

SUPPLEMENTARY INFORMATION

Structural and functional characterization of shaft, anchor, and tip proteins of the Mfa1 fimbria from the periodontal pathogen *Porphyromonas gingivalis*

Michael Hall¹, Yoshiaki Hasegawa^{2*}, Fuminobu Yoshimura² and Karina Persson^{1*}

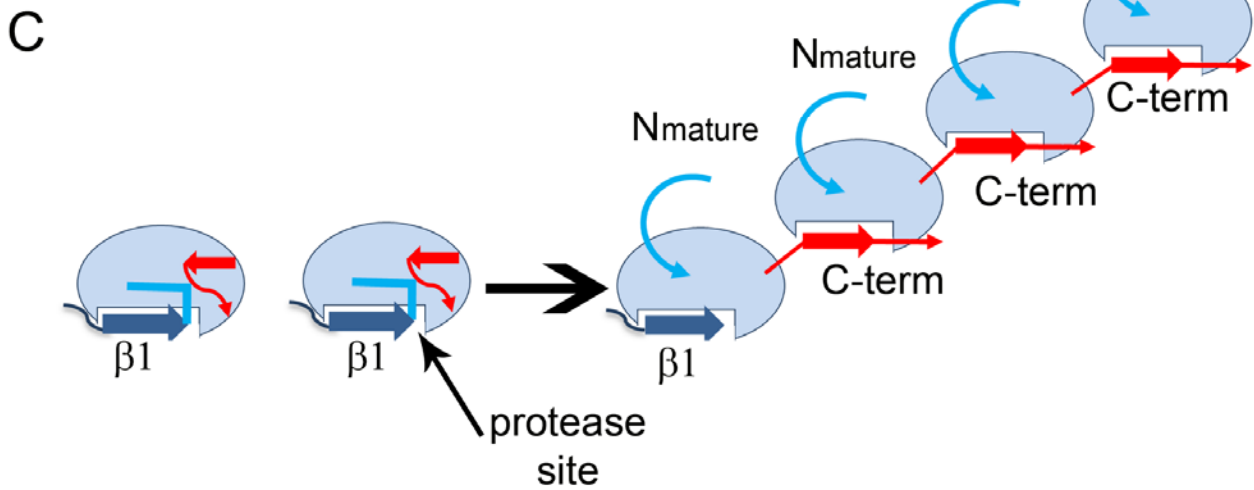
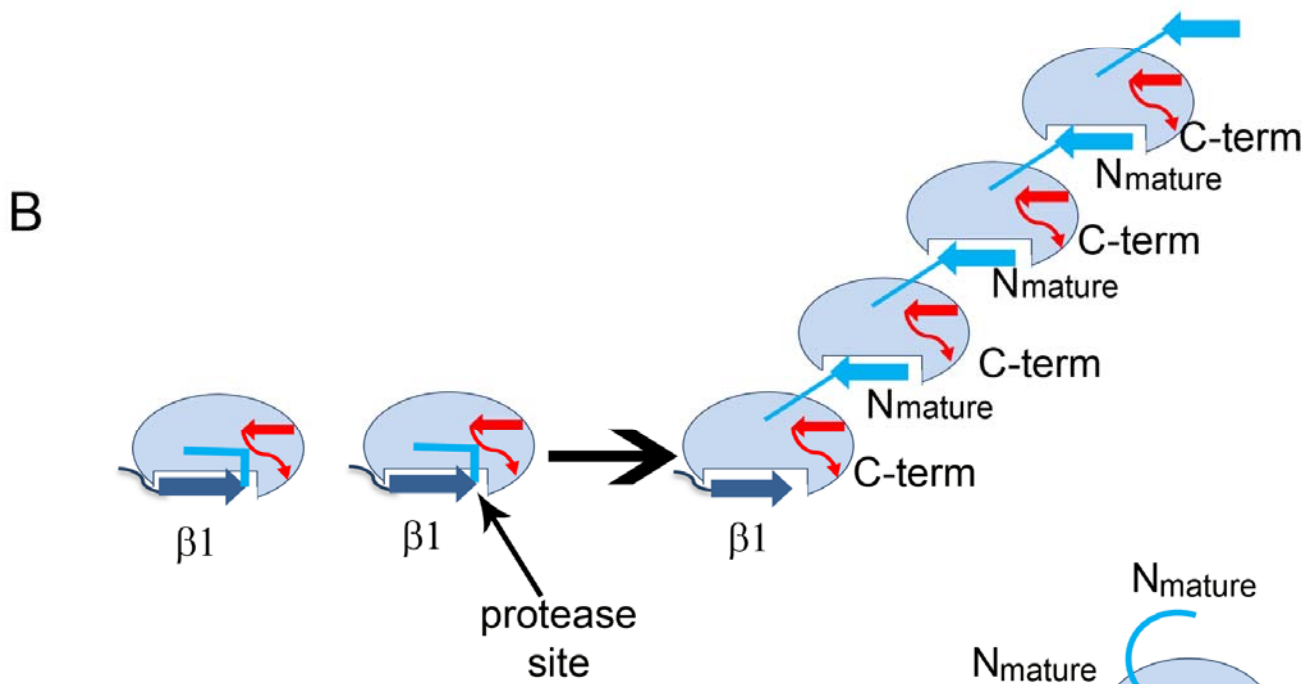
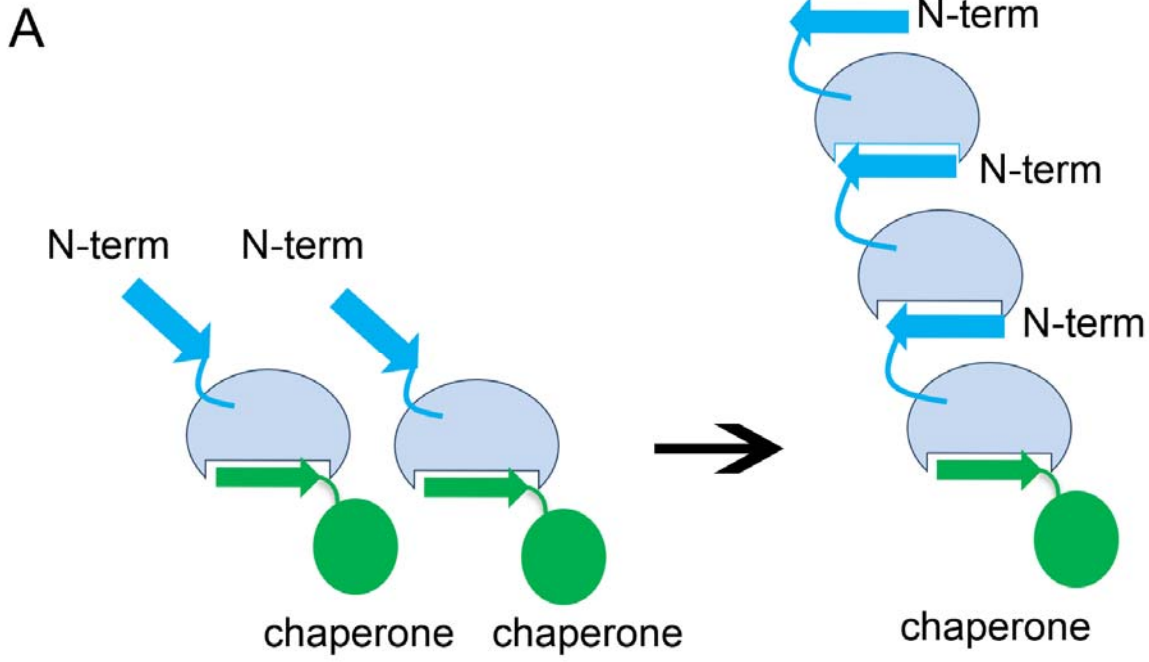
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Supplementary figure S1. Examples of polymerization mechanisms. **(a)** A simplified overview of the chaperone-assisted polymerization of type-I fimbria. Fimbrial proteins are transported to the membrane in complex with a chaperone that donates a β -strand to an incomplete sheet of the fimbrial protein. During polymerization the chaperone β -strand is displaced by an N-terminal β -strand from a neighboring fimbrial protein. Note that the β -strand from the fimbrial protein is oriented in the opposite direction of the chaperone β -strand. The fimbrial protein is depicted in light blue, the chaperone in green and the N-terminal β -strand in turquoise.

(b) Hypothetical type-V polymerization, version 1. The fimbrial proteins are expressed with an internal chaperone, the β 1-strand, located in the N-terminal domain. A protease cleaves the loop that follows, creating a new terminus (N_{mature}). During polymerization the β 1-strand is displaced by N_{mature} from a neighbouring fimbrial protein. Note that the orientation of the donated strand runs in the opposite direction of the internal chaperone, the β 1 strand, similar to the strand arrangement in type-I polymerization. The fimbrial protein is depicted in light blue, β 1 in dark blue, N_{mature} in turquoise and the C-terminus in red.

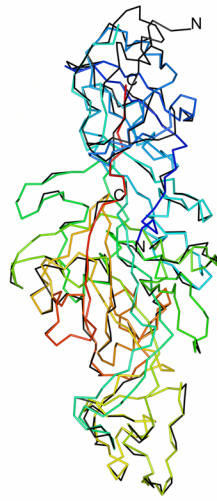
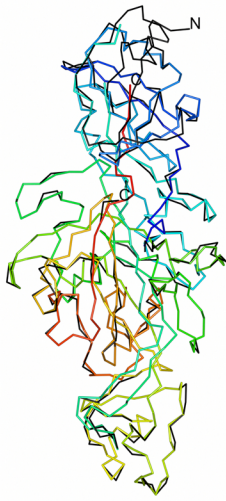
(c) Hypothetical type-V polymerization, version 2. As above the protein is expressed with an internal chaperone, β 1 that is cleaved off by a protease. During polymerization, the C-terminal β -strand and the following extended region elongate and form a long donor strand that reaches over both the N- and C-terminal domain. Note that in this model the displacing β -strand runs in the same direction as the internal chaperone, β 1. Coloring as in (b).

