

**TopA, the *Sulfolobus solfataricus* topoisomerase III, is a decatenase.**

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**SUPPLEMENTARY DATA**

**Figure legends**

**Figure S1.** Purification of *S. solfataricus* recombinant TopA. Coomassie-stained SDS-PAGE of the purified enzyme. Approximately 1 µg of the protein was loaded onto the gel.

**Figure S2.** Activity of *Sso*TopA as a function of temperature. TopA was incubated 8 min with a protein/pTZ18R DNA molar ratio of 2:1 in the standard conditions and at the temperature indicated above each corresponding lane. The DNA control (C) was incubated for 8 min at 75 °C in the absence of TopA. The reaction was performed at 100 mM of NaCl. The reaction products were analysed by one dimensional gel electrophoresis: OC indicates open circular (nicked) DNA, Rel, the relaxed topoisomers and -SC the negatively supercoiled DNA.

**Figure S3.** Activity of *Sso*TopA as a function of the magnesium concentration. TopA was incubated 30 min at 75°C with a protein/pTZ18R DNA molar ratio of 1:1 in the standard conditions with the concentration of MgCl<sub>2</sub> indicated above. The reaction products were analysed by one dimensional gel electrophoresis: OC indicates open circular (nicked) DNA, Rel, the relaxed topoisomers and -SC the negatively supercoiled DNA.

**Figure S4.** Annealing properties of the FI\*\* produced by *SsoTopA*. *SsoTopA* was incubated with pTZ18R DNA (molar ratio of 1:1) for 5 min at 95°C in the standard conditions with 15 mM MgCl<sub>2</sub>. After incubation, the samples were quickly cooled and kept at 4°C except when indicated and finally 0.5% SDS (final concentration) was added (lanes 2-5). SDS was added before the incubation as a control (lane 1). An additional incubation at 65°C for 15 min was realized for lanes 4 and 5. For all the samples, 25 mg/ml bromophenol blue, and 15% sucrose (final concentrations) were added. Samples corresponding to lanes 2 and 4 were further equilibrated at room temperature for 15 min while samples corresponding to lanes 3 and 4 were incubated 2 min at 95°C just prior loading. The reaction products were analysed by agarose gel electrophoresis (2%) in TEP buffer. OC indicates open circular (nicked) DNA and -SC the negatively supercoiled DNA and the particular FI\* and FI\*\* forms.

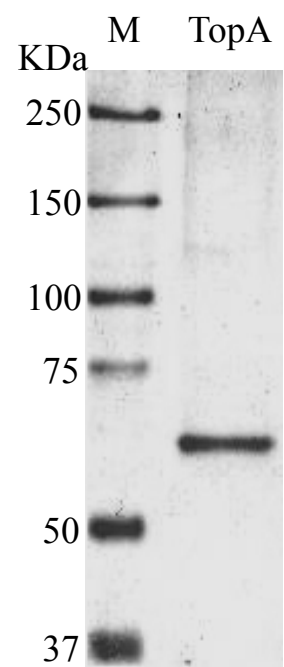
**Figure S5.** Decatenation of multi-linked DNA circles by *SsoTopA*. 600 ng of the multi-linked DNA substrate was incubated with *SsoTopA* for 20 min at 90 °C then cooled at 4°C (lane 1). The samples were further incubated for 30 min at 37°C in the presence of Nb.BbvCI (lane 2) or NdeI (lane 3). The panel B is a zoom of the squared zone from panel A. The DNAs ran in a 2% agarose gel at about 2.5 V.cm<sup>-1</sup> for 3 hours. The different DNA bands were attributed according to their size and topological forms : PP corresponds to the parental plasmid, LC to the decatenated large circle, and mC to the minicircle while OC is the open circular form, Lin the linear, Rel the relaxed and -SC the negatively supercoiled. M corresponds to the GeneRuler 1kb ladders.

