

Supporting Information

**Novel insights into the mechanism of well-ordered
assembly of bacterial flagellar proteins in *Salmonella***

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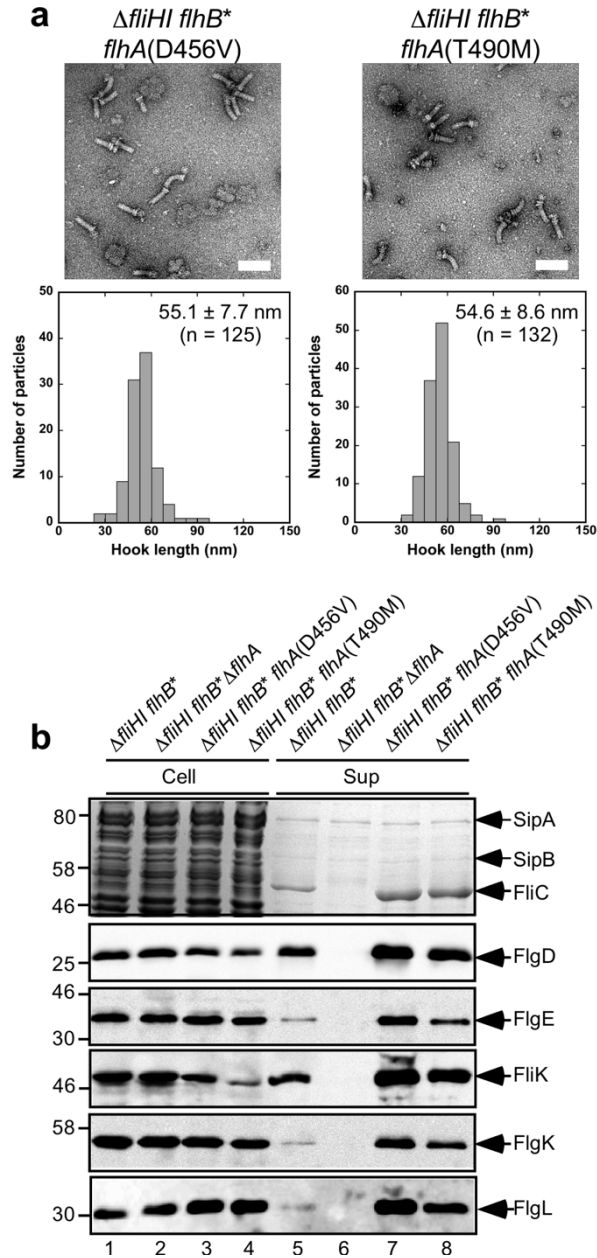


Fig. S1. Effect of the FlhA(D456A) and FlhA(T490M) mutations on flagellar protein export and assembly. (a) Electron micrographs of HBBs and histograms of hook length distribution of MMHI0117-2 [$\Delta fliHI$ *flhB*^{*} *flhA*(D456V)] and MMHI0117-3 [$\Delta fliHI$ *flhB*^{*} *flhA*(T490M)] (b) Flagellar protein export assays. Whole cell proteins (Cell) and culture supernatant fractions (Sup) were prepared from MMHI0117 (indicated as $\Delta fliHI$ *flhB*^{*}), NH004 (indicated as $\Delta fliHI$ *flhB*^{*} $\Delta flhA$), MMHI0117-2 and MMHI0117-3 and were normalized to their optical density of OD₆₀₀ to give a constant number of cells. 8 μ l of each sample was subjected to SDS-PAGE and analyzed by CBB staining (1st row) and immunoblotting, using polyclonal anti-FlgD (2nd row), anti-FlgE (3rd row), anti-FliK (4th row), anti-FlgK (5th row) or anti-FlgL (6th row) antibody. The positions of molecular mass markers (kDa) are indicated on the left. SipA and SipB, which are secreted by the SPI-1 virulence-associated type III secretion apparatus, serve as loading controls.

