

Identification of valid reference genes for mRNA and microRNA normalisation in prostate cancer cell lines

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Supporting Information

Figure S1. Melting peaks of all primer pairs. The specificity of all the primer pairs was confirmed by a single peak in the melting curve.

Figure S2. Relative expression of *PCA3* normalised by different reference genes and their combinations. Fold change in *PCA3* expression of LNCaP cells subjected to different DHT treatments. Stable reference genes *YWHAZ*, *ACTB*, their combination, and least stable reference genes *SDHA* and *ALAS1* were independently used for normalisation. Bars indicate +SD evaluated from three biological replicates. * and ** indicate $P < 0.05$ and $P < 0.01$, respectively.

Figure S3. Full-length unadjusted images of Western blots shown in Figure 1.

Table S1. Information on mRNA and npcRNA reference genes used in this study. * indicates *Homo sapiens* unplaced genomic contig, GRCh37.p5.

Gene symbol	Gene name/RNA Species	Chromosomal Localisation	Molecular function
<i>18S rRNA</i>	18S ribosomal RNA	ChrUn*	Ribosome subunit, translation
<i>ACTB</i>	β -Actin	7p22-p12	Cytoskeleton
<i>ALAS1</i>	aminolevulinate, delta-, synthase 1	3p21.1	Heme synthesis
<i>GAPDH</i>	Glyceraldehyde-3-phosphate dehydrogenase	12p13.31	Glycolysis
<i>HPRT1</i>	Hypoxanthine phosphoribosyl-transferase 1	Xq26.1	Generation of purine nucleotides
<i>K-ALPHA-1</i>	K-alpha-1 tubulin, alpha, ubiquitous	12q13.12	Structural constituent of cytoskeleton
<i>RPL13A</i>	Ribosomal protein L13a	19q13.3	Catalyze protein synthesis
<i>SDHA</i>	Succinate dehydrogenase complex, subunit A	5p15	Glycolysis
<i>TBP</i>	TATA box binding protein	6q27	Transcription initiation
<i>YWHAZ</i>	Tyrosine 3-monooxygenase	8q23.1	Signal transduction
<i>miR-16</i>	miRNA	13q14.2	Negative regulation in chronic lymphocytic leukemia
<i>miR-130b</i>	miRNA	22q11.21	Gene expression regulation in cell growth and cell invasion
<i>miR-1225-3p</i>	miRNA	16p13.3	Gene expression regulation in stem cell self-renewal and differentiation
<i>miR-1228-3p</i>	miRNA	12q13.3	Cellular development and lipid Metabolism
<i>RNU6-2</i>	snoRNA*	19p13.3	Cellular RNAs modification
<i>RNU43</i>	snoRNA	22q13.1	Ribosomal RNA modification

Table S2. Quantity and integrity of total RNA in this study.

	Treatment I					Treatment II			
	DHT (nM)	RNA conc. (ng/ul)	260/280	260/230	RIN	RNA conc. (ng/ul)	260/280	260/230	RIN
RWPE-1	0	760	2.08	1.46	10	712	2.06	2.11	10
	1	692	2.09	1.66	9.9	628	2.09	1.47	10
	10	716	2.06	2.12	10	733	2.10	1.59	10
	100	653	2.08	1.42	10	688	2.10	1.52	9.8
	1000	630	2.08	1.85	10	627	2.09	1.48	10
LNCaP	0	506	2.05	2.09	9.9	519	2.02	1.56	9.8
	1	536	2.07	1.41	9.8	454	2.04	1.89	9.9
	10	559	2.06	2.00	9.7	488	2.02	1.93	10
	100	576	2.08	1.66	9.8	455	2.05	1.53	9.6
	1000	582	2.08	2.04	9.9	410	2.05	1.90	9.7
DU145	0	432	2.06	1.91	10	390	2.10	0.92	9.6
	1	487	2.05	1.58	9.7	356	2.09	1.42	9.8
	10	420	2.05	2.05	9.6	375	2.08	1.90	9.9
	100	402	2.07	2.04	9.7	364	2.08	2.05	9.6
	1000	516	2.02	1.90	9.8	359	2.08	1.82	9.5
22RV1	0	503	2.08	1.98	10	423	2.10	2.03	10
	1	447	2.11	1.80	9.4	446	2.09	1.81	9.9
	10	460	2.10	1.58	9.8	403	2.10	1.80	9.9
	100	483	2.08	1.60	10	438	2.09	2.06	9.8
	1000	476	2.09	1.73	9.7	457	2.09	1.57	10
PC-3	0	301	2.11	0.93	10	290	2.13	1.51	9.8
	1	288	2.10	1.42	10	349	2.08	1.53	9.7
	10	364	2.09	2.09	9.9	364	2.09	1.88	9.7
	100	333	2.10	2.10	9.7	323	2.10	1.34	9.6
	1000	266	2.08	2.08	9.6	301	2.10	1.68	9.5