β1β2-loop

Mfa1	21	SKEGNGPDPDN-AAKSYMSMTLSMFMGSAAGDGGDQANPDYHYV-----GEWAG---KDKIEKVSIIY	79
Mfa2	30	-DKMIYDNYDD--CPRGVYVNFYSQ-----TECA-ENPSY-PAEVARLNVIY	70
Mfa3	22	-DRGVDPQDPDLPQPDVYLLVNAIAAHT-----NGEESINMDAE-----DFEDRVHSLAML	70
Mfa4	20	--SKNNPSEP--VEDRSIEISIRVD-----DFTKTGETVRYEINQGS-AAERLITINLYLL	69
		 	
Mfa1	80	MVPQGGPGLVESAEEDLD-FG---TYENPTIDPATHNAILKPKKGIVNSAVGKTVKVVVLLNDIAGKAK	145
Mfa2	71	AFDKDGIL-RSANVFED-----VQLS--AA-----KEWLIPL--EKDGLYTIFAWGNIDDHYNI	119
Mfa3	71	VFDSNTGEKVAEHFSSS-IG---SGTSTYV-----F'TVKL--KPGQRDFFFVANIPNM--Q	118
Mfa4	70	LFDQSGAN-PAKYI IAGNTFSGGIWLPDD-----MKV--KLDMTQSEAGERKVVVAVNDNAV-K	125
		    	
Mfa1	146	ALLAN-VNAADFDAKFKEIIELSTQAQALGTVADGPNPAT-AAGKIAKNGTDETIMTCLQP-SDALT	212
Mfa2	120	GEIKIGET---TKQQ--VLMRLKQ--DGK-----W-AT--NID-----GTTLWYA-TSPVVELK	162
Mfa3	119	TAMAS-IVNKSMDNHFMQVFRDLDD-----PIHYH--NATNN--NGFPMSR-MYSNQTVT	166
Mfa4	126	TALDA-VA--NESDLQTVKRTTA-----M-PWSTDIA-----SPFLMSG--NKTHDFL	167
		    	
Mfa1	213	-----IEAAVSEANAIAGIKNQAKVTEISVARAMVSTKAQSYEIKATTQIGEIAAGSVLA	268
Mfa2	163	NMEDGAD---QYIH-----TRANLREYTNRVTVSVDS-----	191
Mfa3	167	-----IGGTITQPLPF-----KPDGENNVKLOISVVAKLDVNIIVE-----G	201
Mfa4	168	-----ANRLL-----DNVPLVRAIAKVELNISL-SEKF-----Q	195
		 	
Mfa1	269	TIT-----DIRWVVAQGERRQYL--SKRGTVPENTWVTPGSGFVPTSSTFHTNATEYYDYAGLWED	328
Mfa2	192	LPH-----PENYEIKL-ASSNGSYRFDGTV-----AKA-----DSTYY-----	223
Mfa3	202	VEN-----LQKIELCNANVHYRL--VPNQ-----SEPI-----QFYGPV-----	233
Mfa4	196	IVPIIVNGSLSEFKFRYVNFDKETIV--VKPT-----TKPD-----NLISS-----A	235
		 	
Mfa1	329	HN---TNEAVISGTQVPTLADYQLQDVTGELANAL-----SGKFLLPN-THKSGANAASSDY	381
Mfa2	224	-----PGETKVVG-----DSTCRAFFTTLKLESGH-----	248
Mfa3	234	-----ELRR-----VGAT-----NQWLGYPMEA-IVESTKWWGNTGN	264
Mfa4	236	NGVWPQIT---DWTVWG--ASLNTSPA--PDAGTYTLDANGKVTALRIVTYLNER-----DSK	287
			
Mfa1	382	-KRGNTAYVLVRAKFTPKKEAFIDRGKTYSDNTAVPEYVAGEDDFVGENGQFYVSMKSVTDPKVGGVAGM	450
Mfa2	249	---ENTLSVTHKP-----	258
Mfa3	265	AENKPINFFRLTTRG-----	279
Mfa4	288	---GATVEVALPRV-----DDG	301
			
Mfa1	451	KAHKYVKGVLYYAWLNPST----TSPDSW-----WNSPVVRNNIYHIHIKSIKGLFNWMLVLPD	507
Mfa2	259	-----TGREIFRTDLVGAILS--SQYAQ-----NINLRCINDFDIRLVAHHCNCPD-----	302
Mfa3	280	-----GLVYDVPIITHEGAIPGGQYLPFAKGLLADKPSYTVYRNRHYIYRIKTLPLD-----	330
Mfa4	302	TLPPPEFGPELYRLPLP-----DKILRNHWYKYEVEI-----	333
		 	
		proline rich-region	
Mfa1	508	FDPSNPENPNPNPNPDEPGTVPVPTDPENPLPDDQDTFMSVEVTVLPK-----	555
Mfa2	303	-----DTYVVVQIWING--ML-----	316
Mfa3	331	-----KIEVKYSICDNNIVTNDTYMGYGYNVGVDEQGNV	364
Mfa4		-----	
			
Mfa1	556	-----VHSYEVDL--	563
Mfa2	317	-----IHSYEIEL--	324
Mfa3	365	TITNTMQNCDPHVVRLVAKNGAYFGSQPTDTSVEFTELANGASQTFKVNKDAVAVGSAYLEVYVNPDLNA	434
Mfa4		-----	
Mfa1		-----	
Mfa2		-----	
Mfa3	435	TGVVPDKVFIKK	446
Mfa4		-----	

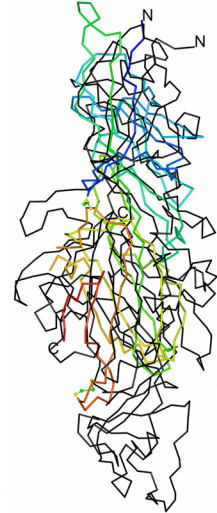
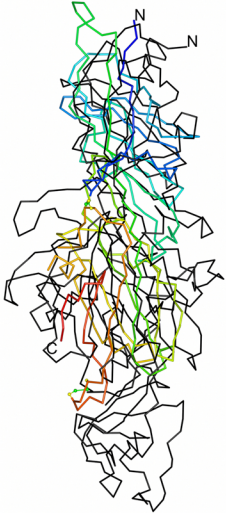
Supplementary figure S2. Structure assisted multiple sequence alignment of proteins building up the Mfa1 fimbria performed using PROMALS3D ¹. The alignment is based on the precursor sequences of each protein and the mMfa1, Mfa2, mMfa3 and Mfa4 (pdb code: 5dhm) structural models. Consensus secondary structure features are indicated as cylinders (α -helices) and arrows (β -strands) below the alignment. Amino acids of specific interest are highlighted red (RgpA/B cleavage site), green (conserved tryptophan), purple (tryptophan interacting residues) and yellow (disulphide forming amino acids in Mfa2).

A



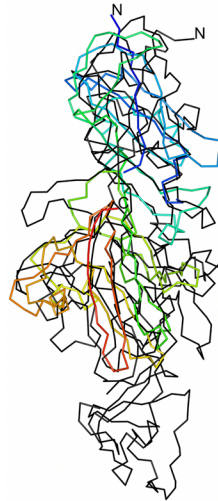
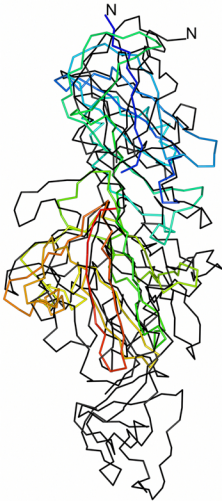
mMfa1/Mfa1 Δ 9

B



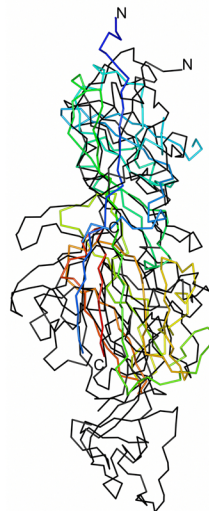
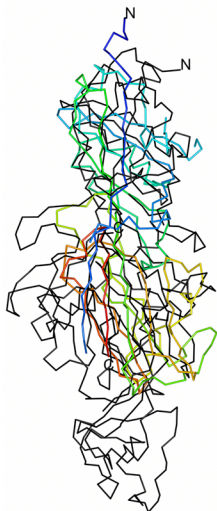
Mfa2/Mfa1 Δ 9

