feasibility of delivery are barriers to community-wide antibiotic distribution strategies.

A short-course of azithromycin given to children with recent severe illness being discharged from hospital may optimize benefit while reducing both individual and population level risks. Azithromycin may reduce post-discharge morbidity and mortality through infection related mechanisms such as treating undiagnosed, incompletely treated or nosocomial infections or by protecting against new or recrudescing infections that occur during recovery. Azithromycin may also act through non-antimicrobial pathways such as by anti-inflammatory and/or immune-modulatory effects.

OBJECTIVE

The primary objective of this double-blind, placebo-controlled, randomized controlled trial (RCT) is to determine whether a 5-day course of azithromycin in children age 1 to 59 months discharged from hospital in western Kenya reduces rates of re-hospitalizations and/or death in the subsequent six months. The secondary objectives are (1) to evaluate possible mechanism(s) by which azithromycin may affect morbidity and mortality, by comparing reasons for re-hospitalization and prevalence of enteric and nasopharyngeal infections between the randomization arms; (2) to determine whether empiric administration of azithromycin at hospital discharge increases risk of antibiotic resistance in commensal *Escherichia coli* (*E. coli*) and *Streptococcus pneumoniae* (*S. pneumoniae*) isolates from treated children and their primary caregiver; (3) to identify correlates and intermediate markers of post-discharge mortality and hospital readmission; (4) to determine the cost-effectiveness of post-discharge administration of a 5-day course of azithromycin in settings of varying antibiotic use, re-hospitalization rates, and mortality rates; and (5) to create a repository of stool, nasopharyngeal, and blood specimens from highly characterized, recently discharged children, half of whom are treated with azithromycin, to be used to address future research questions.

METHODS

Reporting of this study protocol has been verified in accordance with the SPIRIT (Standard Protocol Items for Randomized Trials) recommendations.

Eligibility

Children age 1 to 59 months old weighing at least 2 kg who have been hospitalized, and subsequently discharged, will be eligible for inclusion. Caregivers of potentially eligible children must be at least 18 years of age or classified as an emancipated minor and be willing to participate in the Contact Cohort if randomly selected. Children will be excluded if: azithromycin is contraindicated (children taking or prescribed other macrolide antibiotics, such as erythromycin or clarithromycin, or the protease inhibitor, lopinavir); they were admitted to hospital for a trauma, injury, or a birth defect; they do not plan to remain in the study site catchment area for at least 6 months; the legal guardian does not provide consent; or if a sibling was enrolled in the trial on the same day of discharge.

Recruitment

Children will be recruited from the inpatient wards of health facilities in Kisii and Homa Bay Counties where study staff will accompany hospital staff on ward rounds to identify children being discharged each day. All discharged children, as determined by the onsite hospital clinicians, will be screened by study staff during working hours. If the caregiver is interested in participating and indicates consent for screening, the study staff will screen the child for eligibility, and if eligible, will obtain informed consent for study participation. Informed consent includes an explanation of the potential risks and benefits of the study and additional provision for use of participant data and samples for future studies, and will be conducted in the language of the respondent's choosing (English, Kiswahili, Kisii, or Luo). The parent or guardian (primary caregiver) must sign written informed consent (or provide a witnessed thumbprint if not literate) prior to enrollment.

Enrollment

Children will be enrolled at the time of discharge by the clinical staff. At enrollment, primary caregivers will be interviewed to assess demographic information, medical history, and detailed contact information for the child (Table 1). Medical records will also be used to abstract information from the hospitalization (including presenting diagnosis, medical management, length of stay, procedures performed, relevant medical history, physical examination, and laboratory data). All enrolled participants will undergo a physical examination performed by the study clinician, including measurement of height (in children ≥ 24 months), length (in children <24 months), weight, and mid-upper arm circumference (MUAC), each of which will be measured three times. HIV status will be obtained from medical records or from performed testing if records are not available. Detailed home location and contact information will be collected to enable patient tracing.

Table 1. Summary of data collected among enrolled children at each study visit

Enrollment visit (hospital discharge)	3 month follow up visit	6 month follow up visit	Unscheduled visits
<ul style="list-style-type: none"> Questionnaire of sociodemographic, clinical history, treatments prescribed in hospital and at discharge, hospitalization costs, dietary factors, household factors, and environmental exposures Physical exam Anthropometry Abstraction of medical records (if re-hospitalized) Verbal autopsy (or abstracted medical records) Heel/finger prick (HIV and malaria) Stool collection (<i>Shigella</i>, <i>Salmonella</i>, <i>Campylobacter</i>, <i>Escherichia coli</i>, <i>Cryptosporidium</i>, and <i>Giardia</i>) Nasopharyngeal swab collection (<i>Streptococcus pneumoniae</i>) 	<ul style="list-style-type: none"> Questionnaire of study drug administration, and reported illnesses, hospitalization costs, change in clinical history, and treatments since last visit Physical exam Anthropometry Abstraction of medical records (if re-hospitalized) Verbal autopsy (or abstracted medical records) Heel/finger prick (HIV and malaria) Stool collection (<i>Shigella</i>, <i>Salmonella</i>, <i>Campylobacter</i>, <i>Escherichia coli</i>, <i>Cryptosporidium</i>, and <i>Giardia</i>) Nasopharyngeal swab collection (<i>Streptococcus pneumoniae</i>) 	<ul style="list-style-type: none"> Questionnaire of reported illnesses, hospitalization costs, change in clinical history, and treatments since last visit Physical exam Anthropometry Abstraction of medical records (if re-hospitalized) Verbal autopsy (or abstracted medical records) Heel/finger prick (HIV and malaria, sickle-cell) Stool collection (<i>Shigella</i>, <i>Salmonella</i>, <i>Campylobacter</i>, <i>Escherichia coli</i>, <i>Cryptosporidium</i>, and <i>Giardia</i>) Nasopharyngeal swab collection (<i>Streptococcus pneumoniae</i>) 	<ul style="list-style-type: none"> Questionnaire of reported illnesses since last scheduled visit, change in clinical history, and treatments since last visit Abstraction of medical records (if re-hospitalized) Verbal autopsy (or abstracted medical records)

