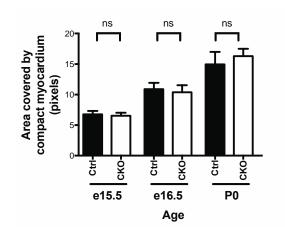


**Supplementary Figure 1.** *Ino80* deletion and expression. (a) Western blot analysis of lysates from primary mouse embryonic fibroblasts (MEFs) prepared from *Ino80 fl/fl* (n=3) and control embryos (n=2) infected with adenovirus (Cre -) or adenovirus expressing Cre recombinase (Cre +). Top panel is blot probed with anti-Ino80 antibody. Bottom panel is blot probed within anti-tubulin antibody. (b) qPCR for *Ino80* and *Vegfr2* and in endothelial and non-endothelial cells isolated from the heart (Sorted cells, n=3 hearts; whole hearts, n=3 hearts at e14.5). Error bars in graphs are standard deviation.

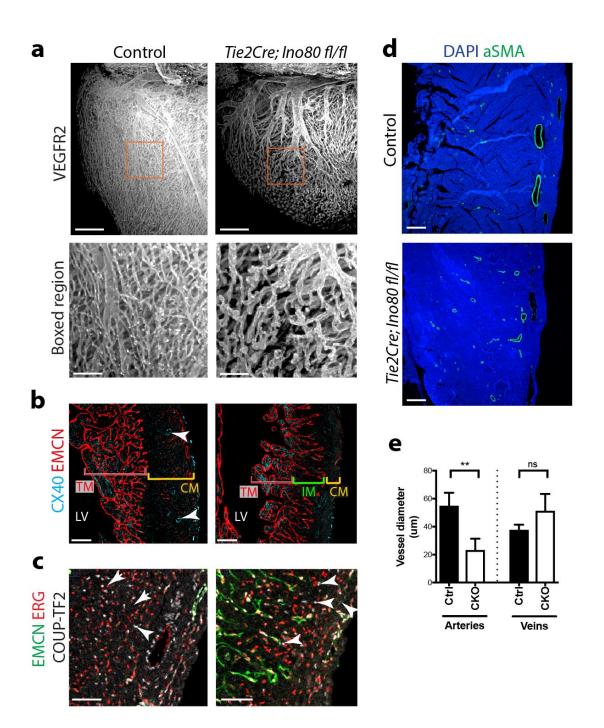
		Wild	type	Het	СКО
Age (embryos)		Tie2Cre- Ino80 fl/+ (25%)	Tie2Cre- Ino80 fl/fl (25%)	Tie2Cre+ Ino80 fl/+ (25%)	Tie2Cre+ Ino80 fl/fl (25%)
	e15.5 (91)	23%	25%	25%	26%
-	e16.5 (32)	34%	22%	31%	13%
_	e17.5 (18)	27%	22%	28%	22%
-	P1 (28)	28%	29%	35%	7%

**Supplementary Figure 2. Embryonic lethality in** *Tie2Cre;Ino80 fl/fl* **embryos**. Red indicates when the percentage of recovered conditional knockout (CKO) embryos was below the expected rates indicated in parentheses below genotypes.



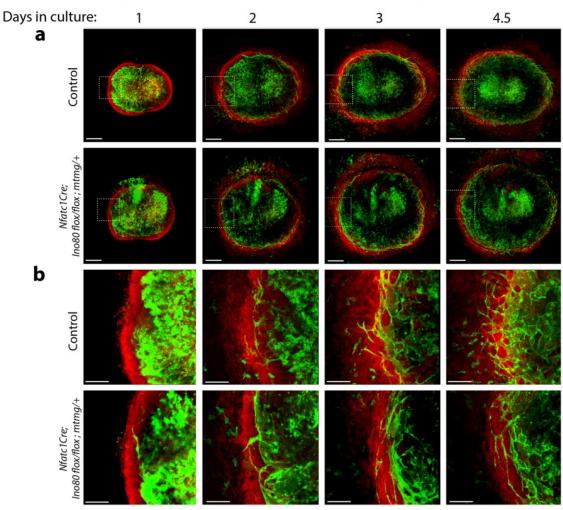
## Supplementary Figure 3. Myocardial deletion of *Ino80* does not alter

**compaction.** Deletion of *Ino80* in cardiomyocytes using the *Myh6Cre* deleter gene does not affect growth of the compact myocardium at indicated developmental stages. Error bars are standard deviation. (ns) Nonsignificant, evaluated by Student's *t*-test.



**Supplementary Figure 4. Postnatal coronary development is defective in** *Ino80* **mutant hearts.** (a) Whole mount confocal images of the coronary vasculature in P0. (control, n=3 hearts; mutant, n=3 hearts). Scale bars: 100 μm (low) and 25 μm (high magnification). (b) Tissue sections through P0 hearts revealed that CX40<sup>+</sup> arteries (arrowheads) are missing in mutants hearts and that the presence of the intermediate myocardium (IM) persists. (control, n=3 hearts; mutant, n=3 hearts). Scale bars: 100 μm. (c) Double labeling with pericyte (COUP-TF2) and endothelial (ERG) markers showed the presence of pericytes near coronary vessels (arrowheads) in control and mutant hearts. (control, n=3 hearts; mutant, n=3