**Figure S6.** DNA extension fluctuation in the absence of *SsoTopA*. (A) Schematic representation of the single molecule assay using magnetic tweezers explaining the

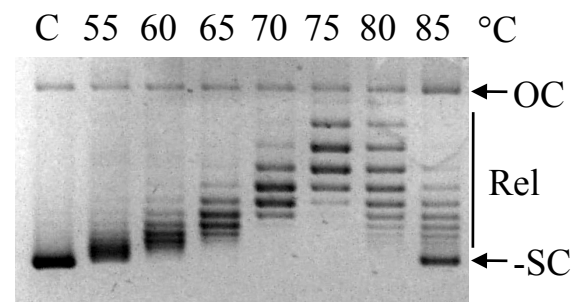
occurrence of DNA extension fluctuation. (B) Time-trace of the 3 kb DNA extension variation at 45 °C at an extended force of 0.45 pN in the absence of *SsoTopA* (control of the figure 6). The variation of extension is indicated as “fluctuation” in green. (C) Same extension fluctuation using the DNA containing the permanent single-stranded bubble region. (D) Corresponding time-trace of the DNA extension variation without *SsoTopA* (control of Figure 7).

**Figure S7.** Decatenation of kDNA by TopA of *S. solfataricus*. (A) TopA is added (0.5 ng) just before the incubation at indicated temperature for 30 min. (B) TopA (0.5 ng) is incubated at 85°C for 30 min in the presence of the indicated amount of SSB protein (ng). The lane T corresponds to kDNA incubated without TopA. The reactions were stopped and kDNA products were separated by a 2% agarose gel electrophoresis.

