1 Scientific Reports

- 2 **Supplemental Materials**
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4 Antibody-mediated enhancement aggravates chikungunya virus 5 infection and disease severity

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Figure S1: Enhanced CHIKV infection in human whole blood. Human whole blood was infected with Zs-Green tagged CHIKV (2×10^7 PFU) either in the presence (enhanced) or absence (non-enhanced) of diluted CHIKV patient plasma containing total IgG at a concentration of 1.8 ± 1.45µg/ml. (a) Level of infection and (b) viral RNA load were determined 24 hours post-infection (hpi). Level of CHIKV infection was determined by the amount of Zs-Green positive cells and is expressed as fold enhancement relative to the non-enhanced infection controls. Data shown are mean ± SD from 5 independent donors by Mann-Whitney *U* test (**P* = 0.0115).

Fig S1_Lum et al., 2017



Figure S2: CHIKV infection enhancement in primary human monocytes. Isolated primary human monocytes (2×10^6 cells) were infected with Zs-Green tagged CHIKV (moi 10) either in the presence (enhanced) or absence (non-enhanced) of diluted CHIKV patient plasma containing total IgG at a concentration of $3.6 \pm 2.9 \mu g/ml$. (**a**) Level of infection and (**b**) viral RNA load were determined at 24 and 48 hpi. Level of CHIKV infection was determined by the amount of Zs-Green positive cells and is expressed as fold enhancement relative to the non-enhanced infection controls. Data shown are mean \pm SD from 4 independent experiments.



Figure S3: Enhanced CHIKV infection in human monocytes-derived macrophages (MDMs) is mediated by FcyRII. MDMs (2×10^6 cells) were treated with 10µg of FcyRII blocking antibody prior to infection with Zs-Green tagged CHIKV (moi 10) in the presence of CHIKV patient plasma containing total IgG at a concentration of $1.8 \pm 1.45\mu$ g/ml. (a) FACs plot showing the decrease in the amount of CHIKV antigen detected from one representative experiment at 24 hours post-infection (hpi). (b) Percentage of infection is expressed as level of CHIKV antigen detected relative to the non-treated-enhanced infection controls. Data are presented as mean \pm SD from 4 independent experiments by Mann-Whitney *U* test (**P* = 0.0143).

Fig S3_Lum et al., 2017



Figure S4: Immunephenotyping of CHIKV infected WT mice. Three-weeks old, C57BL/6 WT female mice were infected via footpad inoculation with 10⁶ PFU of CHIKV. Immediately, these mice were administered intraperitoneally with ~2µg/ml sera from CHIKV-infected mice (enhanced) or PBS (non-enhanced). Gating stratefy of immune-phenotyping illustrated using samples extracted from the CHIKV-infected footpad is shown here. Strategy described is used for all analyzes.

Fig S4_Lum *et al.*, 2017



Figure S5: Gating strategy in determining CHIKV infection in infected primary human cells. Cells were first gated based on their forward and side scatter. Subsequently doublets were excluded from the analysis. Isolated primary cells were then gated based on their specific markers before percentage of CHIKV positive cells were determined from these cells. Illustration is shown using isolated primary human (a) monocytes and (b) B cells from one experiment. Strategy used in (a) was also applied for human monocytes-derived macrophages. Strategy described was used for all replicates of related experiments.

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Genes	Primers sequences ^a	
IL-6	Forward	TTCCATCCAGTTGCCTTCTTG
	Reverse	TTGGGAGTGGTATCCTCTGTGA
iNOS	Forward	CTGCTCACTCAGCCAGGCCCTCACCTACTT
	Reverse	TGATCCCGGGAGGAGCTGATGGAGTAGAAC
SOCS3	Forward	ATGGTCACCCACAGCAAGTTT
	Reverse	TCCAGTAGAATCCGCTCTCCT
IFNα	Forward	GTGACAGGGAACTTAGGGAGC
	Reverse	AGAGTGGGGTGTCTGCCTTA
IFNβ	Forward	AACTTTGACATCCCTGAGGAGATTAAGCAG
	Reverse	GACTATGGTCCAGGCACAGTGACTGTACTC
IFNγ	Forward	ATCTGGAGGAACTGGCAAAA
	Reverse	TGAGCTCATTGAATGCTTGG
IRF3	Forward	GGGGAGCCTCTTGACTGAAAACCGTGGA
	Reverse	TAACCACCAGCCTAGACGCAGTCGACAGCA
IRF9	Forward	GCCGAGTGGTGGGTAAGAC
	Reverse	GCAAAGGCGCTGAACAAAGAG
STAT1	Forward	GCTGGGCGTCTATCCTGTGGT
	Reverse	GCTCAGCTGGTCTGCGTTCA
STAT2	Forward	TCCTGCCAATGGACGTTCG
	Reverse	GTCCCACTGGTTCAGTTGGT
STAT3	Forward	CAATACCATTGACCTGCCGAT
	Reverse	GAGCGACTCAAACTGCCCT
STAT5	Forward	CGATGCCCTTCACCAGATG
	Reverse	AGCTGGGTGGCCTTAATGTTC
STAT6	Forward	CGCTGATAAGCCGTCTGGAT
	Reverse	TGTGTCTTCAGAACCTGCGG
Viperin	Forward	CTTCAACGTGGACGAAGACA
	Reverse	GACGCTCCAAGAATGTTTCA
ISG15	Forward	TGGGACCTAAAGGTGAAGATGCTG
	Reverse	TCAGGCGCAAATGCTTGATCAC
DEF14	Forward	TCCAGGGGACGCATTCCTA
	Reverse	ACCGCTATTAGAACATCGACCTA
mIL-10	Forward	TGGCCTTGTAGACACCTTGG
	Reverse	AGCTGAAGACCCTCAGGATG
GAPDH	Forward	TCGTCCCGTAGACAAAATGG
	Reverse	TTGAGGTCAATGAAGGGGTC

 Table S1. Sequences of primers used for gene expression studies

^a Primers sequences shown are of mouse origin