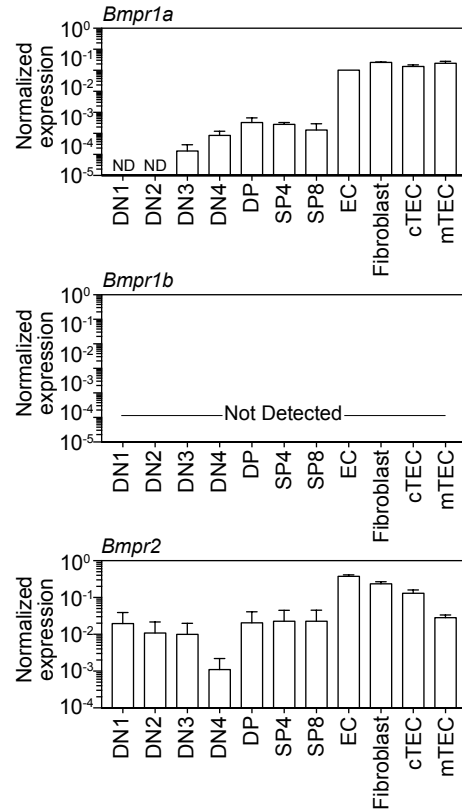


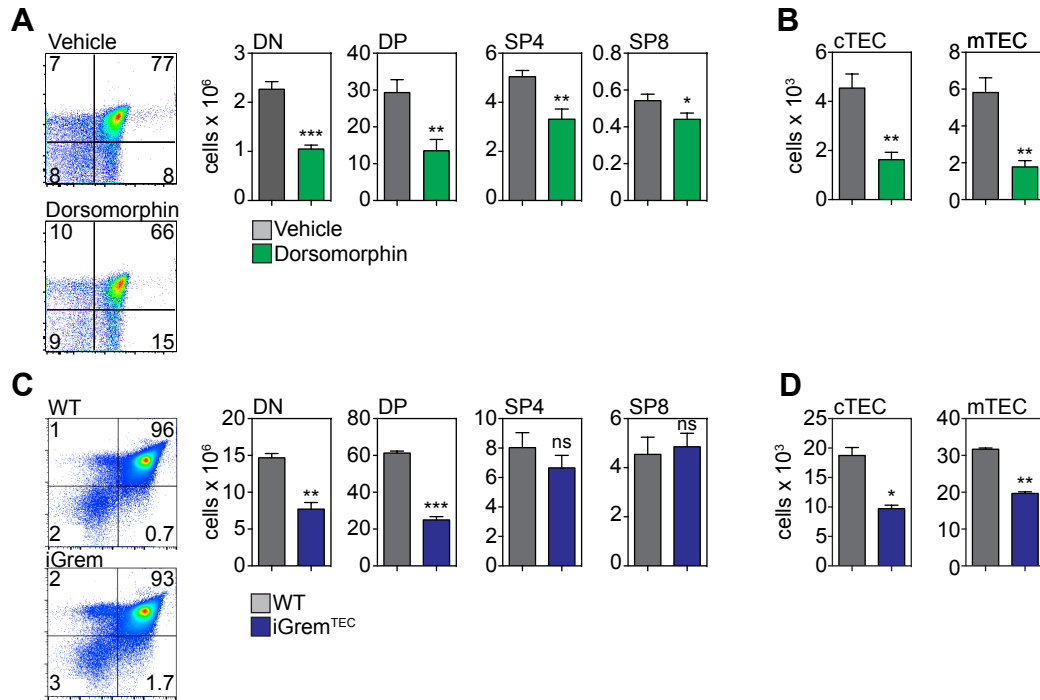
Supplementary Fig. 1: Thymic response to damage

(A) 6 week old female C57BL/6 mice were treated with PBS (n=10), Dexamethasone (Dex, 50mg/kg ip on day 0, n=10), cyclophosphamide (Cyclo, 100mg/kg/day ip on days -1 and 0, n=10) or SL-TBI (550 cGy on day 0, n=10). On day 4 absolute number of CD45⁺ and CD45⁻ cells were calculated and compared to untreated controls. (B) 6 week old female C57BL/6 mice were treated with SL-TBI (550 cGy on day 0) and thymic cellularity was measured at the indicated timepoints (n=5-16). (C) Graph of total thymus cellularity (right axis) and BMP4 measured by ELISA seven days after targeted thymic-irradiation (200, 400 or 800 cGy). Graphs represent combined mean \pm SEM of at least 2 independent experiments. *, p<0.05; **, p<0.01, ***, p<0.001.



