

Figure S1. Related to Figure 1. Localization of aP2-Cre expressing cells in aP2-Cre;R26-mT/mG animals.

(A) Schematic of the *R26-mT/mG* allele and *aP2-Cre* mediated recombination.

(B) Representative whole mount images of BAT (intrascapular brown adipose) and WAT (inguinal white adipose) from *R26-mT/mG* mice (n = 3). Scale bars, 3mm.

(C) Representative whole mount and fixed tissue sections of lung and kidney from *aP2-Cre;mT/mG* mice (n = 3). Scale bars, 3mm (whole mount), 50 μ m (tissue sections). Arrowheads denotes perineal adipose.

(D) Representative fixed tissue sections of testis (a and b) and ovaries (c and d) from *aP2-Cre;mT/mG* mice and percent of global Cre recombination observed in *aP2-Cre* mice (male n = 36; female n = 698). Scale bars, 500 μ m (a and c), 100 μ m (b and d). Arrowhead denotes adipose adjacent to ovary.

(E) Kaplan-Meier Survival curves showing tumor free survival of *Adipoq-Cre;Smo^{M2/+}* mice (n = 6) in comparison to AS mice (n = 49) (Hatley et al., 2012).

(F) Real-time PCR of SmoM2-YFP expression in BAT and WAT from *Adipoq-Cre;Smo^{M2/+}* and *Smo^{M2/+}* mice. Data shown are normalized to *Actb* expression and expressed relative to Cre negative controls (n = 3, mean \pm SEM).

(G) Kaplan-Meier Survival curves showing tumor free survival of *Ucp1-Cre;Smo^{M2/+}* (n = 23) mice in comparison to AS mice (n = 49) (Hatley et al., 2012).

(H) Real-time PCR of SmoM2-YFP expression in BAT and WAT from *Ucp1-Cre;Smo^{M2/+}* and *Smo^{M2/+}* mice. Data shown are normalized to *Actb* expression and expressed relative to Cre negative controls (n = 3, mean \pm SEM).

*** p < 0.001

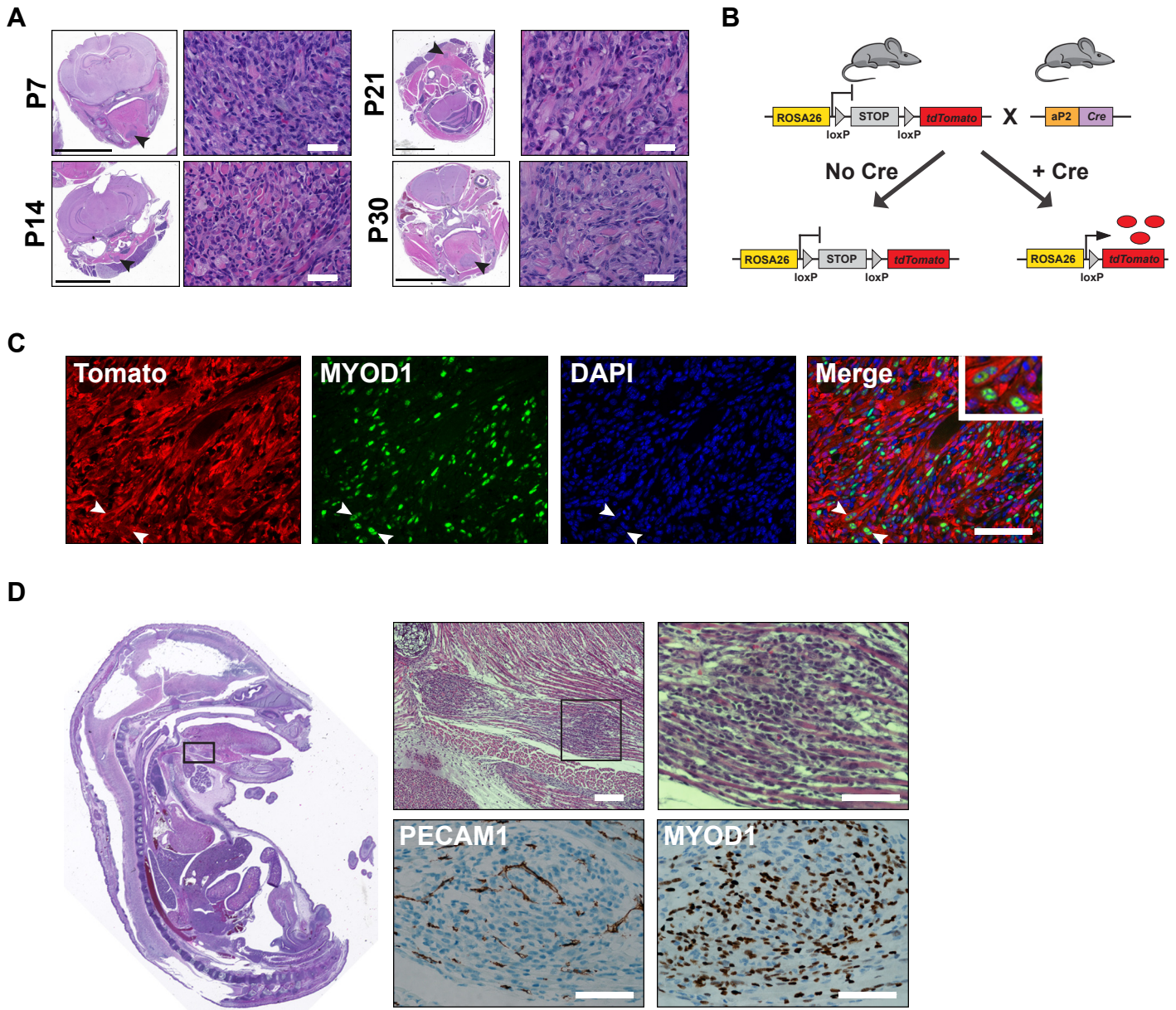


Figure S2. Related to Figure 2. Characterization of tumors from *aP2-Cre; Smo^{M2/+}; R26-tdTom* animals.

(A) Gross cross sections and histology (H & E) of head and neck tumors from P7, P14, P21 and P30 AST mice. Arrows denote tumor localization. Scale bars, 5 mm (left), 25 μ m (right).

(B) Schematic of AT breeding.

(C) Representative immunofluorescence of AST tumors showing MYOD1 (green), Tomato (red) and DAPI (blue)(n = 3). Arrows denotes region of high magnification inset shown on upper right. Scale bars, 50 μ m.

(D) Representative sagittal sections of E16.5 *Fabp4-Cre; Smo^{M2/+}* embryos (n = 3). Insets of boxed regions are shown at low magnification (scale bar, 100 μ m) and high magnification (scale bar, 50 μ m).