Table S3. Ranking of candidate reference genes calculated by NormFinder based on their expression stability values.

Rank	0 nM		1 nM		10 nM		100 nM		1000 nM		Total samples	
	gene	SV	gene	SV	gene	SV	gene	SV	gene	SV	gene	SV
1	<i>GAPDH</i>	0.13	<i>ACTB</i>	0.06	<i>GAPDH</i>	0.06	<i>ACTB</i>	0.07	<i>ACTB</i>	0.03	<i>ACTB</i>	0.08
2	<i>ACTB</i>	0.14	<i>GAPDH</i>	0.13	<i>ACTB</i>	0.09	<i>GAPDH</i>	0.11	<i>GAPDH</i>	0.09	<i>GAPDH</i>	0.09
3	<i>TBP</i>	0.15	<i>TBP</i>	0.14	<i>TBP</i>	0.21	<i>18S rRNA</i>	0.17	<i>TBP</i>	0.24	<i>TBP</i>	0.18
4	<i>HPRT1</i>	0.25	<i>HPRT1</i>	0.25	<i>RPL13A</i>	0.27	<i>K-ALPHA-1</i>	0.21	<i>ALAS1</i>	0.29	<i>HPRT1</i>	0.26
5	<i>SDHA</i>	0.29	<i>SDHA</i>	0.27	<i>SDHA</i>	0.27	<i>TBP</i>	0.23	<i>HPRT1</i>	0.29	<i>SDHA</i>	0.27
6	<i>RPL13A</i>	0.34	<i>RPL13A</i>	0.30	<i>HPRT1</i>	0.28	<i>RPL13A</i>	0.28	<i>YWHAZ</i>	0.30	<i>RPL13A</i>	0.27
7	<i>K-ALPHA-1</i>	0.35	<i>K-ALPHA-1</i>	0.32	<i>K-ALPHA-1</i>	0.29	<i>SDHA</i>	0.29	<i>RPL13A</i>	0.31	<i>K-ALPHA-1</i>	0.28
8	<i>18S rRNA</i>	0.35	<i>YWHAZ</i>	0.32	<i>YWHAZ</i>	0.31	<i>YWHAZ</i>	0.29	<i>K-ALPHA-1</i>	0.32	<i>18S rRNA</i>	0.29
9	<i>ALAS1</i>	0.38	<i>ALAS1</i>	0.32	<i>18S rRNA</i>	0.32	<i>HPRT1</i>	0.33	<i>18S rRNA</i>	0.33	<i>YWHAZ</i>	0.30
10	<i>YWHAZ</i>	0.39	<i>18S rRNA</i>	0.38	<i>ALAS1</i>	0.33	<i>ALAS1</i>	0.33	<i>SDHA</i>	0.33	<i>ALAS1</i>	0.30
1	<i>miR-1228-3p</i>	0.20	<i>miR-16</i>	0.14	<i>miR-1228-3p</i>	0.10	<i>miR-1225-3p</i>	0.12	<i>miR-1225-3p</i>	0.11	<i>miR-1225-3p</i>	0.13
2	<i>miR-1225-3p</i>	0.22	<i>miR-1225-3p</i>	0.22	<i>miR-1225-3p</i>	0.12	<i>miR-16</i>	0.13	<i>miR-16</i>	0.20	<i>miR-16</i>	0.26
3	<i>RNU6-2</i>	0.28	<i>miR-130b</i>	0.33	<i>RNU6-2</i>	0.27	<i>miR-130b</i>	0.13	<i>miR-130b</i>	0.38	<i>miR-1228-3p</i>	0.31
4	<i>miR-16</i>	0.31	<i>RNU43</i>	0.36	<i>miR-130b</i>	0.42	<i>miR-1228-3p</i>	0.44	<i>RNU43</i>	0.43	<i>miR-130b</i>	0.35
5	<i>RNU43</i>	0.40	<i>miR-1228-3p</i>	0.37	<i>miR-16</i>	0.44	<i>RNU43</i>	0.51	<i>miR-1228-3p</i>	0.46	<i>RNU43</i>	0.41
6	<i>miR-130b</i>	0.42	<i>RNU6-2</i>	0.55	<i>RNU43</i>	0.45	<i>RNU6-2</i>	0.59	<i>RNU6-2</i>	0.61	<i>RNU6-2</i>	0.44