C



Mfa3/Mfa1 Δ 9

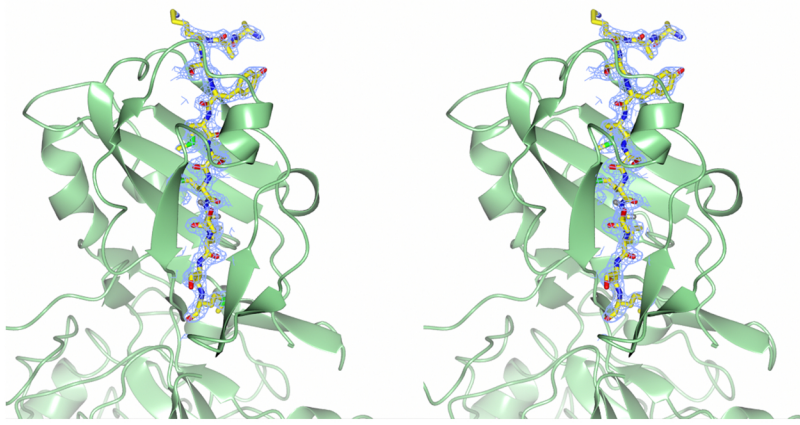
D



Mfa4/Mfa1 Δ 9

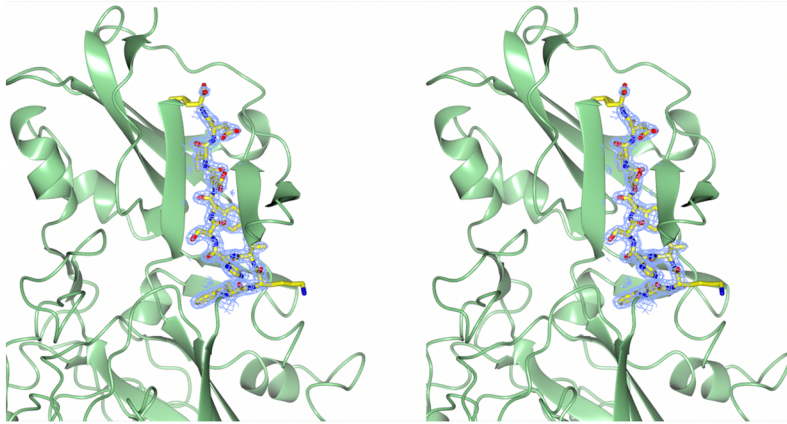
Supplementary figure S3. Structural comparison of the proteins building up the Mfa1 fimbria. The structural model of pMfa1 Δ ₉ (shaft) is superimposed on the structures of **(a)** mMfa1 (shaft); **(b)** Mfa2 (anchor); **(c)** Mfa3 (tip) and **(d)** Mfa4 (tip). pMfa1 Δ ₉ is depicted in black and the other structures are blended from the N-terminus (blue) to the C-terminus (red). The overlays are shown as C α -traces in stereo.

A



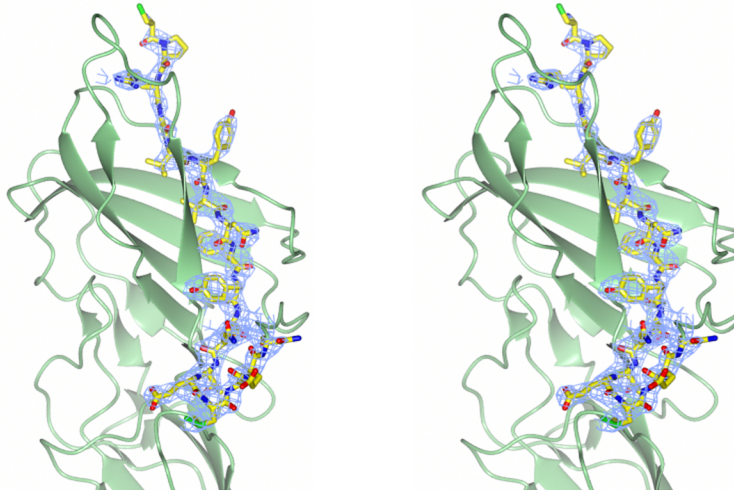
pMfa1 Δ 9

B



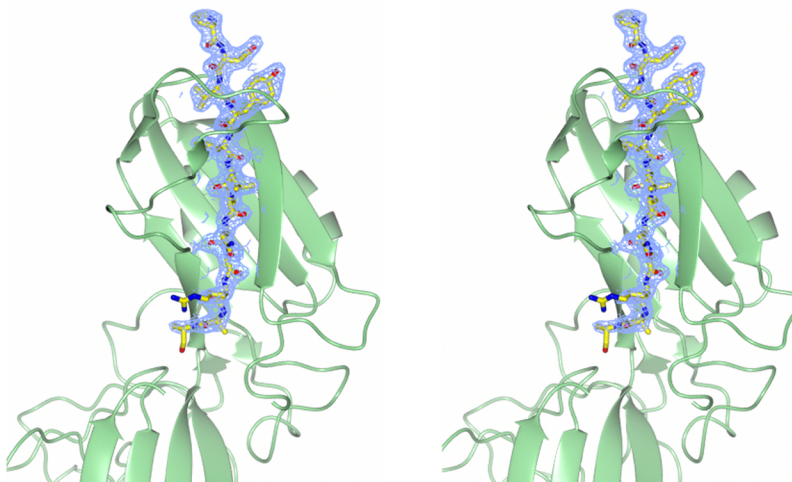
mMfa1

C



Mfa2

D

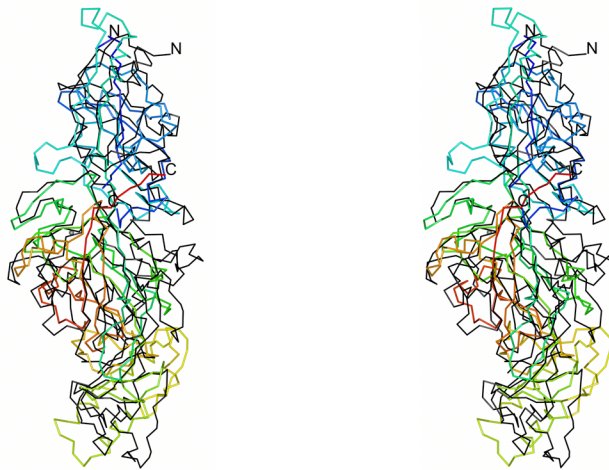


pMfa3

Supplementary figure S4. Highlighted N- and C-terminal structural features in β -sheet 1. **(a)** The N-terminal extension of pMfa1 Δ 9, **(b)** the final strand of mMfa1, **(c)** the N-terminal extension of Mfa2, anchored by a disulphide bond and **(d)** the N-terminal extension of pMfa3. All N- and C-terminal structural features are shown as stick models in a 2fo- f_c map contoured at 1σ in stereo.

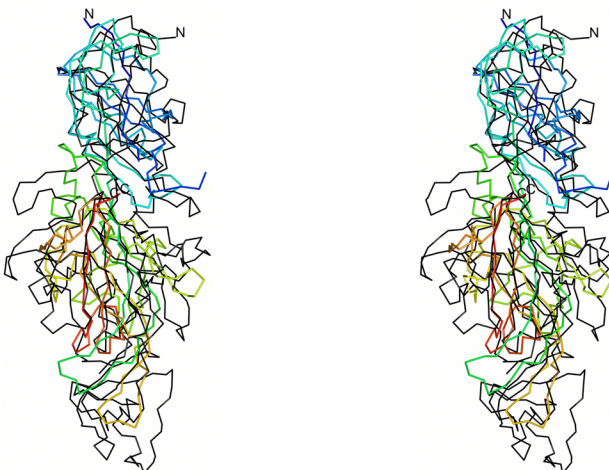
Supplementary figure S5. Structure assisted multiple sequence alignment of Mfa1 and the related shaft proteins BovFim3A from *B. ovatus* (pdb code: 4jrf), *P. distasonis* BdiFim3A (3liu), *B. eggerthii* BegFim1A (4gpv) and FimA from *P. gingivalis* strain W83 (4q98) performed using PROMALS3D. Consensus secondary structure features; α -helices (h) and β -strands (e) are indicated below the alignment.

A



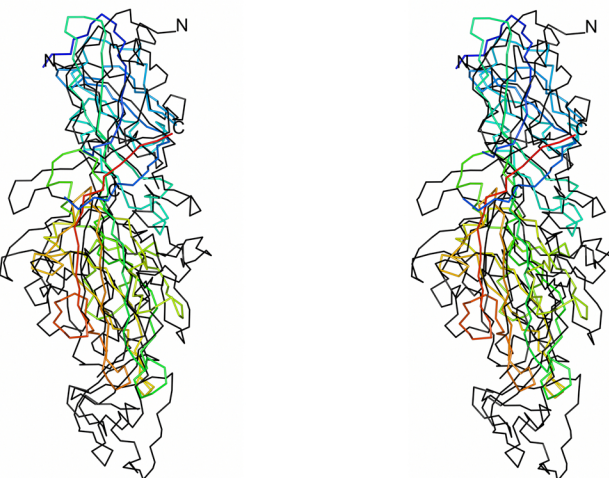
BovFim3A
(*B. ovatus*)
/Mfa1 Δ 9

B



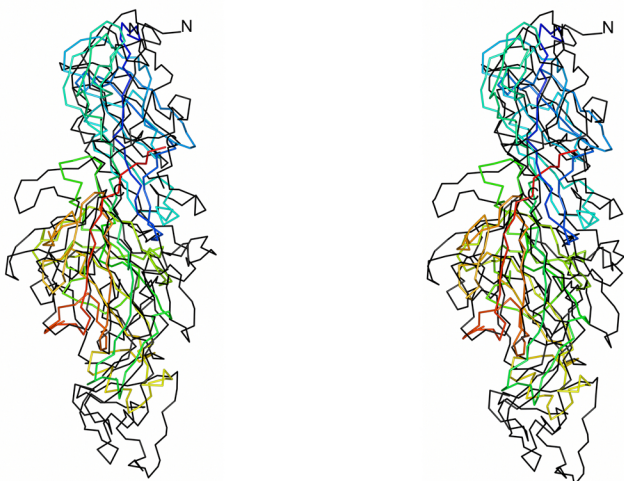
BdiFim3A
(*P. distasonis*)
/Mfa1 Δ 9

C



BegFim1A
(*B. eggerthii*)
/Mfa1 Δ 9

D

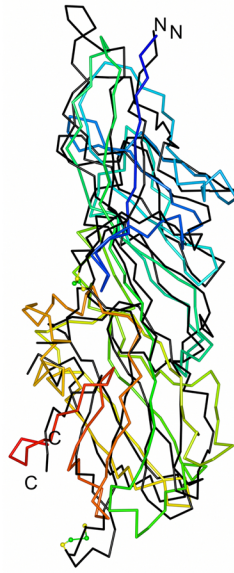
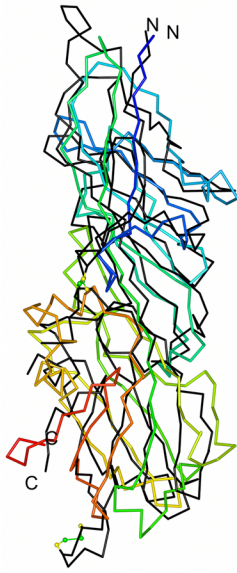


FimA
(*P. gingivalis* W83)
/Mfa1 Δ 9

Supplementary figure S6. Comparison between pMfa1 Δ 9 and structurally related shaft proteins. pMfa1 Δ 9 is superimposed on **(a)** fimbrial shaft protein BovFim3A from *B. ovatus* **(b)** *P. distasonis* BdiFim3A **(c)** *B. eggerthii* BegFim1A and **(d)** FimA from *P. gingivalis* strain W83. pMfa1 Δ 9 is depicted in black and the other structures are blended from the N-terminus (blue) to the C-terminus (red). The overlays are shown as C α -traces in stereo.