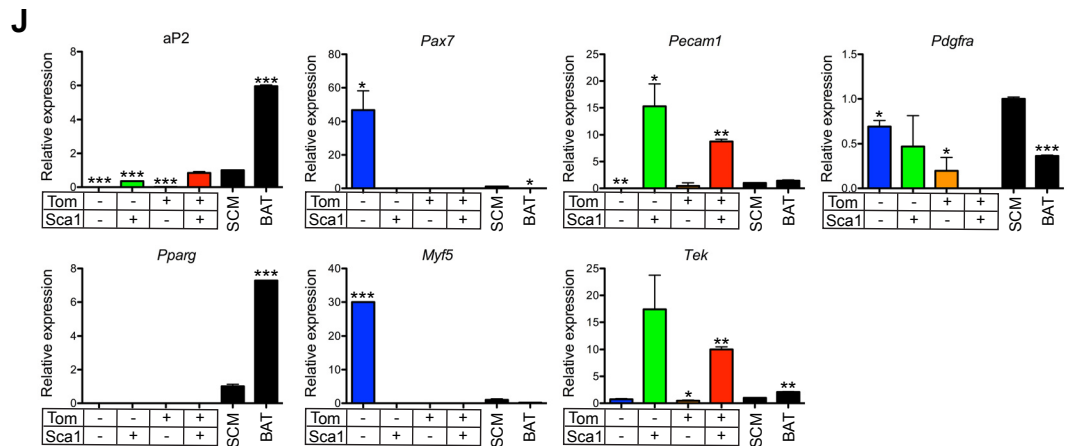
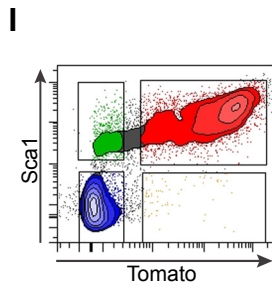
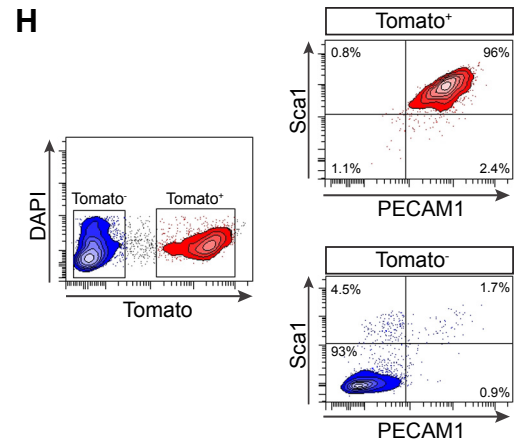
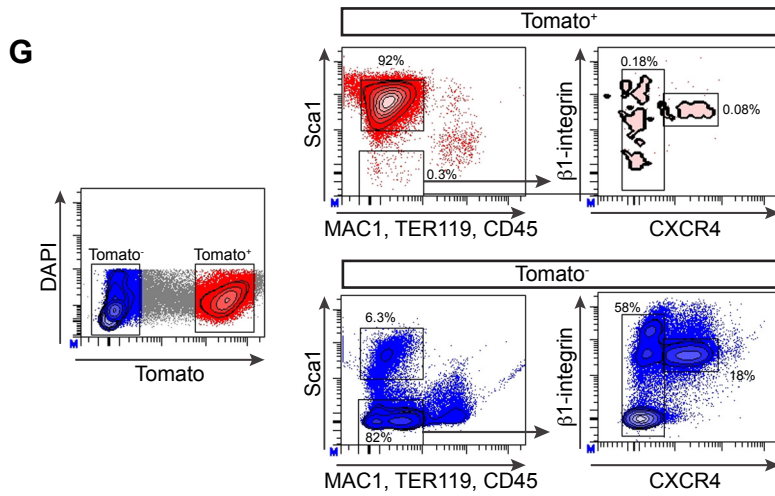
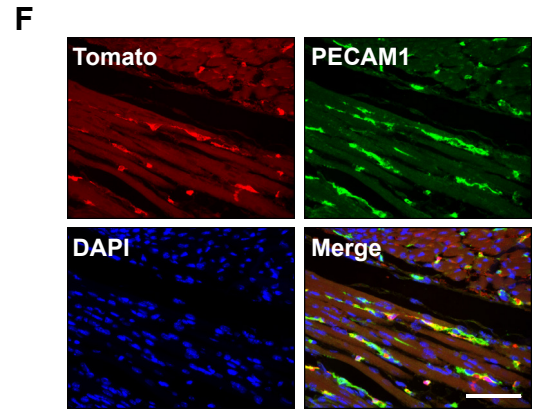
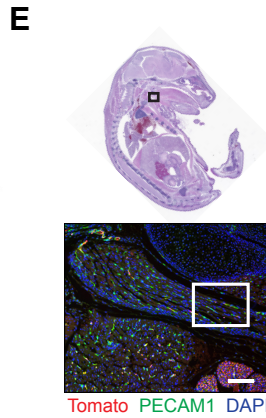
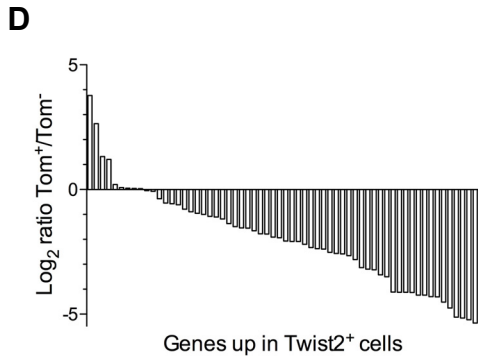
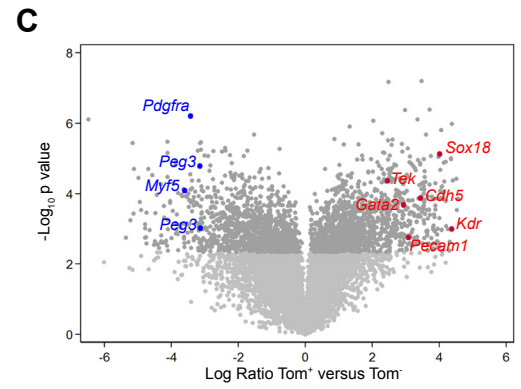
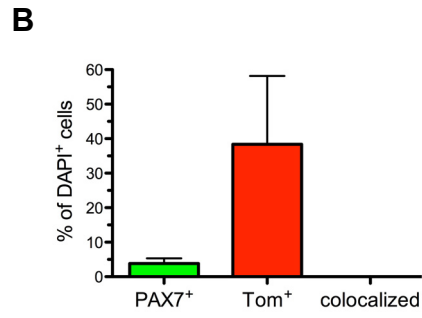
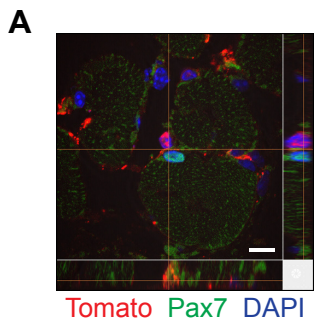


Figure S3. Related to Figure 3. aP2-Cre labels endothelial cells within the muscle interstitium.

(A) Confocal microscopy illustrating 3D cross sections from SCM of AT mice. PAX7 (green), TOMATO (red) and DAPI (blue) shown. Scale bar, 10 μ m.