Specimen collection

Specimens will be collected at enrollment (prior to study medication administration, as well as at 3 and 6-month follow-up visits). All children will also be asked to provide a whole stool for enteric pathogen identification and storage. Stool samples will be divided within one hour of collection for the following purposes: 1) placed in Cary-Blair for eventual bacterial culture (FecalSwab Cary-Blair Collection and Transport System™, Copan Diagnostics), 2) immediately tested for *Giardia* and *Cryptosporidium* using the immunoassay (Quik Chek™, Alere) and 3) placed in -80°C storage for future molecular determination of pathogen or commensal flora and markers of gut function (whole stool for these purposes will be divided into two separate vials). If a child cannot produce whole stool, two flocced rectal swabs (Pediatric FLOQswab™,

Copan Diagnostics) will be collected and one placed in Cary-Blair and the other stored in -80°C for future analyses.

One flocced dry nasopharyngeal swab (Copan Diagnostics) will also be collected from all enrolled children at each time point, immediately placed in skim milk, tryptone, glucose, and glycerine (STGG) media, and frozen (-80°C) within 1 hour of collection for future *S.pneumoniae* culture.^{18 19} Primary caregivers in the Contact Cohort will also be asked to provide a stool sample (or 2 rectal swabs) and nasopharyngeal sample at each visit for testing and storage as described above.

Venous blood (up to 1 teaspoon [5mL]) will be collected from all enrolled children at each time point into EDTA tubes and separated for the following purposes: 1) 0.5mL for immediate HIV-testing (if indicated according to Kenyan Ministry of Health guidelines), 2) 0.4mL for a thin malaria smear which will be stored at room temperature, 3) 0.4mL for a dried blood spot, and 4) 2-4mL for plasma and buffy coat isolation and -80°C storage. Blood will also be collected from primary caregivers for HIV-testing if indicated.

Randomization

Block randomization (1:1) in random sized blocks of no more than 10, stratified by site, will be used. Primary randomization will include allocation to the Contact Cohorts at a ratio of 1:5 (resulting in 150 per treatment arm). Each subject will be assigned a Patient Identification Number (PID), and the randomization code linking each PID to the allocated treatment will be generated by a designated statistician and maintained by the University of Washington Research Pharmacy. Study participants, investigators (other than the statistician), the study staff, hospital clinicians, and persons involved in data management or analysis will remain blinded to the allocation group during all data collection phases of the study.

Intervention

Enrolled children will be prescribed a 5-day course of oral suspension formulation azithromycin (Zithromax® from Pfizer, 10 mg/kg on day 1, followed by 5mg/kg/day on days 2-5) or identically appearing and tasting placebo at discharge. Identically appearing bottles will be pre-labelled with the PID. Dosing ranges were determined such that a given child would never be under-dosed or over-dosed by more than 20% of the weight-specific intended dose (Table 2). The day 1 dose will be split in half and the first half administered by the study clinician (to be observed by the caregiver) followed by the second half administered by the caregiver under careful observation of the study staff. Day 2-5 doses will be administered by caregivers at their home. Caregivers will be provided with visual instructions in the language of their choosing (English, Kiswahili, Luo, Kisii).

Automated daily text message drug administration reminders will be sent for the four days following discharge and caregivers asked to respond with whether or not the child took the daily dose. The response text message will be free of charge to caregivers and caregivers will be reimbursed for each response at the final study visit. Caregivers are also asked to record each administered dose on the bottle and to return bottles at the 3-month follow-up visit. The questionnaire administered during the 3-month follow up visit also includes questions about how many doses of the study drug the child received.

Follow-up Procedures

All enrolled children and primary caregivers will be scheduled to return to the health facility at 3 and 6 months following enrollment to collect clinical information and samples. Anthropometric measurements will be obtained from all children and caregivers at both follow up visits (height/length, weight, MUAC) and caregivers will be asked about any hospitalizations occurring since the last time the child was seen by study staff.

Table 2. Azithromycin dosing chart by child weight

Weight (kg)	Day 1 dose (mL)	Day 2-5 dose (mL)
2.0	0.25 x 2	0.25
2.1-2.4	0.30 x 2	0.30
2.5-2.8	0.35 x 2	0.35
2.9-3.2	0.40 x 2	0.40
3.3-3.6	0.45 x 2	0.45
3.7-4.0	0.50 x 2	0.50
4.1-4.8	0.60 x 2	0.6
4.9-5.6	0.70 x 2	0.7
5.7-6.8	0.85 x 2	0.85
6.9-8.0	1.0 x 2	1.0
8.1-9.6	1.2 x 2	1.2
9.7-11.2	1.4 x 2	1.4
11.3-13.6	1.6 x 2	1.6
13.7-16.0	2.0 x 2	2.0
16.1-19.2	2.4 x 2	2.4
19.3-23.2	2.9 x 2	2.9
23.3-25.0	3.2 x 2	3.2

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Caregivers will be provided with 400KSH (approximately \$4USD) to cover the cost of their round-trip transportation.

Transportation cost will be reimbursed at each follow-up visit. If the participant does not return at their scheduled time, study staff will attempt to make contact with the primary caregiver via cell phone; if no telephone number is provided, or if the participant cannot be reached, study staff will trace the child to the household within 2 weeks of the scheduled follow-up time.

During scheduled follow-up visits, study staff will use a standardized questionnaire to ascertain history of recent illness/morbidity, post-discharge medication use including antibiotic treatment, and current condition of the child (any hospitalizations, admission and discharge date of any hospitalization, vital status, date of death if applicable). If caregivers report a hospitalization, causes of admission, medication administration, cost of admission, and length of stay will be ascertained from both caregivers and medical records, when available.

Caregivers will be encouraged, at enrollment and at each subsequent contact, to bring the child to the study health facility at any time the child is sick. Study staff will triage children to the appropriate health facility staff and will conduct a brief unscheduled visit questionnaire to ascertain adverse event information. If the unscheduled visit leads to a hospitalization, this will trigger the completion of a hospital admission form.

