

## Supplementary information

### **Structural insight into molecular mechanism of poly(ethylene terephthalate) degradation**

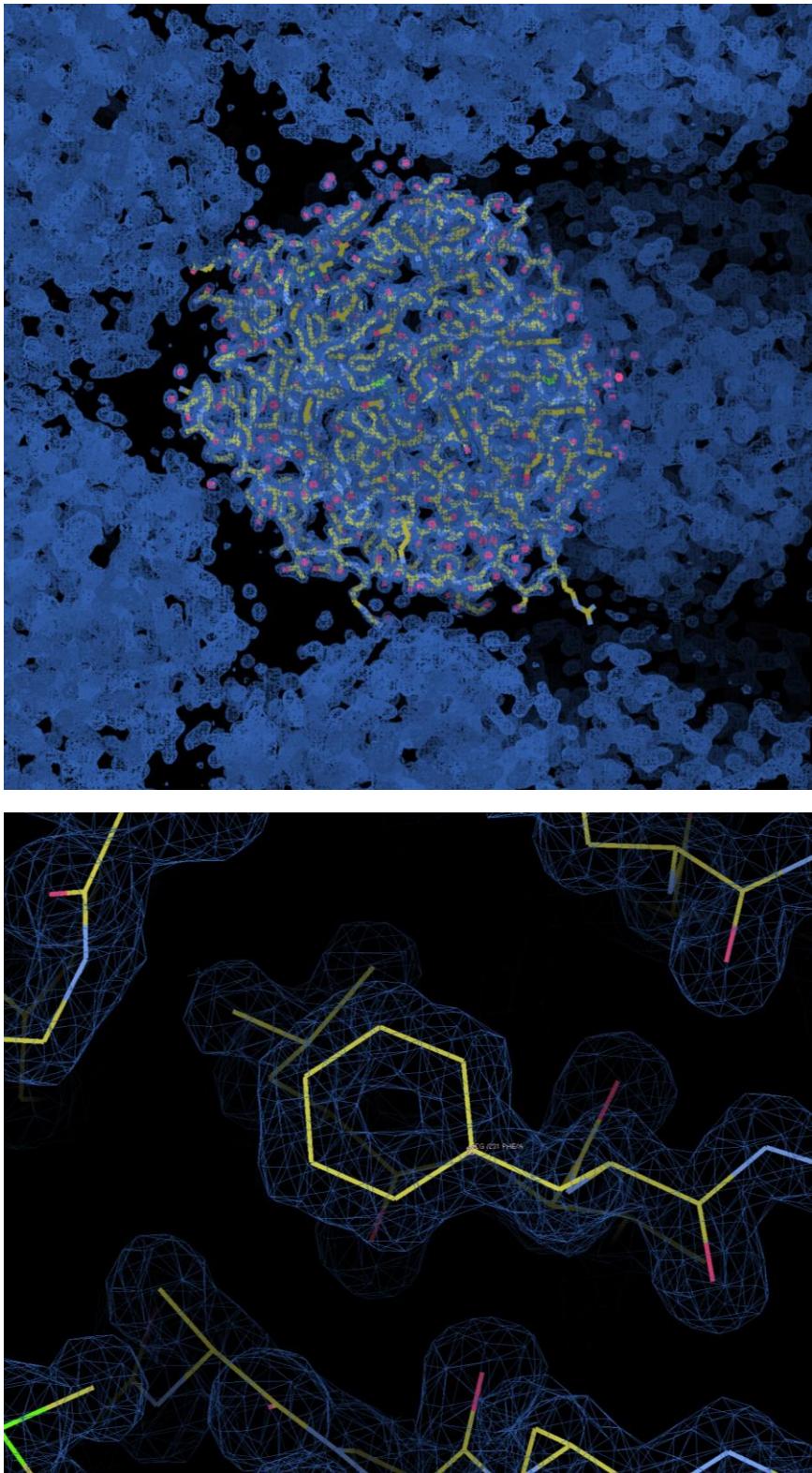
Seongjoon Joo, In Jin Cho, Hogyun Seo, Hyeoncheol Francis Son, Hye-Young Sagong, Tae Joo Shin, So Young Choi, Sang Yup Lee\* & Kyung-Jin Kim\*

\*Correspondence should be addressed to: Email: kkim@knu.ac.kr  
leesy@kaist.ac.kr

