

# APPENDIXES

## Supplemental Materials and Methods

### In Gel Protein Digestion and MALDI-TOF Mass Spectrometry Analysis

Protein bands were excised from the gels, washed with 50 mM ammonium bicarbonate (Fluka Chemie AG), MS-grade acetonitrile (Sigma) (1:1, v/v), washed once with pure acetonitrile, and air-dried. Dried spots were digested for 18 h at 37 °C in 20 µl of 5 mM ammonium bicarbonate and 12 ng/µl sequencing grade modified trypsin (Promega). After digestion, 0.6 µl were spotted onto a matrix PAC target (Prespotted AnchorChip 96, set for proteomics, Bruker Daltonics) and air-dried. Spots were washed with 0.6 µl of 70% ethanol, 0.1% trifluoroacetic acid. Mass spectra were acquired on an Ultraflex MALDI-TOF/TOF mass spectrometer (Bruker Daltonics) in reflectron, positive mode in the mass range of 700–3500 m/z. Spectra were externally calibrated by using a combination of standards prespotted on the target. MS spectra were analyzed with FlexAnalysis (version 2.4, Bruker Daltonics) using default parameters and manually revised. Protein identification was performed using MASCOT search engine (version 2.2.1) by searching in *B. pertussis* Tohama I NCBI protein database (3,425 protein entries, release date: 2001/11/06). Search parameters were for peptide mass fingerprint analysis were: enzyme, trypsin; missed cleavage, 1; peptide tolerance, 100 ppm; variable modifications, methionine and tryptophane oxidation; mass values, [M+H]<sup>+</sup> ions and monoisotopic.

**Table S1. Comparative proteomics data: proteins quantified in OMV from BP536 and BP537 strains.**

With LC-MS/MS analysis, 389 proteins were identified and quantified in three biological replicates of Bvg+ (BP536) and Bvg- (BP537) OMV. For each protein, accession number, proteomic data and protein annotation from *B. pertussis* Tohama I database are listed. In addition, the average of the coverage (n=3 technical replicates), number of detected peptides and peak area with standard deviation are retrieved from PEAKS version 8. Subsequently, theoretical molecular weight (MW) is given with the calculated percentage for each protein in the total sample as described in Materials and Methods. Finally, the fold increase of each protein amount in Bvg+ vs. Bvg- is reported with the PSORTb and DOLOP prediction of subcellular localization.

**FIG. S1. Peptide mass fingerprinting (PMF).** OMV from Bvg+ (lane 1) and Bvg- (lane 2) *B. pertussis* strains were resolved by SDS-PAGE. The most abundant protein band (highlighted in red) was excised, destained and digested with trypsin for PMF identification. Outer membrane porin protein BP0840 was identified with sequence coverage of 27%. MALDI spectrum was reported. For each identified peptide the following information were reported: start-end amino acid position of the identified peptide within the protein sequence; experimental  $m/z$ ; observed  $m/z$ ; calculated mass; difference (error) between the experimental and calculated mass (delta); number of missed cleavage sites; Mowse score from MASCOT; expectation value for the peptide match; MASCOT rank of the peptide match; uniqueness of peptide sequence in the database; amino acid peptide sequence, list on  $m/z$  loaded during the search.

**FIG. S2. Autotransporters sequence coverage of identification.** Sequence coverage of the five autotransporters selected as interesting vaccine candidates: percentage and graphical representation as obtained with Peaks 8 software analyzing the third biological replicate as a representative of the three experiments.