(B) Quantitation of Tomato and PAX7 co-localization from sections in Figure 3A. 909 cells were counted from 22 fields from SCM sections of 5 AT mice. 34 (3.8%) of the cell nuclei were PAX7⁺. There was no co-localization with any of the 335 tomato positive cells and PAX7. Bars are mean \pm SEM.

(C) Volcano plot of the Log of the p value versus the log fold change in expression between the tomato positive (Tom⁺) and negative (Tom⁻) cells isolated from AT SCM by FACS. Solid grey dots indicate FDR > 0.05.

(D) Log₂ ratio of expression in Tom⁺ versus Tom⁻ cells isolated from SCM of AT mice for genes enriched in Twist2 expressing muscle interstitial cells (Liu et al., 2017) illustrating only 4 of 63 genes similarly enriched in Tom⁺ cells from AT SCM.

(E) Gross histology and immunofluorescence of E18.5 AT sagittal section showing PECAM1 (green), TOMATO (red) and DAPI (blue). Scale bars, 100 μ m.

(F) High magnification split channel images of boxed inset from (D). Scale bars, 50 μ m.

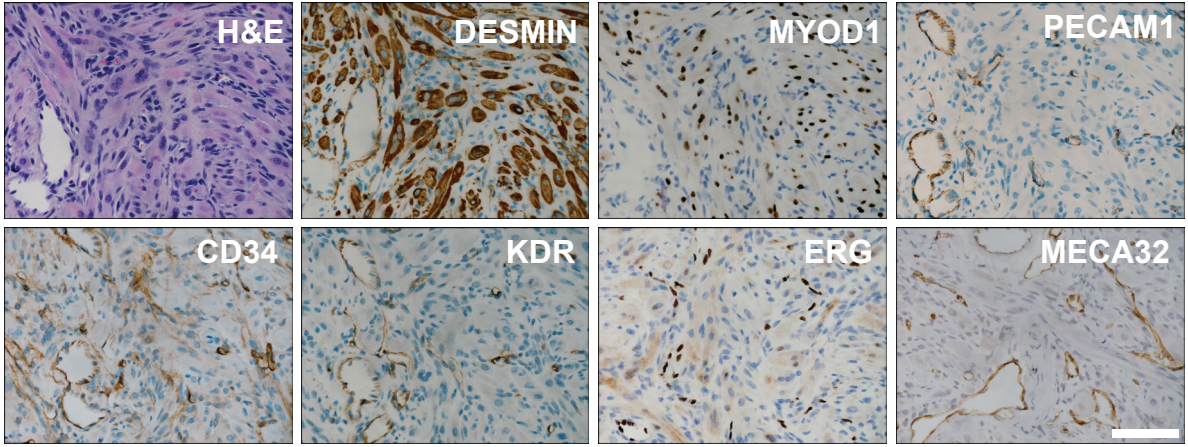
(G-H) Flow cytometric analysis of Tom⁺ and Tom⁻ cells from AT SCM for MAC1, Ter119, CD45, Sca1, β 1integrin and CXCR4 (G) and Sca1 and PECAM1 (H).

(I) FACS plot showing isolation of Tom⁻;Sca1⁻, Tom⁻;Sca1⁺, Tom⁺;Sca1⁻, and Tom⁺;Sca1⁺ cells from AT SCM.

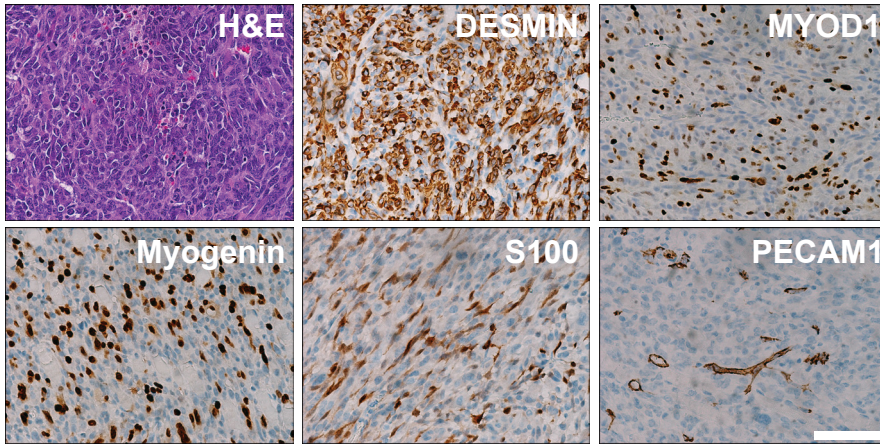
(J) Real-time PCR of isolated populations from (I) compared to mature SCM and BAT from *Smo*^{M2/M2} mice. Data shown are normalized to *Actb* expression and expressed relative to mature SCM. Bars are mean \pm SEM.

*p < 0.05, ** p < 0.01, *** p < 0.001

A



B



C

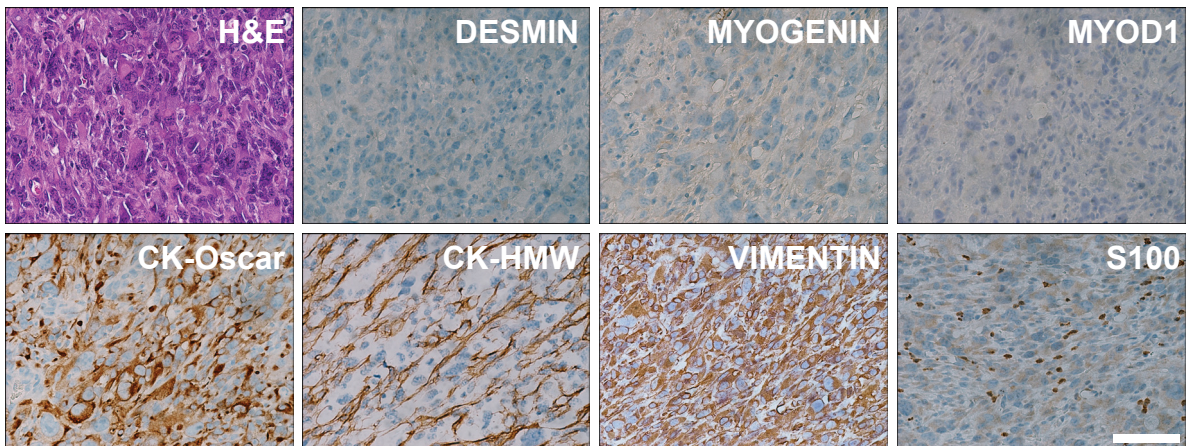


Figure S4: Related to Figure 4. Oncogenic KRAS does not drive FN-RMS in *aP2-Cre* expressing cells

(A) Representative H&E staining and IHC for RMS markers (DESMIN and MYOD1) and angiosarcoma markers (PECAM1, CD34, KDR, ERG, and MECA32) in *aP2-Cre;Smo^{M2/+};R26-tdTom* FN-RMS tumors (n = 3). Scale bars, 50 μ m.

(B) *aP2-Cre;LSL-Kras^{G12D};Cdkn2a^{Flox/Flox}* malignant triton tumor showing H&E staining and IHC (DESMIN, MYOD1, MYOGENIN, S100 and CD31)(n = 1). Scale bars, 50 μ m.