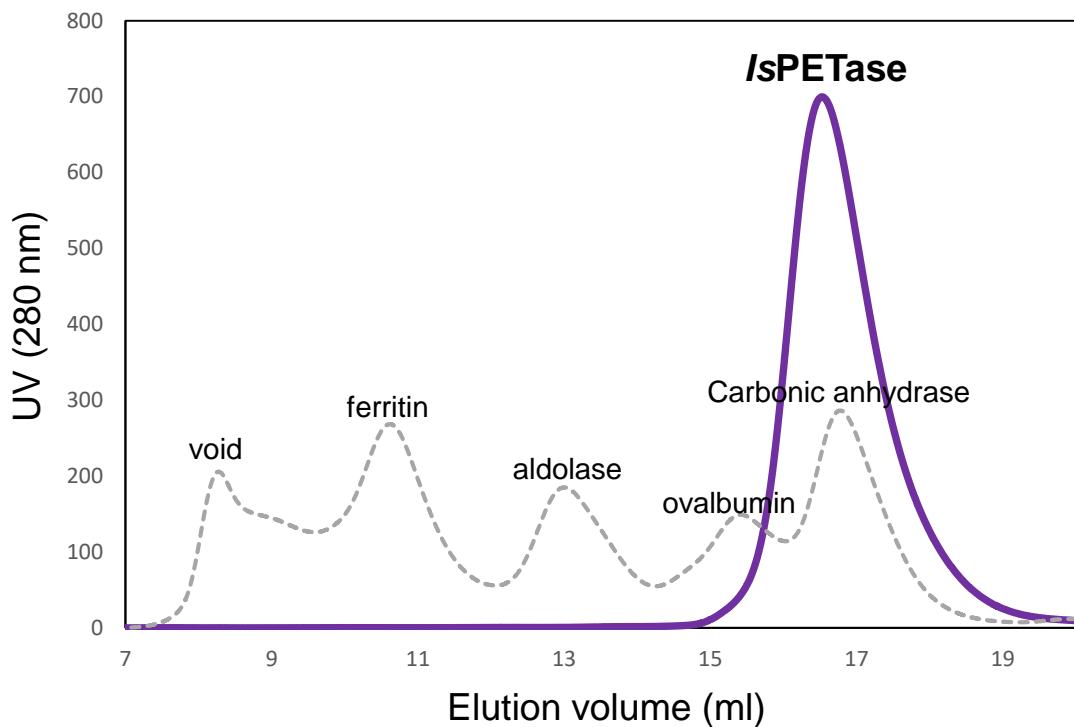
This PDF file includes:

Supplementary Figures 1 to 6.

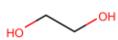
Supplementary Tables 1 and 2



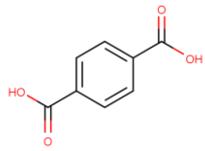
**Supplementary Figure 1.** The electron density map of *IsPETase*. The figures were generated by *WinCoot*. The  $2F_0 - F_c$  map contoured at  $1.5 \sigma$  is shown as blue mesh. The refined atomic model of *IsPETase* are shown as line with different color scheme by atoms; yellow-carbon, red-oxygen, blue-nitrogen, and green-sulfur. No crystallographic symmetry intimates the monomeric structure of *IsPETase* (above). The map represents the high resolution of  $1.5 \text{ \AA}$  of *IsPETase* (below).



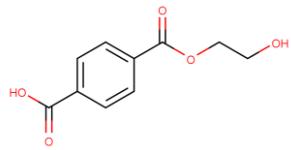
**Supplementary Figure 2.** Size-exclusion chromatographic analysis of *IsPETase*. A elution pick corresponding to a monomeric state of *IsPETase* in Size-exclusion chromatographic analysis. For precise analysis of the molecular weight, standard samples of ferritin (440kDa), aldolase (158kDa), ovalbumin (44kDa), and carbonic anhydrase (29kDa) are used for calibration and labelled.



