

## SUPPLEMENTAL DATA

### Melanocortin 1 Receptor Deficiency Promotes Atherosclerosis in Apolipoprotein E<sup>-/-</sup> Mice

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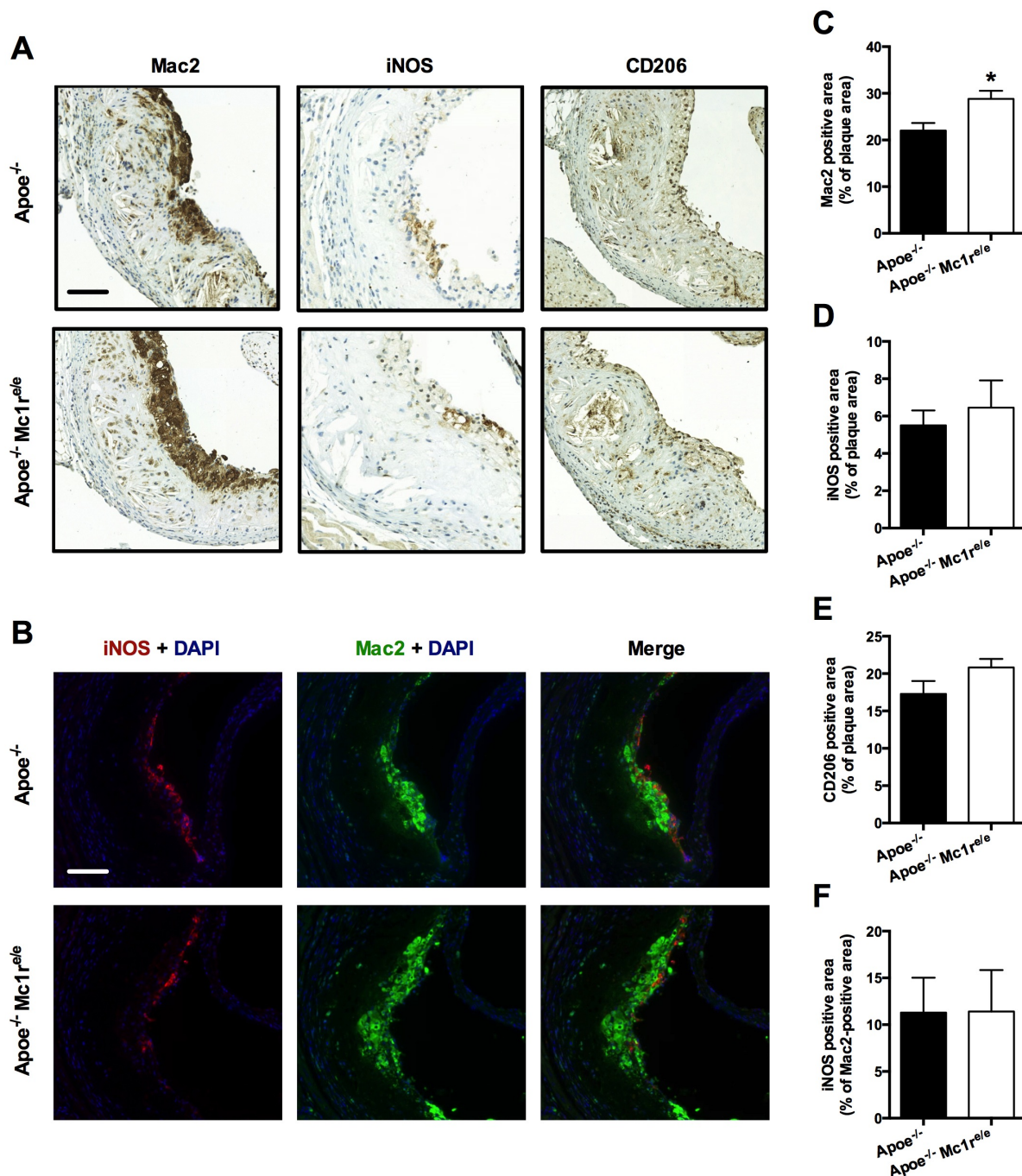
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**Supplemental Table I. Quantitative RT-PCR primers for mouse genes.**