(C) *aP2-Cre;LSL-Kras^{G12D};Cdkn2a^{Flox/Flox}* pleomorphic spindle cell carcinoma showing H&E, and IHC (DESMIN, MYOGENIN, MYOD1, CK-OSCAR, CK-HMV, VIMENTIN and S100)(n = 1). Scale bars, 50 μ m.

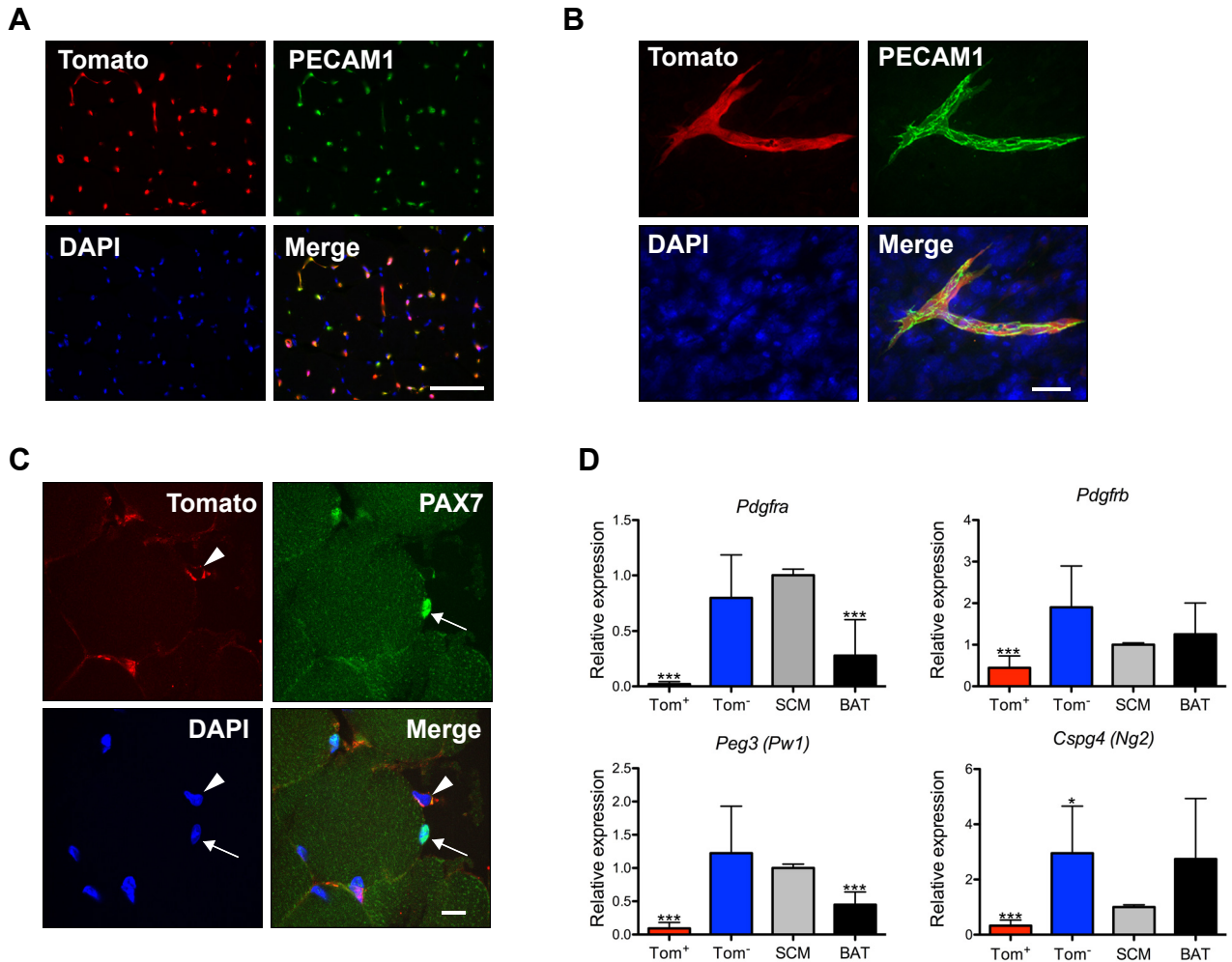


Figure S5. Related to Figure 5. Further characterization of aP2-Cre expressing cells from aP2-Cre;Smo^{M2/+};R26-tdTom SCM.

(A) Immunofluorescence of AST SCM cross sections for PECAM1 (green), TOMATO (red) and DAPI (blue). Scale bars, 50 μ m.

(B) Immunocytochemistry of aP2-Cre expressing cells. Mononuclear cells were isolated from AST SCM, grown until confluence and stained for PECAM1 (green) TOMATO (red) and DAPI (blue). Scale bars, 50 μ m.

(C) Immunofluorescence of AST SCM cross sections for PAX7 (green), TOMATO (red) and DAPI (blue). Scale bars, 10 μ m. Arrowheads denote Tom⁺ cells and arrows denote PAX7⁺ cells.

(D) Real-time PCR of isolated Tom⁺ and Tom⁻ cells from AST SCM in comparison to SmoM2 SCM and BAT. Data shown are normalized to *Actb* expression and expressed relative to mature SCM (n = 3, mean \pm SEM)