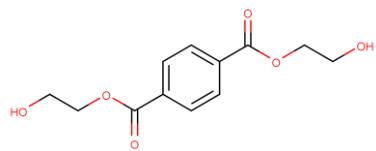
**EG**



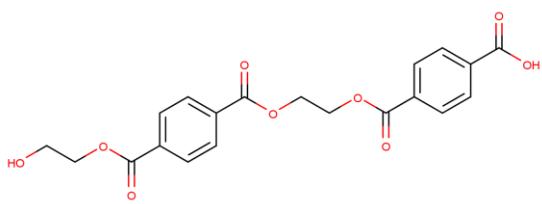
**TPA**



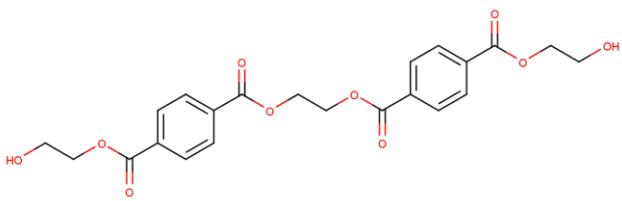
**MHET**



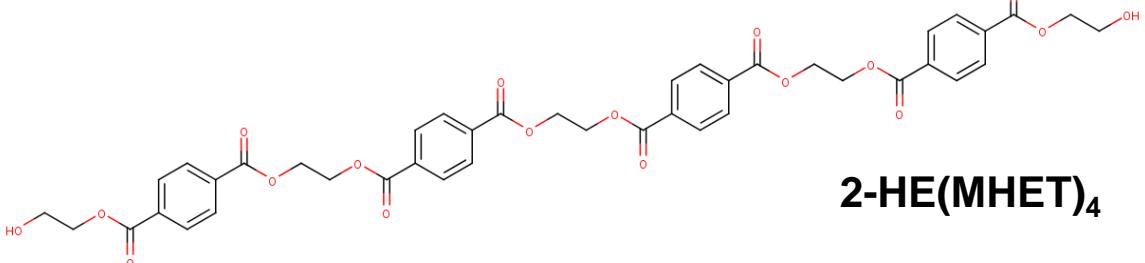
**BHET**



**(MHET)<sub>2</sub>**

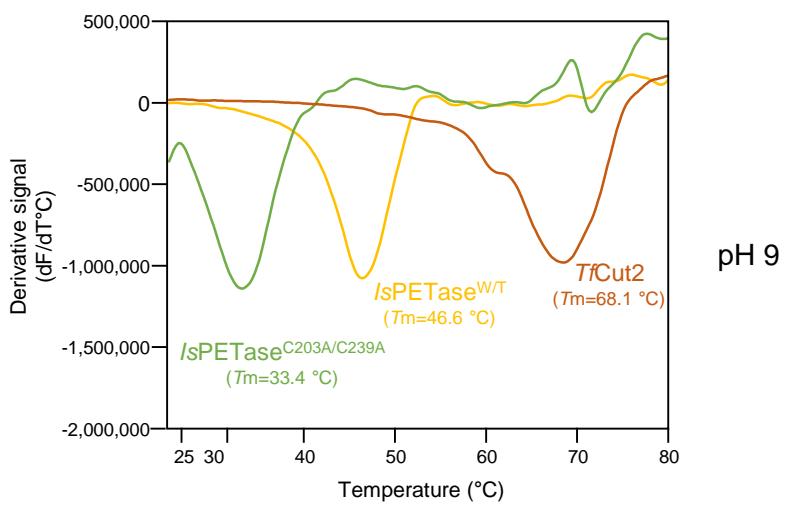
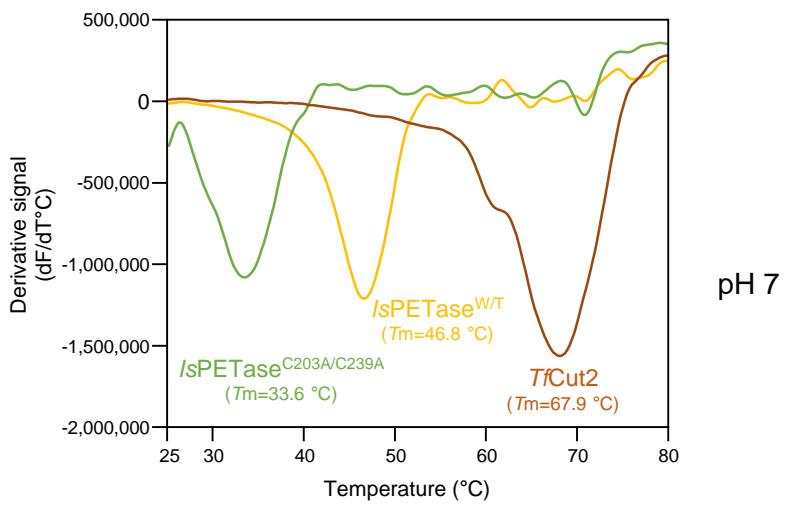