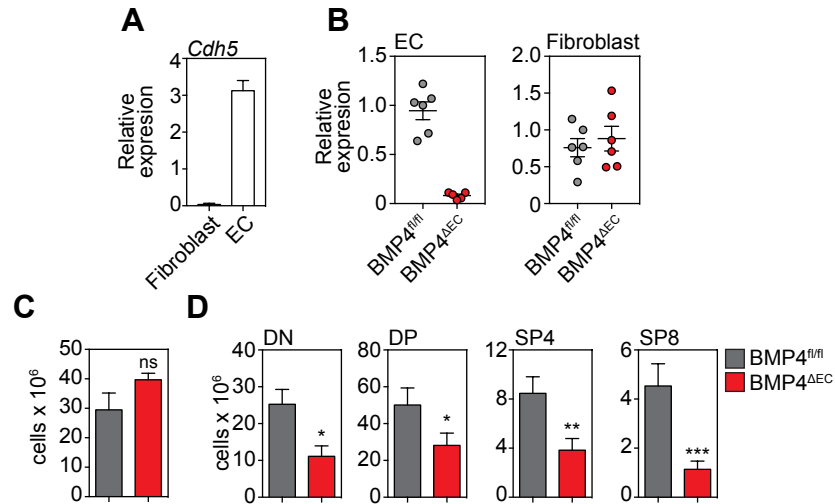
Supplementary Fig. 2: Distribution of BMP receptor subunits on thymic cell populations

(A) DN1 (CD3⁻CD4⁻CD8⁻CD44⁺CD25⁻), DN2 (CD3⁻CD4⁻CD8⁻CD44⁺CD25⁺), DN3 (CD3⁻CD4⁻CD8⁻CD44⁻CD25⁺), DN4 (CD3⁻CD4⁻CD8⁻CD44⁺CD25⁻), DP (CD4⁺CD8⁺), SP4 (CD3⁺CD8⁻CD4⁺), SP8 (CD3⁺CD4⁻CD8⁺), DCs (CD11c⁺MHCII⁺), fibroblasts (CD45⁻EpCAM⁻MHCII⁻CD31⁻VE-Cadherin⁻PDGFR α ⁺), ECs (CD45⁻EpCAM⁻MHCII⁻CD31⁺VE-Cadherin⁺), cTECs (CD45⁻EpCAM⁺MHCII⁺Ly51^{hi}UEA-1^{lo}) and mTECs (CD45⁻EpCAM⁺MHCII⁺Ly51^{lo}UEA-1^{hi}) were FACS purified from untreated 6 week-old female C57BL/6 mice and expression of the BMPR subunits *Bmpr1a*, *Bmpr1b*, and *Bmpr2* was measured by qPCR.



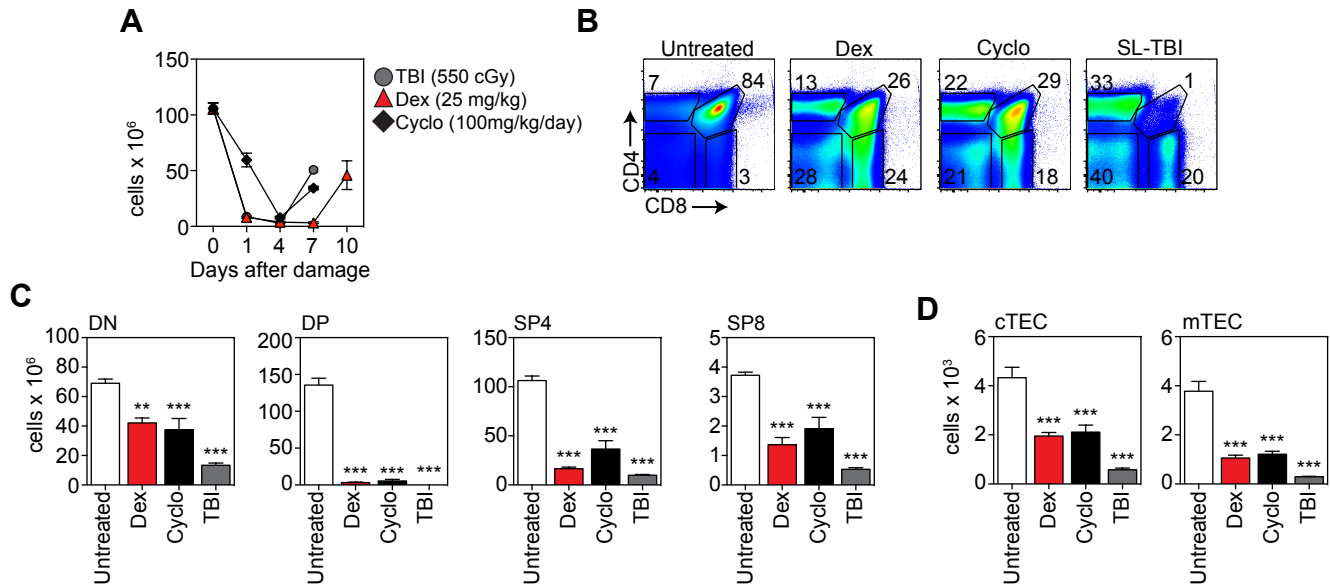
Supplementary Fig. 3: Inhibition of BMP signaling abrogates endogenous thymic regeneration