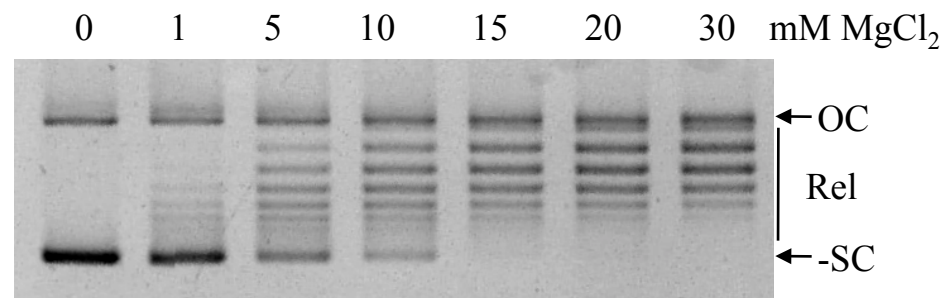
# Supplementary Figure S1



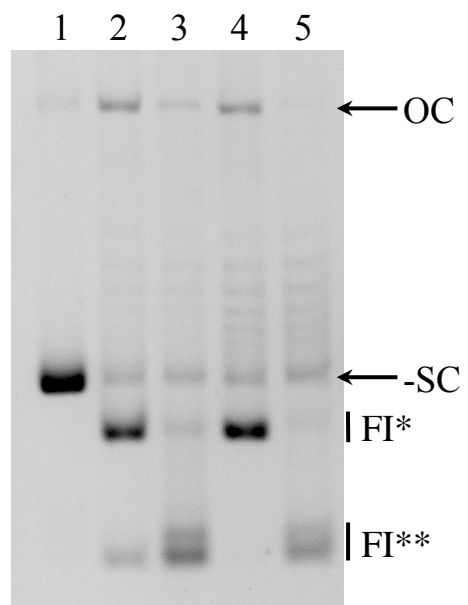
## Supplementary Figure S2



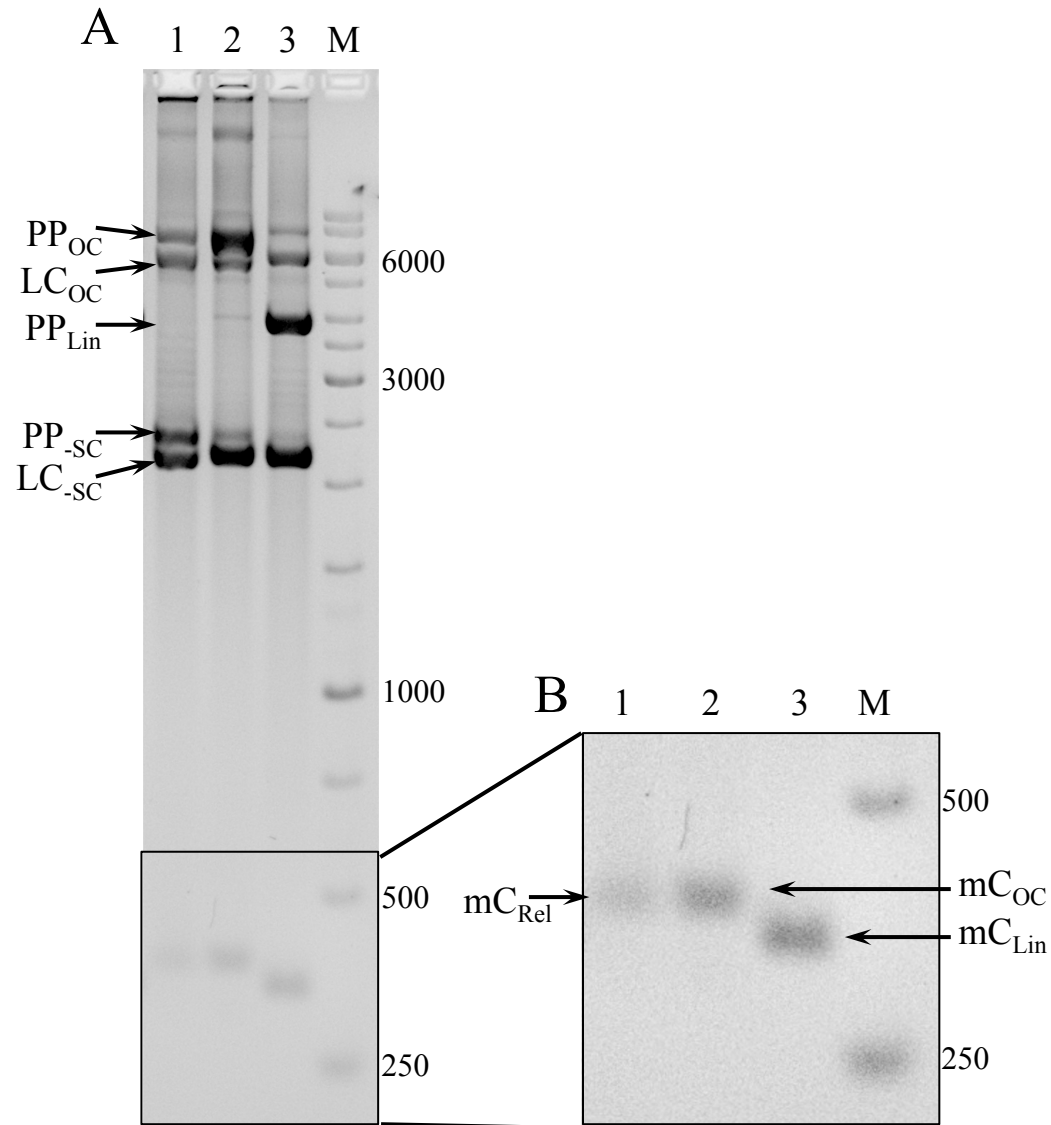
# Supplementary Figure S3



## Supplementary