**2-HE(MHET)<sub>2</sub>**

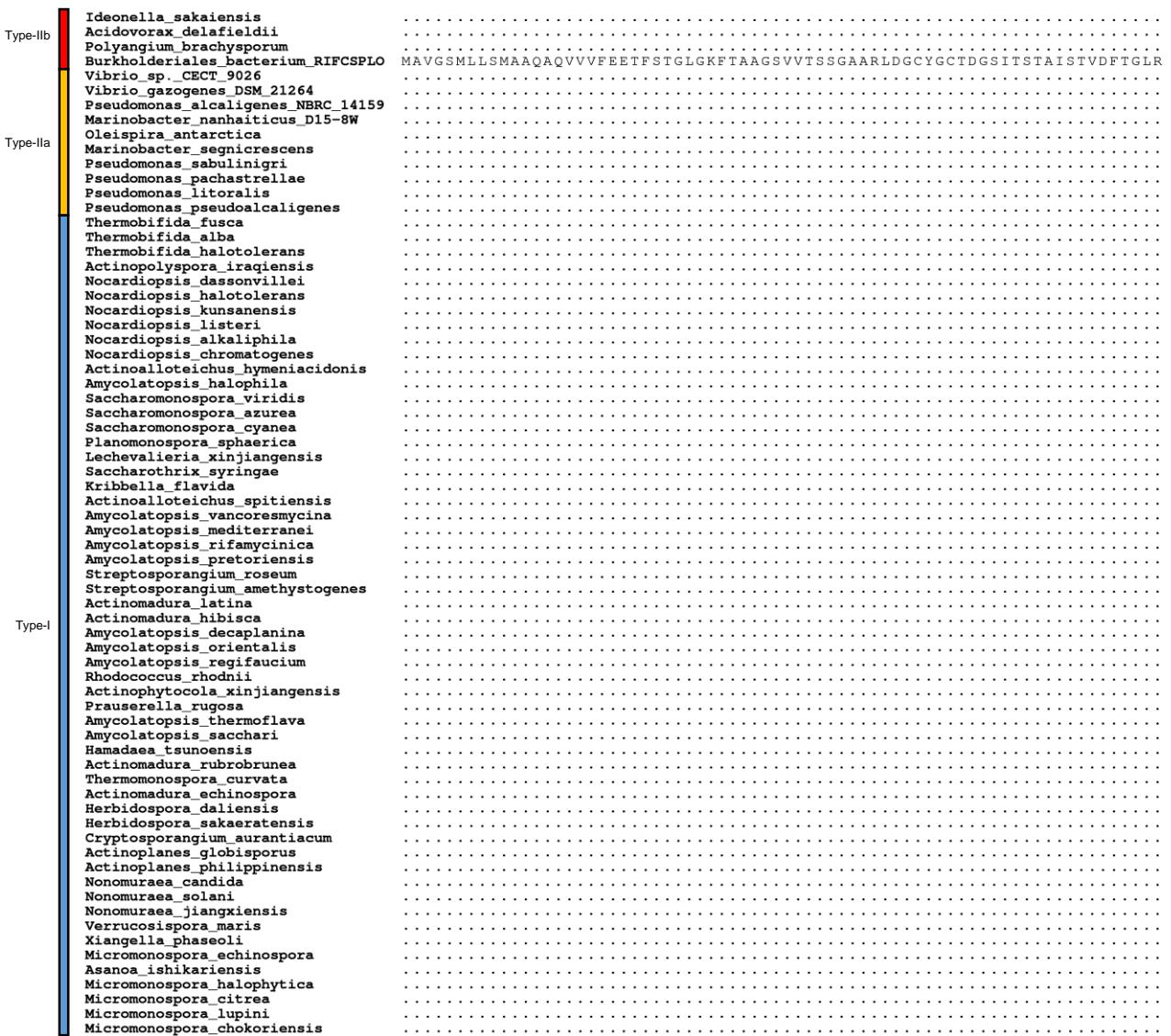


**2-HE(MHET)<sub>4</sub>**

**Supplementary Figure 3.** Chemical structures of the PET-related molecules used in this study.



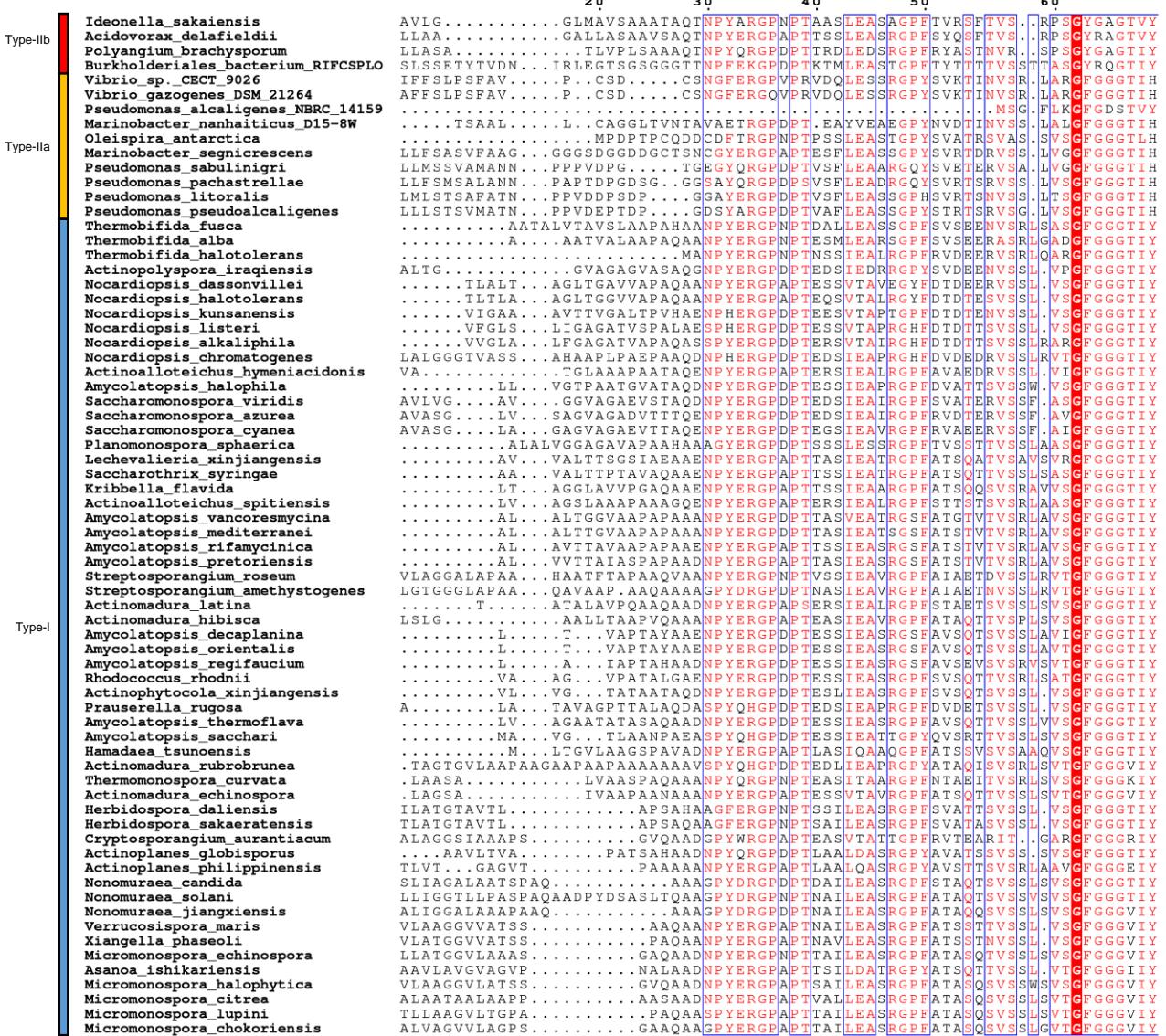
**Supplementary Figure 4.** Thermal stability measurements of  $IsPETase^{W/T}$ ,  $IsPETase^{C203A/C239A}$ , and  $TfCut2$ . The  $T_m$  values at pH 7 and 9 of  $IsPETase^{W/T}$ ,  $IsPETase^{C203A/C239A}$ , and  $TfCut2$  are displayed.



**Supplementary Figure 5.** Amino acid sequence alignment of PET-degrading enzymes. All enzymes involved in the tree generation in Fig. 6a were aligned. Key residues involved in the enzyme catalysis and the constitution of subsite I and subsite II are indicated by red-, blue-, and purple-colored triangles, respectively as same in Fig. 1a. At the left side, types of each enzymes are indicated.