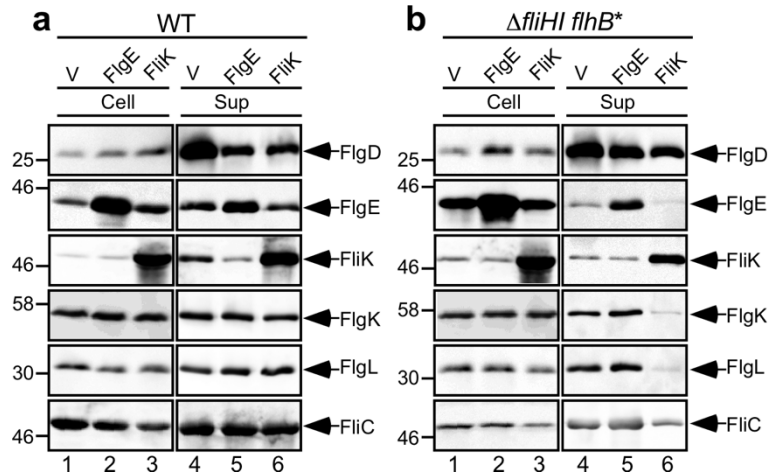


Fig. S2. Effect of over-expression of FlgE and FliK on flagellar protein export in the presence and absence of FliH and FliI. Whole cell proteins (Cell) and culture supernatant fractions (Sup) were prepared from (a) SJW1103 (indicated as WT) carrying pTrc99A (V), pNM001 (FlgE) or pMMIK100 (FliK) and (b) MMHI0117 (indicated as $\Delta fliH fliB^*$) harbouring pTrc99A (V), pNM001 (FlgE) or pMMIK100 (FliK) and were normalized to their optical density of OD₆₀₀ to give a constant number of cells. 8 μ l of each sample was subjected to SDS-PAGE and analyzed by immunoblotting, using polyclonal anti-FlgD (1st row), anti-FlgE (2nd row), anti-FliK (3rd row), anti-FlgK (4th row), anti-FlgL (5th row) or anti-FliC (6th row) antibody. The positions of molecular mass markers (kDa) are indicated on the left.