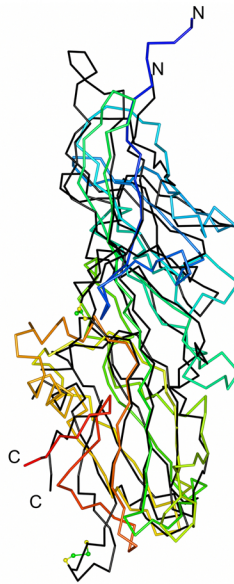
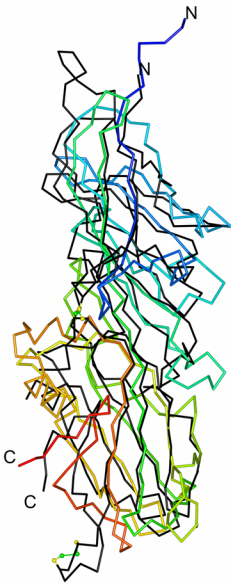
Supplementary figure S7. Structure assisted multiple sequence alignment of Mfa2 and the related anchor proteins BthFim2B and BthFim3B from *B. thetaiotaomicron* (pdb codes: 3gf8 and 4qdg) and BovFim2B from *B. ovatus* (3pay) performed using PROMALS3D. Consensus secondary structure features; α -helices (h) and β -strands (e) are indicated below the alignment.

A



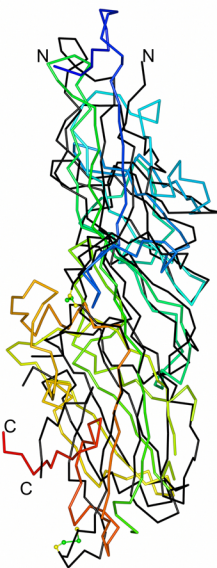
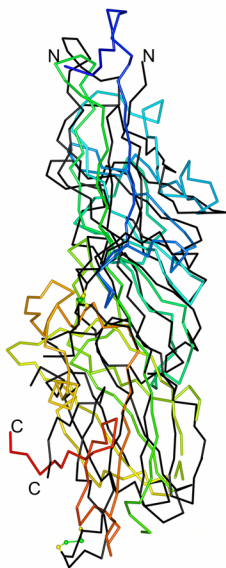
BthFim2B
(*B. thetaiotaomicron*)
/Mfa2

B



BthFim3B
(*B. thetaiotaomicron*)
/Mfa2

C



BovFim2B
(*B. ovatus*)
/Mfa2

Supplementary figure S8. Comparison between Mfa2 and structurally related anchor proteins. Mfa2 is superimposed on anchor proteins from *B. thetaiotaomicron* **(a)** BthFim2B and **(b)** BthFim3B and **(c)** *B. ovatus* (BovFim2B). Mfa2 is depicted in black and the other structures are blended from the N-terminus (blue) to the C-terminus (red). The overlays are shown as $C\alpha$ -traces in stereo.

Mfa3 1 DRGVDPQPDLPDVLV-LVNARA--TNGE--ESINMDAEDFEDRVHSLAMLVFDSN--TGEKVAEHF 63
 FimA 1 -----VVEETNATVVSFIKSGE-----SRAVGDDLTDAKITKLTAMVYAG----QVQEGIKT 49
 BovFim2C 1 -----TRAQL-SIDLNVN-----EQQEKINSMRFIVFGSTPGGVRDLVNEH 41
 BdiFim1A 1 -----VLEKAQQ-LALAIKSET-----TPTDADVNTLTVGVFGV---DGWS-VIYT 41
 BdiFim1C 1 -----PLEGAK-LSVAVKASGTATKAYNPNDVNE-LEEGEAYINNLAUVVFNET--GTELLGYKW 56
Consensus ss: eeeeeee eeeeeee hhh

Mfa3 64 SSSIGSGTSTYV---FTVKLK-PGQ---RDFFFVANIPN-MQTAMASIVNKSDMNHFMQVFRDLDPH- 123
 FimA 50 VEEED-DGGVLK---VEGIPCKSGANNRVLLVVVANHN---YELTGKS---LNEVEALTTSLTAEN- 106
 BovFim2C 42 ILLST-PETATDID-AQLLEVTSSND---ILVVVIANEPQSLTSQLDGIAN---LLTLQEMIIDISSILN 103
 BdiFim1A 42 KDATT-PNSDGTKDVGVPQEVYAGEA---HVVVVVANAAPVIQTELAQAKD---ITDFIETTINLSDE-- 102
 BdiFim1C 57 EALSG-AEHSAI---IADVPTTKAVR---ARIIVLANVPR---DLLSTVST---YDEFQTRLVDLSSQ-- 111
Consensus ss: hh ee ee eeeeeee hhhhhhh hhhhhhhhh

Mfa3 124 -YHNATNNNGF-PMSRMYSNQ--TVTI-GGTITQP-----LFPKPDGENNVKLQ--RNVAK 172
 FimA 107 --QNNAKNNLIIMTGKSAAF--TIKP-GSNHYGYGG-TAS-----DNLVVSSAGTPLAVTTRVHVHAG 164
 BovFim2C 104 SDGQIISATGM-PMTGVIRDI--SIAP-DETKTV-----QMVIE--RAVAR 143
 BdiFim1A 103 ----TLTKGL-TMSSKVLVDV--TLVANTNYIGYDDEVGDIIVKDISGKEVY--GAGPVLV--RDVAS 160
 BdiFim1C 112 ----SQT--NL-TMSSQVIVTKSALSE-EDNYLGYTDLGDQ-----NVDG--ISDPILLT--RVAAR 161
Consensus ss: eee eeee ee e e eeeee eeee ee

Mfa3 173 LDVNIV-EGVEN-----LQKIELCNANVHYRLV-----PN 201
 FimA 165 ISFAGVEEEVNMMATTQYQYQNYSSFKKPA-DAKIAALVAKKDSKIFG-----NSLV 217
 BovFim2C 144 VDV-FIEAIDGGA-----VTGYTAGSTSVTLHNFSDSYFVMGNVNGTRDNADSSKNYKGV 199
 BdiFim1A 161 IALAGA-DIGN--PENANYES---KSFVL---KEVFIASAKGVSSVASTEWEW-----GTIE 207
 BdiFim1C 162 IDLVNI-STRF---AGTPFAG---REVR---IDAVGIYNMKTksyYFSEADW-----GETE 207
Consensus ss: eee ee ee eeeeeee ee

Mfa3 202 QSEPI-----QFYGPV-----ELRRVG-----AT 220
 FimA 218 SNTTNN-----AYLYGVQQ-----TPAGLYTPDAAGETYELEASLNTNYA-----258
 BovFim2C 200 KEDVS-----ESNLLTHS-----WTAATTE-TWAYSSAPGAENRK 233
 BdiFim1A 208 KDFFGDTHFGYLDYKVGLLFLTSNNIDEEGSYKKG---LQTKYDALAKKHVEN-----DPA 261
 BdiFim1C 208 APDA-----VRNSE-----DTSFEDLLVNDGT----SISN 233
Consensus ss: eeeee

Mfa3 221 NQWLGYMPEAIVESTKWWGNTGNAB-NKPIFFF--RLTRG-----G 259
 FimA 259 VGAGFYVLESKYDDASN-----ELRPTILCCIIYGGKLLDKDGNPLTTEPALTDAINAGFCDDGDT 319
 BovFim2C 234 LLCsfyTAERLF--KSD-----YSDRLSIS--MANVLKGPSDV-----T 268
 BdiFim1A 262 LNHEFYVYENTKGEVKSg-ESNVNEAYANHTLL--IVKGDYTYLPQAK---ESIT-----KEN 314
 BdiFim1C 234 TPFVHYVMENMKS-----DDHTMI--AVKATLRGNSSYQ-----DHT 268
Consensus ss: eeeee h ee eeee

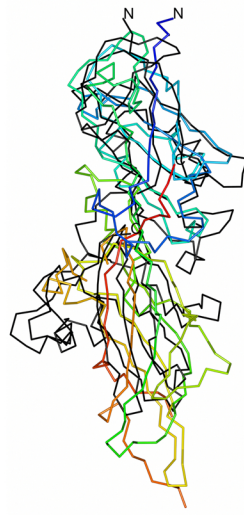
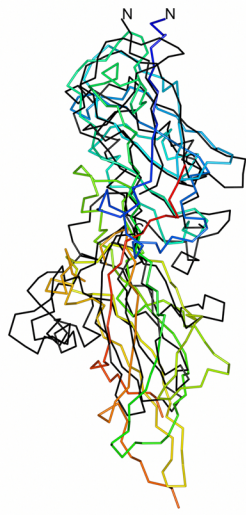
Mfa3 260 LVYDVPIITHEGAIPIGGQYLPFAKGLLA-----DKPSY-TVYRNR-HYI-YRIKTLPD-----309
 FimA 320 TYPVVLVNYD----GNGYI---YSSGA--ITQQGQN-KIVRNNHYKIKSLNITGPGT-----NT 369
 BovFim2C 269 GITGKVIESV-----TKVDGTGSPTAQPFTEIRRNN-VYQ-VTARVGKI-----310
 BdiFim1A 315 CYYAIPVGEE-----VTIDG--TEKRSKF-YVQRNY-KYEEISLTIIGPGS-----EI 358
 BdiFim1C 269 KIFTAVINAG-----GL----QNGYDHN-FIRRY-VY-RLRIYFDGESFDNIPVTPPE 315
Consensus ss: eeeeeee eeee eeeeeeeee

