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Supplemental Information

Widespread Rewiring of Genetic Networks

upon Cancer Signaling Pathway Activation

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Table S2: Gene identifier for the 72 query genes used in the study. Related to Figure 1. For the two independent dsRNA sequences used to target those query genes, please see Table S1.

FBgn ID	CG ID	Symbol
FBgn0001324	CG8491	kto
FBgn0004379	CG10923	Klp67A
FBgn0010380	CG12532	AP-1-2beta
FBgn0010379	CG4006	Akt1
FBgn0034970	CG4005	yki
FBgn0052675	CG32675	lango5
FBgn0039907	CG2041	lgs
FBgn0004597	CG7281	Apcz
FBgn0026597	CG7926	Axn
FBgn0043012	CG6056	AP-2sigma
FBgn0023423	CG3412	slmb
FBgn0015589	CG1451	Арс
FBgn0015391	CG11397	glu
FBgn0250786	CG3733	Chd1
FBgn0005671	CG17369	Vha55
FBgn0025830	CG5859	IntS8
FBgn0000212	CG5942	brm
FBgn0024921	CG7398	Trn
FBgn0030276	CG1938	DIIC
EBgn0002945	CG11614	nkd
FBgn0004378	CG9191	Kin61E
FBgn0003013	CG7467	osa
FBgn0034964	CG3173	IntS1
FBgn0032863	CG2508	Cdc23
FBgn0036038	CG18176	defl
FBgn0031050	CG12235	Arp10
FBgn0036141	CG6210	wls
FBgn0021825	CG8269	Dmn
FBgn0000117	CG11579	arm
FBgn0025781	CG6/59	Cdc16
EBgp0000462	CG2092	
FBgn0085432	CG34403	nan
FBgn0004647	CG3936	N
FBgn0001219	CG4264	Hsc70-4
FBgn0013749	CG11027	Arf102F
FBgn0004009	CG4889	wg
FBgn0053526	CG33526	PNUTS
FBgn0000499	CG18361	dsh
FBgn0003371	CG2621	sgg
EBgn0020238	CG31196	14-3-3ensilon
FBgn0004390	CG6721	RasGAP1
FBgn0003415	CG9936	skd
FBgn0004859	CG2125	ci
FBgn0000119	CG5912	arr
FBgn0261456	CG11228	hpo
FBgn0014020	CG8416	Rho1
FBgn0043900	CG11518	рудо
FBgn0015618	CG10572	Cdk8
FBgn0031030	CG14217	Тао
FBgn00010315	CG9096	CycD
FBgn0035851	CG7999	MED24
FBgn0001085	CG17697	f7
FBgn0001230	CG5436	Hsp68
FBgn0025637	CG16983	skpA
FBgn0016797	CG9739	fz2
FBgn0029709	CG3564	CHOp24
FBgn0005386	CG8887	ash1
FBgn0034708	CG5625	Vps35
FBgn0030093	CG7055	dalao
FBgp0000405	CG10449	Catsup
FBgn0026379	CG5671	Pten
FBgn0037573	CG7483	elF4AIII
FBgn0044028	CG13076	Notum
FBgn0015024	CG2028	Cklalpha
FBgn0086357	CG9539	Sec61alpha
FBgn0014411	CG14804	Vps26



Figure S1. Gene prioritization for genetic interaction analysis in the Wnt signaling pathway. Related to Figure 1. (A) Correlation (Pearson correlation coefficient, PCC) of normalized Wnt signaling activity between the biological replicates of the genome-wide RNAi screen, or the two independent dsRNA designs against 13,369 of the 14,331 targeted genes. (B) Wnt activity z-scores in *Drosophila* S2R+ and 1182-4H cells. Core Wnt pathway components are highlighted in green (positive regulators) and red (negative regulators). (C) Functional groups of the 336 selected genes for Wnt pathway state-dependent quantification of genetic interactions. Genes were assigned to the groups by manual curation. (D) Wnt pathway activity z-scores of the selected genes and per functional group as illustrated in (C). Distribution of all dsRNA designs is shown on top for relative comparison.





G

Pos. genetic interaction



Neg. genetic interaction



Figure S2

Figure S2. Distinct modes of Wnt pathway activation. Related to Figure 1. (A) Foldchange Wnt pathway activity upon increasing concentrations of transfected pAc-wg plasmid. The wildtype pathway activity was measured in 32 wells per concentration and upon transfection of GFP dsRNA (black line). Knockdown of two independent dsRNAs against wg and evi (wls) illustrates that the pathway activity increase was due to Wg expression and secretion (by its cargo receptor Evi). (B) Activity levels of the Wnt-specific dTCF-luc reporter and mRNA levels of the Wnt target gene nkd in the three pathway states in which we screened for genetic interactions. Wnt reporter activity was measured in 576 wells distributed over six screening batches and nkd mRNA levels were quantified by qPCR in biological triplicates. Dashed ellipses illustrate the standard deviation of the data. The data was normalized to the baseline state value. (C) Single and combinatorial knockdown phenotypes of Apc and Apc2 illustrating a positive genetic interaction (FDR of 1%). Error bars correspond to the bootstrapped MAD for the single knockdown effects and SD for the combinatorial knockdown measurements. (D) Comparison of genetic interaction score replicates within and between states. For each pair of a query (Q) and template (T) gene all four combinations of two independent dsRNA designs were scored independently. Q1-T1 represents the 24120 scores measured between the first query and the first template dsRNA design with the scores measured with the other three Q-T combinations. The Pearson correlation coefficient was used to compare replicates. (E) Reproducibility of π -scores with re-screened (four replicates each) query genes *lgs, dsh* and *pygo*, which cover the receptor level and target gene transcription. Pearson correlation coefficients are highly significant (p < 2.2e-16). (F) M-A-plot showing the deviation A between replicates from their mean M for the five query genes that were re-screened. The size of the dots represents 1 – FDR for each genetic interaction (called for replicate 1). (G) Overlap of significant positive (yellow) and negative (blue) genetic interaction (FDR < 1%).