Type-IIb	<i>Ideonella sakaiensis</i> <i>Acidovorax dealfieldii</i> <i>Polyangium brachysporum</i> <i>Burkholderiales bacterium RIFCSPLO</i> <i>Vibrio sp. CECT_9026</i> <i>Vibrio gazogenes DSM_21264</i> <i>Pseudomonas alcaligenes NBRC_14159</i> <i>Marinobacter nanhaiticus D15-8W</i> <i>Oleispira antarctica</i> <i>Marinobacter segniricrescens</i> <i>Pseudomonas sabulinigri</i> <i>Pseudomonas pachastrellaе</i> <i>Pseudomonas litoralis</i> <i>Pseudomonas pseudoalcaligenes</i> <i>Thermobifida fusca</i> <i>Thermobifida alba</i> <i>Thermobifida halotolerans</i> <i>Actinopolyspora iraqiensis</i> <i>Nocardiopsis dassonvillei</i> <i>Nocardiopsis halotolerans</i> <i>Nocardiopsis kunsanensis</i> <i>Nocardiopsis listeri</i> <i>Nocardiopsis alkaliphila</i> <i>Nocardiopsis chromatogenes</i> <i>Actinoalloteichus hymeniacidonis</i> <i>Amycolatopsis halophila</i> <i>Saccharomonospora viridis</i> <i>Saccharomonospora azurea</i> <i>Saccharomonospora cyanea</i> <i>Planomonospora sphaerica</i> <i>Lechevalieria xinjiangensis</i> <i>Saccharothrix syringae</i> <i>Kribbella flavigena</i> <i>Actinoalloteichus spitiensis</i> <i>Amycolatopsis vancoresmycinina</i> <i>Amycolatopsis mediterranei</i> <i>Amycolatopsis rifamycinica</i> <i>Amycolatopsis pretoriensis</i> <i>Streptosporangium roseum</i> <i>Streptosporangium amethystogenes</i> <i>Actinomadura latina</i> <i>Actinomadura hibisca</i> <i>Amycolatopsis decaplanina</i> <i>Amycolatopsis orientalis</i> <i>Amycolatopsis regifaucium</i> <i>Rhodococcus rhodnii</i> <i>Actinophytocula xinjiangensis</i> <i>Prauserella rugosa</i> <i>Amycolatopsis thermoflava</i> <i>Amycolatopsis sacchari</i> <i>Hamadaea tsundensis</i> <i>Actinomadura rubrobrunea</i> <i>Thermomonospora curvata</i> <i>Actinomadura echinospora</i> <i>Herbidospora daliensis</i> <i>Herbidospora sakaeratensis</i> <i>Cryptosporangium aurantiacum</i> <i>Actinoplanes globisporus</i> <i>Actinoplanes philippensis</i> <i>Nonomuraea candida</i> <i>Nonomuraea solani</i> <i>Nonomuraea jiangxiensis</i> <i>Verrucosipora maris</i> <i>Xiangella phaseoli</i> <i>Micromonospora echinospora</i> <i>Asanoa ishikariensis</i> <i>Micromonospora halophytica</i> <i>Micromonospora citrea</i> <i>Micromonospora lupini</i> <i>Micromonospora chokoriensis</i>	MNFPRA.....SR.....LMQA MHLPSRWDP.....FKEE.....TTMT.....HH.FSVR MP.....PDC.....VL.....RR.LAAA MSAENQSLR.....LR.FRINA M.....NVLTCKKL.....A.LGIV M.....NVLTCKKL.....A.LGIV M.....NVLQKLPL.....T.L MSDHYASNPRLR.....SV.VAAAS MI.....NNKQKSTLV.....TL.LASSA MPFNKKSVL.....AL.WGAGA MI.....NNKNLPPSLL.....SM.LAAGA MI.....NRTLPPSLL.....SM.LAAGA MAV.M.....TPRR.ERSSLLSRLAQVTAA MSV.T.....TPRR.EA.SLLSRAVAVAAA MRT.....RVKSGTA..AGRFRTRAL.....GVVTA.AL MRT.YPLSPPE.....TDGARSGFRS.RVRVFAARAGN MRK.YPLSPPEETP..DSHRARRGFRS.RVRSRAARVGVM MRT.....TSPLPBS.....OSHTDTLRSS.WTRFRASRTAA MTE.NP..PSPEG.....RTRPRTRARS.WMSGLAAKSAL MRTD.SPPSPIPAE.....RTRSGRSGRS.WVSAVAKVSAL MSI.ETIP.....HTGGARWA.RAACGTLAATA MASRA.....RVRSGAGRFT.RLAGLVLALT MLFA.TL MRI.....RRQAGTG..ARASMARAI.....GVMTT.AL MARTI.....GVLTA.AL MRI.....RSRSLTR..PHGRVSRTI.....GVLTA.AL M.NHRA.....RTLTKLAL MSLRSPLSR.R.....ALVFLA MSSRSPLSR.R.....ALVFLV MLGL.V.....TTVFA MRS.TLSRPA.....P..SRSLRRLVTS.A.....ALTLG MSA.LTSPPTLSGLG..EKISRRRPWRT.KAAGVVLAA MSA.LTSPPTSSGGSS..EKIPRLRGWRA.KAAGVVLAA MSA.LTSPPTSGPS..GKISRGGWGK.RAAGVVLAA MSV.LTSPPTSSGGSS..EKISRRRPWRA.KAAGVVLAA MLTLAL MHLIPR.....RAAKTLLATAL MHRITTPSGSP..PPARPGRRER..RGPRRARAVAA MCS.....TAN..GDGTMRSRSTTR..ILCGLATVA MRSTTR.....ILCGLATVA MRSTTR.....ILCGLATVA MRRGVRI.....ILTGLTIAAS MRKTLRKGVRI..ILGGVALAAT MTRLLAAL MRRSPR.....NLARLVTALA MRRSAK.....AWSSLVFAC MHTSTKA.GAARVLALAV MRSG.LST.....A..PR..RRALR..AVATTALAL MKRT.L.....KRALS.....LLPA..AA MNTN.L.....ARKLR.....TLPALL MSRLAT..TALAF MRPHFWRSLSRIAT..AALAF MLTIRKPARRGLIGK..LTLAL MTTK.....TRR..FLAA..AAA.. MTP.....VPTRARR..WLVV..ATAI MSREYDVQL..PTPRSGAR..TLTK..LALTV MQL.....STPHPG..LTK..LTLT MQL.....STPQRGAR..TLTK..LTLT MLTIRKPARRGLIGK..LTLAL MTP.....VPTRARR..WLVV..ATAI MPSPTTTRPRRSV..LVAR..LAMAA MPSPTTTRPRSLV..AVAG..LTTAV MKTAIK..L.....LATT MRSPTTTRPRRSV..TMR..LAMAA MSI.CPDPQGVDPVSSPTTTRPRRSV..AA.G..LALAA MPAR.....AVR..LALAA MSSPTTTRPRSV..AAR..LGLSA
Type-IIa		
Type-I		

