

## Supplementary Online Content

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**eAppendix.** Additional Methods

**eFigure.** Schematic diagram of the customized sequencing and data analysis pipeline

**eTable 1.** Broad copy number aberrations (Broad Regions) regions identified by GISTIC analysis with a q-value significance score  $< 0.2$ .

**eTable 2.** Specific amplified (Amp\_Genes) regions identified by GISTIC analysis with a q-value significance score  $< 0.2$ .

**eTable 3.** List of specific amplified genes identified by GISTIC analysis.

**eTable 4.** Specific deleted gene (Del\_Genes) regions identified by GISTIC analysis with a q-value significance score  $< 0.2$ .

This supplementary material has been provided by the authors to give readers additional information about their work.

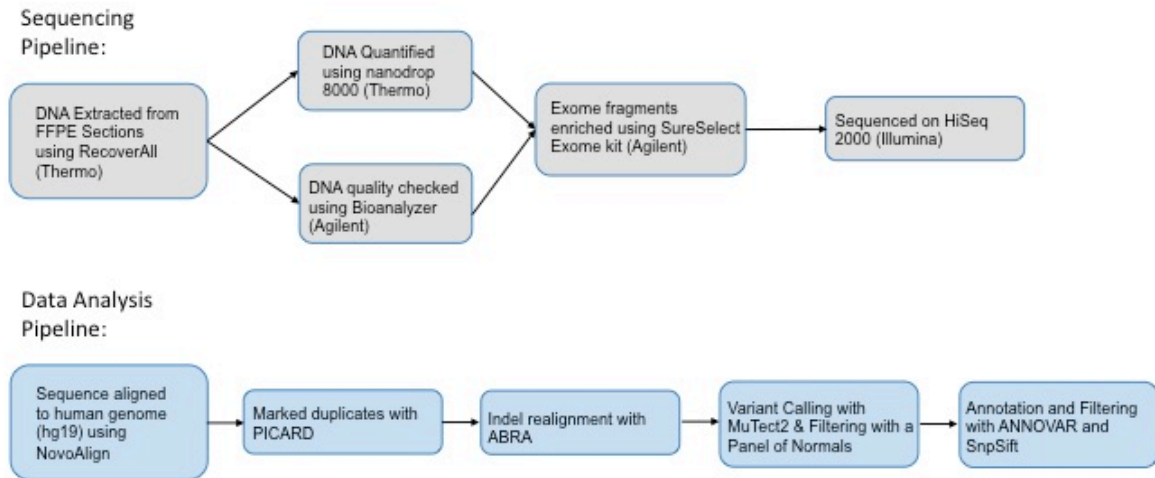
## **eAppendix.** Additional Methods

Genomic DNA from FFPE sections was purified by RecoverAll Total Nucleic Acid Isolation Kit (Thermo, Rockland, DE) following the manufacturer's instructions. The quantity and quality of DNA was evaluated by a nanodrop 8000 spectrophotometer (Thermo, Rockland, DE) and a Bioanalyzer 2000 (Agilent, Palo Alto, CA), respectively. Whole exome sequencing was conducted in the Sequencing Core facility at the John P. Hussman Institute for Human Genomics in the University of Miami. Briefly, DNA samples were sheared using an E210 sonicator (Covaris, Woburn, MA) and the whole exome was captured using the SureSelect XT Human All Exon V5 kit (Agilent, Palo Alto, CA). To sequence the enriched 50Mb exomes, a 3-plex strategy per lane was conducted on the HiSeq 2000 sequencer (Illumina, San Diego, CA) using 125-bp paired-end reads, which yielded an average ~100× coverage depth at targeted regions.