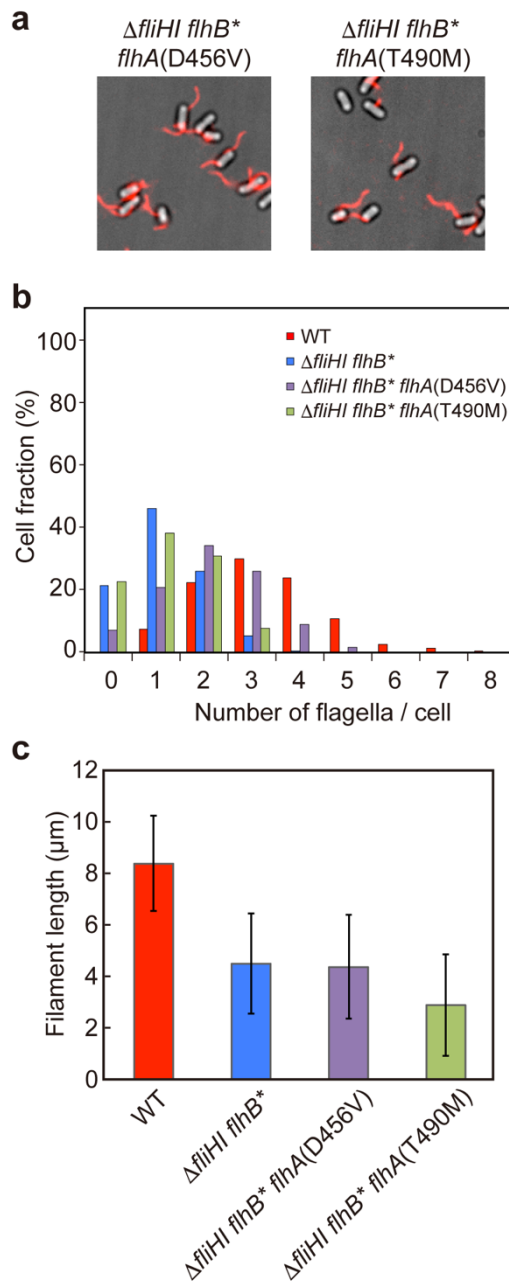


Fig. S3. Effect of the FlhA(D456V) and FlhA(T490M) mutations on filament assembly in the $\Delta fliH$ -*fliB*(P28T) mutant background. (a) Fluorescent images of MMHI0117-2 [$\Delta fliH$ *fliB*^{*} *fliA*(D456V)] and MMHI0117-3 [$\Delta fliH$ *fliB*^{*} *fliA*(T490M)]. Flagellar filaments were labeled with Alexa Fluor 594. The fluorescence images of the filaments labeled with Alexa Fluor 594 (red) were merged with the bright field images of the cell bodies. (b) Distribution of the number of the flagellar filaments. More than 200 cells for each strain were counted. (c) Measurements of the length of the flagellar filaments. Filament length is the average of more than 100 cells for each strain, and vertical lines are standard deviations.

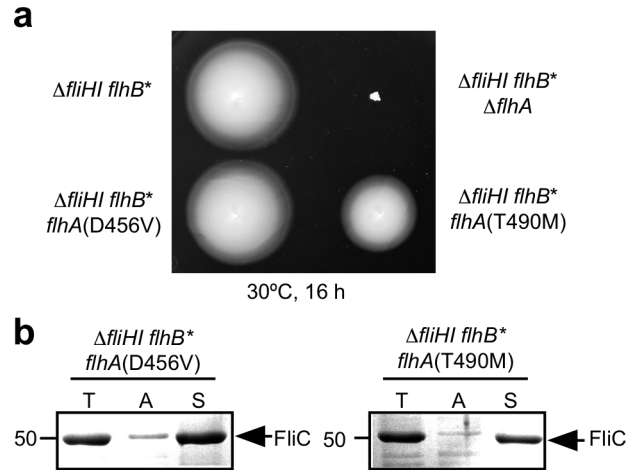


Fig. S4. Effect of the FlhA(D456V) and FlhA(T490M) mutations on motility of the $\Delta fliH$ - $fliI$ $flhB$ (P28T) mutant. (a) Motility of MMHI0117 (indicated as $\Delta fliHI flhB^*$), NH004 (indicated as $\Delta fliHI flhB^* \Delta flhA$), MMHI0117-2 [$\Delta fliHI flhB^* flhA(D456V)$] and MMHI0117-3 [$\Delta fliHI flhB^* flhA(T490M)$] in soft agar. Plates were incubated at 30°C for 16 hours. (b) Measurements of FliC leakage during flagellar filament assembly. CBB-staining gels of total extracellular FliC (indicated as T), polymerized FliC (indicated as A), and secreted FliC (indicated as S) of MMHI0117-2 and MMHI0117-3. A position of the 50 kDa molecular mass marker is indicated on the left. The position of 50 kDa molecular mass marker is indicated on the left.

