

**Figure S1. UV exposure causes formation of H4K20me2 foci that are independent of SETD8.** (A) DICER does not change H4K14ac distribution at chromatin. Immunofluorescence images showing distribution of H4K14ac in U2OS 2–6–3 cells containing a chromatin array tethered with either mCherry-LacR or mCherry-LacR-DICER. Bars, 5  $\mu$ m. (B) UV exposure results in colocalization of H4K20me2 with ZRF1 foci. Immunofluorescence images showing nuclear distribution of H4K20me2 in UV-unexposed and UV-exposed U2OS cells expressing mCherry-ZRF1. Bars, 25  $\mu$ m. (C) There is no global increase of H4K20me2 levels after UV damage. Western blots showing levels of the indicated proteins in whole-cell extracts at various time points after UV exposure. Ponceau is used as a loading control. (D) Increasing UV dose leads to an increase in H4K20me2 levels. Western blot showing levels of H4K20me2 in the chromatin fraction 2 h after UV exposure at the indicated dose. Ponceau is used as a loading control. (E) Western blot showing the siRNA-mediated DICER knockdown. Ponceau is used as a loading control. (F) MMSET is not recruited to a control LacO array. Immunofluorescence images showing distribution of EGFP-MMSET in undamaged cells with mCherry-LacR arrays. Bar, 5  $\mu$ m. (G) DICER does not recruit SETD8 to chromatin. Immunofluorescence images showing distribution of EGFP-SETD8 in cells with DICER-LacR–tethered arrays in both UV-unexposed and exposed cells. Mild recruitment was observed in 3 out of 20 cells in both conditions. (H) Immunofluorescence images showing H4K20me2 distribution in U2OS 2–6–3 cells, expressing mCherry-LacR-SETD8 (SETD8-LacR; left). The LacO array is visualized by binding of SETD8-LacR. Bars, 5  $\mu$ m. The graph on the right shows a quantification of the number of arrays showing an H4K20me2 mark in unexposed cells (–UV) and 2 h