Mfa3 310 -----KIEVKYS--ICDWNIVTNDTYMGYGYNVGVDEQGNVITITNTMQNCDPHVRLVAKNGA 365
 FimA 370 PENFQQPPVQANLNNVTCCQVTPW-----394
 BovFim2C 311 -----GIQILTI--SVED-----321
 BdiFim1A 359 PYD--PMI--STNV-SASVK-VEPW-----377
 BdiFim1C 316 PEV-----DTN-LNIAVQ--VVGW-----331
Consensus ss: eeeeeee eeee

Mfa3 366 YFGSQPTDTSVEFTELANGASQTFKVNKDAVAVGSAYLEVYVNPDLNATGVVPDKVFIKK 425
 FimA 395 -----VVVNQ-----399
 BovFim2C 322 -----W-----322
 BdiFim1A 378 -----N-----378
 BdiFim1C 332 -----GPVMQH-----337
Consensus ss: ee

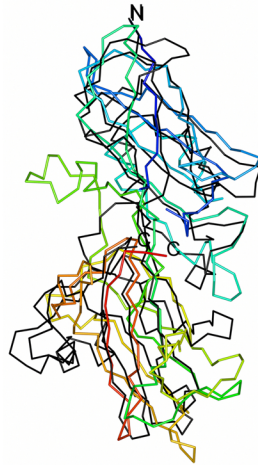
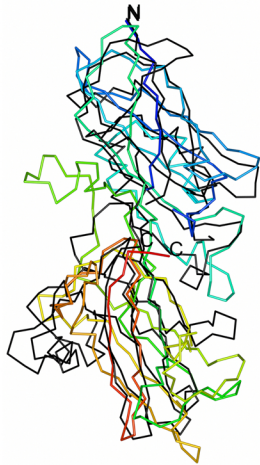
Supplementary figure S9. Structure assisted multiple sequence alignment of Mfa3 and the structurally related proteins BdiFim1C from *P. distasonis* (pdb code: 4jg5), BovFim2C from *B. ovatus* (3up6), BdiFim1A from *P. distasonis* (3liu) and FimA from *P. gingivalis* W83 (4q98) performed using PROMALS3D. Consensus secondary structure features; α -helices (h) and β -strands (e) are indicated below the alignment.

A



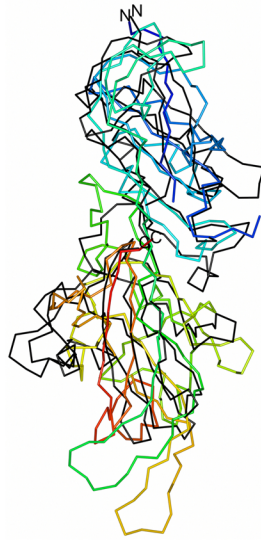
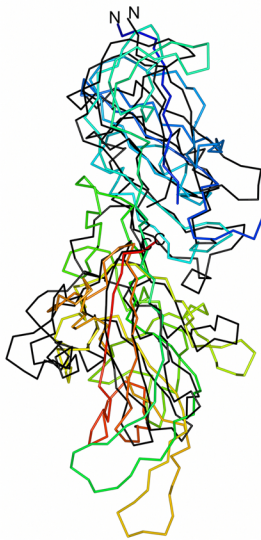
BdiFim1C
(*P. distasonis*)
/Mfa3

B



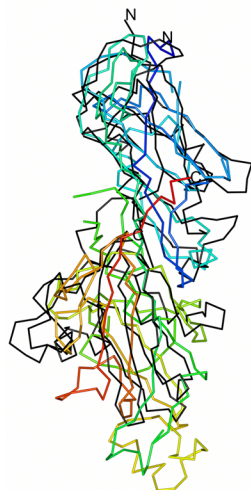
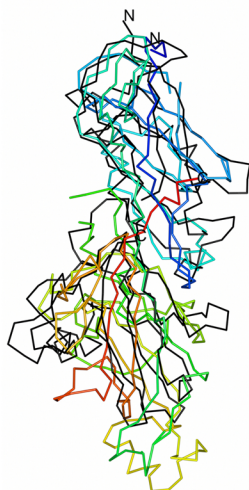
BovFim2C
(*B. ovatus*)
/Mfa3

C



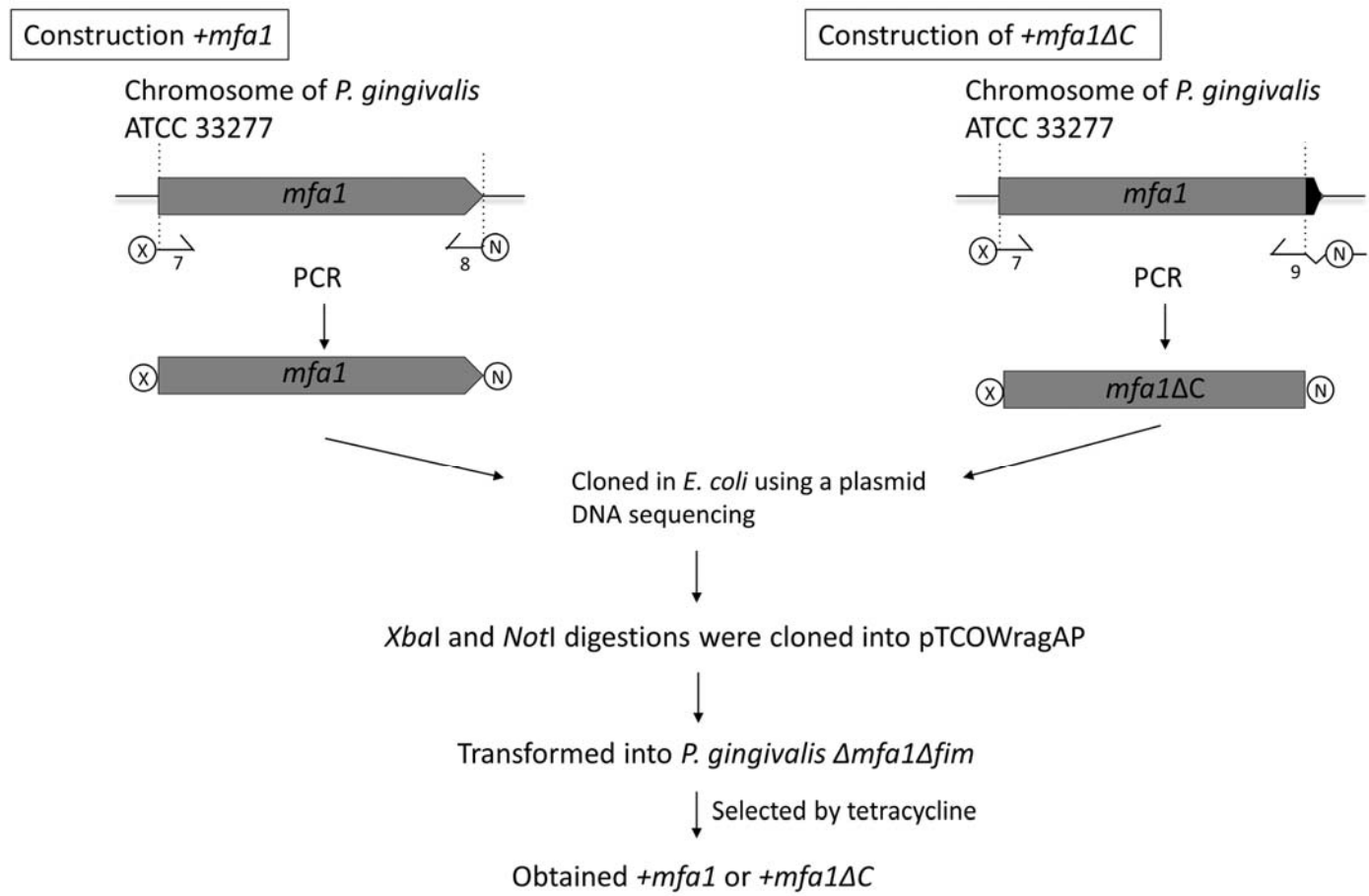
BdiFim1A
(*P. distasonis*)
/Mfa3

D



FimA
(*P. gingivalis* W83)
/Mfa3

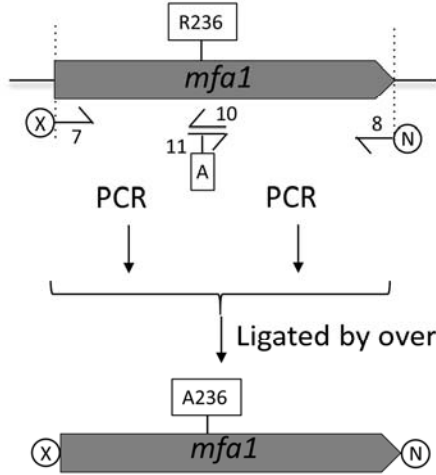
Supplementary figure S10. Comparison between Mfa3 and structurally related proteins. Mfa3 is superimposed on **(a)** a tip protein from *P. distasonis* (BdiFim1C), **(b)** a shaft protein from *B. ovatus* (BovFim2C), **(c)** a fimbrial protein of unknown function from *P. distasonis* (BdiFim1A) and **(d)** the shaft protein FimA from *P. gingivalis* W83. Mfa3 is depicted in black and the other structures are blended from the N-terminus (blue) to the C-terminus (red). The overlays are shown as C α -traces in stereo.