Sequences were aligned to the human genome hg19 using NovoAlign (<http://www.novocraft.com/products/novoalign/>). Quality control and file manipulation was performed with FastQC, PICARD and SAMtools.<sup>1</sup> Variant calling was performed using MuTect2.<sup>2</sup> Variants predicted to be germline by the algorithms in MuTect2 or present in a panel of normal samples (n = 117, samples from blood to represent germline) were removed. To minimize artifact introduced by the specimen archiving process,<sup>3</sup> we filtered out all variants present in less than 20% of sequencing reads. Additionally, variants were excluded if they were outside of coding or splicing regions, had fewer than 3 alternate reads, were present in greater than 0.5% of the population, or were predicted to be non-damaging using ANNOVAR.<sup>4</sup>

1. Li H, Handsaker B, Wysoker A, et al. The Sequence Alignment/Map format and SAMtools. *Bioinformatics* 2009; 25(16):2078-9.
2. McKenna A, Hanna M, Banks E, et al. The Genome Analysis Toolkit: a MapReduce framework for analyzing next-generation DNA sequencing data. *Genome Res* 2010; 20(9):1297-303.
3. Wong SQ, Li J, Tan AYC et al. Sequence artefacts in a prospective series of formalin-fixed tumours tested for mutations in hotspot regions by massive parallel sequencing. *BMC Medical Genomics* 2014; 7:23.
4. Wang K, Li M, Hakonarson H. ANNOVAR: functional annotation of genetic variants from high-throughput sequencing data. *Nucleic Acids Res* 2010; 38(16):e164.

**eFigure 1.** Schematic diagram of the customized sequencing and data analysis pipeline.



**eTable 1.** Broad copy number aberrations (Broad Regions) regions identified by GISTIC analysis with a q-value significance score < 0.2. Chromosome 6p gain was noted in all samples.

Arm	# Genes	Amp frequency	Amp frequency score	Amp z-score	Amp q-value	Arm	# Genes	Del frequency	Del frequency score	Del z-score	Del q-value
1q	1955	0.4	0.4	1.97	0.155	16p	872	0.6	0.6	2.47	0.0502
6p	1173	1	1	5.1	7.43E-06	16q	702	0.6	0.6	2.36	0.0573
6q	839	0.6	0.6	2.44	0.0532	19q	1709	0.4	0.4	1.78	0.182
8q	859	0.6	0.6	2.46	0.0532	21p	13	1	1	4.13	0.000264
						21q	509	0.6	0.6	2.25	0.0672

**eTable 2.** Specific amplified (Amp\_Genes) regions identified by GISTIC analysis with a q-value significance score < 0.2. Chromosome 6p gain was noted in all samples.

cytoband	1p36.13	4p15.33	5q13.2	6p22.1	6p22.1	8q12.1	9p13.1	11q13.1	12p13.31	15q26.3	16p11.2	17q21.2
q value	4.43E-05	0.12939	0.003965	0.00032494	0.00032494	0.03588	0.14622	4.43E-05	4.43E-05	0.02189	0.00053898	0.10191
residual q value	4.43E-05	0.12939	0.003965	0.00032494	1	0.03588	0.14622	4.43E-05	4.43E-05	0.02189	0.00053898	0.10191
wide peak boundaries	chr1:168534-17100800	chr4:14884836-15076917	chr5:68856260-70096675	chr6:27765466-27870329	chr6:1-171115067	chr8:59404889-59555610	chr9:39010498-40513014	chr11:65164168-65273902	chr12:9446959-9605858	chr15:102291831-102355571	chr16:29321829-29641162	chr17:39174123-39353070