after UV exposure (+UV). Colocalization was measured from three independent experiments, counting 30 cells per experiment. (I and J) Western blots showing the esiRNA-mediated knockdowns of the indicated proteins. Ponceau is used as a loading control. (K) SETD8 is not required for DICER-dependent setting of H4K20me2. The graph shows a quantification of the number of DICER-LacR arrays showing an H4K20me2 mark in cell lines with the indicated esiRNA-mediated knockdowns 2 h after UV exposure (+UV; mean  $\pm$  SEM). Colocalization was measured from three independent experiments, with 30 cells per experiment.

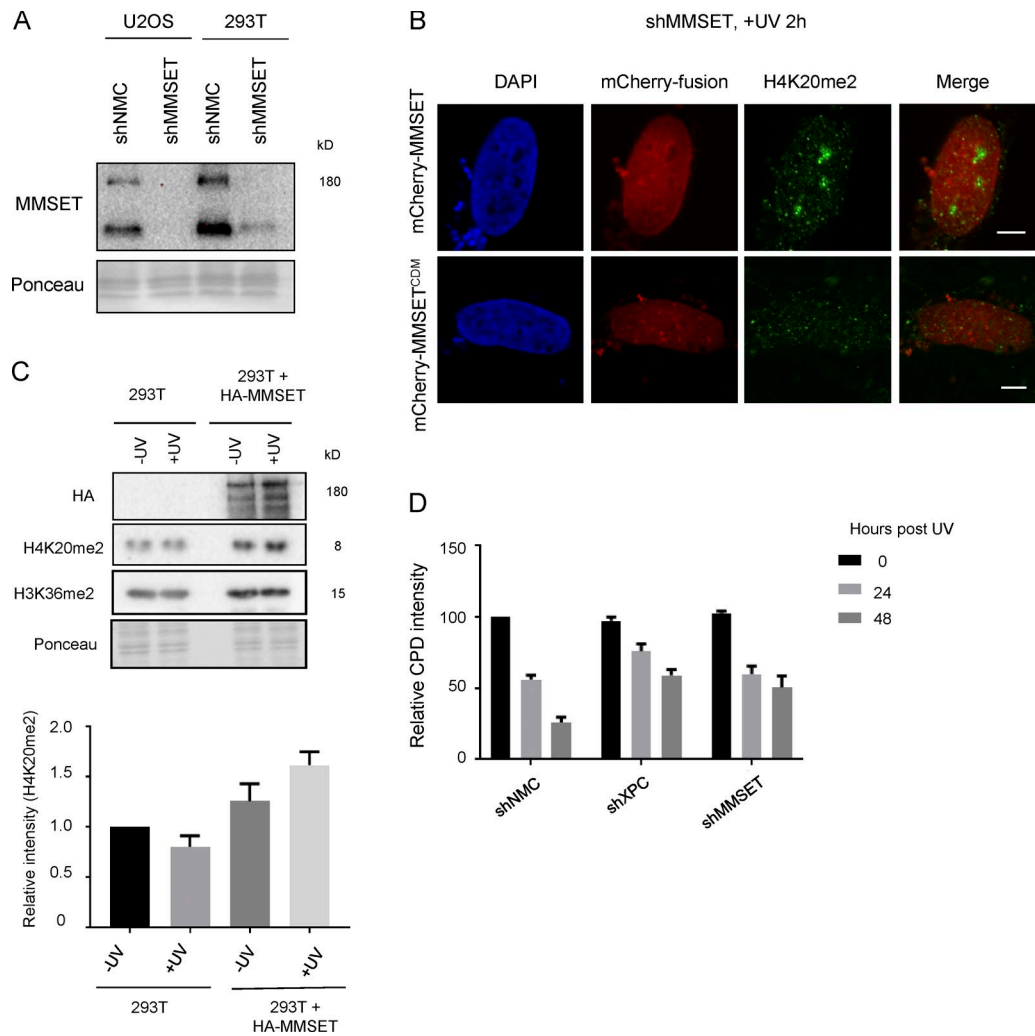
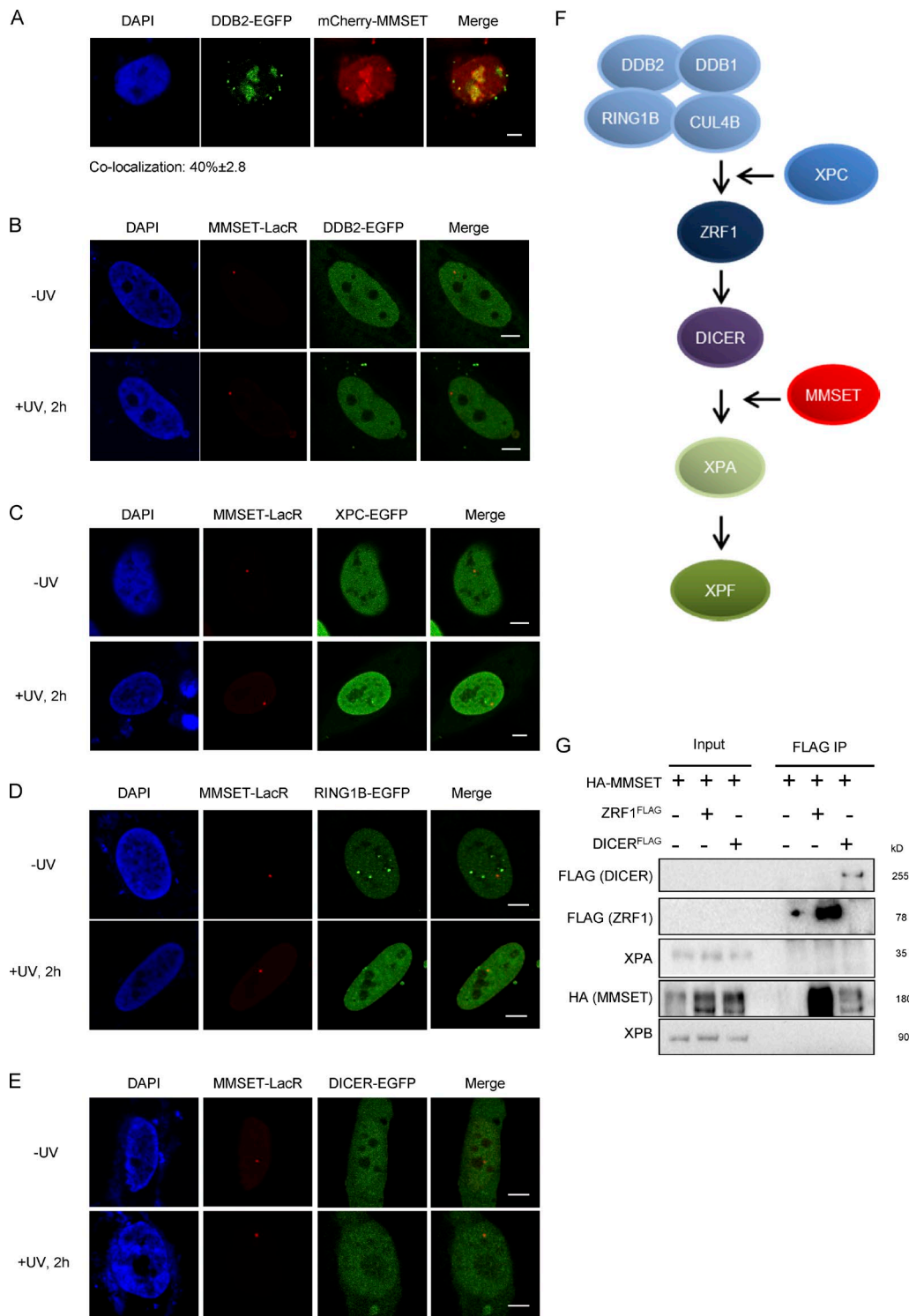


Figure S2. **MMSET interacts with ZRF1 and DICER and is required for CPD removal.** (A) Western blots showing the levels of MMSET in stable shRNA-mediated knockdown cell lines generated in the indicated cells. Ponceau was used as a loading control. (B) MMSET can rescue formation of H4K20me2 foci. Immunofluorescence images showing the distribution of H4K20me2 in shMMSET U2OS cells 2 h after UV exposure. Cells were transfected with either mCherry-MMSET or mCherry-MMSET<sup>CDM</sup>. Bars, 5  $\mu$ m. (C) Overexpression of MMSET can lead to an increase in H4K20me2 levels. Western blot showing the levels of the indicated proteins in cells transfected with either empty plasmid or HA-MMSET in both UV conditions (top). Ponceau was used as a loading control. The graph on the bottom shows the relative band intensity for H4K20me2 in the indicated conditions. Intensity was measured from three independent experiments. (D) The graph shows the relative CPD intensity at various time points after UV exposure in U2OS cells with stable knockdowns of the indicated proteins (mean  $\pm$  SEM). CPD intensity was measured from three independent experiments, with intensities quantified for 500–1,000 cells per experiment.