(A-B) 6 wo C57BL/6 mice were administered with the BMP type I receptor inhibitor Dorsomorphin dihydrochloride (12.5mg/kg) i.p. at day -1 before SL-TBI (550cGy single dose) and twice daily after SL-TBI. Thymus was harvested on day 7 and absolute number of (A) DN, DP, SP4, and SP8 as well as (B) cTEC and mTEC was assessed by flow cytometry. (C-D) iGremlin::K5-CreER (Grem^{iTEC}) mice were generated and tamoxifen was administered on days -1, 0 and +1 surrounding SL-TBI (550cGy). Thymus was harvested on day 9 and absolute number of (c) DN, DP, SP4, and SP8 as well as (D) cTEC and mTEC was assessed by flow cytometry. *, p<0.05; **, p<0.01, ***, p<0.001.



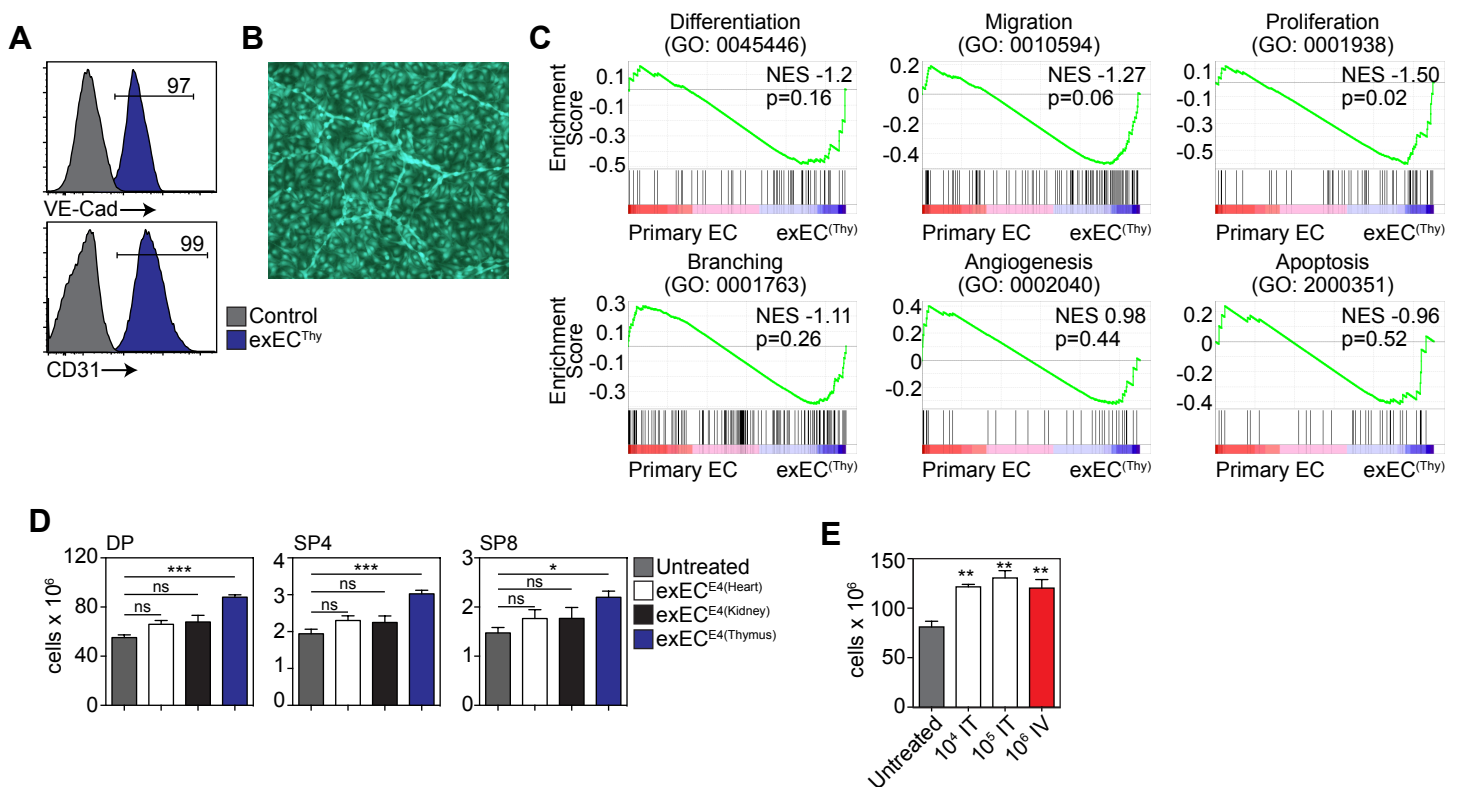
Supplementary Fig. 4: Deletion of BMP4 in ECs abrogates endogenous thymic regeneration