*p < 0.05, ** p < 0.01, *** p < 0.001

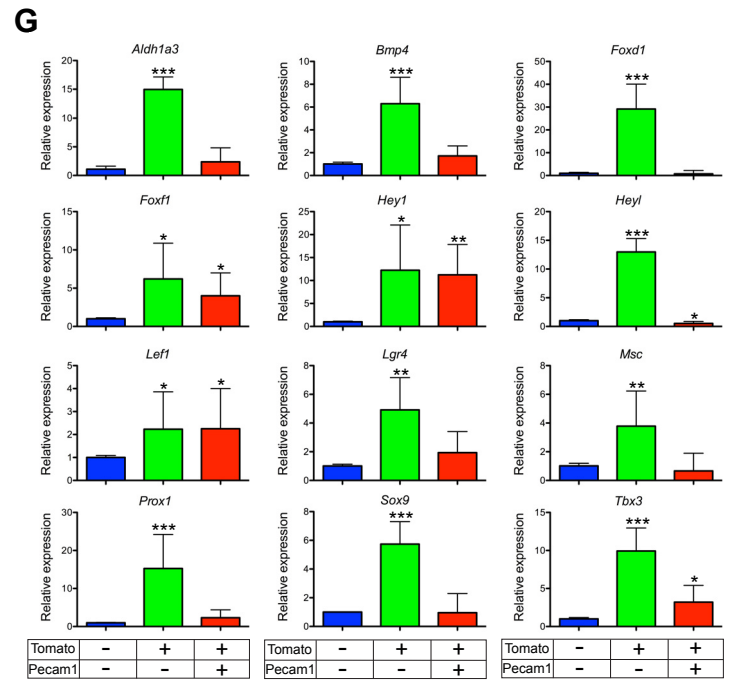
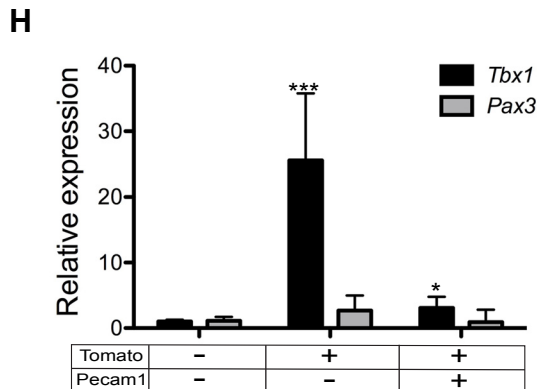
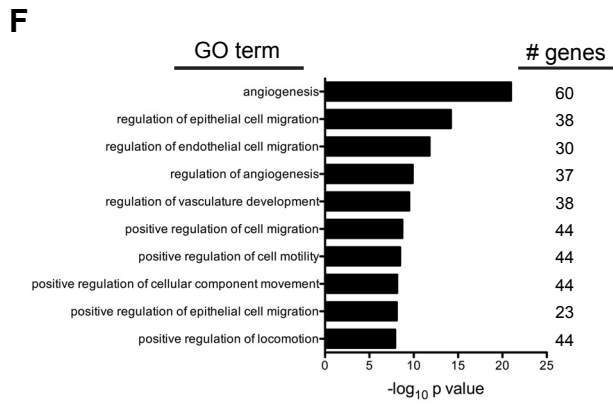
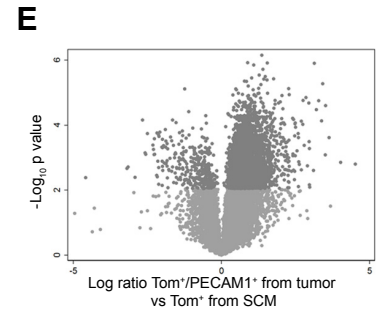
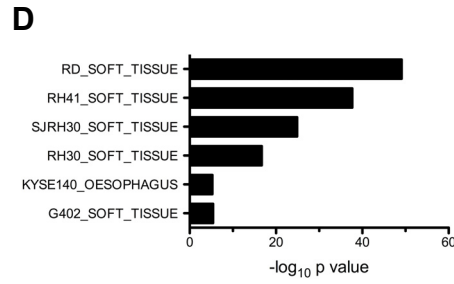
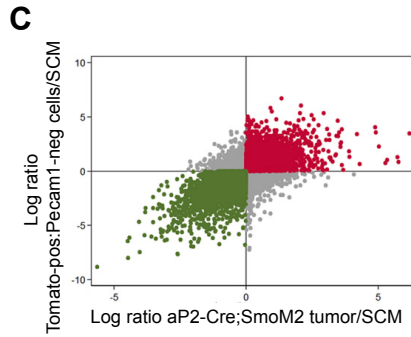
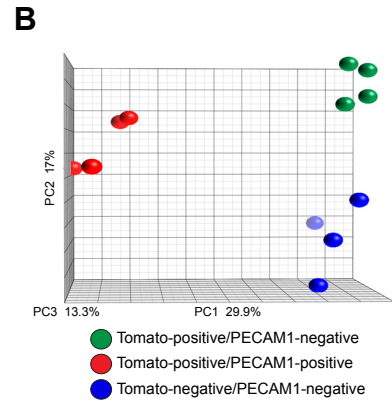
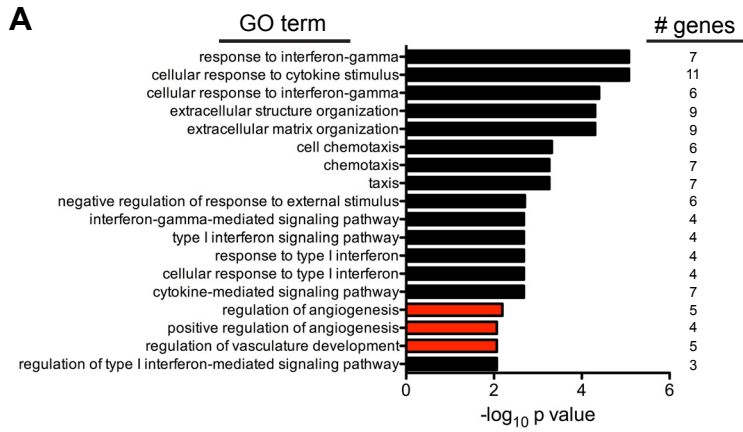


Figure S6. Related to Figure 6. Characterization of cell populations isolated from *aP2-Cre;Smo^{M2/+};R26-tdTom* tumors by FACS

(A) GO term analysis of genes upregulated in *aP2-Cre;Smo^{M2/+}* tumors in comparison to *Myog-Cre;Smo^{M2/+}* tumors (GSE 40359) (Hatley et al., 2012).

(B) Principal component analysis of isolated tumor cell populations from AST animals. Coordinates describe 60.2% of the data (PC1 = 29.9%, PC2 = 17% and PC3 = 17%).

(C) Comparison of isolated Tom⁺PECAM1⁻ tumor cells to whole tumors from *aP2-Cre;Smo^{M2/+}* animals. 55% of 16,957 gene pairs showed agreement in genes upregulated or downregulated in comparison to normal SCM. Pearson correlation = 0.56 (GSE 40359 and 85834).

(D) Comparison of genes enriched in isolated Tom⁺PECAM1⁻ with respect to Tom⁻PECAM1⁻ to genes enriched in cancer cell lines from cancer cell line encyclopedia.

(E) Volcano plot of the Log of the p value versus the log fold change in expression between the Tom⁺ cells isolated from AT SCM and Tom⁺PECAM1⁺ cells isolated from AST tumors by FACS. Solid grey dots indicate 4663 genes with FDR > 0.05. Of total 4663 genes 4518 (97%) were similarly expressed.

(F) GO term analysis of genes upregulated in Tom⁺PECAM1⁺ cells with respect to Tom⁻PECAM1⁻ cells.

(G) Real-time PCR of populations isolated from AST tumors by FACS. Data shown are normalized to *Actb* expression and expressed relative to Tom⁻PECAM1⁻ cells (n = 3, mean ± SEM).

(H) Real-time PCR analysis of *Tbx1* and *Pax3* expression in populations isolated from AST tumors. Data shown are normalized to *Actb* expression and expressed relative to Tom⁻PECAM1⁻ cells (n = 3, mean ± SEM).

*p < 0.05, ** p < 0.01, *** p < 0.001

Table S1. Related to Figure 6. Genes upregulated in *aP2-Cre;Smo^{M2/+}* tumors when compared to *Myogenin-Cre;Smo^{M2/+}* tumors (GSE 40359) (≥ 2 fold; p value < 0.05).