If at any point during follow-up a child dies, a verbal autopsy using the Population Health Metrics Research Consortium Shortened Verbal Autopsy Questionnaire.²⁰ If the death occurred in a hospital, data from the hospital records, including cause of death, if available, will be abstracted. If a death certificate is available, cause(s) and timing of death will be abstracted.

Final causes of re-hospitalization and death will be determined after data collection is complete by an independent adjudication committee comprised of clinicians specializing in pediatrics and infectious disease. Sources of cause of re-hospitalization (medical records and caregiver report) and causes of death (causes automatically assigned from the verbal autopsy using SmartVA-Analyze [Tariff 2.0 Method]²¹, hospital records, or death certificates) will be presented to the adjudication committee for final cause assignment.

Laboratory Procedures

Stool (rectal swabs), nasopharyngeal swabs and blood will be collected as described above and undergo either immediate or future laboratory testing as described in Table 3. All biological samples will be collected by staff trained in biosafety and Good Clinical Laboratory Practice (GCLP). Samples will be processed in Kenya when technology is available at the Kenya Medical Research Institute (KEMRI) Wellcome Trust or Centre for Microbiology Research [CMR]. Metagenomic analyses and/or analyses that require technology not available in Kenya will be performed at the University of Washington. If stool culture results report *Shigella* or *Salmonella* infection, the study staff will contact the child's caregiver and encourage the caregiver to bring the child back for an evaluation and potential treatment if the child is symptomatic.

Table 3. Sample storage and processing descriptions

Specimen Collected	Purpose	Tests Performed
Stool/ flocked rectal swabs	Bacterial ID and storage for AST	Fresh samples/rectal swabs will be cultured to identify <i>Shigella</i> , <i>Salmonella</i> , <i>Campylobacter</i> , and <i>Escherichia coli</i> (<i>E.coli</i>) using standard microbiologic methods and biochemically confirmed using bioMérieux's API [®] strips. All <i>Shigella</i> , <i>Salmonella</i> , and <i>Campylobacter</i> isolates, as well as a random subset of <i>E.coli</i> isolates will undergo antibiotic resistance testing using disc diffusion for the following antibiotics: amoxicillin-clavulanic acid (augmentin), ampicillin, azithromycin, chloramphenicol, ciprofloxacin, ceftriaxone, ceftazidime, ceftazidime/ceftiofur, gentamicin, imipenem, trimethoprim-sulfamethoxazole, ceftazidime/clavulanate (ESBL), cefotaxime/clavulanate (ESBL). Categorizations of susceptible, intermediate, and resistant will be determined using zone-size cut-offs outlined in Clinical and Laboratory Standards Institute (CLSI) interpretive standards.
	Parasite detection	Fresh stool and rectal swabs will be tested for <i>Giardia</i> and <i>Cryptosporidium</i> using the immunoassay Giardia/ Cryptosporidium QUIK CHEK [™] .
	Storage	Stool/ flocked swabs and colonies of <i>E.coli</i> , <i>Shigella</i> spp., <i>Salmonella</i> spp., and <i>Campylobacter</i> spp. will be stored at -80°C.
Nasopharyngeal Swabs	Bacterial isolation, storage, and resistance testing	<i>Streptococcus pneumoniae</i> (<i>S. pneumoniae</i>) colonies will be isolated using standard microbiologic or molecular diagnostic protocols and susceptibility testing performed using standard microbiologic or molecular techniques. A random subset of <i>S. pneumoniae</i> isolates will undergo antimicrobial resistance testing using disc diffusion for the following antibiotics: amoxicillin-clavulanic acid (Augmentin), ampicillin, azithromycin, chloramphenicol, ciprofloxacin, ceftriaxone, ceftazidime, ceftiofur, gentamicin, imipenem, and trimethoprim-

		sulfamethoxazole. Categorizations of susceptible, intermediate, and resistant will be determined using zone-size cut-offs outlined in CLSI interpretive standards.
	Storage	Back-up sample and <i>S.pneumoniae</i> colonies will be will be stored at -80°C.
Blood	HIV and malaria testing	HIV testing will be performed per Kenyan National Guidelines and malaria microscopy performed using standard methods.
	Storage	Plasma and buffy coat will be stored at -80°C. Dried blood spots will be stored at room temperature.

Data Management and Confidentiality

Personal information about the participants, including medical records and data ascertained per caregiver interview, will be securely stored in files in the study offices at the study sites. Only pre-designated study staff will have access to the files. Data will be entered into an electronic database (Dacima® Electronic Data Capture) regularly by study staff. Access to the electronic database will be secured using password protected accounts for study staff. Data reports of screening, enrollment, and exclusion totals will be disseminated to the study team on a weekly basis; reports including baseline demographic characteristics, laboratory results, adherence data, and serious adverse event summaries will be distributed to study co-investigators and data monitors quarterly. Data will be regularly queried to facilitate ongoing data cleaning.

Data Analysis

Primary endpoints

The primary study endpoint is a combined outcome of mortality and hospital readmission, as re-hospitalization is highly associated with risk of subsequent poor outcome.² Re-hospitalizations that are a continuation of management from the previous hospitalization (such as elective blood transfusion) or that occur during enrollment procedures, due to a clinical deterioration post-discharge, will be excluded from the analysis. Loss to follow-up will be defined as non-attendance at both follow-up visits despite one month of active tracing and no clear evidence of death.