Supplementary figure S11. Construction of +*mfa1* and +*mfa1*ΔC *P. gingivalis* strains. Small arrows indicate the annealing sites of primers. X and N show *Xba*I and *Not*I restriction sites, respectively.

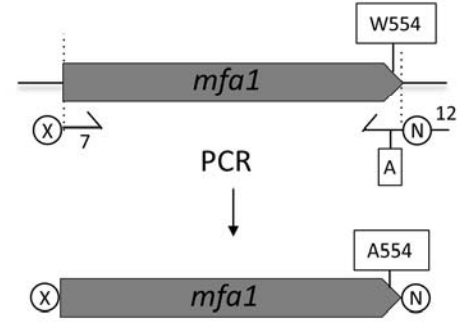
Construction of *+mfa1R236A*

Chromosome of *P. gingivalis*
ATCC 33277



Construction of *+mfa1W554A*

Chromosome of *P. gingivalis*
ATCC 33277

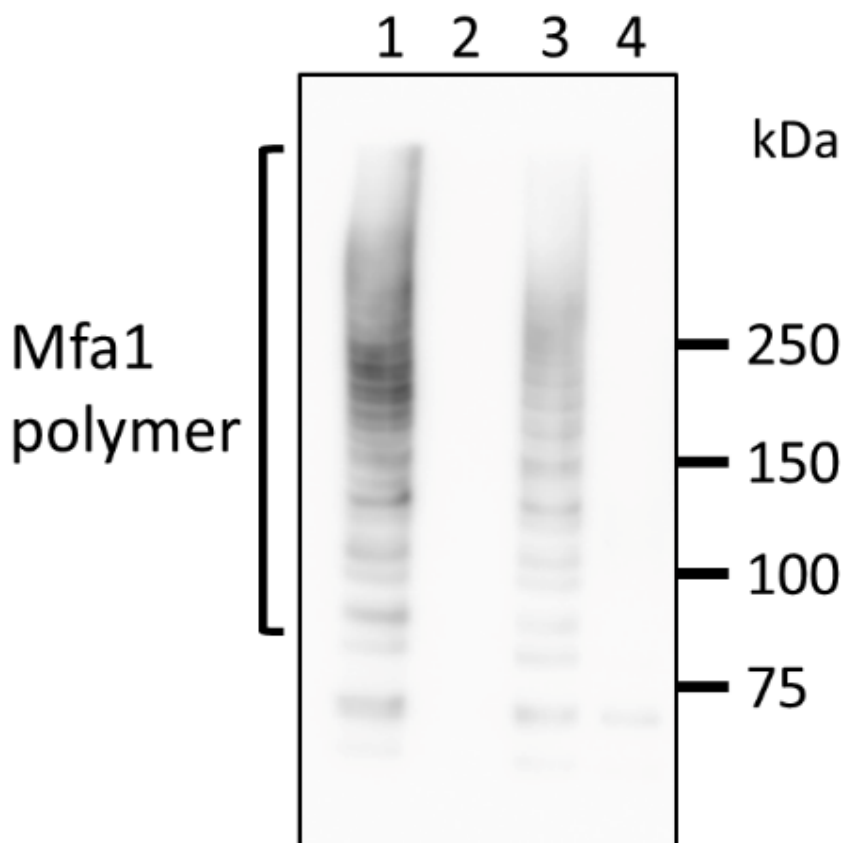


Cloned in *E. coli* using a plasmid
DNA sequencing

*Xba*I and *Not*I digestions were cloned into pTCOWragAP
and transformed into *P. gingivalis* $\Delta mfa1\Delta fim$
Obtained *+mfa1R236A* or *+mfa1W554A*

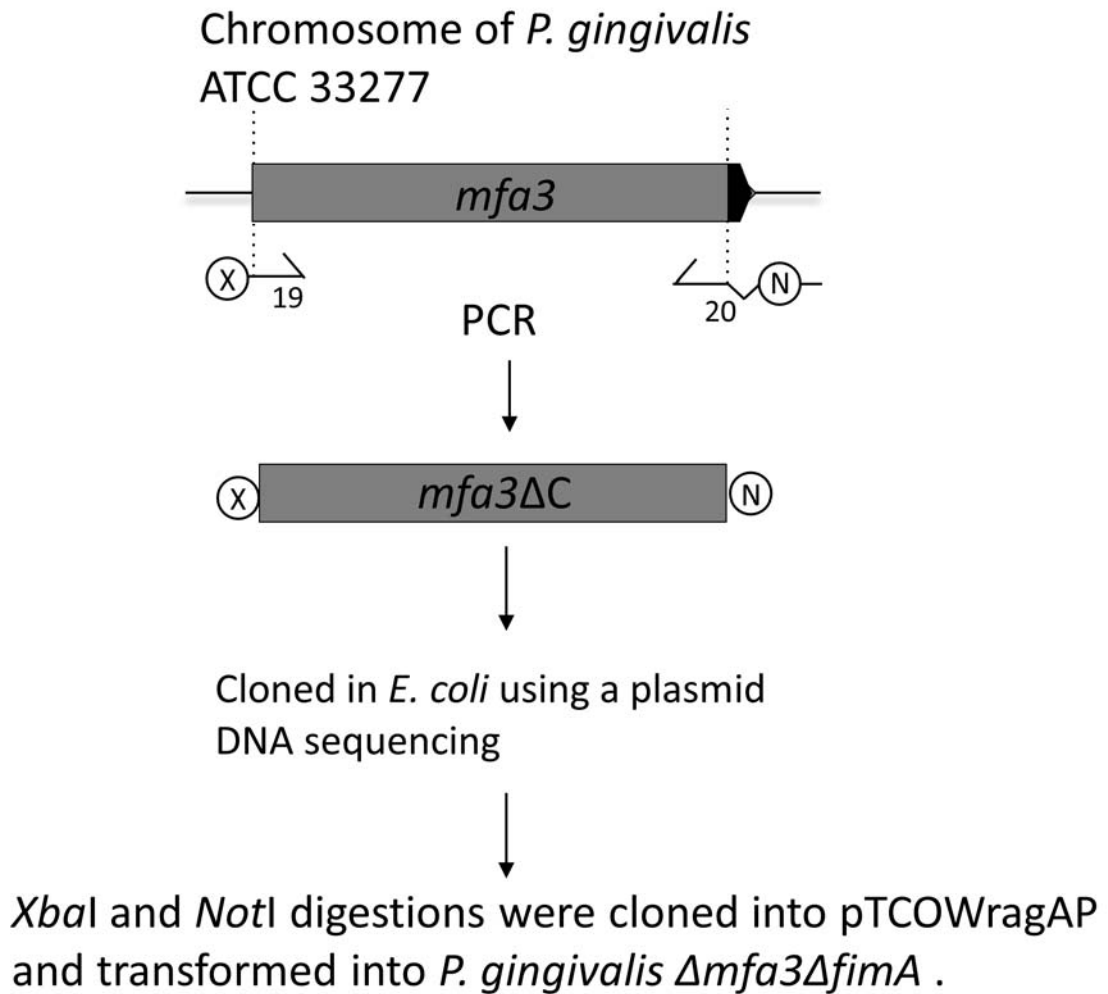
Supplementary figure S12. Construction of *+mfa1R236A* and *+mfa1W554A* *P. gingivalis* strains.

Small arrows indicate the annealing sites of primers. X and N show *Xba*I and *Not*I restriction sites, respectively



Supplementary figure S13. Analysis of Mfa1 denaturation at 42°C. Whole cell lysates were solubilized in SDS-buffer (containing 2-mercaptoethanol), heated to 42°C for 10 min, separated by SDS-PAGE, blotted to a PVDF membrane and probed with a polyclonal Mfa1 fimbriae antibody. Lanes: 1, JI-1; 2, $\Delta mfa1\Delta fim$; 3, +*mfa1*; 4, +*mfa1* ΔC .

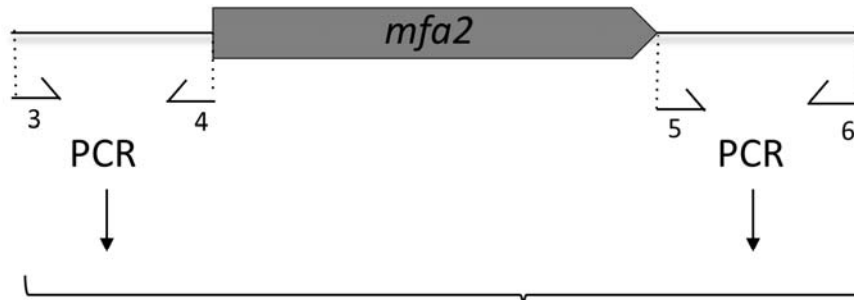
Construction of *+mfa3ΔC*



Supplementary figure S14. Construction of *+mfa3ΔC* *P. gingivalis* strain. Small arrows indicate the annealing sites of the primers. X and N show *Xba*I and *Not*I restriction sites, respectively.

Construction of $\Delta mfa2\Delta fimA$

Chromosome of *P. gingivalis*
ATCC 33277



Ligated by overlap extension PCR



Cloned in *E. coli* using a plasmid

Transformed *P. gingivalis* KDP98

Selected by chloramphenicol
DNA sequencing

Obtained *mfa2*-deletion mutant of $\Delta mfa2\Delta fimA$

Supplementary figure S15. Construction of $\Delta mfa2\Delta fimA$ *P. gingivalis* strain. Small arrows indicate the annealing sites of primers. *cat* confers chloramphenicol resistance to *P. gingivalis*.