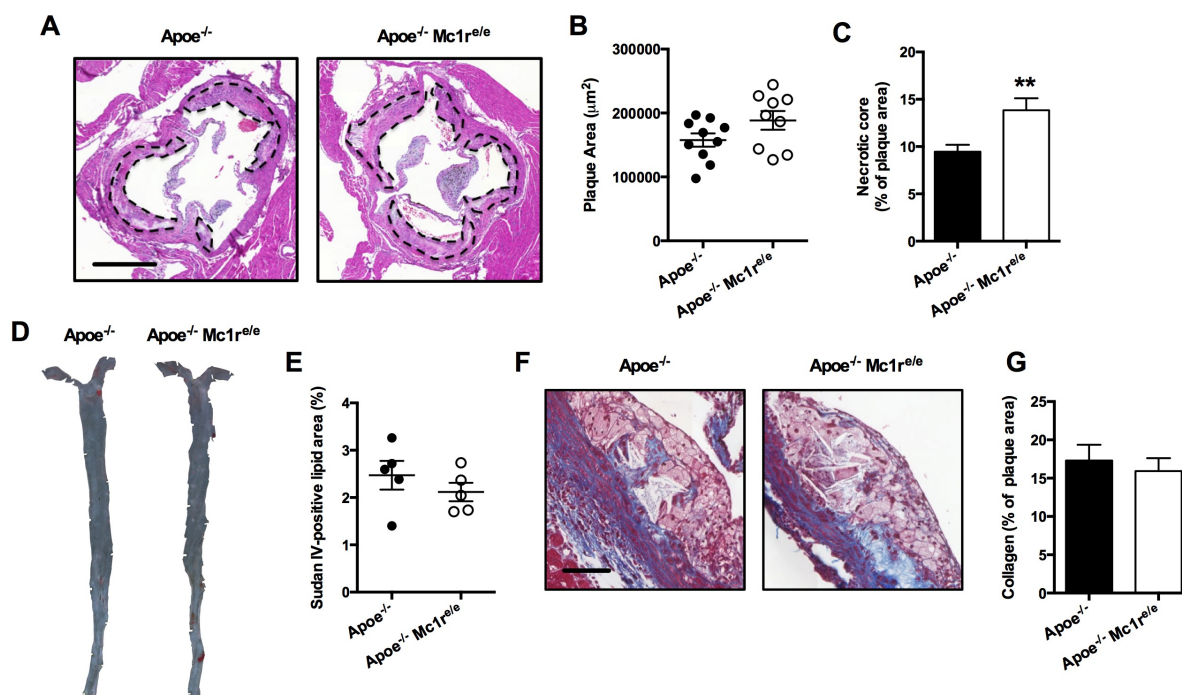
<b>Gene name</b> Accession number	<b>5'-3' primer sequence</b>
<b>ABCA1</b> NM_013454.3	Forward: gcagatcaagcatcccaact Reverse: ccagagaatgttcattgtcca
<b>ABCG1</b> NM_009593.2	Forward: gggctgaactgccctacct Reverse: tactcccctgatgccacttc
<b>ABCG5</b> NM_031884.2	Forward: tggatccaacacctctatgctaaa Reverse: ggcaggtttctcgatgaactg
<b>ABCG8</b> NM_026180.3	Forward: tgcccacctccacatgtc Reverse: atgaagccggcagtaaggtaga
<b>ACTA2</b> NM_007392.3	Forward: agattgtgcgcatcaaag Reverse: gcagactccataccgataaagga
<b>ACTB</b> NM_007393.5	Forward: tccatcatgaagtgtgacgt Reverse: gagcaatgatcttgatctca
<b>BAAT</b> NM_007519.3	Forward: tagagcacaccacgttctctg Reverse: gcacaggctcatcaacaaga
<b>BAL (Slc27a5)</b> NM_009512.2	Forward: tgtgtgtaaggaacctgga Reverse: acccgacaactttgtgaag
<b>BSEP</b> NM_021022.3	Forward: aagctacatctgccttagacacagaa Reverse: caatacaggctccgacctctct
<b>CCR2</b> NM_009915.2	Forward: agagagctgcagcaaaaagg Reverse: ggaaagaggcagttgcaaag
<b>CCR5</b> NM_009917.5	Forward: gagacatccgttccccctac Reverse: gtcggaactgaccttgaaa
<b>COL1A1</b> NM_007742.4	Forward: gtcctcttaggggacct Reverse: ccacgtctcaccattgggg
<b>COL1A2</b> NM_007743.3	Forward: tgcaacttcgtgcctagc Reverse: acgtggtcctctgtctcca
<b>COL3A1</b> NM_009930.2	Forward: ctaaaattctgccacccccgaa Reverse: aggatcaaccagattctccactc
<b>COL4A1</b> NM_009931.2	Forward: ctggcacaagaggacgag Reverse: acgtggccgagaattcacc
<b>CD36</b> NM_001159558.1	Forward: ccaagctattgcgacatgatt Reverse: tctcaatgtccgagactttca
<b>CD62P</b> NM_011347.2	Forward: gaggaagaaagccagacg Reverse: ggcgtccaggaaccttt
<b>CX3CR1</b> NM_009987.4	Forward: cagcatcgaccggtacctt Reverse: gctgcactgtccggtgtt

<b>Gene name</b> Accession number	<b>5'-3' primer sequence</b>
<b>CYP7A1</b> NM_007824.2	Forward: gatcctctgggcatctcaag Reverse: agaggctgctttcattgctt
<b>CYP7B1</b> NM_007825.4	Forward: gaaaactcttcaaaggcaacatgg Reverse: actggaaagggttcagaacaaatg
<b>CYP8B1</b> NM_010012.3	Forward: gcctcaagtatgatcggttcct Reverse: gatcttcttggcccgactgtaga
<b>CYP27A1</b> NM_024264.5	Forward: gcctcacctatgggatcttca Reverse: tcaaagcctgacgcagatg
<b>DHCR7</b> NM_007856.2	Forward: gaggcgtccaagaagggtg Reverse: gcagcccattcacctcatalc
<b>FXR</b> NM_001163700.1	Forward: tccggacattcaaccatcac Reverse: tcactgcacatcccagatctc
<b>HMGCR</b> NM_008255.2	Forward: tgattggagttggcaccat Reverse: tggccaacactgacatgc
<b>HNF4a</b> NM_008261.3	Forward: accaagagggtccatgggtgtt Reverse: gtgccgagggacgatgtag
<b>ICAM1</b> NM_010493.3	Forward: tggccctggtcaccggttgat Reverse: aacagttcacctgcacggacca
<b>LDLR</b> NM_010700.3	Forward: gcgtaaagaggaggacactgtt Reverse: ccaatctgtccagttacatgaagc
<b>LRH-1</b> NM_030676.3	Forward: tgggaaggaagggacaatctt Reverse: cgagactcaggaggtgttgaa
<b>LXR<math>\alpha</math></b> NM_013839.4	Forward: tgggatgtccacgagtgactgtt Reverse: tccctaatgtctacggaaggctct
<b>NTCP</b> NM_011387.2	Forward: gaagtccaaaaggccacactatgt Reverse: acagccacagagaggagaaag
<b>PECAM1</b> NM_008816.3	Forward: cgggtttcagcgagatcc Reverse: actcgacaggatggaaatcac
<b>S29</b> NM_009093.2	Forward: atgggtcaccagcagctcta Reverse: agcctatgtccttcgcgtact
<b>SR-A</b> NM_031195.2	Forward: gggagtgtaggcggatca Reverse: tcacagatttgccccact
<b>SR-BI</b> NM_016741.2	Forward: gccatcatctgccaaact Reverse: tcctgggagcccttttact
<b>SREBP-2</b> NM_033218.1	Forward: ccaaagaaggagagaggcgg Reverse: cgccagactgtgcatcttg
<b>VCAM1</b> NM_011693.3	Forward: ggtcttgggagcctcaacggt Reverse: agggccatggagtcaccgatt