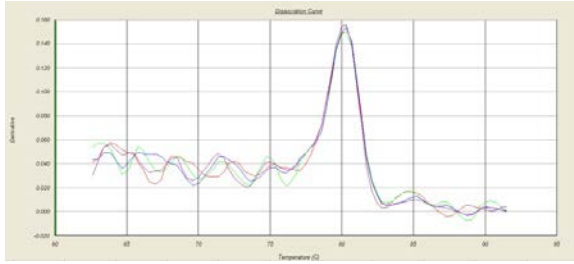
Table S4. Ranking of candidate reference genes calculated by BestKeeper based on their standard deviation.

Rank	0 nM		1 nM		10 nM		100 nM		1000 nM		Total samples	
	gene	SD	gene	SD	gene	SD	gene	SD	gene	SD	gene	SD
1	<i>ACTB</i>	0.23	<i>ACTB</i>	0.16	<i>ACTB</i>	0.17	<i>ACTB</i>	0.12	<i>ACTB</i>	0.15	<i>ACTB</i>	0.17
2	<i>GAPDH</i>	0.25	<i>GAPDH</i>	0.22	<i>YWHAZ</i>	0.19	<i>18S rRNA</i>	0.16	<i>YWHAZ</i>	0.20	<i>YWHAZ</i>	0.23
3	<i>TBP</i>	0.29	<i>YWHAZ</i>	0.23	<i>RPL13A</i>	0.19	<i>GAPDH</i>	0.21	<i>RPL13A</i>	0.22	<i>RPL13A</i>	0.24
4	<i>RPL13A</i>	0.30	<i>RPL13A</i>	0.25	<i>GAPDH</i>	0.24	<i>YWHAZ</i>	0.22	<i>ALAS1</i>	0.25	<i>GAPDH</i>	0.25
5	<i>YWHAZ</i>	0.32	<i>TBP</i>	0.26	<i>ALAS1</i>	0.32	<i>RPL13A</i>	0.25	<i>18S rRNA</i>	0.29	<i>18S rRNA</i>	0.30
6	<i>18S rRNA</i>	0.33	<i>ALAS1</i>	0.31	<i>18S rRNA</i>	0.34	<i>K-ALPHA-1</i>	0.30	<i>GAPDH</i>	0.31	<i>ALAS1</i>	0.31
7	<i>ALAS1</i>	0.34	<i>SDHA</i>	0.32	<i>SDHA</i>	0.34	<i>SDHA</i>	0.32	<i>TBP</i>	0.39	<i>TBP</i>	0.33
8	<i>SDHA</i>	0.36	<i>18S rRNA</i>	0.39	<i>TBP</i>	0.38	<i>ALAS1</i>	0.32	<i>SDHA</i>	0.43	<i>SDHA</i>	0.35
9	<i>HPRT1</i>	0.39	<i>K-ALPHA-1</i>	0.43	<i>K-ALPHA-1</i>	0.45	<i>TBP</i>	0.33	<i>K-ALPHA-1</i>	0.47	<i>K-ALPHA-1</i>	0.42
10	<i>K-ALPHA-1</i>	0.44	<i>HPRT1</i>	0.45	<i>HPRT1</i>	0.49	<i>HPRT1</i>	0.48	<i>HPRT1</i>	0.48	<i>HPRT1</i>	0.46
1	<i>miR-16</i>	0.31	<i>miR-16</i>	0.19	<i>miR-1228-3p</i>	0.09	<i>miR-1225-3p</i>	0.22	<i>miR-16</i>	0.19	<i>miR-16</i>	0.29
2	<i>miR-1228-3p</i>	0.36	<i>miR-1225-3p</i>	0.28	<i>RNU43</i>	0.29	<i>miR-16</i>	0.33	<i>miR-1225-3p</i>	0.25	<i>miR-1228-3p</i>	0.32
3	<i>RNU43</i>	0.42	<i>miR-1228-3p</i>	0.32	<i>miR-1225-3p</i>	0.30	<i>miR-1228-3p</i>	0.38	<i>miR-1228-3p</i>	0.38	<i>miR-1225-3p</i>	0.32
4	<i>miR-130b</i>	0.45	<i>miR-130b</i>	0.43	<i>miR-16</i>	0.40	<i>miR-130b</i>	0.41	<i>miR-130b</i>	0.44	<i>RNU43</i>	0.53
5	<i>miR-1225-3p</i>	0.47	<i>RNU43</i>	0.45	<i>RNU6-2</i>	0.56	<i>RNU43</i>	0.59	<i>RNU6-2</i>	0.63	<i>miR-130b</i>	0.54
6	<i>RNU6-2</i>	0.50	<i>RNU6-2</i>	0.56	<i>miR-130b</i>	0.68	<i>RNU6-2</i>	0.82	<i>RNU43</i>	0.64	<i>RNU6-2</i>	0.63