Construction of *+mfa2*

Chromosome of *P. gingivalis*
ATCC 33277



PCR

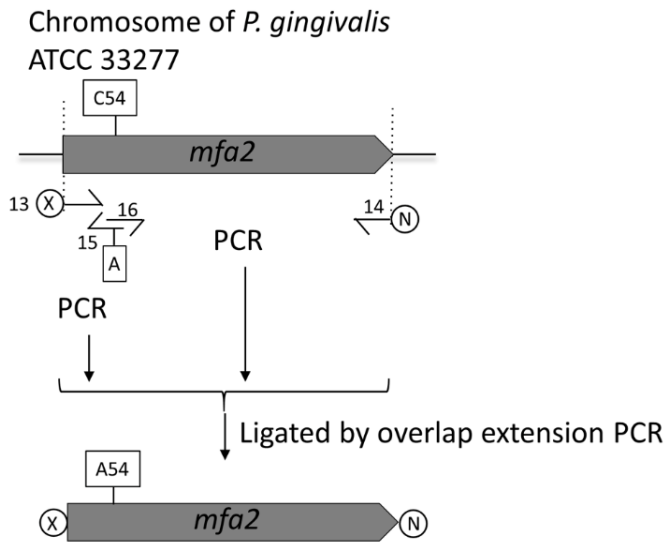


Cloned in *E. coli* using a plasmid
DNA sequencing

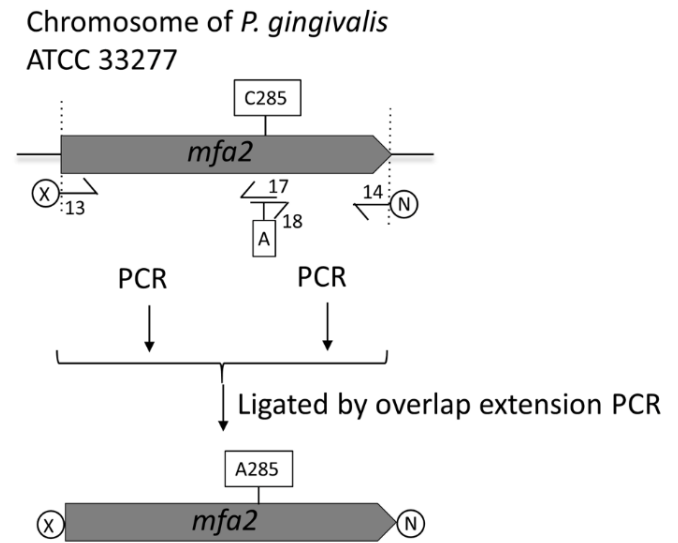
*Xba*I and *Not*I digestions were cloned into pTCOWragAP
and transformed into *P. gingivalis* $\Delta mfa2\Delta fimA$
Obtained *+mfa2*

Supplementary figure S16. Construction of *+mfa2* *P. gingivalis* strain. Small arrows indicate the annealing sites of the primers. X and N show *Xba*I and *Not*I restriction sites, respectively.

Construction of +*mfa2C54A*



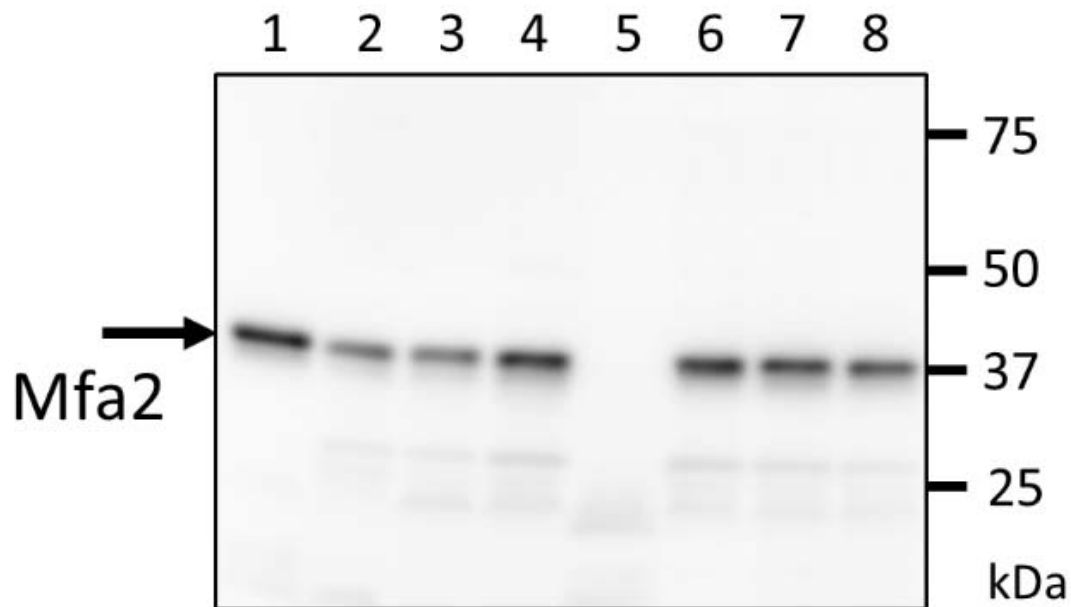
Construction of +*mfa2C285A*



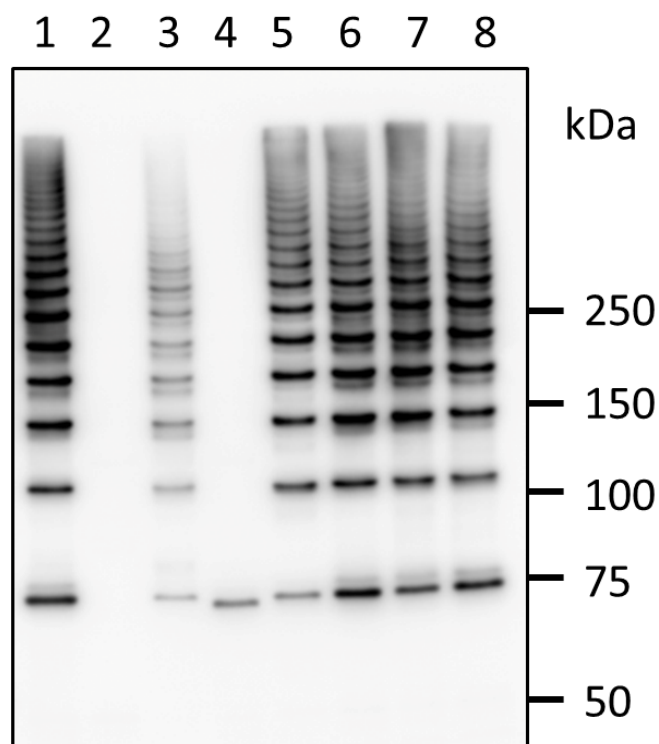
Cloned in *E. coli* using a plasmid
DNA sequencing

*Xba*I and *Not*I digestions were cloned into pTCOWragAP
and transformed into *P. gingivalis* Δ *mfa2* Δ *fimA*

Supplementary figure S17. Construction of +*mfa2C54A* and +*mfa2C285A* *P. gingivalis* strains. Small arrows indicate the annealing sites of the primers. X and N show *Xba*I and *Not*I restriction sites, respectively.



Supplementary figure S18. Confirmation of Mfa2 protein expression in mutant strains. Whole cell lysates were solubilized in SDS-buffer (+2BME), heated at 100°C for 5 min, separated by SDS-PAGE, blotted to a PVDF membrane and probed with a polyclonal antibody against Mfa2. Lanes: 1, JI-1; 2, $\Delta mfa1\Delta fim$; 3, +*mfa1*; 4, +*mfa1* ΔC ; 5, $\Delta mfa2\Delta fimA$ (negative control); 6, +*mfa2*; 7, +*mfa2C54A*; 8, +*mfa2C285A*.



Supplementary figure S19. Immunoblot analysis of Mfa1 polymerization under non-reducing conditions. Whole cell lysates solubilized in SDS buffer (-2BME) were heated to 80°C for 5 min, separated by SDS-PAGE, blotted to a PVDF membrane and probed with a polyclonal Mfa1 fimbriae antibody. Lanes: 1, JI-1; 2, $\Delta mfa1\Delta fim$; 3, +*mfa1*; 4, +*mfa1* ΔC ; 5, $\Delta mfa2\Delta fimA$; 6, +*mfa2*; 7, +*mfa2C54A*; 8, +*mfa2C285A*.

Supplementary Table S1: Pairwise structural comparison between Mfa1 and other fimbrial proteins building up the Mfa1 fimbria of *P. gingivalis* ATCC33277. The structures of Mfa1, Mfa2 and Mfa3 and Mfa4².

Structure/PDB aligned with Mfa1Δ9 (shaft)	z-score	lali	nres	r.m.s.d.	% identity
Shaft mMfa1	55.7	470	497	0.4	100
Mfa2, anchor	9.4	211	277	4.5	10
Mfa3, tip	16.3	255	295	3.7	11
Mfa4 (5dhm), tip	13.6	228	287	3.4	15

Supplementary Table S2: Pairwise structural comparison between Mfa2 and other fimbrial proteins building up the Mfa1 fimbria of *P. gingivalis* ATCC33277. Statistics were obtained by DALI³.

Structure/PDB aligned with Mfa2 (anchor)	z-score	lali	nres	r.m.s.d.	% identity
Mfa1Δ9 (shaft)	9.4	211	277	4.5	10
Mfa3, tip	11.8	212	277	4.9	11
Mfa4 (5dhm), tip	10.3	187	268	4.1	10

Supplementary Table S3: Pairwise structural comparison between Mfa3 and other fimbrial proteins building up the Mfa1 fimbria of *P. gingivalis* ATCC33277. Statistics were obtained by DALI³.

Structure/PDB aligned with Mfa3 (tip)	z-score	lali	nres	r.m.s.d.	% identity
Mfa1Δ9 (shaft)	16.3	255	295	3.7	11
Mfa2 (anchor)	11.8	212	277	4.9	11
Mfa4 (5dhm), tip	14.4	191	266	2.5	20

Supplementary Table S4. Structural comparison between Mfa1 and other fimbrial proteins⁴ as obtained by DALI³.