Gene	<i>aP2-Cre;Smo^{M2/+}/ Myogenin-Cre;Smo^{M2/+}</i>		Gene	<i>aP2-Cre;Smo^{M2/+}/ Myogenin-Cre;Smo^{M2/+}</i>	
	Fold change	p value		Fold change	p value
<i>Mmp10</i>	9.1	0.048	<i>C1qb</i>	2.5	0.006
<i>li</i>	8.6	0.013	<i>H2-DMa</i>	2.5	0.007
<i>li</i>	8.6	0.008	<i>H2-Dmb1</i>	2.5	0.010
<i>H2-Ab1</i>	6.9	0.010	<i>D11Ert759e</i>	2.5	0.017
<i>H2-Ab1</i>	6.8	0.020	<i>Fbln1</i>	2.4	0.020
<i>H2-Eb1</i>	6.6	0.004	<i>Rgs5</i>	2.4	0.036
<i>H2-Ab1</i>	5.9	0.009	<i>1500015O10Rik</i>	2.4	0.001
<i>H2-Ab1</i>	5.8	0.005	<i>Fcgr3</i>	2.4	0.002
<i>Comp</i>	4.6	0.023	<i>Xlr4a</i>	2.4	0.014
<i>Ppfibp1</i>	4.4	1.99E-06	<i>Cfd</i>	2.4	0.002
<i>2310061N23Rik</i>	4.1	0.006	<i>C1qg</i>	2.3	0.021
<i>Cxcl14</i>	3.9	0.037	<i>lbsp</i>	2.3	0.010
<i>H2Aa</i>	3.7	0.005	<i>Psemb8</i>	2.3	1E-04
<i>Spp1</i>	3.4	0.024	<i>Laptm5</i>	2.2	0.021
<i>li</i>	3.3	0.002	<i>Aif1</i>	2.2	0.002
<i>Fcrl3</i>	3.3	0.003	<i>H2-Q2</i>	2.2	0.008
<i>Dbp</i>	3.2	0.002	<i>Ms4a6d</i>	2.2	0.015
<i>Lama1</i>	3.2	0.008	<i>Col181a</i>	2.2	0.043
<i>H2-DMa</i>	3.2	0.006	<i>Psemb9</i>	2.2	0.003
<i>Gbp2</i>	3.1	0.048	<i>Mpeg1</i>	2.2	0.006
<i>Ccl8</i>	3.0	0.017	<i>Ly86</i>	2.2	0.012
<i>H2-L</i>	2.9	0.001	<i>Parp14</i>	2.1	0.032
<i>Oasl2</i>	2.9	0.003	<i>Hsp105</i>	2.1	0.014
<i>Wnt5a</i>	2.9	0.042	<i>LOC381292</i>	2.1	0.004
<i>H2-Dmb2</i>	2.9	0.002	<i>Ctsh</i>	2.1	0.023
<i>B2m</i>	2.8	0.003	<i>Scd1</i>	2.1	9.36E-05
<i>Gbp4</i>	2.8	0.026	<i>Wisp2</i>	2.1	0.023
<i>Col8a1</i>	2.8	0.038	<i>Tmepai</i>	2.1	0.021
<i>C1qtnf3</i>	2.8	0.049	<i>Wnt5a</i>	2.1	0.011
<i>Gvin1</i>	2.7	0.037	<i>Cx3cr1</i>	2.1	0.016
<i>Usp18</i>	2.7	0.005	<i>Cxcl16</i>	2.1	0.02
<i>H2-D1</i>	2.7	0.006	<i>ApoE</i>	2.1	0.025
<i>Cd52</i>	2.7	0.032	<i>Bcl2a1d</i>	2.1	0.048
<i>Cd72</i>	2.6	0.002	<i>Ptgis</i>	2.1	0.017
<i>Vcam1</i>	2.6	0.035	<i>Stat1</i>	2.1	0.008
<i>Slamf9</i>	2.6	0.014	<i>Sla</i>	2.0	0.018
<i>B2M</i>	2.6	0.013	<i>Chad</i>	2.0	0.04
<i>LOC56628</i>	2.6	0.005	<i>Oas1g</i>	2.0	0.001
<i>H2-L</i>	2.6	0.003	<i>Col81a</i>	2.0	0.033
<i>Ms4a7</i>	2.5	0.005			

Table S2. Related to Figure 6. Gene ontology functional analysis of 382 genes upregulated in sorted Tom⁺PECAM1⁺ cells with respect to Tom⁻PECAM1⁻ cells isolated from AST tumors (≥ 2 fold; p value < 0.05).

GO Biological Process	Genes	Overlap	Adjusted p value
Actin-myosin filament sliding	<i>Mybpc1; Actn3; Actn2; Tnnc1; Tnnc2; Neb; Myl6b; Ttn; Acta1; Actc1; Des; Myh3; Myl1; Tnnt1; Tnnt2; Tnnt3; Myh8; Tnni1</i>	18/38	9.29E-16
Muscle filament sliding	<i>Mybpc1; Actn3; Actn2; Tnnc1; Tnnc2; Neb; Myl6b; Ttn; Acta1; Actc1; Des; Myh3; Myl1; Tnnt1; Tnnt2; Tnnt3; Myh8; Tnni1</i>	18/38	9.29E-16
Muscle contraction	<i>Ryr1; Chrna1; Smpx; Tshz3; Myl6b; Ttn; Csrp3; Sgca; Mb; Cacng1; Chrnb1; Mybpc1; Dtna; Actn2; Tnnc1; Tnnc2; Pgam2; Khlh41; Trdn; Acta1; Myh1; Des; Actc1; Myh3; Myl1; Tnnt1; Myot; Tnnt3; Myh8; Sntb1</i>	30/195	9.29E-16
Muscle system process	<i>Ryr1; Chrna1; Tshz3; Smpx; Atp2a1; Myl6b; Ttn; Csrp3; Sgca; Mb; Cacng1; Myog; Chrnb1; Mybpc1; Dtna; Actn2; Tnnc1; Tnnc2; Pgam2; Khlh41; Trdn; Acta1; Myh1; Actc1; Des; Myh3; Myl1; Tnnt1; Myot; Tnnt3; Myh8; Sntb1</i>	32/237	1.46E-15
Actin-mediated cell contraction	<i>Mybpc1; Actn3; Actn2; Tnnc1; Tnnc2; Neb; Myl6b; Ttn; Acta1; Actc1; Myh3; Des; Myl1; Tnnt1; Tnnt2; Tnnt3; Myh8; Tnni1</i>	18/46	5.76E-15
Tissue morphogenesis	<i>Lama5; Hhip; Lef1; Tcf21; Ttn; Sall1; Hey1; Sox18; Tnni1; Gpc3; Sostdc1; Sox9; Pitx2; Tbx1; Foxd1; Foxf1; Tnnc1; Ptch1; Wnt5a; Prox1; Bmp7; Pgf; Tbx3; Bmp4; Aldh1a3; Heyl; Actc1; Pax7; Tnnt2; Myf6; Met; Lgr4; Fgfr2; Frem2; Myf5</i>	35/358	2.27E-13
Actin filament-based movement	<i>Mybpc1; Actn3; Actn2; Tnnc1; Tnnc2; Neb; Myl6b; Ttn; Acta1; Actc1; Myh3; Des; Myl1; Tnnt1; Tnnt2; Tnnt3; Myh8; Tnni1</i>	18/61	2.51E-13
Regulation of muscle cell differentiation	<i>Tbx1; Myog; Cav3; Megf10; Tshz3; Mamstr; Foxf1; Prox1; Tbx3; Khlh41; Bmp4; Myod1; Myf6; Smyd1; Sox9; Cdh15; Rbm24; Fgfr2; Cdon; Myf5</i>	20/122	8.82E-11
Striated muscle contraction	<i>Chrna1; Dtna; Smpx; Tnnc1; Pgam2; Tnnc2; Khlh41; Ttn; Csrp3; Myh3; Actc1; Myl1; Tnnt1; Mb; Tnnt3; Myh8</i>	16/71	2.90E-10
Tube development	<i>Robo2; Tgfb2; Foxf2; Tshz3; Foxf1; Wnt5a; Tcf21; Cobl; Prox1; Gata2; Bmp7; Bmp4; Heyl; Calb1; Sall1; Sox18; Plxna2; Sox9; Fgfr2; Stra6</i>	20/164	9.35E-09