Secondary endpoints

1. *Cause-specific re-hospitalizations* assessed by questionnaire (maternal recall of diagnosis) at month 3 and month 6 follow-up visits and medical record review (discharge diagnosis). In cases when both sources are available, information from the medical record will be considered as the primary source. Separate analyses will be performed for each diagnosis: diarrhea, acute respiratory infection, malnutrition, or malaria.
2. *Mild, moderate, and severe events* that did not result in re-hospitalization, including diarrhea, vomiting, skin rash, lip swelling, difficulty breathing/wheeze, and seizure will be ascertained by caregivers identified by the study clinicians during clinical exams at scheduled follow-up visits or during unscheduled visits. Severity (grade 1-3) will be defined according to 2014 Division of AIDS (DAIDS) Table for Grading the Severity of Adult and Pediatric Adverse Events.
3. *Enteric pathogen carriage*, operationalized as presence of a bacterial pathogen-*Shigella* species (spp.), *Campylobacter* spp., or *Salmonella* spp. - or parasite- *Giardia* or *Cryptosporidium*-in stool or rectal swabs assessed at month 3 and month 6 follow-up visits.
4. *Streptococcus pneumoniae* (*S. pneumoniae*) isolated from nasopharyngeal swab cultures at month 3 and month 6 follow-up visits.
5. *Antibiotic resistance*, specifically resistance to azithromycin, ampicillin, augmentin, ciprofloxacin, trimethoprim-sulfamethoxazole, in *E.coli* and *S. pneumoniae* isolates, and presence of ESBL in *E.coli* isolates, from month 3 and month 6 follow-up visits.

Statistical analysis

To compare rates of re-hospitalization and mortality in the 6 months following hospital discharge among Kenyan children receiving 5-day azithromycin vs. placebo

Primary analyses will be modified intent-to-treat (mITT) based on randomization allocation to the 5-day course of azithromycin versus placebo. Cumulative incidence of death or first re-hospitalization will be compared between treatment groups using Cox proportional hazards regression. Participants will be censored at the date of their first re-hospitalization, or at the date of death. Median time to hospitalization-free survival will be

1
2 compared between randomization groups using Kaplan-Meier (K-M) survival analysis and associated log-rank
3 test. If the baseline assessment of randomization reveals an imbalance in characteristics between the
4 treatment groups, we will evaluate these variables as potential confounders in a sub-analysis secondary to the
5 mITT. Potential baseline confounders will be added stepwise in a multivariable Cox model and maintained in
6 the model if adjustment changes the hazard ratio by more than 10%. In per protocol analyses also secondary
7 to the mITT, we will compare treatment effects in groups defined by self-reported adherence to the 5-day
8 course of azithromycin (5 doses vs. < 5 doses; ≥3 doses vs. <3 doses; >1 dose vs. 1 dose only). In addition,
9 we will conduct Cox regression and K-M survival analyses for time to mortality and time to re-hospitalization as
10 separate endpoints to understand intervention effects on these outcomes individually. The assumption of
11 proportional hazards will be checked in all models using graphical methods including plotting a $\ln(-\ln(S(t)))$ plot
12 for each treatment group and assessing the parallelism of the two lines and by plotting Schoenfeld residuals
13 over time. If there is substantial missing covariate data, multiple imputation using the Markov chain Monte
14 Carlo (MCMC) method will be used to impute covariate information. Missing outcome data (death or re-
15 hospitalization) will not be imputed, but participants will be censored at the last follow-up visit therefore
16 contributing some person-time to the analysis. In sensitivity analyses, we will compare treatment effects in
17 children whose caregivers report no additional antibiotic use over follow-up and separately, who report no
18 additional azithromycin use specifically, and in subsets of children defined by age, site, and discharge
19 diagnosis.

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23 *To evaluate possible mechanism(s) by which azithromycin may affect morbidity and mortality, by comparing*
24 *reasons for re-hospitalization and change in prevalence of pathogen carriage between the randomization arms*

25 To evaluate the association between azithromycin and the rates of cause-specific re-hospitalizations
26 (hospitalization due to diarrhea, acute respiratory infection, malnutrition, or malaria) we will use Anderson-Gill
27 proportional hazards modeling with previous re-hospitalizations included as time-dependent covariates in the
28 model to capture the dependent structure of recurrence times. Because we will not have granularity in the time
29 points other than 3 months and 6 months for assessment of pathogen carriage, we will compare the
30 prevalence of a bacterial and parasitic pathogens (*Shigella*, *Salmonella*, *Campylobacter*, *Cryptosporidium*,
31 *Giardia*) at 3 and 6 months by randomization arm using generalized estimating equations (GEE) with a Poisson
32 link, exchangeable correlation structure, and will adjust for baseline presence of a bacterial pathogen. To
33 determine whether an observed association between the intervention and pathogen carriage wanes over time,
34 we will test the hypothesis that the prevalence ratios comparing carriage in intervention arms are the same at
35 the two follow-up time points using a chi-squared test.

36
37
38 *To determine whether empiric administration of azithromycin at hospital discharge increases risk of antibiotic*
39 *resistance in commensal E. coli and pneumococcal isolates from treated children and their household contacts*

40 Among children and adult household contacts in whom commensal *E. coli* and/or *S. pneumoniae* are
41 isolated, we will compare the proportion of isolates resistant to azithromycin, ampicillin, augmentin,
42 ciprofloxacin, and trimethoprim-sulfamethoxazole, between randomization arms and Contact Cohorts for each
43 arm, at 3 and 6 months using GEE with a Poisson link and exchangeable correlation structure. A chi-squared
44 test will be used to determine whether the association between intervention arm and antibiotic resistance
45 wanes over time. Because the likelihood of having a bacterial pathogen isolated may depend on baseline
46 factors, including intervention arm, we will conduct secondary analyses utilizing propensity scores to account
47 for the potential differential likelihood of having antibiotic susceptibility testing performed, which will allow us to
48 make inference to the entire study population and their contacts. Also we will compare resistance proportions
49 among children (as opposed to among isolates) where absence of an isolated bacteria is considered not
50 resistant.

51
52
53 *To identify correlates and intermediate markers of post-discharge mortality and hospital-readmission among*
54 *hospitalized children*

55 Enrollment hospital admission diagnosis, indicators of malnutrition, age, HIV-exposure and HIV-
56 infection status, sickle cell anemia, and randomization arm will be assessed in a multivariable Cox regression
57 model to identify correlates of the primary endpoint of death and/or hospital-readmission independent of the
58 treatment effect. In addition, Cox regression models will also be built for correlates of mortality and correlates
59 of re-admission individually to understand distinct cofactors for each of these outcomes.