Structure/PDB aligned with Mfa1Δ9	organism	z-score	lali	nres	rmsd	% identity
BovFim3A shaft, 4jrf	<i>Bacteroides ovatus</i>	22.6	325	479	3.3	22
BdiFim3A shaft, 3liu	<i>Parabacteroides distasonis</i>	16.0	262	373	3.7	16
BegFim1A shaft, 4gpv	<i>Bacteroides eggerthii</i>	14.6	254	328	3.8	14
FimA, shaft 4q98	<i>Porphyromonas gingivalis</i> W83	14.7	256	357	3.4	18

Supplementary Table 5: Proteins structurally related to Mfa2 as obtained by DALI³.

Structure/PDB aligned with Mfa2	organism	z-score	lali	nres	rmsd	% identity
BthFim2B anchor, 3gf8	<i>Bacteroides thetaiotaomicron</i>	24.4	262	284	2.7	21
BthFim3B anchor, 4qdg	<i>Bacteroides thetaiotaomicron</i>	23.2	258	302	3.0	16
BovFim2B anchor, 3pay	<i>Bacteroides ovatus</i>	22.7	260	313	2.9	17

Supplementary Table 6: Proteins structurally related to Mfa3 as obtained by DALI³.

Structure/PDB aligned with Mfa3 (tip)	organism	z-score	lali	nres	rmsd	% identity
BdiFim1C tip, 4jg5	<i>Parabacteroides distasonis</i>	17.8	241	336	3.5	15
BovFim2C unknown,3up6	<i>Bacteroides ovatus</i>	17.6	248	324	3.0	12
BdiFim1A, shaft, 3liu	<i>Parabacteroides distasonis</i>	17.3	246	372	3.5	15
FimA, shaft, 4q98	<i>Porphyromonas gingivalis</i> W83	16.7	231	357	2.7	15

Supplementary Table S7. Primers used in this study.

Primer	Sequence (5'-3')	Description of underline	Primer number in supplemental Figs.
Construction of recombinant proteins			
mMfa1F	AAA <u>ACCATGG</u> CGGGTGACGGACAGGAT	<i>NcoI</i> site	
pMfa1F	AAA <u>ACCATGG</u> GTAAGAGGGCAATGGC	<i>NcoI</i> site	
Mfa1R	AAA <u>AGGTACCT</u> TAGAGATCAACCTCATA	<i>Acc65I</i> site	
Mfa1Δ8R	AAA <u>AGGTACCT</u> TACCAAGGCAAAACTGTAAC	<i>Acc65I</i> site	
Mfa2F	AAA <u>ACCATGG</u> ATAAGATGATTTATGAC	<i>NcoI</i> site	
Mfa2R	TTTT <u>GGTACCT</u> TAAAGTTCTATTTTCGTAAC	<i>Acc65I</i> site	
pMfa3F	AAA <u>ATCATG</u> AGAGGAGTTGATCCACAG	<i>PagI</i> site	
Mfa3R	AAAA <u>AGGTACC</u> CTATTTCTTGATAAAAAC	<i>Acc65I</i> site	
Construction of complemented strains of <i>mfa1</i>			
Mfa1WTxbF	TTAATATTAATCCTTTTAAACATTT <u>CTAGA</u> ATGAAG TTAAACAAAATGTTTTG	<i>XbaI</i> site	7
Mfa1WTnoR	CACCCTCAAAAAGAAAATTTTACAG <u>CGGCCG</u> CTT AGAGATCAACCTCATAGGAATG	<i>NotI</i> site	8
Mfa1ΔCnoR	CCCACCCTCAAAAAGAAAATTTTACAG <u>CGGCCG</u> C TTAATGAACCTTCCAAGGCAAAACTGTAACC	<i>NotI</i> site	9
Mfa1R236AR	<u>ACCATCGCACGTGCTACAGACGCCTCCACCGTAA</u> <u>CCTTGGCCT</u>	Ala mutation and overlapping region of Mfa1R236AR	10
Mfa1R236AF	<u>AGGCCAAGGTTACGGTGGAGGCGTCTGTAGCACG</u> <u>TGCGATGGT</u>	Ala mutation and overlapping region of Mfa1R236AF	11
Mfa1W554AR	CACCCTCAAAAAGAAAATTTTACAG <u>CGGCCG</u> CTT AGAGATCAACCTCATAGGAATGAACCTT <u>CGCAGG</u> CAAAACTGTAACCTCAA	<i>NotI</i> site and Ala mutation	12
Construction of <i>Δmfa2ΔfimA</i>			
AGU01	ATGGAGAAAAAATCACTGGA		1
AGU02	TTACGCCCCGCCCTGCCACTC		2
Mfa2upF	GTCAGGTGCTAATGCTGCCTCG		3
Mfa2upR	<u>CCAGTGATTTTTTCTCCAT</u> TGTTTTAAAAAATATA GAGGG	Overlapping region of 5' end of <i>cat</i>	4
Mfa2downF	<u>GCAGGGCGGGCGTA</u> AGAGAAAAAAGACCGGTT CTTC	Overlapping region of 3' end of <i>cat</i>	5
Mfa2downR	GGCCATGGCAGTCTGCATATTG		6

Construction of complemented strains of *mfa2*

Mfa2WTxbF	TTCTCCCACCCTCTATATTTTTTCTAGAAATGAACA AACGGAAGCATATGG	<i>Xba</i> I site	13
Mfa2WTnoR	GAAAGCGAGTGAAGAACCGGTCTGCGGCCGCTTA AAGTTCTATTTTCGTA ACTATG	<i>Not</i> I site	14
Mfa2C54AR	<u>ATTTTCGGCAGCCTCAGTCTGAGAATAGAAGTTG</u>	Ala mutation and overlapping region of Mfa2C54AF	15
Mfa2C54AF	<u>GACTGAGGCTGCCGAAAATCCTTCTTATCCTGCG</u>	Ala mutation and overlapping region of Mfa2C54AR	16
Mfa2C285AR	<u>AAGTCATTGATAGCGCGCAAGTTGATATTTTGAGC</u> ATAC	Ala mutation and overlapping region of Mfa2C285AF	17
Mfa2C285AF	<u>CAACTTGCGCGCTATCAATGACTTCGATATAAGGT</u> TGG	Ala mutation and overlapping region of Mfa2C285AR	18

Construction of +*mfa3*ΔC

cMfa3F	ATACACTTCTAGAAATGATGCAGCTTAAAAAGAGAT	<i>Xba</i> I site	19
Mfa3ΔCnoR	CACTCGTTAACAACGAGGCATATAACAAATACTTT TTCATGCGGCCGCCTAGACCCCTGTTGCATTCAGA TCCGGGTTATAATAAACCTCC	<i>Not</i> I site	20

Supplementary Table S8. *Porphyromonas gingivalis* strains used in this study.

Strain	Genotype and Relevant Description ¹	Reference
Jl-1	<i>fimA</i> deletion mutant from ATCC 33277, Cp ^r	5
KDP98	<i>fimA</i> deletion mutant from ATCC 33277, Em ^r	6
$\Delta mfa1\Delta fim$	<i>mfa1</i> and <i>fimA-E</i> deletion mutant, Cp ^r Em ^r	7
$\Delta mfa2\Delta fimA$	<i>mfa2</i> and <i>fimA</i> deletion mutant, Cp ^r Em ^r	This study
$\Delta mfa3\Delta fimA$	<i>mfa3</i> and <i>fimA</i> deletion mutant, Cp ^r Em ^r	8
+ <i>mfa1</i>	$\Delta mfa1\Delta fim$ complemented with intact <i>mfa1</i> through pTCOWragAP:: <i>mfa1</i> , Cp ^r Em ^r Tc ^r . A control strain for + <i>mfa1</i> ΔC , + <i>mfa1R236A</i> and + <i>mfa1W554A</i>	This study
+ <i>mfa1</i> ΔC	$\Delta mfa1\Delta fim$ complemented with <i>mfa1</i> deleting C-terminal region (SYEVDL), Cp ^r Em ^r Tc ^r	This study
+ <i>mfa1R236A</i>	$\Delta mfa1\Delta fim$ complemented with <i>mfa1</i> point mutation R236A, Cp ^r Em ^r Tc ^r	This study
+ <i>mfa1W554A</i>	$\Delta mfa1\Delta fim$ complemented with <i>mfa1</i> point mutation W554A, Cp ^r Em ^r Tc ^r	This study
+ <i>mfa2</i>	$\Delta mfa2\Delta fimA$ complemented with intact <i>mfa2</i> through pTCOWragAP:: <i>mfa2</i> , Cp ^r Em ^r Tc ^r . A control strain for + <i>mfa2</i> C54A and C285A.	This study
+ <i>mfa2</i> C54A	$\Delta mfa2\Delta fimA$ complemented with <i>mfa2</i> point mutation C54A, Cp ^r Em ^r Tc ^r	This study
+ <i>mfa2</i> C285A	$\Delta mfa2\Delta fimA$ complemented with <i>mfa2</i> point mutation C285A, Cp ^r Em ^r Tc ^r	This study
+ <i>mfa3</i>	$\Delta mfa3\Delta fimA$ complemented with intact <i>mfa3</i> through pTCOWragAP:: <i>mfa3</i> , Cp ^r Em ^r Tc ^r . A control strain for + <i>mfa3</i> ΔC .	8
+ <i>mfa3</i> ΔC	$\Delta mfa3\Delta fimA$ complemented with <i>mfa3</i> deleting C-terminal region (VPDKVFIKK), Cp ^r Em ^r Tc ^r	This study

¹ Cp^r, chloramphenicol resistance; Em^r, erythromycin resistance; Tc^r, tetracycline resistance.

² ATCC, American Type Culture Collection.

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