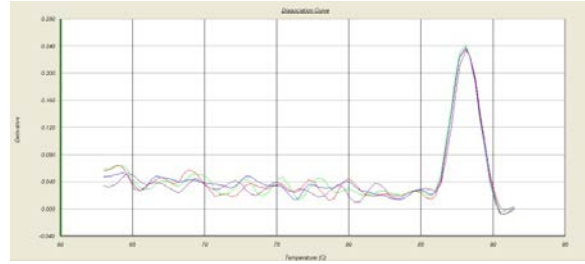
Table S5. Information on primers for amplification mRNA reference genes and target gene. *Not available.

Gene	Forward and Reverse Primer	Primer concentration (nM)	Product (bp)	Intron spanning	Reference
<i>18S rRNA</i>	GGATGTAAAGGATGGAAAATACA	50	72	NA*	[1]
	TCCAGGTCTTCACGGAGCTTGTT	50			
<i>ACTB</i>	TGACGTGGACATCCGCAAAG	50	205	Yes	[1]
	CTGGAAGGTGGACAGCGAGG	50			
<i>ALAS1</i>	AGTGTGAAAACCGATGGAGG	200	140	Yes	[2]
	CGATCATACTGAAAAGTGGAAACAG	200			
<i>GAPDH</i>	GACAGTCAGCCGCATCTTCT	50	127	Yes	[1]
	TTAAAAGCAGCCCTGGTGAC	50			
<i>HPRT1</i>	GACCAGTCAACAGGGGACAT	50	132	Yes	[3]
	CCTGACCAAGGAAAGCAAAG	50			
<i>K-ALPHA-1</i>	TGGAACCCACAGTCATTGATGA	200	135	Yes	[4]
	TGATCTCCTTGCCAATGGTGTA	200			
<i>RPL13A</i>	CGGACCGTGCGAGGTAT	200	114	Yes	[4]
	CACCATCCGCTTTTTCTTGTC	200			
<i>SDHA</i>	TGGTTGTCTTTGGTCGGG	50	85	Yes	[5]
	GCGTTTGGTTTAATTGGAGGG	50			
<i>TBP</i>	GAGAGTTCTGGGATTGTACCG	200	143	Yes	[5]
	ATCCTCATGATTACCGCAGC	200			
<i>YWHAZ</i>	ATGCAACCAACACATCCTATC	50	178	Yes	[6]
	GCATTATTAGCGTGCTGTCTT	50			