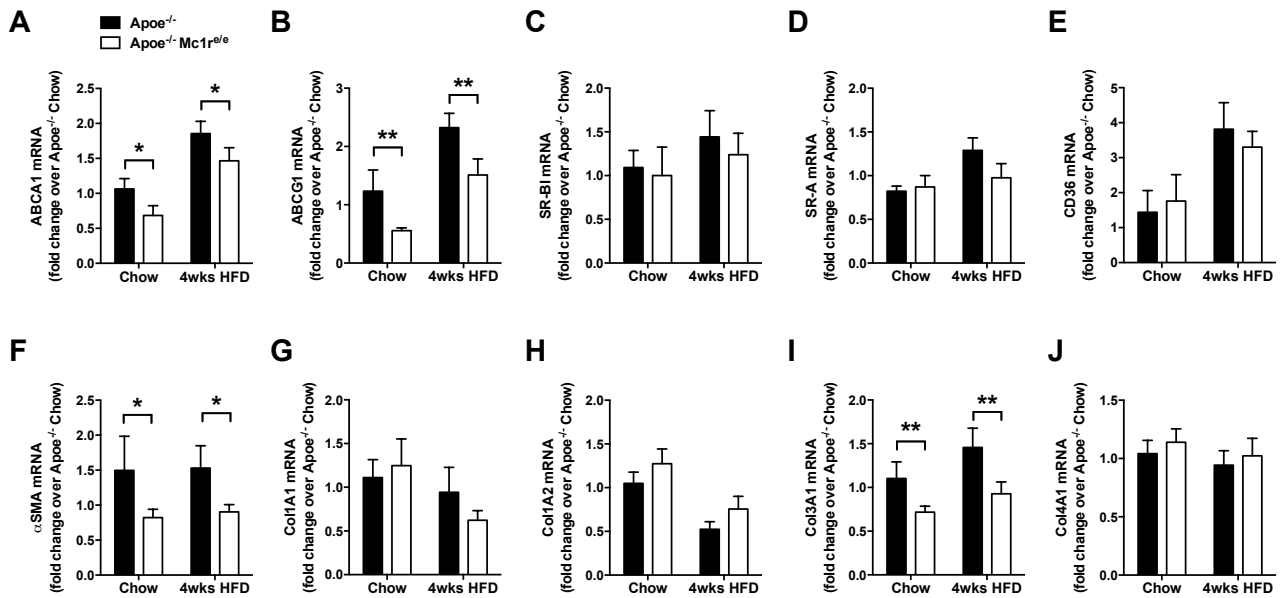


**Supplemental Figure I. Melanocortin 1 receptor deficiency enhances macrophage content in aortic plaques of high-fat diet (HFD)-fed Apoe<sup>-/-</sup> mice without affecting macrophage polarization.** **A**, Representative images of Mac2, iNOS and CD206 immunostaining of the aortic sinus of Apoe<sup>-/-</sup> and Apoe<sup>-/-</sup> Mc1r<sup>e/e</sup> mice fed a high-fat diet (HFD) for 12 wks. Scale bar, 100  $\mu$ m. **B**, Representative immunofluorescent stainings of iNOS and Mac2 in the aortic roots of HFD-fed Apoe<sup>-/-</sup> and Apoe<sup>-/-</sup> Mc1r<sup>e/e</sup> mice. Scale bar, 100  $\mu$ m. **C** through **F**, Quantification of Mac2-, iNOS- and CD206 positive areas (expressed as % of plaque area) as well as iNOS-positive macrophage area (% of Mac2-positive area) in the aortic sinuses. n=5-10 mice per group in each graph. \* P<0.05 versus Apoe<sup>-/-</sup> mice by Student's t-test. Values are mean  $\pm$  SEM.