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To determine the cost-effectiveness of post-discharge administration of a 5-day course of azithromycin in settings of varying antibiotic use, re-hospitalization rates, and mortality rates

Costs analysis: We will assess the costs of all supplies, services and equipment necessary to implement the intervention (direct medical costs). The perspective will be that of the healthcare provider, i.e. Kenya's Ministry of Health. Using WHO guidelines and its ingredients approach, we will quantify the resources and associated unit costs required to deliver a 5-day course of azithromycin, organized in standard expenditure categories: personnel (salaries), supplies including drugs, equipment, services, space and overhead. We will also measure the costs of severe child hospitalizations, the costs for the different types of personnel employed (e.g. nurses/doctors) and the time demanded from them for conducting the intervention.²² When data are missing, they will be complemented by data extracted from the literature and other available sources. Full incremental costs will be derived, with estimation of the potential healthcare cost-offset realized in avoiding severe hospitalizations. Costs will be measured in local currency (Kenyan Shilling) and converted into US\$. Our main metric will be cost per child treated. Cost-effectiveness analysis (CEA): we will develop a CEA mathematical model, and estimate incremental costs and cost-effectiveness for implementation of the intervention. The model will include two components: costs (described immediately above) and health benefits. The study will provide clinical outcomes (mortality/morbidity) over a 6-month follow-up period. Subsequently, deaths averted, life-years saved and disability-adjusted life years (DALYs) averted by the intervention will be estimated. We will estimate: a) incremental costs and b) incremental cost-effectiveness of the intervention vs. status quo. *Incremental costs* are the net sum of the costs to implement the intervention compared with status quo, and the costs averted due to the decrease in severe child hospitalizations. *Incremental cost-effectiveness ratios (ICERs)* will be estimated as cost per death averted, cost per life-year saved, and cost per DALY averted. We will use recent estimates of disability weights for estimation of DALYs.^{23 24} Short-term (over study follow-up i.e. 6 months) and longer-term time horizons (extrapolated to 1, 5, and 10 years) will be used. DALYs and costs will be discounted at 3% per year, consistent with CEA guidelines (undiscounted results will also be presented). Sources of uncertainty in the results will be explored in univariate and probabilistic sensitivity analysis.^{25 26} Finally, we will compare our findings to CEA estimates for other health interventions in sub-Saharan Africa.^{27 28}

Data and Safety Monitoring

A Data Safety and Monitoring Committee (DSMC) will be established at study initiation to monitor severe adverse events (SAEs) and to evaluate the statistical analysis plan and associated stopping rules. The DSMC includes expertise in clinical trials, statistics, child mortality assessment, ethics, and pediatric care in resource limited settings. Adverse events will be monitored by the DSMC. Monthly adverse event summaries will be sent to the DSMC safety officer and individual-child SAE forms, which include detailed medication history to evaluate possible drug interactions, will be sent to the safety officer per request. Each SAE will be assigned the plausibility of relatedness to study drug by study PIs. The data will not be presented by intervention group unless requested by DSMB safety officer. These reports will be descriptive (no statistical analyses). The DSMC will make recommendations regarding any imbalances in safety outcomes.

A single interim analysis for re-hospitalization-free survival will be prepared by the study statistician using O'Brien-Fleming boundaries for benefit and harm when 50% of expected person time (350 child-years) has been accrued. Assuming 157 events will be available at half of the person-time accrual, a z-score critical value of 2.797, or p -value < 0.005, from a Kaplan Meier log-rank test will determine the cut-off of statistical significance. A symmetric boundary will be used for benefit and harm. The DSMC will review this analysis and make a determination about study continuation. Futility will not be a basis for stopping rules because of the trials' value in understanding mechanisms of post-discharge worsening and antibiotic resistance. Assuming the DSMC decides to continue the trial after the interim analysis, an alpha of 0.045 will be used as the statistical significance boundary at the final analysis.

Statistical Power

To compare rates of re-hospitalization and mortality in the 6 months following hospital discharge among Kenyan children receiving 5-day azithromycin vs. placebo
The total sample size required was calculated for the primary endpoint of time to death or hospital re-admission within the 6 month post-discharge period, assuming an alpha level of 0.05, power of 0.80, and a

ratio of treatment to placebo random assignment of 1:1. In SSA, it is estimated that 2-15% of children aged less than 5 years died within 6 months of hospital discharge and 15.5% of children who survived discharge from the district hospital were re-admitted with the same diagnosis within 6-months.^{2 4 9} Assuming that an additional 5-10% of children are re-admitted for other conditions, we expect that re-hospitalizations will occur in 20.5 to 30.5% of children enrolled in the study. Combined with our expected fatality rate (2-15%), we expect the cumulative incidence of the combined endpoint to range from 22.5 to 45.5%.⁹ Based on a previous trial of mass drug administration of a single dose of azithromycin in which a single dose of the antibiotic was associated with a 49% reduction in risk of death, we calculated sample sizes using estimates of reduction in risk ranging from 30-50% with the cumulative incidence range of 22.5 to 45.5% in the placebo-treated group, and found the sample size required ranged from 90 to 550 children per treatment arm.¹⁶ Using the most conservative estimates of a hazard ratio of 0.70 and 22.5% prevalence of re-admission/death, we need to enroll 1100 children in the study (550 per arm) to achieve adequate power. We will recruit an additional 300 children ($\approx 20\%$) to account for possible loss to follow-up, resulting in a total planned enrollment of 1400 children, or 700 per treatment group. When considering mortality alone, and estimated mortality ranges of 2-17% among placebo-treated children, we will have $>80\%$ power to detect hazard ratios ≤ 0.5 for mortality rates of $\geq 8\%$ and hazard ratios ≤ 0.6 for mortality rates $\geq 11\%$ (Figure 1).