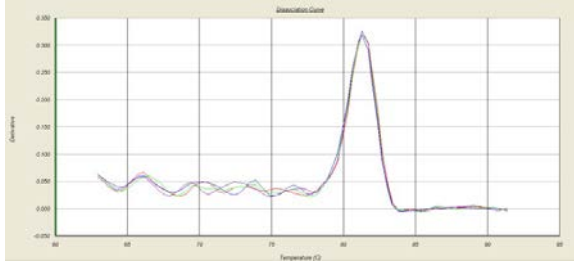
18S rRNA



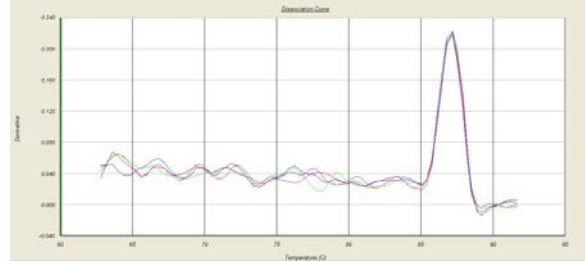
ACTB



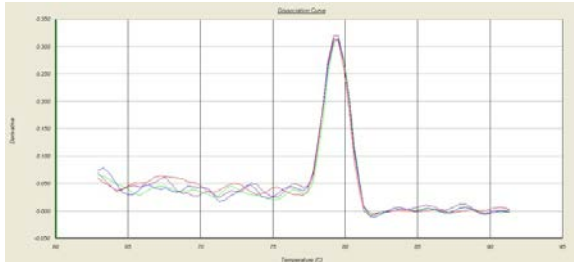
ALAS1



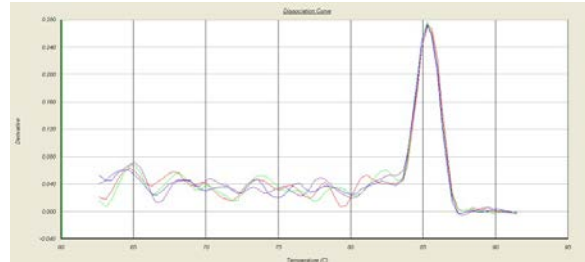
GAPDH



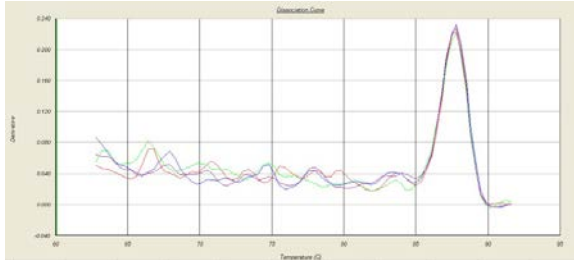
HPRT1