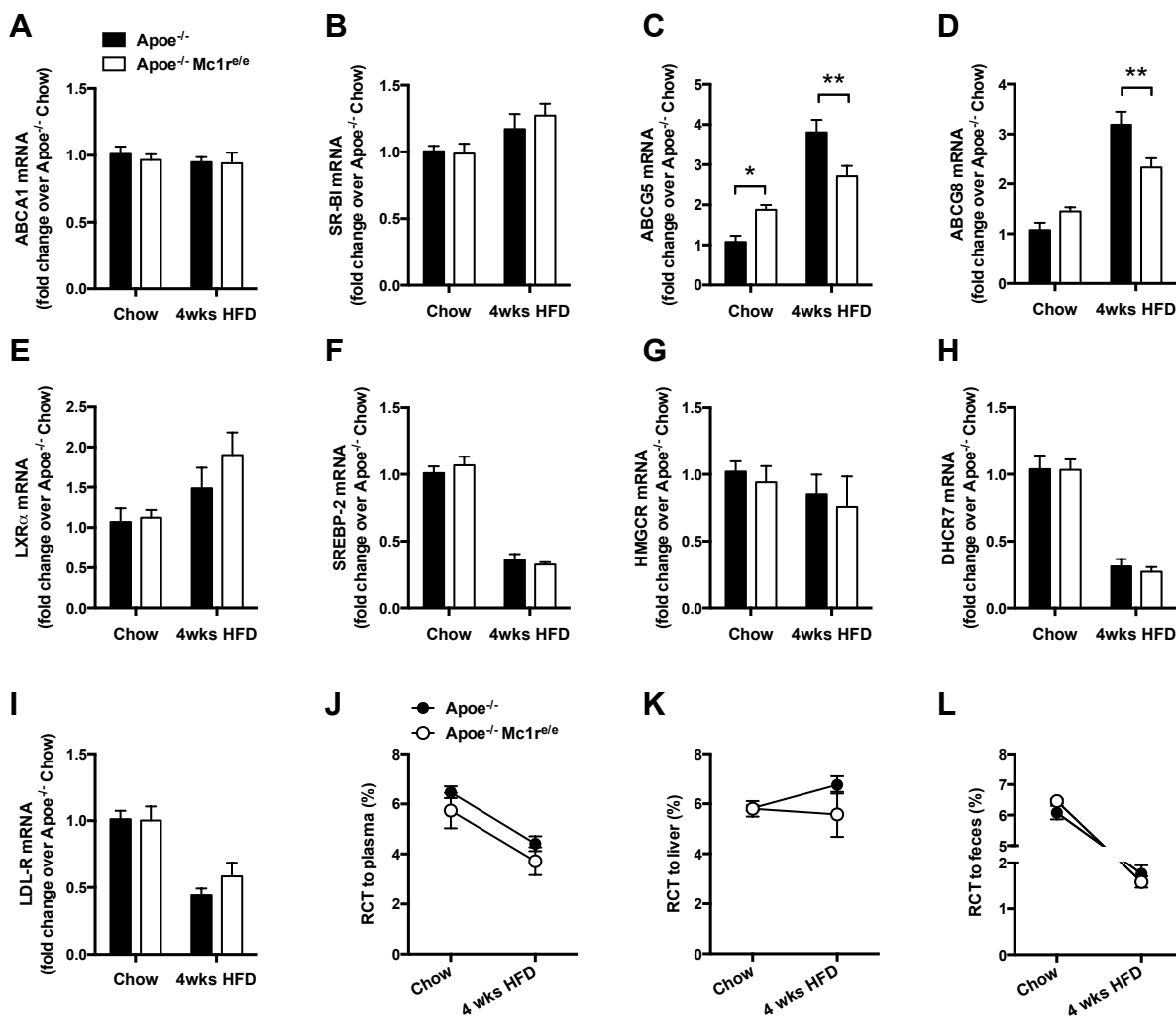




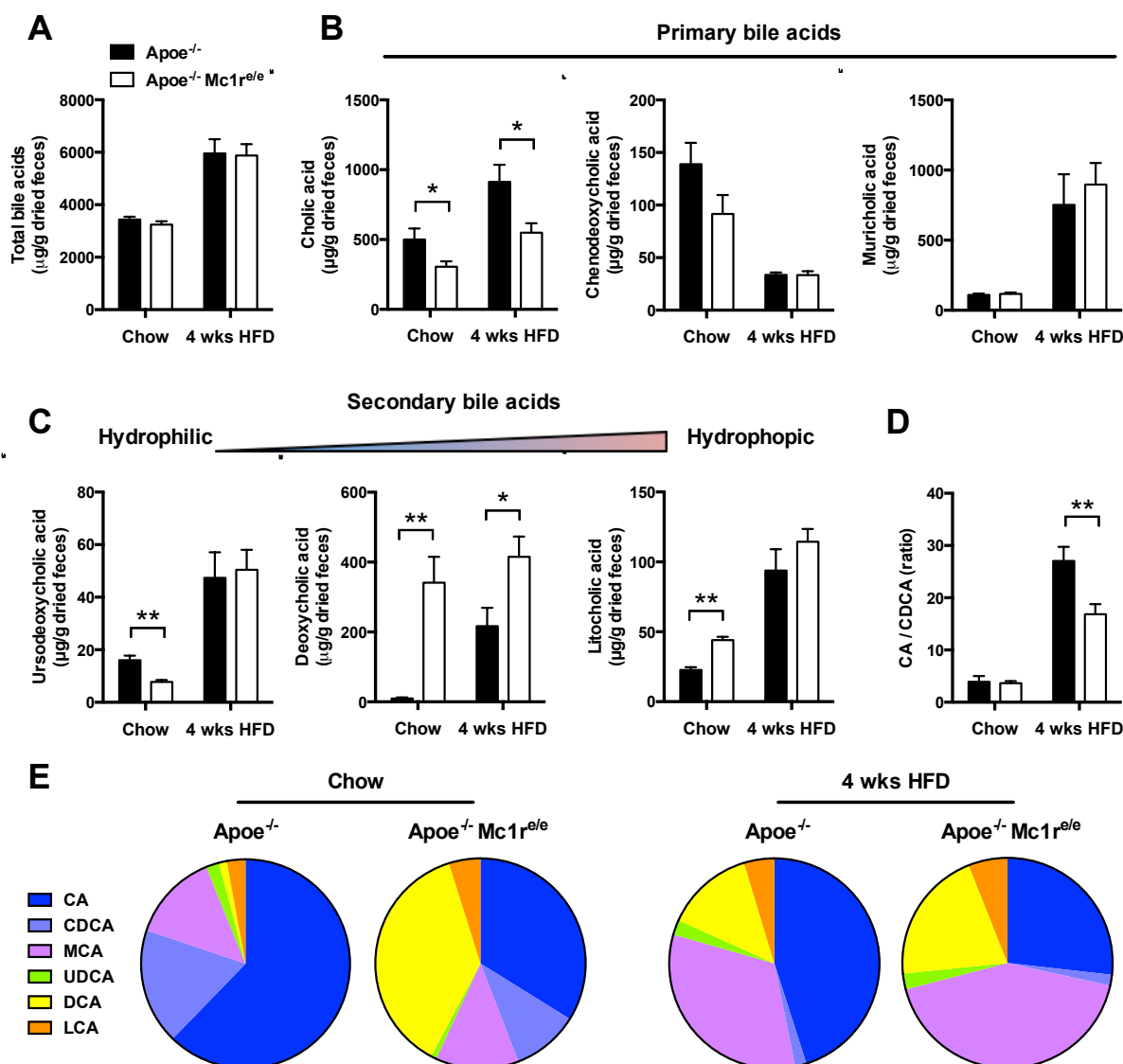
**Supplemental Figure II. Atherosclerotic plaque size and composition in chow-fed *Apoe*<sup>-/-</sup> and *Apoe*<sup>-/-</sup> *Mc1r*<sup>e/e</sup> mice.** **A**, Representative images of hematoxylin and eosin (H&E) staining of the aortic sinus of *Apoe*<sup>-/-</sup> and *Apoe*<sup>-/-</sup> *Mc1r*<sup>e/e</sup> mice fed a normal chow diet. Mice were euthanized at the age of 6 months and were age-matched with the 12 wks HFD mice. Scale bar, 500  $\mu\text{m}$ . **B**, Quantification of plaque area in aortic sinuses. **C**, Quantification of acellular necrotic core areas as percentage of total plaque area. **D** and **E**, Representative en face Sudan IV stainings and quantification of Sudan IV-positive lipid area in the aorta. **F**, Representative images of Masson's trichrome staining in the aortic sinus. **G**, Quantification of plaque collagen content as percentage of total plaque area. Scale bar, 100  $\mu\text{m}$ .  $n=8-10$  mice per group in each graph, except for E ( $n=5$  per group). \*\*  $P<0.01$  versus *Apoe*<sup>-/-</sup> mice by Student's t-test. Values are mean  $\pm$  SEM.