**eTable 3.** List of specific amplified genes identified by GISTIC analysis in eTable 2.

cytoband	1p36.13	4p15.33	5q13.2	6p22.1	6p22.1	8q12.1	9p13.1	11q13.1	12p13.31	15q26.3	16p11.2	17q21.2
	MST1P2	CPEB2	SMN1	HIST1H1B	hsa-mir-1913	CYP7A1	CNTNAP3	hsa-mir-612	DDX12P	OR4F6	SULT1A3	KRTAP1-3
	MST1P9	LOC441009	SMN2	HIST1H2AI	hsa-mir-1202	SDCBP	FAM74A1	FRMD8	LOC642846		SLX1B	KRTAP1-1
	NBPF1		SERF1A	HIST1H2AK	hsa-mir-1273c	NSMAF	ZNF658B	NEAT1			SLC7A5P1	KRTAP4-6
	CROCCP2		SMA4	HIST1H2AJ	hsa-mir-3145		FAM75A2	MALAT1			LOC388242	KRTAP2-1
	ESPNP		SMA5	HIST1H2AL	hsa-mir-548a-2		FAM75A1	MIR612			SNX29P2	KRTAP4-12
	MIR3675		GUSBP3	HIST1H2AM	hsa-mir-588		LOC653501				LOC440354	KRTAP1-5
			GTF2H2B	HIST1H2BL	hsa-mir-3144						SULT1A4	KRTAP4-4
			GTF2H2C	HIST1H2BN	hsa-mir-548b						SLX1A	KRTAP4-1
			SERF1B	HIST1H2BM	hsa-mir-587						BOLA2	KRTAP4-5
			GTF2H2D	HIST1H2BO	hsa-mir-2113						LOC606724	KRTAP4-3
			GUSBP9	HIST1H3I	hsa-mir-4282						BOLA2B	KRTAP4-2

			LOC100170939	HIST1H3J	hsa-mir-30a						SLX1A-SULT1A3	KRTAP2-4
			LOC100272216	HIST1H3H	hsa-mir-30c-2						SLX1B-SULT1A4	KRTAP4-11
				HIST1H4K	hsa-mir-548u							KRTAP4-8
				HIST1H4J	hsa-mir-133b							KRTAP2-2
				HIST1H4L	hsa-mir-586							KRTAP9-1
					hsa-mir-1275							LOC730755
					hsa-mir-219-1							KRTAP4-9
					hsa-mir-1236							KRTAP4-7
					hsa-mir-877							
					hsa-mir-3143							
					hsa-mir-548a-1							
					ABCF1							
					ACAT2							
					CRISP1							
					AGER							
					AIF1							
					AIM1							
					AMD1							
					ARG1							
					ATP6V1G2							
					BAI3							
					BAK1							
					BCKDHB							
					CFB							
					PRDM1							
					BMP5							
					BMP6							
					DST							
					BPHL							
					BTN1A1							
					BYSL							
					C2							
					C4A							
					C4B							
					DDR1							
					RUNX2							
					CCNC							
					CCND3							
					CDC5L							
					CDKN1A							
					CDSN							
					CGA							
					CLIC1							
					CLPS							
					CCR6							
					CNR1							
					COL9A1							

					COL10A1									
					COL11A2									
					COL12A1									
					COL19A1									
					COX7A2									
					ATF6B									
					MAPK14									
					CSNK2B									
					CTGF									
					CYP21A2									
					CYP21A1P									
					DAXX									
					DNAH8									
					DOM3Z									
					DSP									
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					EEF1A1									
					SERPINB1									
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					FABP7									
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					GSTA2									
					GSTA3									
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					HIST1H1D									
					HIST1H1E									
					HIST1H1B									
					HIST1H1T									
					HIST1H2AE									
					HIST1H2AD									
					HIST1H2BD									
					HIST1H2BB									
					HIST1H1A									
					HCRTR2									
					HDAC2									
					HFE									
					HIVEP1									
					HIVEP2									
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					HLA-B									
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					HLA-DPA1									
					HLA-DPB1									
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					HLA-DQA1									
					HLA-DQA2									
					HLA-DQB1									
					HLA-DQB2									
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					HLA-DRB1									
					HLA-DRB5									
					HLA-DRB6									
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					HLA-G									
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					HLA-J									
					HLA-L									
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					HSF2									