**Supplementary Figure 5.** Amino acid sequence alignment of PET-degrading enzymes (continued).



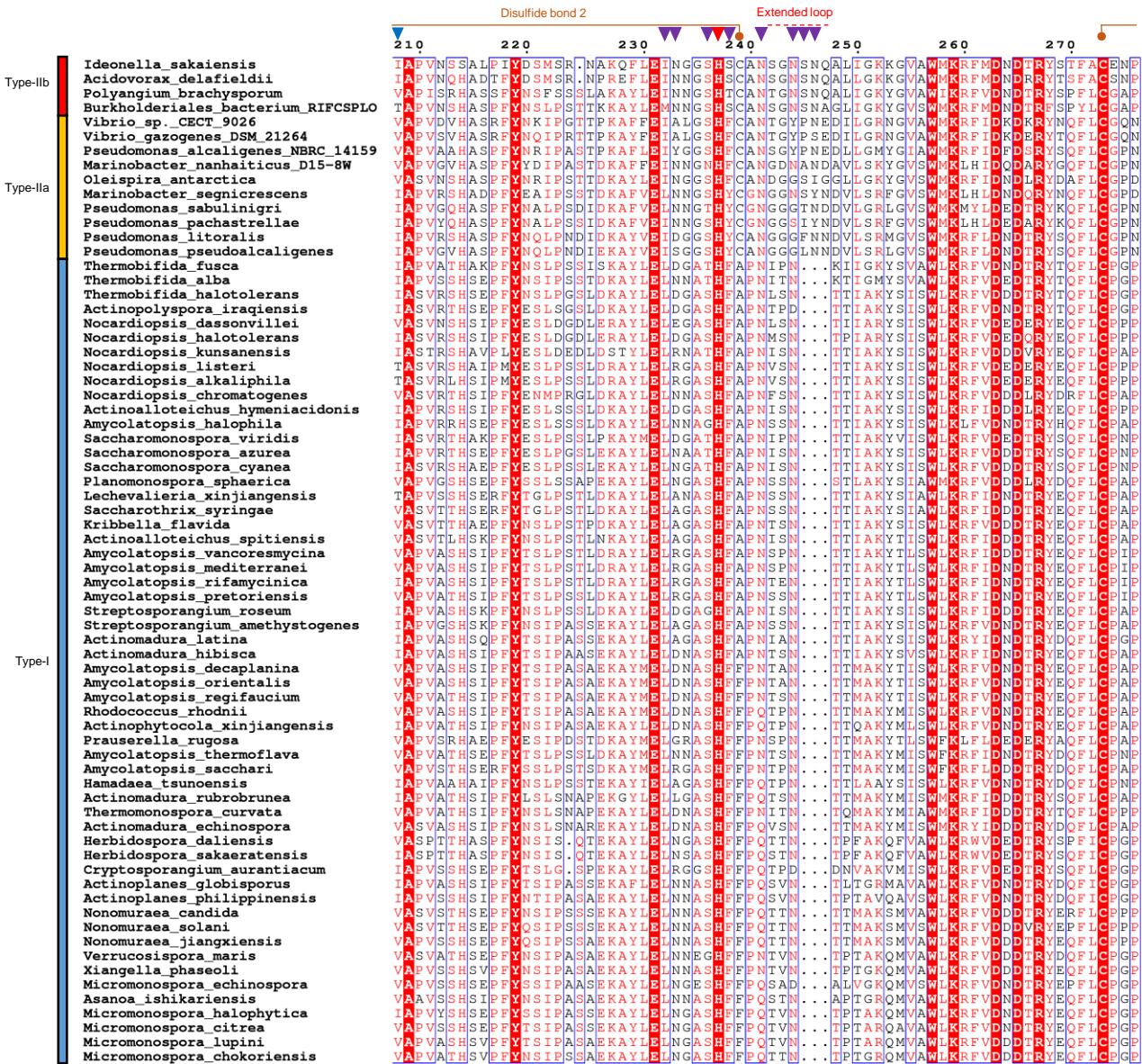
**Supplementary Figure 5.** Amino acid sequence alignment of PET-degrading enzymes (continued).