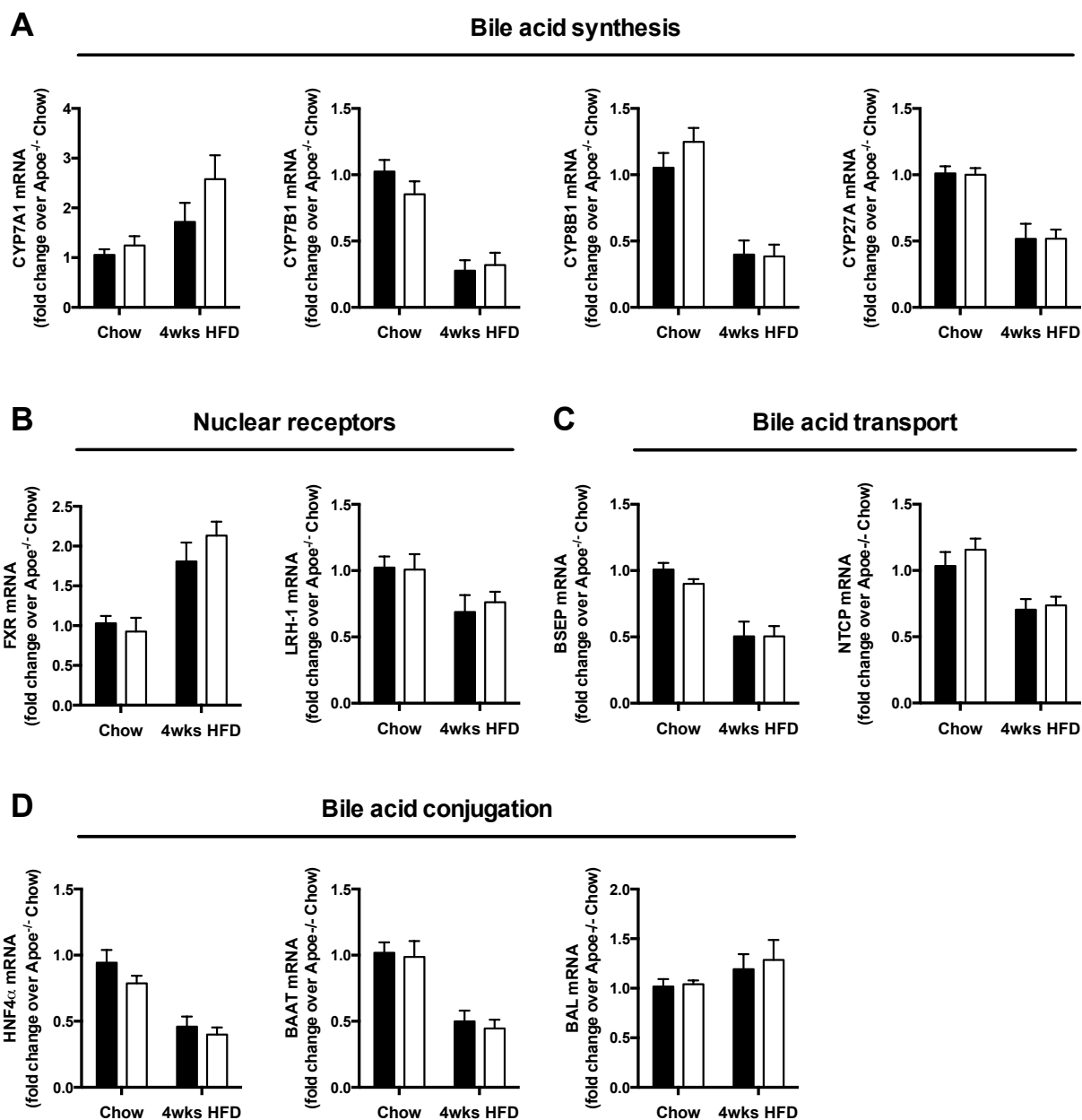
**Supplemental Figure III. Expression of cholesterol transport and stability-related genes in the aorta of Apoe<sup>-/-</sup> and Apoe<sup>-/-</sup> Mc1r<sup>e/e</sup> mice.** Quantitative RT-PCR analysis of the indicated genes in the aorta of Apoe<sup>-/-</sup> and Apoe<sup>-/-</sup> Mc1r<sup>e/e</sup> mice fed a normal chow diet or high-fat diet (HFD) for 4 weeks. **A**, ABCA1, ATP-binding cassette sub-family A member 1. **B**, ABCG1, ATP-binding cassette sub-family G member 1. **C**, SR-BI, scavenger receptor class B member 1. **D**, SR-A, macrophage scavenger receptor 1. **E**, CD36, scavenger receptor class B member 3. **F**, αSMA, alpha smooth muscle actin. **G** through **J**, Collagen, type I, alpha 1 (Col1A1) and alpha 2 (Col1A2), type III, alpha 1 (Col3A1) and type IV, alpha 1 (Col4A1). \* P<0.05 and \*\* P<0.01 for genotype effect by 2-way ANOVA. Interaction between the genotype and diet as well as pairwise comparisons within diet groups by Bonferroni *post hoc* tests were non-significant. Data are mean ± SEM, n=7-8 mice per group.