Figure S4



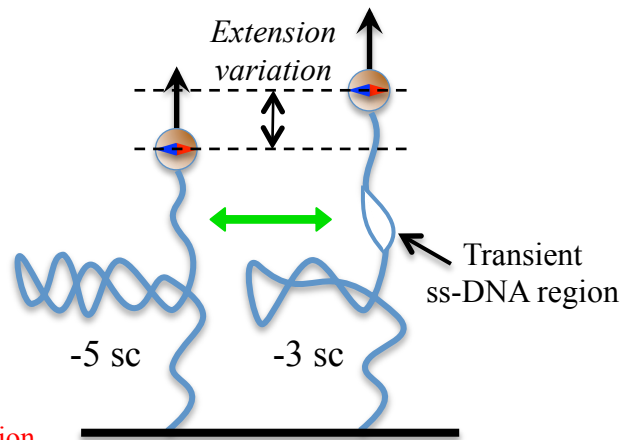
# Supplementary Figure S5



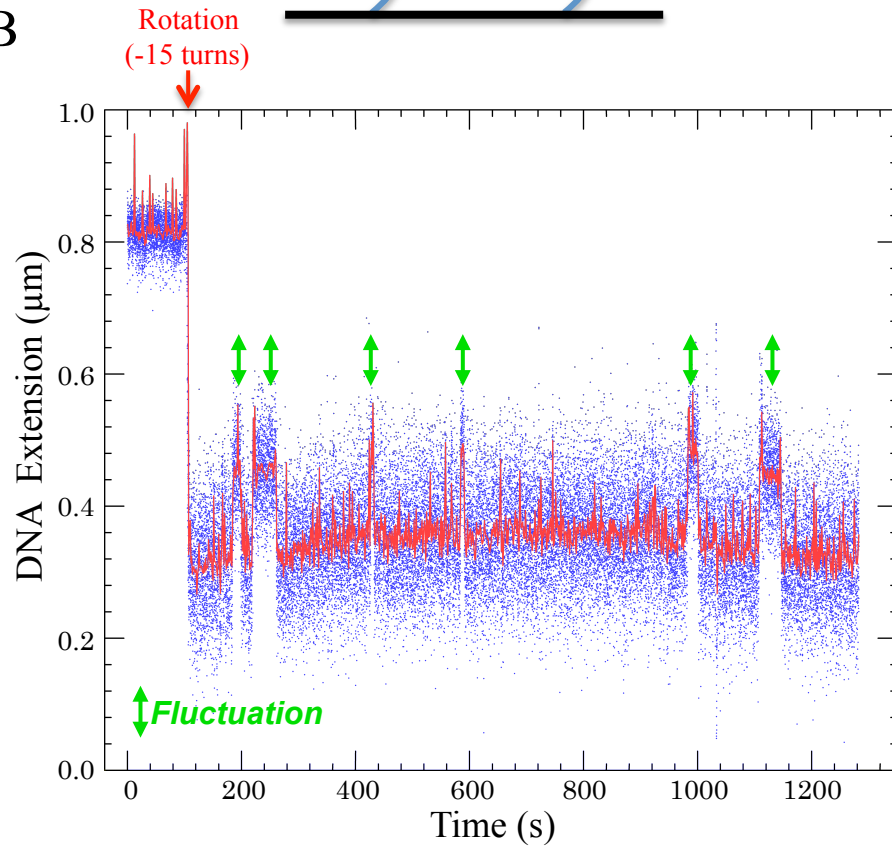


# Supplementary Figure S6

A



B



C



D

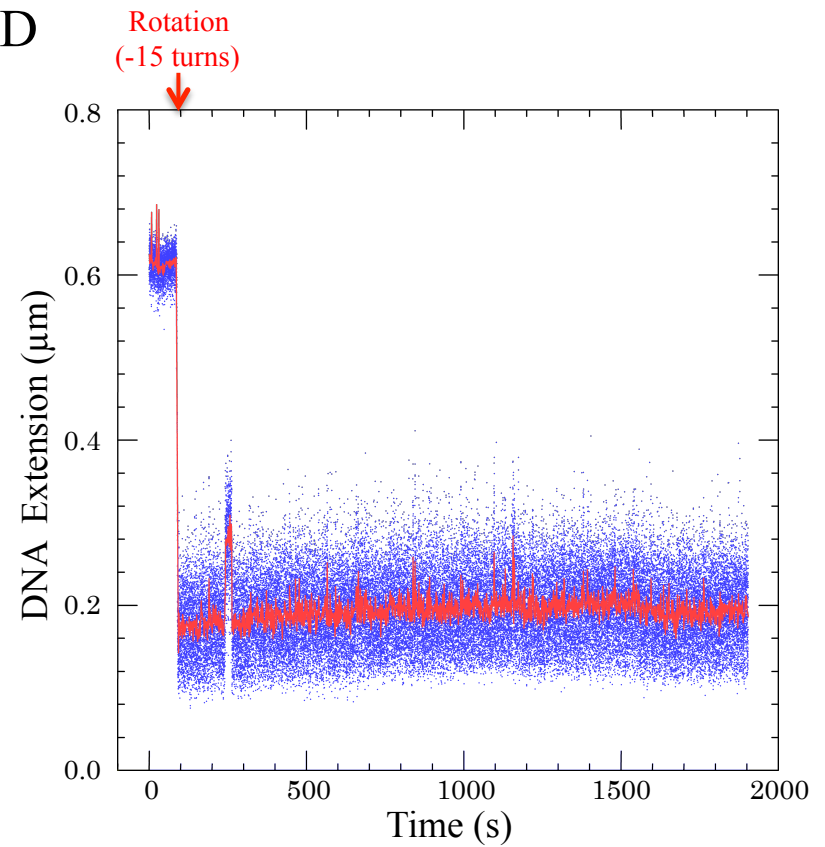


Figure S7