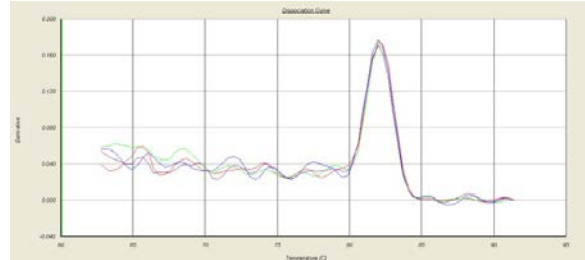
K-ALPHA-1



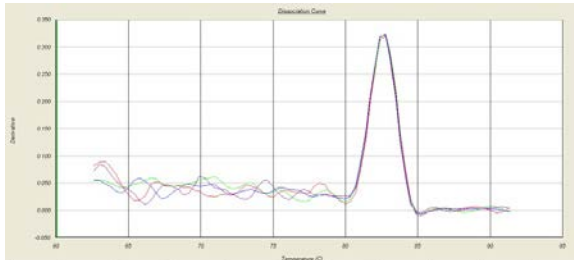
RPL13A



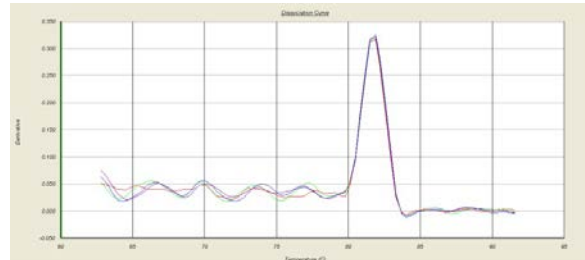
SDHA



TBP



YWHAZ



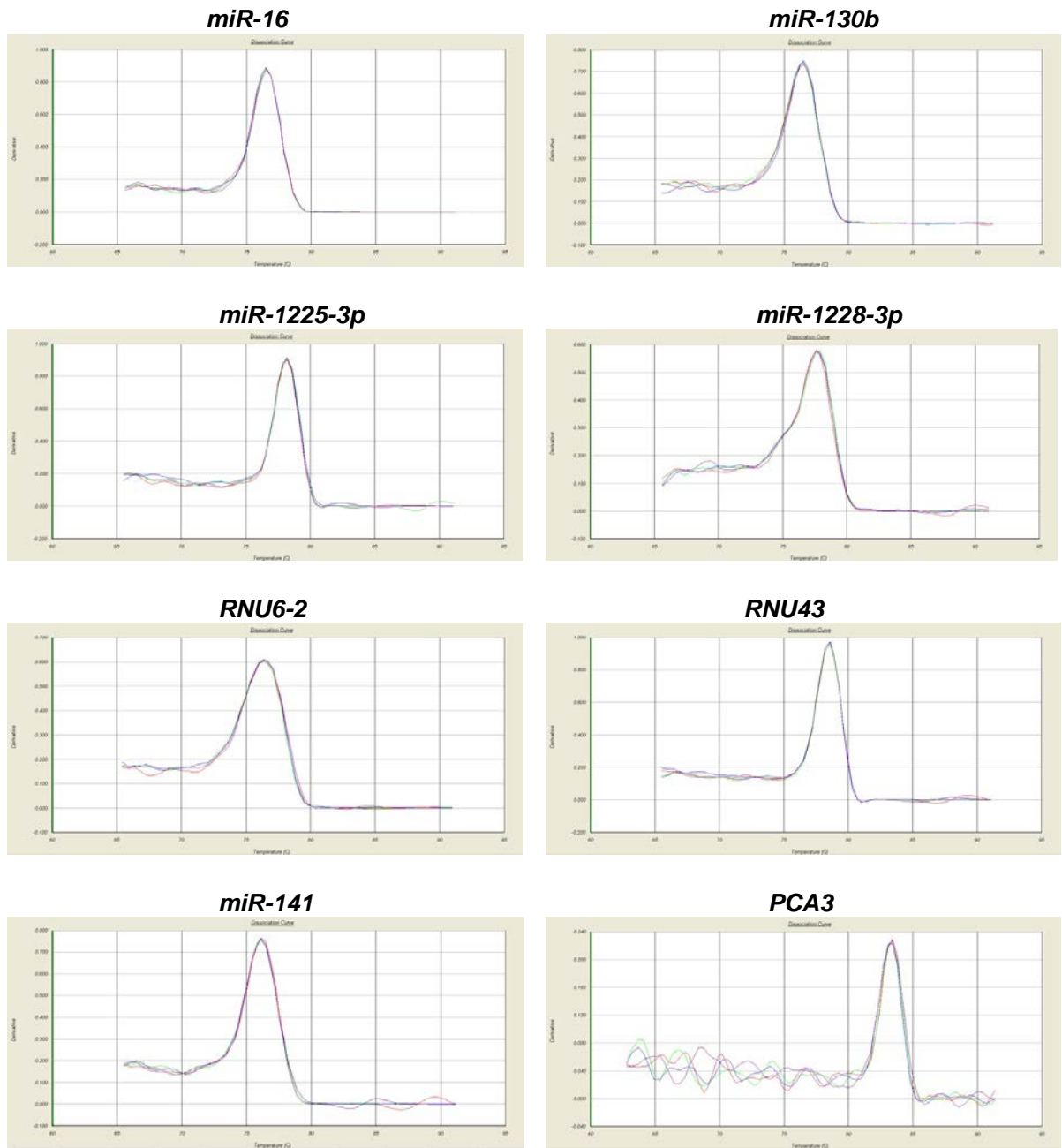


Fig. S1

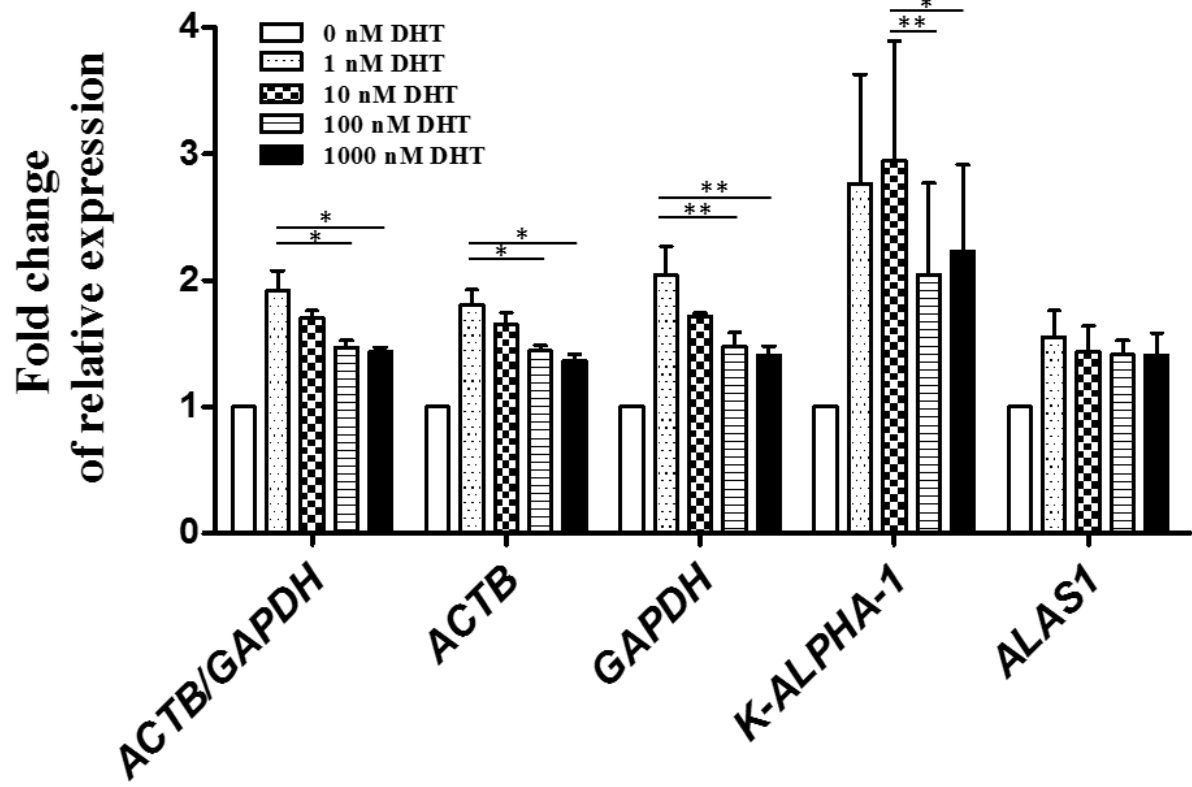


Fig. S2