To evaluate possible mechanism(s) by which azithromycin may affect morbidity and mortality, by comparing reasons for re-hospitalization and prevalence of pathogen carriage between the randomization arms

We calculated the minimum detectable association between treatment arm and cause-specific re-hospitalizations (hospitalization due to diarrhea, acute respiratory infection, malnutrition, or malaria vs. any other) among enrolled children, assuming an alpha level of 0.05, power of 0.80, and a ratio of treatment to placebo of 1:1. Based on data from Kenya, re-hospitalization rates due to specific causes ranged from approximately 0.5% to 5.7% in the 6 month post-discharge period.⁴ By not conditioning on the child having the same diagnosis as the initial hospitalization, we expect the cumulative incidence of cause-specific re-hospitalizations to range from 2.5% to 10%. With this range of outcome rates, we will be able to detect hazard ratios of 0.48 to 0.70 for the effect of azithromycin on specific severe morbidities.

We expect 56% of children in the placebo group to have *S. pneumoniae* isolated from nasopharyngeal swabs, providing $\geq 80\%$ power to detect a prevalence ratio of 0.85 (or 1.15) between the two treatment arms at each time point.²⁹⁻³¹ Based on prevalences of *Shigella*, *Salmonella*, *Campylobacter*, *Cryptosporidium*, *Giardia* among asymptomatic children in Western Kenya, we expect 10% of children in the placebo group to have a bacterial pathogen isolated at each time point, resulting in $\geq 80\%$ power to detect differences in enteric pathogen prevalences of 0.67 (1.49) at each time point.³²

To determine whether empiric administration of azithromycin at hospital discharge increases risk of antibiotic resistance in commensal *E. coli* and *S. pneumoniae* isolates from treated children and their household contacts

We will select a random selection of 400 *E. coli* and 400 *S. pneumoniae* isolates (200 per arm) for β -lactam and macrolide resistance testing at each timepoint. We will also store all *S. pneumoniae*, *E. coli* isolates and other isolated bacteria from stool for potential future testing in the event that resistance prevalence is lower than expected. As shown in Table 4, we will have $> 80\%$ power to detect prevalence ratios > 1.1 , with an ability to detect the smallest effect sizes when the prevalence of resistance in the placebo group is highest. We will enroll 300 adults in the Contact Cohort for *E. coli* and *S. pneumoniae* isolation. We expect *E. coli* to be isolated from all adults and *S.pneumoniae* isolated from between 5-55%.^{29 33 34} Assuming an alpha of .05, a 1:1 ratio of testable isolates, and a prevalence of resistance of 50% in the placebo arm, we will have 80% power to detect a 1.4-fold higher prevalence to 1.9-fold higher resistance prevalence in the contacts of azithromycin-treated children.

Table 4. Power (%) to detect prevalence ratios of macrolide and β -lactamase resistance in 200 *E.coli* and 200 *S.pneumoniae* isolates per treatment group

		Resistance prevalence (%) in placebo group						
		10	20	30	40	50	60	70
Resistance prevalence (%) in azithromycin group	10							
	20	80						
	30	>99	64					
	40	>99	99	55				
	50	>99	>99	98	48			
	60	>99	>99	>99	>99	52		
	70	>99	>99	>99	>99	98	55	
	80	>99	>99	>99	>99	>99	99	64

To identify correlates and intermediate markers of post-discharge mortality and hospital-readmission among hospitalized Kenyan children

Conservatively estimating a 20% loss-to-follow-up rate in the RCT and a cumulative incidence of death or re-hospitalization of 22.5%, we will have >80% power to detect hazard ratios ≥ 1.3 between correlates and the outcome with exposure prevalences of $\geq 20\%$ or more and hazard ratios ≥ 1.5 for exposure prevalences $< 20\%$.

Study timeline

The trial began on June 28, 2016 and participant recruitment and follow-up will continue over a 36-month period, with anticipated final follow-up visit(s) occurring in June 2019. Primary analyses will be complete by February 2020.

Potential Challenges and Limitations

In order to ensure adequate power to detect a discernable clinically relevant difference between study groups in the primary outcome, we have combined hospital readmission with death. Preliminary studies suggest that sufficient numbers of children will reach this combined outcome. However, we have incorporated an interim analysis by the DSMB to review the accrued data and an adapted sample size could be considered if the combined event frequency is less than predicted. It is possible that since most children receive antibiotics during hospitalization, the benefit anticipated with the use of azithromycin based on previous trials of mass drug administration will not be observed. However, most hospitalized children are treated with penicillins, cephalosporins, gentamicin, or cotrimoxazole while in hospital and the broad spectrum of activity (including malaria prevention) and long half-life of azithromycin suggest that there may be additive treatment and/or prophylactic benefit. Similarly, children may receive azithromycin during follow up - either as treatment for an illness or because the caregiver sought out azithromycin upon learning of the hypothesis - and this azithromycin use may lead to contamination in the placebo-arm. After discharge, it is difficult to ensure adherence with the full 5-day treatment course. We will measure adherence using three different measures (text message responses, bottle check boxes, and caregiver-report at follow-up visits) although all are limited by caregiver-report. In addition, the mortality benefit of azithromycin observed in Ethiopia was from a single dose and in this study the first dose will be directly observed.¹⁶ While relying on caregiver report of mortality and morbidity may lead to bias due to outcome misclassification, this misclassification should not differ between randomization arms and therefore will be non-differential. Further hospital records will be used when available to determine diagnoses. Finally, resistance prevalence may be lower than predicted, limiting power to detect clinically relevant differences in resistance prevalence between the intervention arms. We will store all isolates in the event that a greater number of isolates are needed for antibiotic resistance testing.

Ethics and Dissemination

This study has received IRB approval by the University of Washington Human Subjects Division (HSD), KEMRI Scientific and Ethics Review Unit (SERU), and the Kenya Pharmacy and Poisons Board. The clinical trial is also registered with clinicaltrials.gov (NCT02414399). Any modifications to the study protocol or consent materials will be submitted for approval all regulatory authorities before implementation. The study is being externally monitored and a data safety and monitoring committee has been assembled to monitor patient safety and to evaluate the efficacy of the intervention. Results of this study will be disseminated by publication in a peer-reviewed scientific journal, presented at relevant academic conferences, and amongst participating partners and health facilities in Kenya.