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				HSPA1B											
				HSPA1L											
				HSP90AB1											
				HTR1B											
				HTR1E											
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				IFNGR1											
				IGF2R											
				IL17A											
				IMPG1											
				IRF4											
				ITPR3											
				JARID2											
				KIFC1											
				KIF25											
				KPNA5											
				LAMA2											
				LAMA4											
				LPA											
				LTA											
				LTB											
				MARCKS											
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				MAN1A1											
				MAS1											
				MCM3											
				MDF1											
				ME1											
				MEA1											
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				MAP3K5											
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				MICB											
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				NEDD9											
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				NFKBIE											
				NFKBIL1											
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				NMBR											
				NQO2											



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					NT5E								
					OPRM1								
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					PCMT1								
					PDCD2								
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					PEX6								
					PEX7								
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					PGK2								
					PGM3								
					PHF1								
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					SERPINB9								
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					PRIM2								
					PKIB								
					MAPK13								
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					PRPH2								
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				SIM1								
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				SLC22A1								
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				SLC22A2								
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				SNRPC								
				SOD2								
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				SSR1								
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				MAP3K7								
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				TAPBP								
				TBCC								
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				TCF21								
				TCP1								
				TCP10								
				TCP11								
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				TNXB								
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				TPMT								

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						TUBB2A								
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						RDBP								
						LST1								
						PLA2G7								
						TFEB								
						STL								
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						HIST1H2AI								
						HIST1H2AK								
						HIST1H2AJ								
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						HIST1H2AC								
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				WISP3									
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				IER3									
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				VNN2									
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				HIST1H2AG									
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					GCM2								
					WDR46								
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					TAAR2								
					TAAR3								
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					PPT2								
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					TRIM10								
					FLOT1								
					RCAN2								

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					BTN2A2							
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					UBD							
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					SCGN							
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					C6orf10							
					FARS2							
					FUT9							
					CNPY3							
					TRAF3IP2							
					HBS1L							
					SLC17A3							
					RPP40							
					FRS3							
					PDE10A							
					SLC22A7							
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					PNRC1							
					ASCC3							
					RAB32							
					TRIM31							
					KATNA1							
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					BTN3A2							
					BTN3A1							
					BTN2A1							
					CAPN11							
					BVES							
					NUDT3							
					SEC63							
					NRM							
					STK38							
					SCAF8							



					SLC17A5									
					OR12D2									
					OR11A1									
					RGS17									
					OR2W1									
					OR2J2									
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					PDE7B									
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					GNMT									
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					SESN1									
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					DLL1									
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					TMEM14A									
					MRPL18									
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					C6orf15									
					MYLIP									
					ABT1									
					DSE									
					PRICKLE4									
					PACSIN1									
					ZNRD1									
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					CYB5R4									
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					C6orf203									
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					CYP39A1									
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					RWDD1									
					AIG1									
					NOL7									
					SNX9									



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					UBE2J1								
					DCDC2								
					ETV7								
					TMEM14C								
					VTA1								
					LGSN								
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					BRP44L								
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					CLIC5								
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					TREM1								
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					HCG4								
					GFOD1								
					HMGCLL1								
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					BTN2A3P								
					AHI1								
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					AKIRIN2-AS1								
					DDX43								
					FAM46A								
					LRRC16A								
					TBC1D22B								
					TMEM30A								
					EXOC2								

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					ECHDC1									
					APOM									
					BTNL2									
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				ULBP1							
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**eTable 4.** Specific deleted gene (Del\_Genes) regions identified by GISTIC analysis with a q-value significance score < 0.2. Chromosome 6p gain was noted in all samples.

cytoband	3q11.1	12q14.2	16q24.2	21q11.2	Xp22.33
q value	0.0029269	0.089361	0.19893	0.1887	0.0048605
residual q value	0.0029269	0.089361	0.19893	0.1887	0.0048605
wide peak boundaries	chr3:89521354-93709932	chr12:63544058-64181655	chr16:88104957-88524995	chr21:11095865-14991078	chrX:1652540-2533608
genes in wide peak	PROS1	DPY19L2	ZNF469	hsa-mir-3156-3	ASMT
cytoband				ANKRD30BP2	AKAP17A
q value				MIR3156-3	ZBED1
residual q value					DHRX
wide peak boundaries					LINC00102