Table S3. Related to Figure 7. Genes upregulated in both Tom⁺PECAM1⁺ and Tom⁺PECAM1⁻ cells with respect to Tom⁻PECAM1⁻ cells isolated from AST tumors by FACS. Data shown as fold change from expression arrays.

Gene	Tom ⁻ ;PECAM1 ⁻ /Tom ⁺ ; PECAM1 ⁻		Tom ⁻ ; PECAM1 ⁻ /Tom ⁺ ; PECAM1 ⁺		Tom ⁺ ; PECAM1 ⁻ /Tom ⁺ ; PECAM1 ⁺	
	Fold change	p value	Fold change	p value	Fold change	p value
<i>9530026P05Rik</i>	2.0	0.005	2.1	0.004	-1.1	0.741
<i>9630013A20Rik</i>	2.1	0.023	4.0	0.001	-1.9	0.013
<i>Ablim3</i>	2.3	0.020	16	1.1E-04	-7.0	8.2E-04
<i>Adamts9</i>	2.0	0.003	4.7	3.6E-05	-2.3	7.0E-05
<i>Cdh13</i>	2.3	0.001	4.8	2.1E-05	-2.1	0.001
<i>Cdh5</i>	2.1	0.005	36	1.8E-05	-17	3.8E-05
<i>Chp2</i>	2.4	0.008	5.1	2.3E-04	-2.1	0.015
<i>D930048N14Rik</i>	2.1	0.002	2.6	4.4E-04	-1.2	0.186
<i>Dach1</i>	2.1	0.018	11	1.3E-04	-5.4	1.6E-05
<i>Dock6</i>	2.0	0.007	3.8	1.8E-04	-1.9	0.003
<i>Enpp3</i>	2.5	0.008	8.8	1.5E-04	-3.6	8E-04
<i>Enpp6</i>	4.4	0.002	4.7	0.003	-1.1	0.720
<i>Gata2</i>	2.0	0.006	4.7	5E-04	-2.3	0.010
<i>Gt(ROSA)26Sor</i>	2.4	1.1E-04	3.3	4.3E-05	-1.4	0.004
<i>Itga3</i>	2.0	0.003	4.4	5.1E-05	-2.2	0.002
<i>Lama5</i>	2.1	0.002	3.4	1.4E-04	-1.7	0.010
<i>Lepr</i>	3.7	1.5E-04	3.2	3.6 E-04	1.2	0.348
<i>Lmo7</i>	2.6	0.002	4.3	3.7E-04	-1.7	0.010
<i>Mamstr</i>	2.4	0.002	4.1	7.2E-04	-1.7	0.042
<i>Mpzl1</i>	2.1	0.004	2.0	0.004	1.0	0.870
<i>Nqo1</i>	2.5	0.002	2.6	0.005	-1.0	0.860
<i>Pcdh17</i>	2.9	0.017	4.8	0.004	-1.6	0.060
<i>Plxna2</i>	2.3	0.011	3.6	0.001	-1.6	0.050
<i>Ptprk</i>	2.3	0.015	4.3	8.1E-04	-1.9	0.014
<i>Samd5</i>	2.1	7.7E-04	5.4	1.5E-06	-2.6	1.9E-04
<i>Sncaip</i>	2.1	4.8E-04	7.9	1.5E-05	-3.8	4.7E-05
<i>Sox18</i>	3.1	8.7E-05	19	7.7E-06	-6.1	6.8E-05
<i>Tanc1</i>	2.1	8.1E-04	2.5	0.003	-1.2	0.393
<i>Thsd7a</i>	2.0	0.007	3.1	6.8E-04	-1.5	0.009
<i>Tspan12</i>	2.1	0.007	3.7	3.8E-04	-1.8	0.004
<i>Tspan7</i>	2.2	0.017	11	1.2E-04	-5.1	5.1E-05
<i>Wipf3</i>	2.5	0.034	2.3	0.032	1.1	0.736

Table S4. Related to STAR Methods. Antibodies used for Immunostaining.

Antigen	Sections	Catalog/ Vendor	Concentration	Antigen Retrieval	Secondary	Detection
CD34	FFPE	553731 BD bioscience	1:50	CC1 950-500 Roche	Omap Rat, 760-4457, Roche	DAB
Desmin	FFPE	RB-9014, ThermoFisher	1:500	CC1 950-500 Roche	OmniMap Rabbit, 760-4311, Roche	DAB
ERG	FFPE	ab133264, Abcam	1:1700	ER2 AR9640 Leica	Bond Polymer DS9800, Leica	DAB
CK- Oscar	FFPE	465-01 Covance	1:250	CC1, 950-500 Roche	Rabbit Anti-Mouse OmniMap Rabbit, 760-4311, Roche	DAB
CK-HMV	FFPE	M063029 DAKO	1:750	Proteinase K S302030 DAKO	Rabbit Anti-Mouse OmniMap Rabbit, 760-4311, Roche	DAB
KDR	FFPE -IF	AF644, R&D systems	1:50	pH6 Citrate Buffer C9999, Sigma	Anti-Goat HRP A16005, ThermoFisher	TSA – Fluorescein
	FFPE-IHC	AF644, R&D systems	1:20	Target Retrieval pH6 S1699 DAKO	Anti-Goat HRP SC-2042, Santa Cruz	Strep-HRP, DAB
Laminin	Frozen	L9393, Sigma	1:500	N/A	Anti-Rabbit Cy5 A10523, ThermoFisher	N/A
Meca32	FFPE	553849, BD Bioscience	1:50	Target Retrieval pH6 S1699 DAKO	Rat Polymer RT517L, Biocare Medical	DAB
MHC	Frozen	MF20, DSHB	1:25	N/A	Anti-Mouse AlexaFluor488 A11029, ThermoFisher	N/A
	FFPE	MF20, DSHB	1:25	pH6 Citrate Buffer C9999, Sigma	Anti-Mouse AlexaFluor488 A11029, ThermoFisher	N/A
MyoD1	FFPE-IF	M3512, DAKO	1:50	Target Retrieval pH9 S2367 DAKO	Goat Anti Mouse IgG1HRP S2969 SantaCruz	TSA – Fluorescein
	FFPE-IHC	386R-18, Cell Marque	undiluted	ER2 AR9640, Leica	Bond Polymer DS9800, Leica	DAB
Myogenin	FFPE-IHC	M3559, DAKO	1:200	Target Retrieval pH9 S2367 DAKO	Mouse on Mouse polymer HRP MM510L, Biocare Medical	DAB
Pax7	Frozen	DSHB	1:10	pH6 Citrate Buffer C9999, Sigma	Anti mouseIgG1 AlexaFluor488 A21121, ThermoFisher	N/A
Pecam1	Cultures/ Frozen	550274 BD bioscience	1:50	N/A	Anti Rat AlexaFluor488 A11006, ThermoFisher	N/A
	FFPE - IF	Dianova Dia 310	1:20	pH6 Citrate Buffer C9999, Sigma	Anti rat HRP AP136P, Millipore	TSA – Fluorescein
	FFPE-IHC	Dianova Dia 310	1:100	CC1 950-500 Roche	OmniMap Rat 760-4457 Roche	DAB
S100	FFPE-IHC	Z031129 Dako	1:2,000	CC1 950-500 Roche	OmniMap Rabbit, 760-4311, Roche	DAB
Tomato	Frozen (Pax7 only)	600-401-379 Rockland	1:100		Anti Rabbit AlexaFluor568, A11056, A10042 ThermoFisher	N/A
	FFPE-IHC	600-401-379 Rockland	1:100		Anti Rabbit AlexaFluor568, A11056, ThermoFisher	N/A
	RNAscope/ IHC	LS-C340696 LS Bio	1:20,000	CC1,950-500 Roche	OmniMap Rabbit, 760-4311, Roche	DAB
Vimentin	FFPE-IHC	NBP1-40730, Novus	1:1000	ER1, AR9961 Leica	Bond Polymer DS9800, Leica	DAB