Figure S3. State-dependent genetic interactions of Wnt signaling. Related to Figure 1. (A-C) Combinatorial RNAi enables quantification of genetic interactions representing epistatic relations in the Wnt pathway. Single knockdown phenotypes were estimated from phenotypes of two independent dsRNAs in 144 different genetic backgrounds each (error bars show standard error of median determined by bootstrapping). The white dashed bar indicates the expected combinatorial knockdown effect for each gene pair using a multiplicative neutrality function. The measured combinatorial phenotype illustrates the median of the four possible combinations of two independent dsRNA designs against each gene. The difference between the expected and measured combinatorial phenotype is quantified as a π -score and quantifies the genetic interaction (yellow if positive, blue if negative at FDR < 1%). The data is presented at log₂ scale. Examples from the Wnt active state are shown in Figure 1C, D. (D, E) Complete genetic interactions measured in the baseline state versus ligand-mediated (D) or Apc loss-mediated (G) induction. Genetic interactions between members of known selected functional modules are highlighted. (F-H) Comparison of genetic interaction profile correlation and protein-protein interactions. Density of genetic interaction profile correlation of gene pairs that share a protein-protein interaction as reported previously (Guruharsha et al., 2011) (darker line) and those that do not share an interaction in this study (ligher grey line). (I) Overlap of significant genetic interactions (FDR 1%) in the baseline, Wnt active and Apc-loss state as well as protein-protein interactions.



Figure S4. Genetic interaction profile self-correlation between pathway states and state-specific gene-gene profile correlation. Related to Figure 3. (**A**, **C**, **E**) Change of genetic interaction profiles of receptor complex components arr and dsh (A), transcriptional regulators trx and pnt (C) or ash1 (E) from baseline to Wnt-active state. Grey lines connect genetic interaction scores between the gene indicated at the axis and a given query gene. (**B**, **D**, **F**) Self-correlation of genetic interaction profiles of the above-mentioned genes between baseline and Wnt-active state.



Figure S5. Functional plasticity of destruction complex components. Related to Figure 4. (A) Distribution of the 112,560 gene-gene between-state relations between baseline and Wnt-active (left) or Apc-loss (right) state. (B) Comparison of baselinespecific similarity (Pearson correlation coefficient (PCC) between genetic interaction profiles) with between baseline and Wnt-active state similarity of baseline state Axn with other destruction complex components. The relative differential PCC (Rel. dPCC) shows the slope (baseline-specific PCC minus between-state PCC) relative to the respective baseline-specific PCC. The violin plots show all PCCs with Axn. (C) Comparison of baseline-specific similarity with between baseline and Wnt-active state similarity of baseline state sgg with other destruction complex components. (D) Relative differential PCC of remaining destruction complex components. (E) sgg knockdown effect on Wnt signaling activity in the baseline and Wnt-active state. The single knockdown phenotype was estimated from phenotypes of two independent dsRNAs in 144 different genetic backgrounds each (error bars show standard error of median determined by bootstrapping). (F) Change of the single knockdown phenotype upon ligand-mediated pathway activation. Gene depletion lead to more negative (green) or positive (red) pathway activity. The order of genes (y axis) was determined by the between-state cumulative rank distance shown in (Figure 4B). (G) Comparison of PCC and CSI illustrating pair-wise similarity in the Wnt active state. The dotted line represents the threshold applied for identifying functional modules in Figure 4D, E. (H) Number of members of a given functional module that showed genetic interaction profile similarity with each of the module members when considering the PCC (left) or the CSI (right).



Catsup⁴⁷

Catsup47

Ng tota

Figure S6

Figure S6. *Catsup* is required for Wg secretion. Related to Figure 5. (A) Knockdown of *Catsup* depletes Wnt target gene *sens* expression in the *Drosophila* wing disc. (B) Quantification of *sens* expression along the A/P axis of the wing disc. enGal4/UAS-*Catsup*RNAi, posterior to the right. *sens* staining intensity was quantified using FIJI (n > 4 wing discs). (C) Knockout of *Catsup* in the *Drosophila* wing disc leads to Wg accumulation in the Wg producing cells. (D) Wg accumulates at the apical side of the Wg secreting cells in the wing disc. This cross section was reconstructed using z-stacks. *Catsup*⁴⁷ clones in the *Drosophila* wing disc are marked by the absence of GFP expression.