Author's contributions

JLW, PBP, GJS, BAR, BOS, and RN conceived of this trial and developed of this study protocol. JLW and BOS are study co-Principal Investigators and PBP is the Project Director; BAR oversaw the statistical analyses plans; JBB developed the CEA plan; KDT developed procedures for ascertaining and reporting SAEs; CJM developed procedures related to blood specimen procedures and drug adherence measurement. GJS, CJM, RN, and PBP provide scientific expertise. RLB and MA are involved in collection and management of the data. MA and PBP coordinate and oversee implementation of all clinical study procedures and SK, with assistance

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from DR, oversees all laboratory procedures. All authors contributed to the development of this manuscript and/or study procedures, and to reading and approving the final version for publication.

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Competing interests statement

None of the authors or study co-investigators have any competing interests to declare.

Figure Legend

Figure 1. Power and detectable hazard ratios assuming a range of mortality rates from 2-17%

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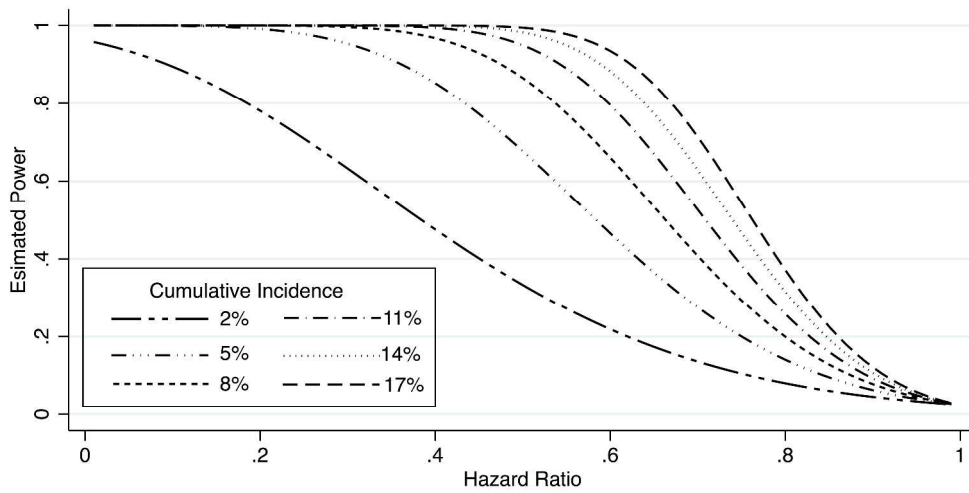


Figure 1. Power and detectable hazard ratios assuming a range of mortality rates from 2-17%

644x332mm (300 x 300 DPI)

review only



SPIRIT 2013 Checklist: Recommended items to address in a clinical trial protocol and related documents*

Section/item	Item No	Description	Addressed on page number
Administrative information			
Title	1	Descriptive title identifying the study design, population, interventions, and, if applicable, trial acronym	_____
Trial registration	2a	Trial identifier and registry name. If not yet registered, name of intended registry	_____
	2b	All items from the World Health Organization Trial Registration Data Set	_____
Protocol version	3	Date and version identifier	_____
Funding	4	Sources and types of financial, material, and other support	_____
Roles and responsibilities	5a	Names, affiliations, and roles of protocol contributors	_____
	5b	Name and contact information for the trial sponsor	_____
	5c	Role of study sponsor and funders, if any, in study design; collection, management, analysis, and interpretation of data; writing of the report; and the decision to submit the report for publication, including whether they will have ultimate authority over any of these activities	_____
	5d	Composition, roles, and responsibilities of the coordinating centre, steering committee, endpoint adjudication committee, data management team, and other individuals or groups overseeing the trial, if applicable (see Item 21a for data monitoring committee)	_____

1 **Introduction**

2

3 Background and 6a Description of research question and justification for undertaking the trial, including summary of relevant
4 rationale studies (published and unpublished) examining benefits and harms for each intervention _____

5

6 6b Explanation for choice of comparators _____

7

8 Objectives 7 Specific objectives or hypotheses _____

9

10 Trial design 8 Description of trial design including type of trial (eg, parallel group, crossover, factorial, single group),
11 allocation ratio, and framework (eg, superiority, equivalence, noninferiority, exploratory) _____

12

13

14 **Methods: Participants, interventions, and outcomes**

15

16 Study setting 9 Description of study settings (eg, community clinic, academic hospital) and list of countries where data will
17 be collected. Reference to where list of study sites can be obtained _____

18

19 Eligibility criteria 10 Inclusion and exclusion criteria for participants. If applicable, eligibility criteria for study centres and
20 individuals who will perform the interventions (eg, surgeons, psychotherapists) _____

21

22 Interventions 11a Interventions for each group with sufficient detail to allow replication, including how and when they will be
23 administered _____

24

25 11b Criteria for discontinuing or modifying allocated interventions for a given trial participant (eg, drug dose
26 change in response to harms, participant request, or improving/worsening disease) _____

27

28 11c Strategies to improve adherence to intervention protocols, and any procedures for monitoring adherence
29 (eg, drug tablet return, laboratory tests) _____

30

31 11d Relevant concomitant care and interventions that are permitted or prohibited during the trial _____

32

33 Outcomes 12 Primary, secondary, and other outcomes, including the specific measurement variable (eg, systolic blood
34 pressure), analysis metric (eg, change from baseline, final value, time to event), method of aggregation (eg,
35 median, proportion), and time point for each outcome. Explanation of the clinical relevance of chosen
36 efficacy and harm outcomes is strongly recommended _____

37

38 Participant timeline 13 Time schedule of enrolment, interventions (including any run-ins and washouts), assessments, and visits for
39 participants. A schematic diagram is highly recommended (see Figure) _____