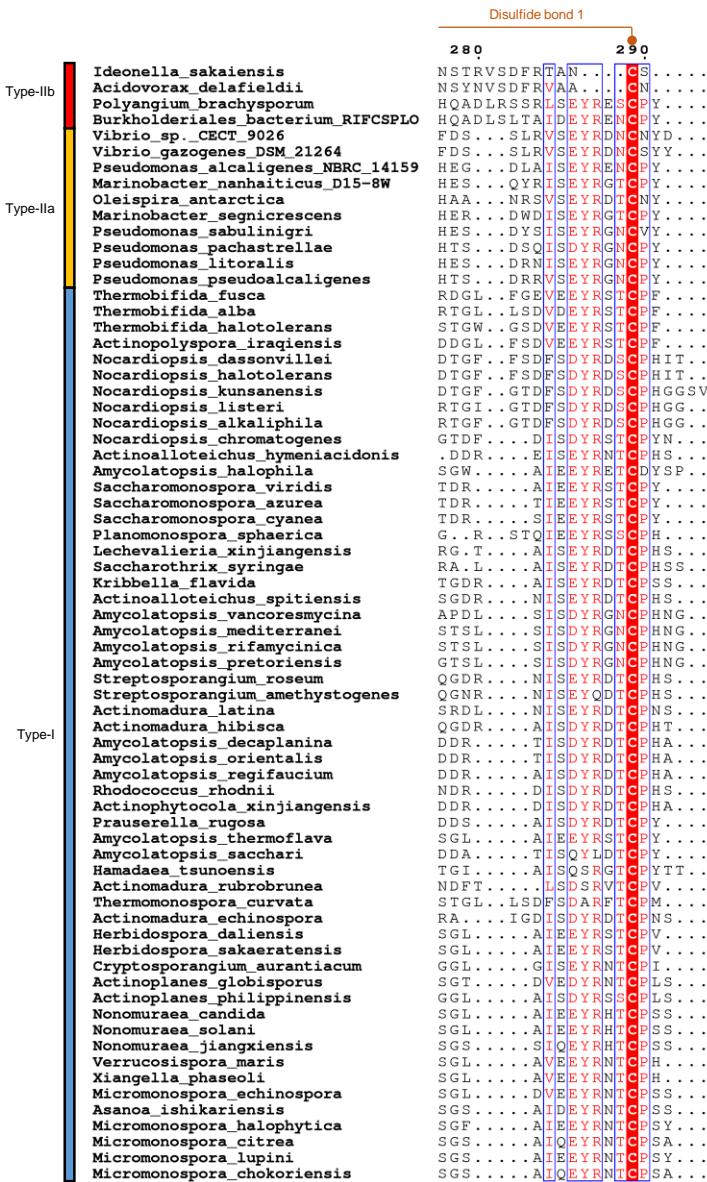
**Supplementary Figure 5.** Amino acid sequence alignment of PET-degrading enzymes (continued).



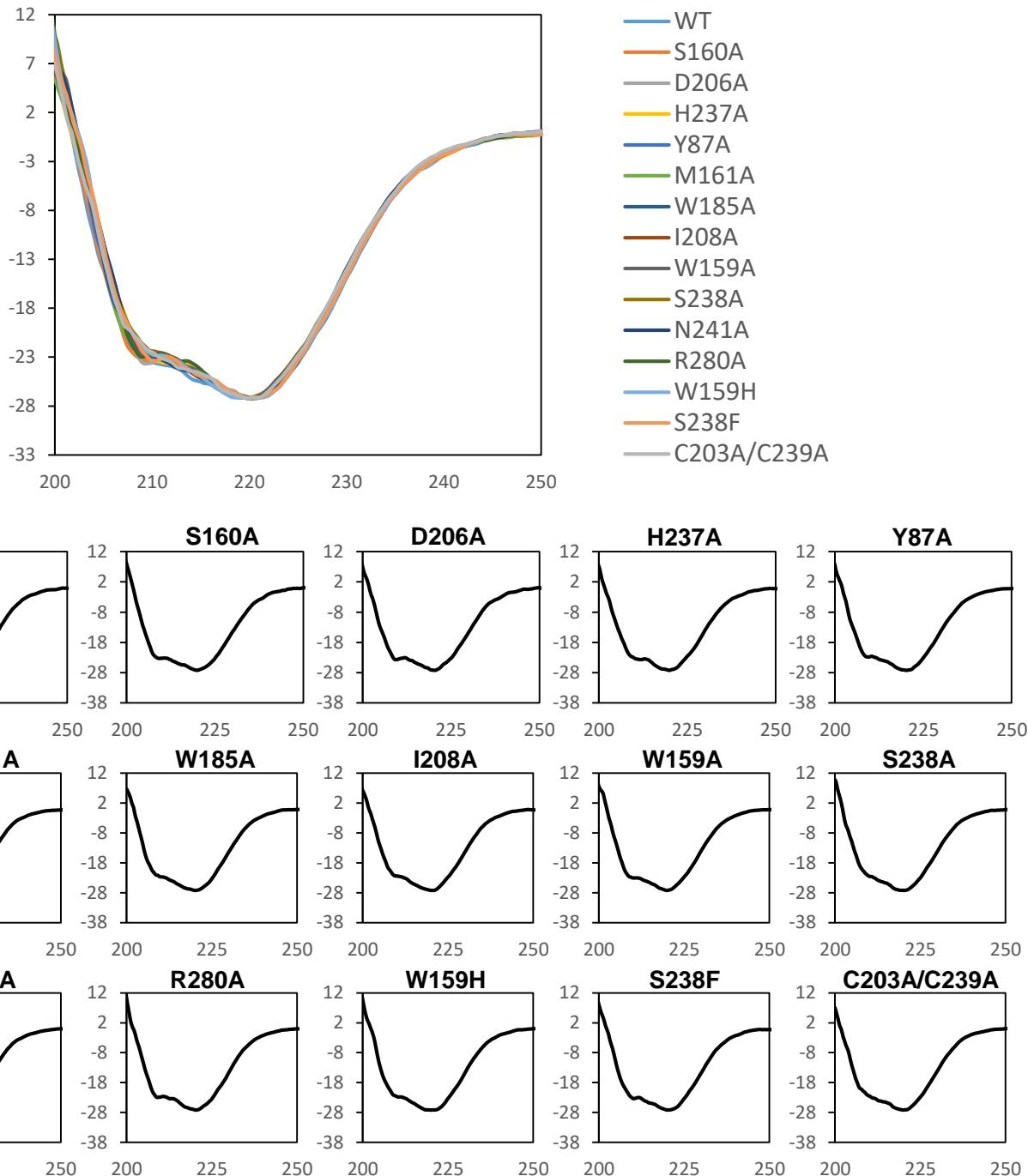
**Supplementary Figure 5.** Amino acid sequence alignment of PET-degrading enzymes (continued).



