Mitochondrial dysfunction activates lysosomal-dependent mitophagy selectively in cancer cells

SUPPLEMENTARY MATERIALS

Conjugated and unconjugated redox active agents



Supplementary Figure 1: Chemical Structures of redox active molecules conjugated and unconjugated to the mitochondrial targeting moiety. (a) The structure of the mitochondria-targeted redox active molecules: mitoquinone, mitotempol, mitochromanol acetate and mitoapocynin. (b) The structure of the redox active molecules: ubiquinone, tempol, chromanol acetate, and apocynin unconjugated to a carbon linker chain and triphenylphosphonium.

Mitochondria membrane potential in MCF-12A cells



Supplementary Figure 2: Autophagic flux and mitochondrial depolarization in MDA-MB-231 and MCF-12A cells. JC-1 analysis of mitochondrial membrane potential in MCF-12A cells treated with 1 μ M of ubiquinone, MitoQ, tempol, MitoT, chromanol acetate, MitoCA, apocynin, and MitoApo for 24 hours. Bar represents mean \pm SEM. (*n*=3). **P*<0.05 indicates statistical significance.

MTA induced mitochondrial dysfunction



Supplementary Figure 3: MTAs induce a loss in mitochondria function but not mitochondrial content at 6 hours in MDA-MB-231 cells as compared to MCF-12A cells. (a) Mitochondria membrane potential was determined using JC-1 fluorometrics in MDA-MB-231 and MCF-12A cells treated with 1 μ M of MitoQ, MitoT, MitoCA and MitoApo for 6 hours. Bar represents mean ± SEM. (*n*=3). (b) Mitochondria protein turnover was determined by fluorescent emission in MitoTracker red preloaded MDA-MB-231 and MCF-12A cells treated with 1 μ M of MitoQ, MitoT, MitoCA and MitoApo for 6 hours Bar represents mean ± SD. (*n*=3). (c) Representative confocal images of MitoTracker red preloaded MDA-MB-231 and MCF-12A cells following 6 hours treatment with each MTA at 1 μ M. Scale bar is 40 μ M. Yellow box is area selected for magnification. (d) Large (3x3 tiles stitched) merged confocal images of MTA treated MDA-MB-231 cells stained with MitoTracker green, MitoSox and Hoechst. Scale bar is 100 μ m. (e) Oxygen consumption rate (pmol/min) was measured using the Seahorse XF96 Flux analyzer, while Hoechst fluorescent emission was determined to normalize the mitochondrial function per cell in MDA-MB-231 and MCF-12A cells after 6 hours of 1 μ M MTA treatment. Bars represent mean ± SEM (*n*=3). **P*<0.05 and ***P*<0.01 indicates statistical significance.



CCCP induced mitochondrial acidification

Supplementary Figure 4: Mitochondrial pH changes in stably expressing mt-mKeima MDA-MB-231 and MCF-12A cells treated CCCP. (a) Representative cell population dot plots for MDA-MB-231 and MCF-12A cells expressing mt-mKeima treated with 30 μ M CCCP for 3 hours. (b) Quantification of three individual experiments for the percentage of cells that shifted into the upper (4>pH<6) quadrant. Bar is mean \pm SD (n=3). **P<0.01 indicates statistical significance.



MTA induced lysosomal dependent mitochondrial degradation

Supplementary Figure 5: MTAs induce mitophagy. (a) Representative cropped LC3 immunoblot of MDA-MB-231 and MCF-12A cells without any MTA treatments in the presence and absence of 5 nM Baf for 2 hours. Bars represent mean \pm SD. (n=3). (b) Representative MDA-MB-231 cell population analyses in the presence and absence of 100 nM Rapamycin or serum starvation for 24 hours with and without 5 nM Baf. for the final 2 hours. Graph depicts the cell population shift from three independently performed experiments. Bar is the \pm and SD (*n*=3). Upper and lower quadrants were set on the non-Baf. treated cells. (c) Representative MDA-MB-231 and MCF-12A cell population analyses after 12 hours of DMSO or MTA treatment in the presence or absence of 5 nM Baf. for the final 2 hours. (d) Confocal image of mt-mKeima expressing MDA-MB-231 cells treated with the indicated MTAs at 1 μ M for 24 hours in the presence of lysotracker green (100 nM). Scale bar is 20 μ M. (e) Kinetic confocal analysis of MTA treated MDA-MB-231 cells expressing mt-mKeima in the presence of Lysotracker in the presence of Baf. Scale bar is 10 μ M. **P*<0.05 indicates statistical significance.

Supplementary Table 1: MDA-MB-231 stable expressing mt-mKeima FACS analysis

Table describes the percentage of the MDA-MB-231 cell population in the upper and lower quadrants from three independently performed FACs analyses. Cell count was 50,000 events per condition per experiment. (i-iii) MDA-MB-231 cells were treated with DMSO (control) or 1 μ M of different MTAs for (i) 6, (ii) 12 and (iii) 24 hours prior to FACS analysis. (iv) Upper and lower quadrants were set using control cells. Cells were treated with or without 30 μ M CCCP for 3 hours. (v) MDA-MB-231 cells were treated with or without 30 μ M of CCCP for 3 hours in the presence or absence of 5 nM Baf. Non-Baf. treated cells were used to set the upper and lower quadrants. (vi) Cells were treated with 1 μ M MTA for 10 hours followed by 5 nM Baf. for 2 hours prior to analysis. Non-Baf. treated cells were used to set the upper and lower quadrants. (vii) Cells were treated with and without 100 nM rapamycin or (viii) serum starved for 24 hours in the presence and absence of 5 nM Baf. for the final 2 hours. Non-Baf. treated cells were used to establish the upper and lower quadrants.

See Supplementary File 1

Supplementary Table 2: MCF-12A stable expressing mt-mKeima FACS analysis

Table describes the percentage of the MCF-12A cell population in the upper and lower quadrants from three independently performed FACS analyses. Cell count was 50,000 events per condition per experiment. (i-iii) MCF-12A cells were treated with DMSO (control) or 1 μ M of different MTAs for (i) 6, (ii) 12 and (iii) 24 hours prior to FACS analysis. (iv) Upper and lower quadrants were set using control cells. Cells were treated with or without 30 μ M CCCP for 3 hours. (v) MCF-12A cells were treated with or without 30 μ M of CCCP for 3 hours in the presence or absence of 5 nM Baf. Non-Baf. treated cells were used to set the upper and lower quadrants. (vi) Cells were used to set the upper and lower guadrants.

See Supplementary File 2