# **Supplemental Information**

Inwardly Rectifying Potassium (Kir) Channels Represent a Critical Ion Conductance Pathway in the Nervous Systems of Insects

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### **Supplemental Methods**

#### Adult emergence

Fifteen adult female and 5 adult male *Drosophila* flies were released into a *Drosophila* vial containing standard medium and were maintained at 25°C, 12 hour-12 hour photoperiod and 55% relative humidity. Flies were removed from the vial 72 hours after being released, which was ample time to ensure oviposition occurred. Adult emergence was determined by counting the number of adult flies that emerged daily for 28 days after the removal of the parental flies. After the daily count, flies were removed to prevent recounting of flies. Total emergence was performed on five replicates for the control (OR), GFP knockdown (knockdown control), and *irk*2 knockdown flies. Percent emergence of OR fly strain, which was standardized to unity. Data were plotted using GraphPad Prism (La Jolla, CA) software. Statistical analysis was performed using an unpaired *t*-test of the RNAi strains compared to OR.

## Neurophysiological recordings of Drosophila CNS

The methods for determining the influence of diazoxide and glybenclamide were the same for the other electrophysiological recordings of the CNS, which is described in section 2.4.

## **Supplemental Figures**



Figure S1. **Percent adult emergence of** *irk***2 RNAi-knockdown flies.** Bars represent average (n=5) percent emergence of control (GFP) and *irk***2** RNAi-mediated knockdown when compared to OR flies. Error bars represent SEM. Asterisks represent statistical significance at P<0.05.



Figure S2. Neurophysiological recordings from the CNS of third instar larvae of *D. melanogasgter* after exposure to  $K_{ATP}$  modulators. Representative nerve discharge traces of the  $K_{ATP}$  opener diazoxide (A) and the  $K_{ATP}$  inhibitor glibenclamide (B). Initial firing frequencies in spikes/second (Hz) for each experiment are given to the left of each trace.