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**Supplementary Figure 6.** Far-UV Circular dichroism. Above figure includes superposed CD spectra of the *IsPETase* variants used in this study. Each spectrum of the variants is also shown in below. The spectra indicate that there are no significant difference among the variants and *IsPETase*<sup>WT</sup>.

	<i>IsPETase</i>	<i>IsPETase</i> <sup>R280A</sup>
<b>PDB code</b>	5XJH	5YNS
<b>Data collection</b>		
Wavelength (Å)	0.97934	0.97934
Unit cell ( <i>a</i> , <i>b</i> , <i>c</i> ; $\gamma$ ) (Å; °)	43.48, 50.40, 129.49; 90.0	43.61, 50.59, 129.58; 90.0
Space group	<i>P</i> 2 <sub>1</sub> 2 <sub>1</sub> 2 <sub>1</sub>	<i>P</i> 2 <sub>1</sub> 2 <sub>1</sub> 2 <sub>1</sub>
Solvent content (%)	53.38	52.66
Protein chains in AU	1	1
Resolution range (Å)	50.00-1.55	50.00-1.36
Highest resolution shell (Å)	1.58-1.55	1.38-1.36
Unique reflections	42939	62434
Redundancy	6.5(5.9)	9.3(5.7)
Completeness (%)	99.9(99.9)	98.7(96.9)
<i>R</i> <sub>merge</sub> (%)	6.8(29.8)	6.7(30.1)
Average I/σ(I)	31.4(5.2)	40.8(4.6)
B from Wilson plot (Å <sup>2</sup> )	14.2	10.8
<b>Refinement</b>		
<i>R</i> (%)	15.9	16.6
<i>R</i> <sub>free</sub> (%)	19.2	19.2
Mean B value (Å <sup>2</sup> )*	16.1	15.0
RMS deviation bond lengths (Å)	0.026	0.025
RMS deviation bond angles (°)	2.286	2.249
Number of amino acid residues	272	264
Number of water molecules	150	294
<b>Ramachandran plot</b>		
Most favored regions (%)	97.3	97.6
Additional allowed Regions (%)	2.7	2.4

\*Mean B value is for both protein atoms and the solvent molecules.

**Supplementary Table 1.** Data collection and refinement statistics of *IsPETase*.

Extra amino acids at N-terminus	MGSSHHHHHSSGLVPRGSHM
Codon optimized DNA sequence used for expression of <i>IsPETase</i>	CGCGGTCCGAATCCGACAGCCGCCAGTTGGAAG CGAGCGCTGGTCCATTACCGTTCGCTCCTTACC GTGAGTAGACCGAGCGGTTATGGCGCTGGCACCG TTTACTATCCAACAAATGCTGGGGTACCGTGGC GCCATAGCCATAGTTCCGGGTATACGGCACGGCA GTCATCAATTAAATGGTGGGGACCGCGTCTGGCAT CCCACGGTTCTGTAGTAATTACAATTGACACAAATT CCACGTTAGACCAGCCATCAAGTCGGAGTTCGCAA CAAATGGCCGCCTGCCAGGTGGCGTCGTTAA ACGGTACAAGTAGCAGCCCCGATTACGGAAAGGTC GATACCGCTCGTATGGGTGTTATGGGGTGGAGTAT GGGAGGGTGGAGGCTCCCTGATCTCTGCTGCTAAC AACCCTTCGCTGAAAGCAGCGCGCCTCAAGCAC CATGGGATTCTCGACAAATTAGTTAGTTCTGTAAGT TGCCCACGCTGATCTCGCATGTGAAAACGATACT ATAGCCCCGGTCAACTCTTCAGCACTTCTATCTAT GATTCTATGTCACGCAACGCTAACGAGTTCTCGA AATTAAATGGTGGCTCACATTCCGTGCGAATAGCG GCAATTCTAACCAAGCATTAAATCGGAAAAAAAAGGC GTTGCATGGATGAAACGTTTATGGACAATGATACT AGGTATTCTACTTTGCCTGCGAGAACCCGAATAG CACCAAGAGTGTCTGATTTCGTACAGCGAATTGCA GC
Extra amino acids at C-terminus	LEDPAANKARKEAELAAATAEQ

**Supplementary Table 2.** Codon-optimized DNA sequence used for expression of *IsPETase* and extra amino acids at the N- and the C-termini.