Table S5. Related to STAR Methods. Antibodies used for Flow Cytometry.

	Clone	Catalog # /Company	Concentration
β -1 integrin-FITC	Ha2/5	561796 BD Biosciences	1:50
CD11b-APC	M1/70	553312 BD Biosciences	1:50
CD45-APC	30-F11	561018 BD Biosciences	1:50
CXCR4-Biotin	2B11/CXCR4	551968 BD Biosciences	1:50
Streptavidin PE-Cy7		25-431782 eBioscience	1:50
Pecam1-APC	Mec13.3	551262 BD Biosciences	1:50
Pecam1-FITC	Mec13.3	561813 BD Biosciences	1:50
Sca1-PerCP-Cy5.5	D7	25-5981-82 eBioscience	1:50
Ter119-APC		557909 BD Biosciences	1:50

Table S6. Related to STAR Methods. SYBR primers used for quantitative real-time PCR.

	Right Primer	Left Primer
<i>Actb</i>	ATGGAGGGGAATACAGCCC	TTCTTTGCAGCTCCTTCGTT
<i>Aldh1A3</i>	GACGAAAAGGCATGAAGGA	TCAGCTGGCTGACCTTGTAG
<i>aP2</i>	CACTTTCCTTGTGGCAAAGC	AATGTGTGATGCCTTTGTGG
<i>Bmp4</i>	AGCCCGCTTCTGCAGGA	AAAGGCTCAGAGAAGCTGCG
<i>Cdh5</i>	CGTTGGACTTGATCTTTCCC	CGCCAAAAGAGAGACTGGAT
<i>Cspg4</i>	ACTCTGGTCAGAGCTGAGGG	TCTTACCTTGGCCTTGTGG
<i>Foxd1</i>	GTCTGGTCCAAGAATCCGAA	GGGAGAGCGAAGGTAGGACT
<i>Foxf1</i>	TGGGCGACTGTGAGTGATAC	ACTCCCTGGAGCAGCCATAC
<i>Gata2</i>	GCCATAAGGTGGTGGTTGTC	CACTACCTGTGCAATGCCTG
<i>Gli1</i>	ATTGGATTGAACATGGCGTC	GGATGAAGAAGCAGTTGGGA
<i>Hey1</i>	TCCGATAGTCCATAGCCAGG	TTGCAGATGACTGTGGATCA
<i>Heyl</i>	TCTCTGCGCCGTTTCTCTAT	CCATCGATGTGGGTCAAGAG
<i>Kdr</i>	TCCAGAATCCTCTTCCATGC	CCAGAGACCCTCGTTTTTCAG
<i>Lef1</i>	AAATGGGTCCCTTTCTCCAC	TCGTCGCTGTAGGTGATGAG
<i>Myf5</i>	GACAGGGCTGTTACATTCAGG	TGAGGGAACAGGTGGAGAAC
<i>Myod1</i>	GTCTAGCCATTCTGCCG	AGCACTACAGTGGCGACTCA
<i>Pax7</i>	GTCCGGTTCTGATTCCACAT	GCGAGAAGAAAGCCAAACAC
<i>Pdgfra</i>	AGAAAATCCGATACCCGGAG	AGAGGAGGAGCTTGAGGGAG
<i>Pdgfrb</i>	TGGTATCACTCCTGGAAGCC	AACAGAAGACAGCGAGGTGG
<i>Pecam1</i>	AGTTGCTGCCATTCATCAC	CTGGTGCTCTATGCAAGCCT
<i>Peg3</i>	TCAACTGATCTCCCCTTGCT	TTTGGAGACAACTGGCAAGA
<i>Pparg</i>	TCTTCCATCACGGAGAGGTC	GATGCACTGCCTATGAGCAC
<i>Prox1</i>	GGCATTGAAAACTCCCGTA	TTGTGAGCTATACCGAGCCC
<i>Ptch1</i>	AATTCTCGACTCACTCGTCCA	CTCCTCATATTTGGGGCCTT
<i>Sox9</i>	TCCACGAAGGGTCTCTTCTC	AGGAAGCTGGCAGACCAGTA
<i>Sox18</i>	AAGTGCTGCACACGCAAC	CACAACGCAGTACTGAGCAAG
<i>SmoM2-YFP</i>	GGGATCCATTCAATCCCGCA	AAACGGCCACAAGTTCAGC
<i>Tbx3</i>	CATTGCCAGTGTCTCGAAAA	TCCCGGAAACAGAATTCATC
<i>Tcf7l2</i>	GGGATCATGATGAAGGGGTA	TAAATCCCGGGAAAGTTTGG
<i>Tcf21</i>	GGCTGGCGTCCAGCTACATCG	TGCCGGCCACCATAAAGGGC
<i>Tomato</i>	AGTTCATGTACGGCTCCAAGG	AGCGCGTGATGAACTTCGA
<i>Tomato2</i>	GGCATTAAAGCAGCGTATCC	CTGTTCTGTACGGCATGG
<i>Tek</i>	TTTCGGCATCAGACACAAGA	CCGGCTTAGTTCTCTGTGGA
<i>Vwf</i>	CCGTCTTCAGTAGCTGGCAT	GTGTAAACGGGCATCTCCTC
<i>Ucp1</i>	AGAAGCCACAAACCCTTTGA	TCTCTGCCAGGACAGTACC