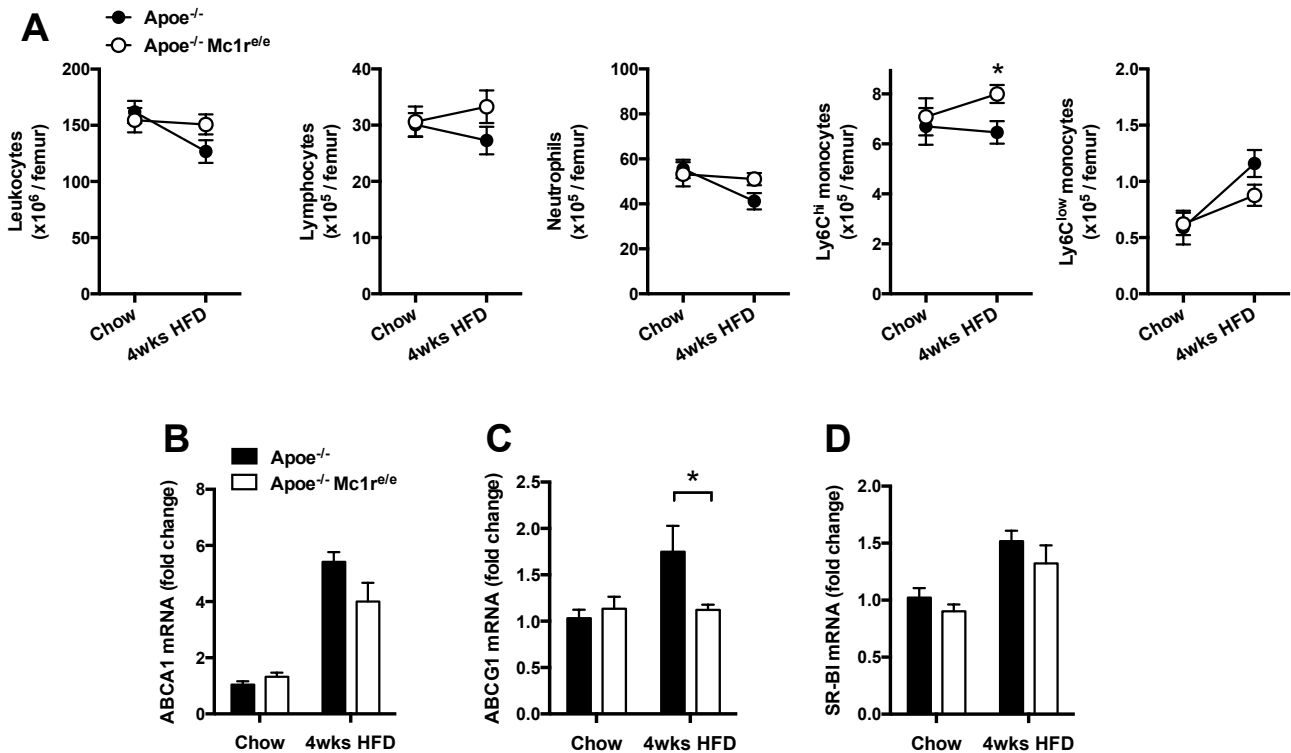
**Supplemental Figure IV. Hepatic expression of cholesterol transport and synthesis genes in Apoe<sup>-/-</sup> and Apoe<sup>-/-</sup> Mc1r<sup>e/e</sup> mice.** Quantitative RT-PCR analysis of genes involved in cholesterol transport and synthesis in the liver of Apoe<sup>-/-</sup> and Apoe<sup>-/-</sup> Mc1r<sup>e/e</sup> mice fed a normal chow diet or high-fat diet (HFD) for 4 weeks. **A**, ABCA1, ATP-binding cassette sub-family A member 1. **B**, SR-BI, scavenger receptor class B member 1. **C**, ABCG5, ATP-binding cassette sub-family G member 5. **D**, ABCG8, ATP-binding cassette sub-family G member 8. **E**, LXR $\alpha$ , liver X receptor alpha. **F**, SREBP-2, sterol regulatory element-binding protein 2. **G**, HMGCR, HMG-CoA reductase. **H**, DHCR7, 7-dehydrocholesterol reductase. **I**, LDL-R, low-density lipoprotein receptor. n=8 mice per group. **J** through **L**, Plasma, liver and fecal radioactivities at 48 hours after injection of <sup>3</sup>H-cholesterol-labelled macrophages. Data are expressed as percentage of total injected <sup>3</sup>H-radioactivity. Samples for chow and 4 wks HFD groups were derived from different sets of mice. n=4-6 mice per group. \* P<0.05 and \*\* P<0.01 versus Apoe<sup>-/-</sup> mice by 2-way ANOVA and Bonferroni *post hoc* tests. Data are mean  $\pm$  SEM, n=7-10 per group.