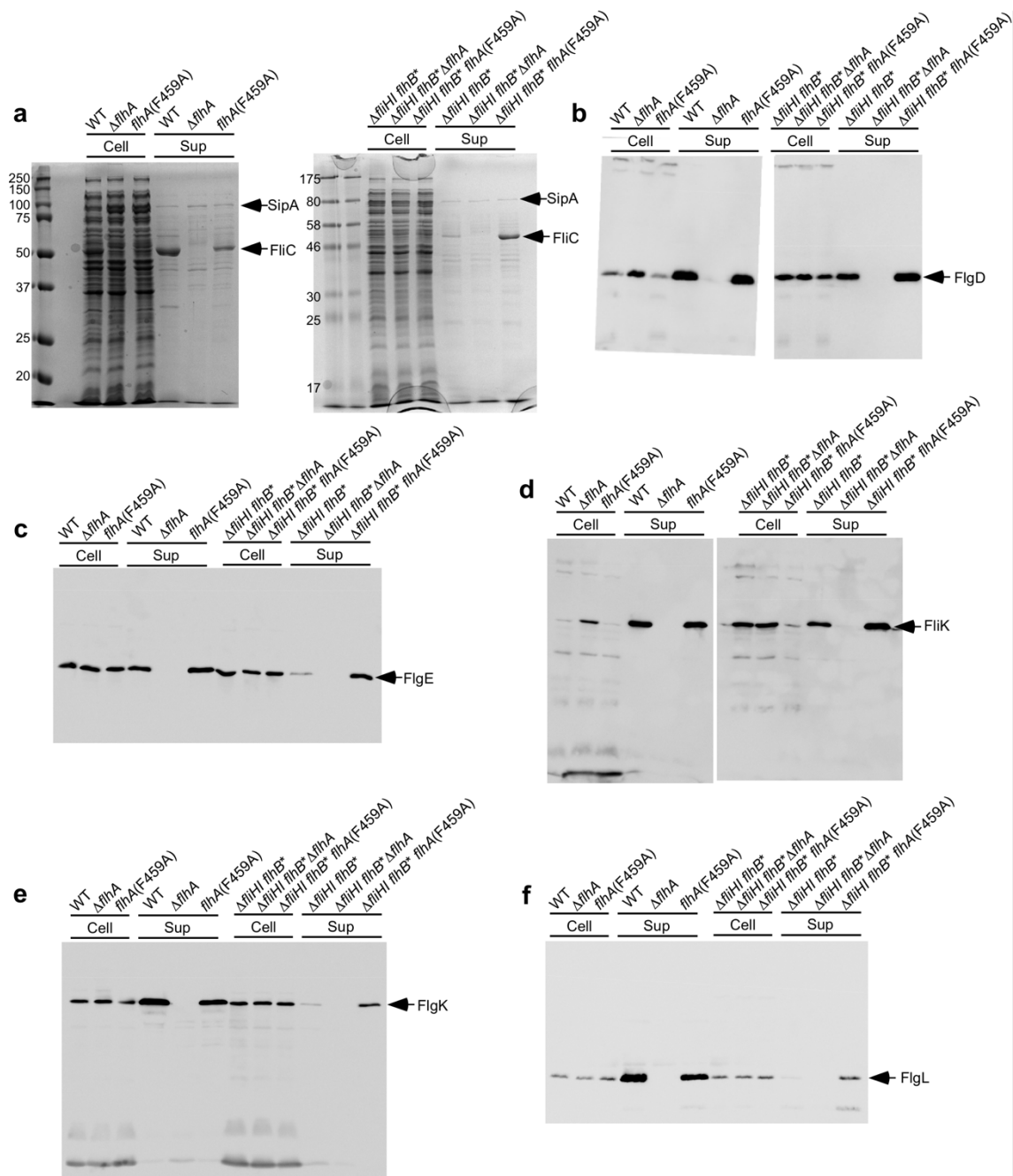


Fig. S5. Original image data of Figure 3b. Whole cell proteins (Cell) and culture supernatant fractions (Sup) were prepared from SJW1103 (indicated as WT), NH001 (indicated as $\Delta flhA$), MMA459 [indicated as $flhA(F459A)$], MMHI0117 (indicated as $\Delta fliHI flhB^*$), NH004 (indicated as $\Delta fliHI flhB^* \Delta flhA$) and MMHI0117-1 [$\Delta fliHI flhB^* flhA(F459A)$]. 8 μ l of each protein sample, which was normalized to an optical density of OD₆₀₀, was subjected to SDS-PAGE and was analyzed by (a) CBB staining and immunoblotting with polyclonal (b) anti-FlgD, (c) anti-FlgE, (d) anti-FliK, (e) anti-FlgK or (f) anti-FlgL antibody. All image data were captured by a Luminoimage analyser LAS3000. The positions of molecular mass markers (kDa) are indicated on the left.