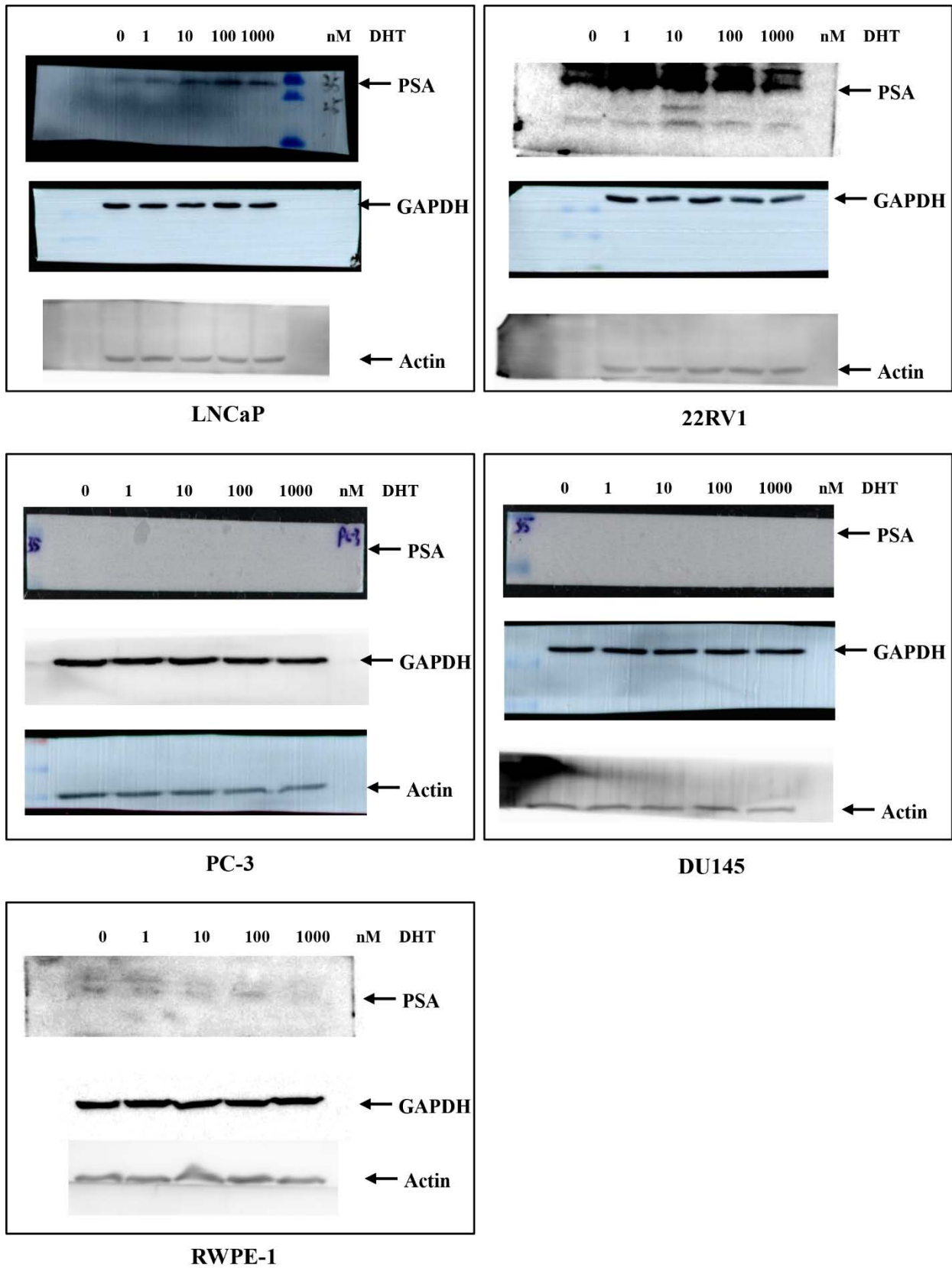


Fig. S3

Supplementary References

1. Liu, L. L. *et al.* Identification of valid reference genes for the normalization of RT-qPCR expression studies in human breast cancer cell lines treated with and without transient transfection. *PLoS One.* **10**, e0117058 (2015).
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6. De Spiegelaere, W. *et al.* Elimination of amplification artifacts in real-time reverse transcription PCR using laser capture microdissected samples. *Anal. Biochem.* **382(1)**, 72-74 (2008).