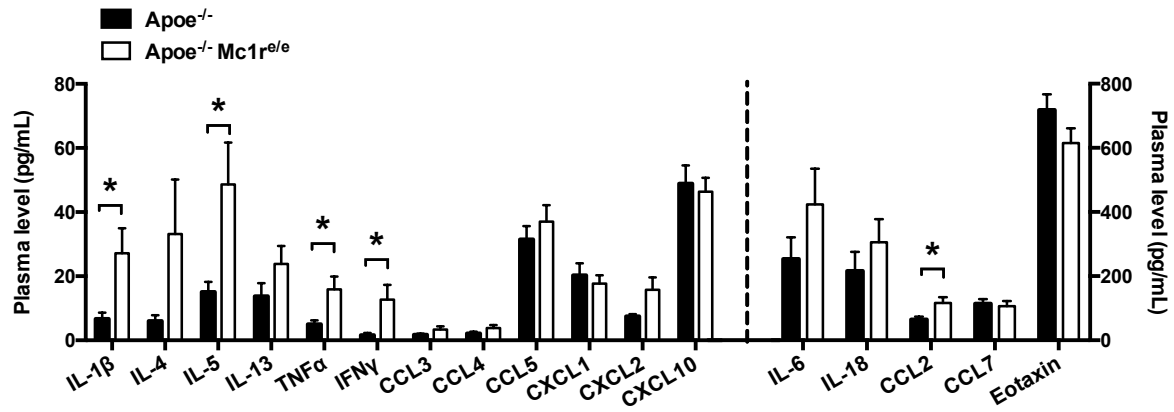
**Supplemental Figure V. Composition of the fecal bile acid pool in  $Apoe^{-/-}$  and  $Apoe^{-/-} Mc1r^{e/e}$  mice.** Feces were collected over 48 h and quantified for total bile acids (A), the primary bile acids; cholic acid, chenodeoxycholic acid and murocholic acid (B), and the secondary bile acids; ursodeoxycholic acid, deoxycholic acid and lithocholic acid (C). The results are presented as  $\mu\text{g}$  per g of dried feces. D, The ratio of cholic acid to chenodeoxycholic acid was calculated as a measure of bile acid hydrophilicity. E, Relative distribution of individual bile acids in the fecal samples of  $Apoe^{-/-}$  and  $Apoe^{-/-} Mc1r^{e/e}$  mice. CA, cholic acid; CDCA, chenodeoxycholic acid; MCA, muricholic acid; UDCA, ursodeoxycholic acid; DCA, deoxycholic acid; LCA, lithocholic acid. \*  $P < 0.05$  and \*\*  $P < 0.01$  versus  $Apoe^{-/-}$  mice by 2-way ANOVA and Bonferroni *post hoc* tests. Data are mean  $\pm$  SEM,  $n = 4-6$  mice per group.



**Supplemental Figure VI. Hepatic expression of genes that govern bile acid synthesis, transport and conjugation.** Quantitative RT-PCR analysis of genes involved in the bile acid synthesis (**A**), transport (**B**) and conjugation (**C**) in the liver of  $Apoe^{-/-}$  and  $Apoe^{-/-}$   $Mc1r^{e/e}$  mice fed a normal chow diet or high-fat diet (HFD) for 4 weeks. CYP7A1, cholesterol 7  $\alpha$ -hydroxylase; CYP7B1, 25-hydroxycholesterol 7- $\alpha$ -hydroxylase; CYP8B1, sterol 12- $\alpha$ -hydroxylase; CYP27A1, sterol 27-hydroxylase; FXR, farnesoid X receptor; LRH-1, liver receptor homologue 1; BSEP, bile-salt export pump; NTCP, Na<sup>+</sup>-taurocholate cotransporting polypeptide; HNF4 $\alpha$ , hepatocyte nuclear factor 4  $\alpha$ ; BAAT, bile acid-CoA:amino acid N-acyltransferase; BAL, bile acid-CoA ligase. \*  $P < 0.05$  and \*\*  $P < 0.01$  versus  $Apoe^{-/-}$  mice by 2-way ANOVA and Bonferroni *post hoc* tests. Data are mean  $\pm$  SEM,  $n = 8$  mice per group.

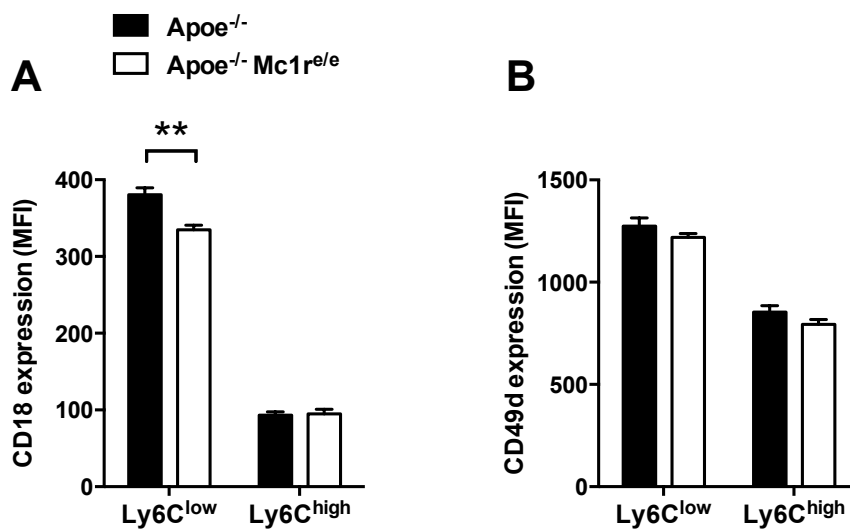


**Supplemental Figure VII. Melanocortin 1 receptor deficiency accelerates monocytois in HFD-fed Apoe<sup>-/-</sup> mice.** **A**, Quantification of total leukocytes (CD45<sup>+</sup>), lymphocytes (CD45<sup>+</sup>, CD11b<sup>-</sup>), neutrophils (CD45<sup>+</sup>, CD11b<sup>+</sup>, Ly6G), Ly6C<sup>high</sup> monocytes (CD45<sup>+</sup>, CD11b<sup>+</sup>, CD115<sup>+</sup>, Ly6C<sup>high</sup>) and Ly6C<sup>low</sup> monocytes (CD45<sup>+</sup>, CD11b<sup>+</sup>, CD115<sup>+</sup>, Ly6C<sup>low</sup>) in the bone marrow of Apoe<sup>-/-</sup> mice and Apoe<sup>-/-</sup> Mc1r<sup>e/e</sup> mice fed a normal chow diet or high-fat diet (HFD) for 4 weeks. Samples for chow and 4 wks HFD groups were derived from different sets of mice. **B**, Quantitative RT-PCR analysis of ATP-binding cassette transporter A1 (ABCA1), G1 (ABCG1) and scavenger receptor class B member 1 (SR-BI) expression in the bone marrow of Apoe<sup>-/-</sup> mice and Apoe<sup>-/-</sup> Mc1r<sup>e/e</sup> mice. \* P<0.05 versus Apoe<sup>-/-</sup> mice by 2-way ANOVA and Bonferroni *post hoc* tests. Data are mean  $\pm$  SEM, n=7-10 per group.