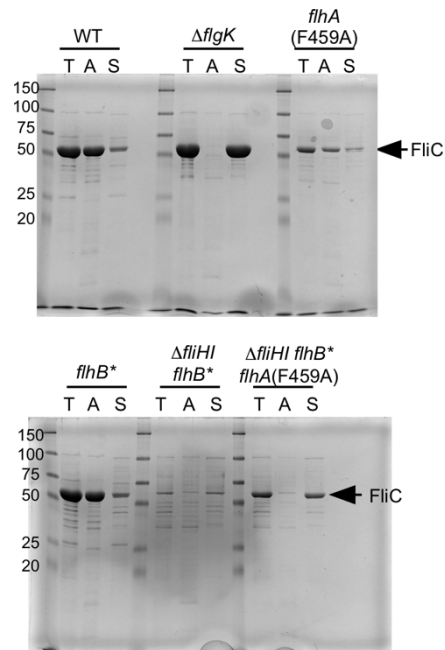


Fig. S6. Original image data of panels a–f in Figure 6. CBB-staining SDS gels of total extracellular FliC (indicated as T), polymerized FliC (indicated as A), and FliC leaked into the culture media (indicated as S) of SJW1103 (indicated as WT), MM1103gK (indicated as $\Delta flgK$), MMA459 [indicated as *flhA*(F459A)], MMB017 (*flhB*^{*}), MMHI0117 (indicated as $\Delta fliHI$ *flhB*^{*}) and MMHI0117-1 [$\Delta fliHI$ *flhB*^{*} *flhA*(F459A)]. Gel images were captured by a Luminoimage analyser LAS3000.

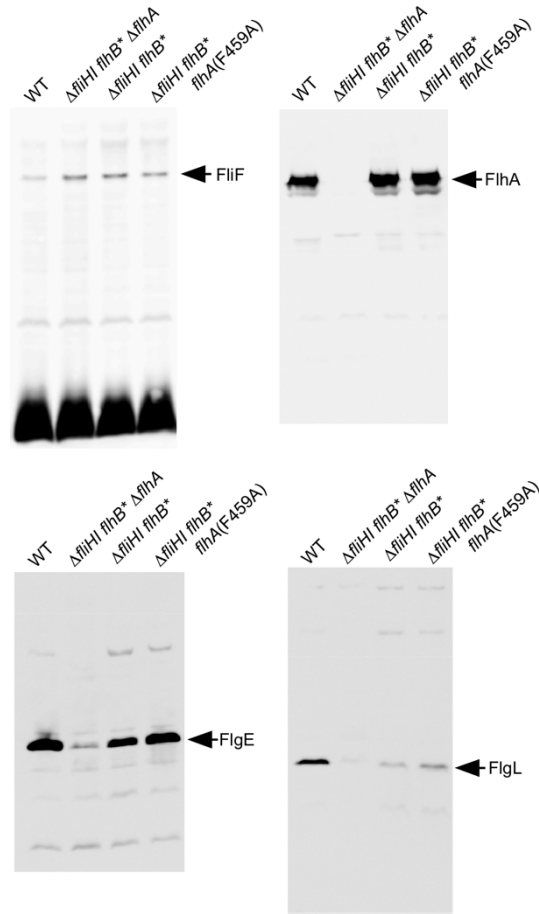


Fig. S7. Original image data of Figure 6g. The membrane fractions of SJW1103, NH004 (indicated as $\Delta fliHI flhB^* \Delta flhA$), MMHI0117, or MMHI0117-1 were prepared after sonication and ultracentrifugation. Then, the membrane fractions were subjected to SDS-PAGE, and analyzed by immunoblotting with polyclonal anti-FliF, anti-FlhA, or anti-FlgL antibody. Chemiluminescence signals were detected by a Luminoimage analyzer LAS-3000.

Table S1. Plasmids and *Salmonella* strains used in this study

Plasmids/ Strains	Relevant characteristics	Source or reference
Plasmids		
pTrc99AFF4	Modified pTrc vector	1
pMMIK100	pTrc99AFF4/ FliK	2
pNM001	pTrc99AFF4/ FlgE	3
<i>Salmonella</i>		
SJW1103	Wild type for motility and chemotaxis	4
MM1103gK	<i>flgK::Tn10</i>	5
MMHI0117	Δ <i>fliH-flil flhB</i> (P28T)	6
MMB017	<i>flhB</i> (P28T)	6
MMA459	<i>flhA</i> (F459A)	This study
MMHI0117-1	Δ <i>fliH-flil flhB</i> (P28T) <i>flhA</i> (F459A)	This study
MMHI0117-2	Δ <i>fliH-flil flhB</i> (P28T) <i>flhA</i> (D456V)	This study
MMHI0117-3	Δ <i>fliH-flil flhB</i> (P28T) <i>flhA</i> (T490M)	This study
NH001	Δ <i>flhA</i>	7
NH004	Δ <i>fliH-flil flhB</i> (P28T) Δ <i>flhA</i>	7

References

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