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1 Sample size 14 Estimated number of participants needed to achieve study objectives and how it was determined, including _____
 2 clinical and statistical assumptions supporting any sample size calculations

3
 4 Recruitment 15 Strategies for achieving adequate participant enrolment to reach target sample size _____
 5

6 7 **Methods: Assignment of interventions (for controlled trials)**

8 9 Allocation:

10 Sequence 16a Method of generating the allocation sequence (eg, computer-generated random numbers), and list of any _____
 11 generation factors for stratification. To reduce predictability of a random sequence, details of any planned restriction
 12 (eg, blocking) should be provided in a separate document that is unavailable to those who enrol participants
 13 or assign interventions
 14

15
 16 Allocation 16b Mechanism of implementing the allocation sequence (eg, central telephone; sequentially numbered, _____
 17 concealment opaque, sealed envelopes), describing any steps to conceal the sequence until interventions are assigned
 18 mechanism
 19

20 Implementation 16c Who will generate the allocation sequence, who will enrol participants, and who will assign participants to _____
 21 interventions
 22

23 Blinding (masking) 17a Who will be blinded after assignment to interventions (eg, trial participants, care providers, outcome _____
 24 assessors, data analysts), and how
 25

26
 27 17b If blinded, circumstances under which unblinding is permissible, and procedure for revealing a participant's _____
 28 allocated intervention during the trial
 29

30 31 32 **Methods: Data collection, management, and analysis**

33
 34 Data collection 18a Plans for assessment and collection of outcome, baseline, and other trial data, including any related _____
 35 methods processes to promote data quality (eg, duplicate measurements, training of assessors) and a description of
 36 study instruments (eg, questionnaires, laboratory tests) along with their reliability and validity, if known.
 37 Reference to where data collection forms can be found, if not in the protocol
 38

39
 40 18b Plans to promote participant retention and complete follow-up, including list of any outcome data to be _____
 41 collected for participants who discontinue or deviate from intervention protocols
 42

1	Data management	19	Plans for data entry, coding, security, and storage, including any related processes to promote data quality	_____
2			(eg, double data entry; range checks for data values). Reference to where details of data management	
3			procedures can be found, if not in the protocol	
4				
5	Statistical methods	20a	Statistical methods for analysing primary and secondary outcomes. Reference to where other details of the	_____
6			statistical analysis plan can be found, if not in the protocol	
7				
8		20b	Methods for any additional analyses (eg, subgroup and adjusted analyses)	_____
9				
10		20c	Definition of analysis population relating to protocol non-adherence (eg, as randomised analysis), and any	_____
11			statistical methods to handle missing data (eg, multiple imputation)	
12				
13				
14				

Methods: Monitoring

15				
16				
17	Data monitoring	21a	Composition of data monitoring committee (DMC); summary of its role and reporting structure; statement of	_____
18			whether it is independent from the sponsor and competing interests; and reference to where further details	
19			about its charter can be found, if not in the protocol. Alternatively, an explanation of why a DMC is not	
20			needed	
21				
22		21b	Description of any interim analyses and stopping guidelines, including who will have access to these interim	_____
23			results and make the final decision to terminate the trial	
24				
25				
26	Harms	22	Plans for collecting, assessing, reporting, and managing solicited and spontaneously reported adverse	_____
27			events and other unintended effects of trial interventions or trial conduct	
28				
29	Auditing	23	Frequency and procedures for auditing trial conduct, if any, and whether the process will be independent	_____
30			from investigators and the sponsor	
31				
32				

Ethics and dissemination

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34				
35	Research ethics	24	Plans for seeking research ethics committee/institutional review board (REC/IRB) approval	_____
36	approval			
37				
38	Protocol	25	Plans for communicating important protocol modifications (eg, changes to eligibility criteria, outcomes,	_____
39	amendments		analyses) to relevant parties (eg, investigators, REC/IRBs, trial participants, trial registries, journals,	
40			regulators)	
41				
42				
43				

1	Consent or assent	26a	Who will obtain informed consent or assent from potential trial participants or authorised surrogates, and	_____
2			how (see Item 32)	
3				
4		26b	Additional consent provisions for collection and use of participant data and biological specimens in ancillary	_____
5			studies, if applicable	
6				
7	Confidentiality	27	How personal information about potential and enrolled participants will be collected, shared, and maintained	_____
8			in order to protect confidentiality before, during, and after the trial	
9				
10	Declaration of	28	Financial and other competing interests for principal investigators for the overall trial and each study site	_____
11	interests			
12				
13				
14	Access to data	29	Statement of who will have access to the final trial dataset, and disclosure of contractual agreements that	_____
15			limit such access for investigators	
16				
17	Ancillary and post-	30	Provisions, if any, for ancillary and post-trial care, and for compensation to those who suffer harm from trial	_____
18	trial care		participation	
19				
20	Dissemination policy	31a	Plans for investigators and sponsor to communicate trial results to participants, healthcare professionals,	_____
21			the public, and other relevant groups (eg, via publication, reporting in results databases, or other data	
22			sharing arrangements), including any publication restrictions	
23				
24				
25		31b	Authorship eligibility guidelines and any intended use of professional writers	_____
26				
27		31c	Plans, if any, for granting public access to the full protocol, participant-level dataset, and statistical code	_____
28				
29				
30	Appendices			
31				
32	Informed consent	32	Model consent form and other related documentation given to participants and authorised surrogates	_____
33	materials			
34				
35	Biological	33	Plans for collection, laboratory evaluation, and storage of biological specimens for genetic or molecular	_____
36	specimens		analysis in the current trial and for future use in ancillary studies, if applicable	
37				

*It is strongly recommended that this checklist be read in conjunction with the SPIRIT 2013 Explanation & Elaboration for important clarification on the items. Amendments to the protocol should be tracked and dated. The SPIRIT checklist is copyrighted by the SPIRIT Group under the Creative Commons "[Attribution-NonCommercial-NoDerivs 3.0 Unported](https://creativecommons.org/licenses/by-nc-nd/3.0/)" license.