Figure S1

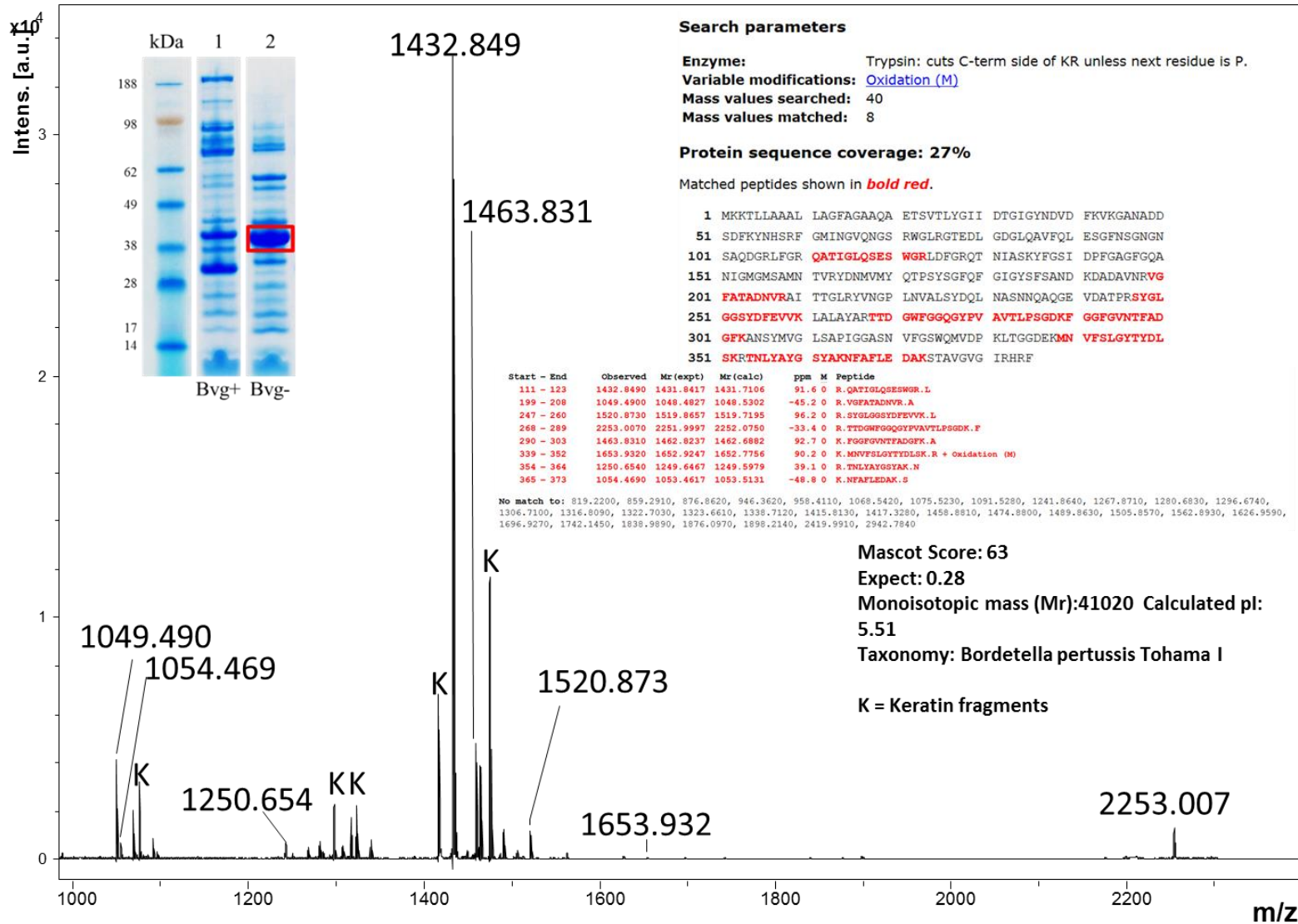







Figure S2

Accession	Protein name [Bordetella pertussis Tohama I]	Sequence coverage	%
NP_882013.1	<b>BrkA</b> autotransporter		85
NP_879104.1	<b>SphB1</b> autotransporter subtilisin-like protease		69
NP_879893.1	<b>BipA</b> outer membrane ligand binding protein		81
NP_880953.1	<b>Vag8</b> autotransporter		87
NP_879974.1	<b>TcfA</b> tracheal colonization factor		82