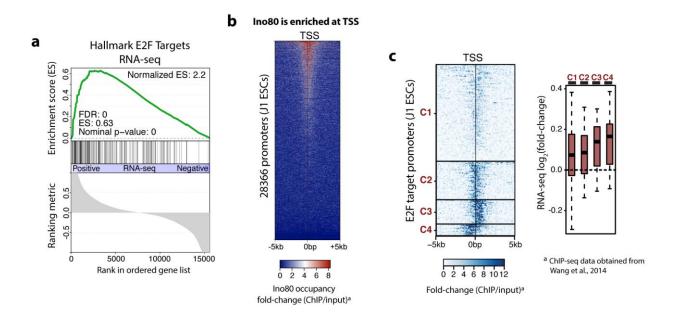
hearts). Scale bars: 100  $\mu$ m. (**d**, **e**) Immunofluorescence (**d**) and quantification (**e**) of arterial smooth muscle in P22 hearts indicate that surviving mutants have small arteries. Error bars in graphs are standard deviation. (control, n=4 hearts; mutant, n=5 hearts). (ns) Nonsignificant; (\*\*) P < 0.01, evaluated by Student's *t*-test. Scale bars: 50  $\mu$ m.



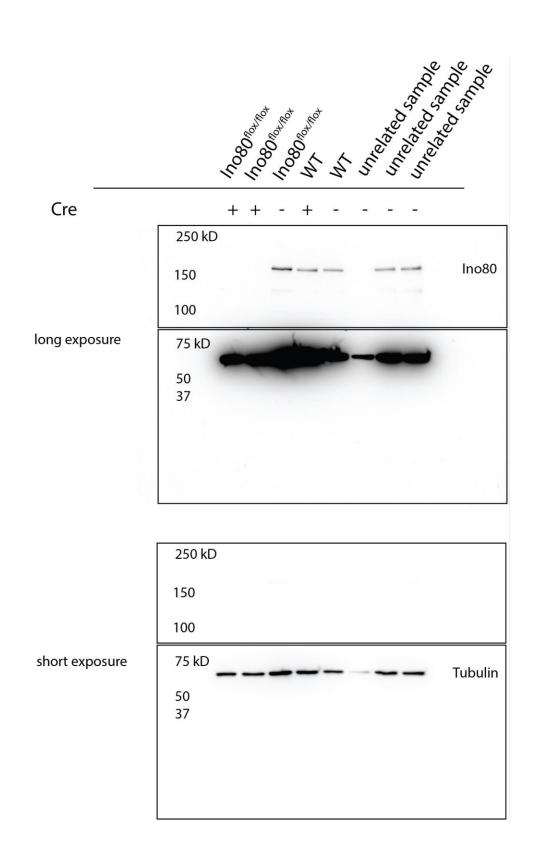
Nfatc1Cre; Rosa<sup>mTmG</sup>: endocardial cells and derived vessels, non-endocardial cells

## Supplementary Figure 5. Time lapse analysis of ventricle/endocardial cell

**cultures.** Two representative ventricle cultures (one control and one *Nfatc1Cre;Ino80 flox/flox* mutant) imaged after the indicated number of days in cultures. Endocardial cells and their derivatives are labeled in green through *Nfatc1Cre*-mediated recombination of the Rosa<sup>mTmG</sup> reporter allele. All other cells are labeled in red. Scale bars: 100  $\mu$ m (a) and 25  $\mu$ m (b).



**Supplementary Figure 6. Ino80 regulates E2F-target genes.** (a) RNA-seq enrichment of hallmark E2F-target genes in *Tie2Cre;Ino80 fl/fl* hearts. False discovery rate (FDR) is adjusted p-value of enrichment. (b) Ino80 ChIP occupancy values (fold-change, IP/input) across 28,336 promoters within a 10kb window surrounding the transcription start site (TSS) of J1 embryonic stem cells (ESCs). (c) Left panel, Ino80 occupancy across individual genes are indicated by heatmap with k-means clustering. Right panel, boxplot shows RNA-seq expression trends for each of the 4 E2F-target clusters (C1-C4) in *Tie2Cre;Ino80 fl/fl* hearts. Wilcoxon rank sum p-values are <0.01 for each cluster compared to the genome-wide RNA-seq log<sub>2</sub> fold change.



**Supplementary Figure 7**. Original immunoblot used for Supplementary Figure 1 indicated within the main manuscript.

**Supplementary Table 1. Summaries of antibodies used for Immunostaining** Anybody information and working condition for immunostaining are provided in the table

		Dilutions/ Immunostaining condition			
Antibody	Manufacturer/Cat#	Whole	Paraffin	Frozen	Cell/
		mount	section	section	Explant
ENDOMUCIN	NDOMUCIN Santa Cruz (sc-65495)		1:250		
VE-cadherin	E-cadherin BD Biosciences			1:500	
СТИТ	Developmental Studies Hybridoma Bank (CT3-c)	1:1000	1:1000	1:1000	
DACH1	DACH1 Proteintech (10914-1-AP)		1:1000	1:1000	
CX40	Alpha Diagnostic Intl. Inc., (CX40-A)	1:500	1:1000	1:1000	
hPROX1	hPROX1 R&D Systems (AF2727)		1:250		
COUP-TF2	Perseus Proteomics (PP-H7147-00)		1:1000		
Myomesin	Developmental Studies Hybridoma Bank, (mMaC myomesin B4)		1:1000	1:1000	
Myosin Heavy Chain	Alta Aesar I BI 56/1				
NKX2.5	NKX2.5 Santa Cruz (sc-8697)		1:250	1:250	1:250
ERG	ERG Abcam (ab92513)		1:1000	1:1000	1:1000
VEGFR2	VEGFR2 R&D Systems (AF644)			1:250	
DAPI	DAPI Sigma Aldrich		1:2000	1:2000	1:2000