(A) Fibroblasts or ECs were FACS purified from from untreated 6 week-old female C57BL/6 mice and expression of *Cdh5* (VE-Cadherin) was measured by qPCR. (B-D) BMP4^{fl/fl} mice were crossed with *Cdh5*-CreERT2 mice. (B-C) Tamoxifen was administered over 5 days to induce deletion of BMP4 in ECs. (B) Expression of *Bmp4* was assessed in FACS purified ECs and fibroblasts. (C) Total thymus cellularity. (D) Tamoxifen was administered on days -2, -1, 0, 1 and 2 surrounding SL-TBI (550cGy). Thymus was harvested on day 7 and absolute number of (DN, DP, SP4, and SP8) were assessed by flow cytometry. Graphs represent combined mean ± SEM. *, p<0.05; **, p<0.01, ***, p<0.001.



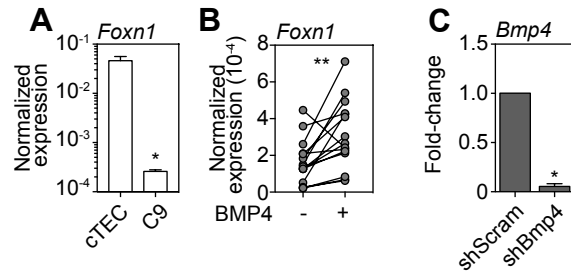
Supplementary Fig. 5: Damage response in the thymus to corticosteroids, chemotherapy and TBI

6 week old female C57BL/6 mice were treated with PBS (n=10), Dexamethasone (Dex, 50mg/kg ip on day 0, n=10), cyclophosphamide (Cyclo, 100mg/kg/day ip on days -1 and 0, n=10) or SL-TBI (550 cGy on day 0, n=10). On day 4, mice were perfused with 25 μ g anti-VE-cadherin antibody (BV13) conjugated to Alexa 647 and thymus harvested. **(A)** Total thymic cellularity at day 1, 4, 7, 10, and 14. **(B)** Concatenated flow cytometric plots detailing the proportion of thymocyte subsets. **(C-D)** Absolute number of **(C)** DN, DP, SP4, and SP8 as well as **(D)** cTEC and mTEC were assessed by flow cytometry. Graphs represent combined mean \pm SEM. *, p<0.05; **, p<0.01, ***, p<0.001.



Supplementary Fig. 6: exECs can be propagated ex vivo and maintain an EC phenotype

ECs were FACS sorted from the thymus, heart or kidney based on expression of VE-cadherin and transduced with the viral gene E4ORF1. **(A)** Expression of VE-Cadherin and CD31 on exECs derived from thymus. **(B)** Phase contrast image of exECs derived from thymus. **(C)** Microarray analysis was performed on freshly isolated thymic ECs and on exEC^(Thymus). GSEA analysis was performed looking signatures for EC Differentiation (GO: 0045446), EC Migration (GO: 0010594), EC Proliferation (GO: 0001938), EC Branching (0001763), Angiogenesis (GO: 0002040), and EC Apoptosis (2000351). **(D)** In order to model immune injury we exposed 6-8 weeks old C57/B6 mice to a single dose of sublethal TBI (550cGy) and 1 x 10⁶ exEC were administered iv at day 3 after SL-TBI (n=10-15/group). Thymus was harvested on day 9 and absolute number of DP, SP4, and SP8 were assessed by flow cytometry. **(E)** On day 3 after SL-TBI, exEC^(Thymus) were transplanted into mice either IV (3 x 10⁶ cells), or via ultrasound guided intrathymic injection (3 x 10⁴ or 3 x 10⁵ cells). Total thymus cellularity was measured on day 9. Graphs represent combined mean ± SEM. *, p<0.05; **, p<0.01, ***, p<0.001.



Supplementary Fig. 7: Validating methods of inducing *Foxn1* and silencing *Bmp4*

(A) Expression of *Foxn1* was measured by qPCR in freshly isolated FACS purified cTECs or the C9 (cTEC) cell line. (B) C9 cells were incubated for 24 hours with recombinant BMP4 (100ng/ml) and *Foxn1* expression was measured by qPCR. (C) Expression of *Bmp4*, measured by qPCR in thymus-derived exEC transduced to express either a *Bmp4* shRNA (shBMP4) or scrambled (shScram) control.