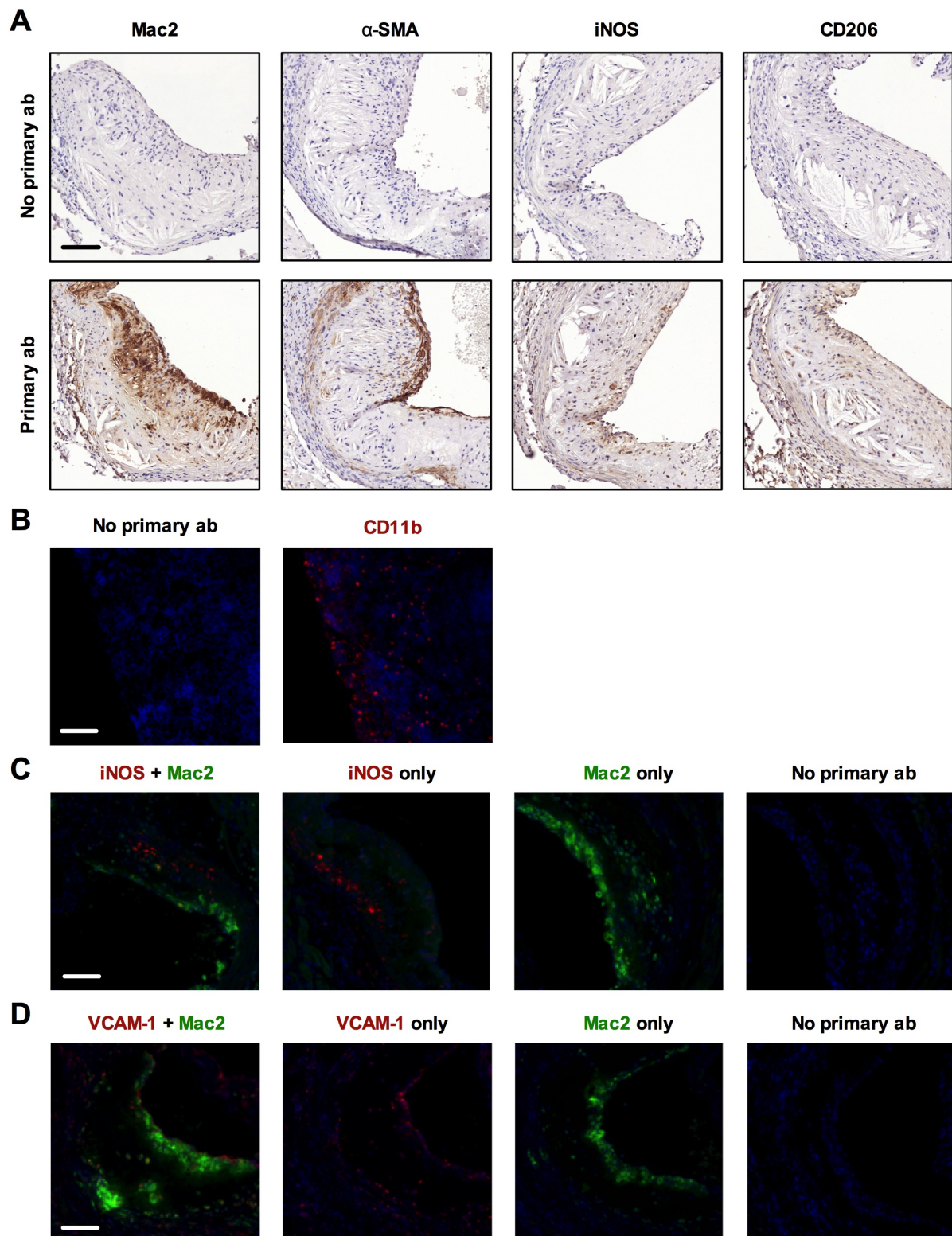


**Supplemental Figure VIII. Melanocortin 1 receptor deficiency elevates pro-inflammatory cytokine levels in chow-fed Apoe<sup>-/-</sup> mice.** Plasma cytokine levels in chow-fed Apoe<sup>-/-</sup> and Apoe<sup>-/-</sup> Mc1r<sup>e/e</sup> mice. \* P<0.05 versus Apoe<sup>-/-</sup> mice by Student's t test. Data are mean ± SEM, n=12-13 per group.





**Supplemental Figure IX. Integrin expression in blood monocytes from chow-fed Apoe<sup>-/-</sup> mice.** Mean fluorescence intensity (MFI) of CD18 (**A**) and CD49d (**B**) in the Ly6C<sup>low</sup> and Ly6C<sup>high</sup>-gated blood monocytes from chow-fed Apoe<sup>-/-</sup> and Apoe<sup>-/-</sup> Mc1r<sup>e/e</sup> mice. \*\* P<0.01 versus Apoe<sup>-/-</sup> mice by 2-way ANOVA and Bonferroni *post hoc* tests. Data are mean ± SEM, n=8 mice per group.



**Supplemental Figure X. Controls for immunohistochemistry and immunofluorescence.** Negative controls for Mac2,  $\alpha$ -SMA, iNOS and CD206 (**A**) and CD11b (**B**) were obtained by staining consecutive sections with the appropriate secondary antibodies only. **C-D**, Appropriate single stain and no primary antibody controls are presented for iNOS+Mac2 and VCAM-1+Mac2 immunofluorescence staining. Scale bar, 100  $\mu$ m in each panel.