**Figure S3. MMSET is required for NER and interacts with the NER machinery.** (A) MMSET is recruited to sites of UV damage. Immunofluorescence images showing distribution of DDB2-EGFP and mCherry-MMSET in U2OS cells subjected to damage through a 3- $\mu$ m micropore membrane. The damage sites are marked by DDB2-EGFP. MMSET is observed at  $\approx$ 40% of lesions 30 min after UV exposure. Bar, 5  $\mu$ m. (B) DDB2 is not recruited to MMSET-LacR-tethered arrays. Immunofluorescence images showing distribution of DDB2-EGFP in cells with MMSET-LacR-tethered arrays in UV-unexposed and exposed cells. No recruitment was seen in 20 out of 20 cells. Bars, 5  $\mu$ m. (C) XPC is not recruited to MMSET-LacR-tethered arrays. Immunofluorescence images showing distribution of XPC-EGFP in cells with MMSET-LacR-tethered arrays in UV unexposed and exposed cells. No recruitment was seen in 20 out of 20 cells. Bars, 5  $\mu$ m. (D) RING1B is not recruited to MMSET-LacR-tethered arrays. Immunofluorescence images showing distribution of RING1B-EGFP in cells with MMSET-LacR-tethered arrays in UV unexposed and exposed cells. No recruitment was seen in 20 out of 20 cells. Bars, 5  $\mu$ m. (E) DICER is not recruited to MMSET-LacR-tethered arrays. Immunofluorescence images showing distribution of DICER-EGFP in cells with MMSET-LacR-tethered arrays in UV-unexposed and exposed cells. No recruitment was seen in 20 out of 20 cells. Bars, 5  $\mu$ m. (F) Schematic showing sequential recruitment of NER factors. The figure shows the hierarchy of recruitment for specific NER factors and indicates a possible position for MMSET in the hierarchy. (G) Both DICER and ZRF1 interact with MMSET. Western blot shows levels of select proteins in FLAG purifications performed from cells transfected with the indicated plasmids.

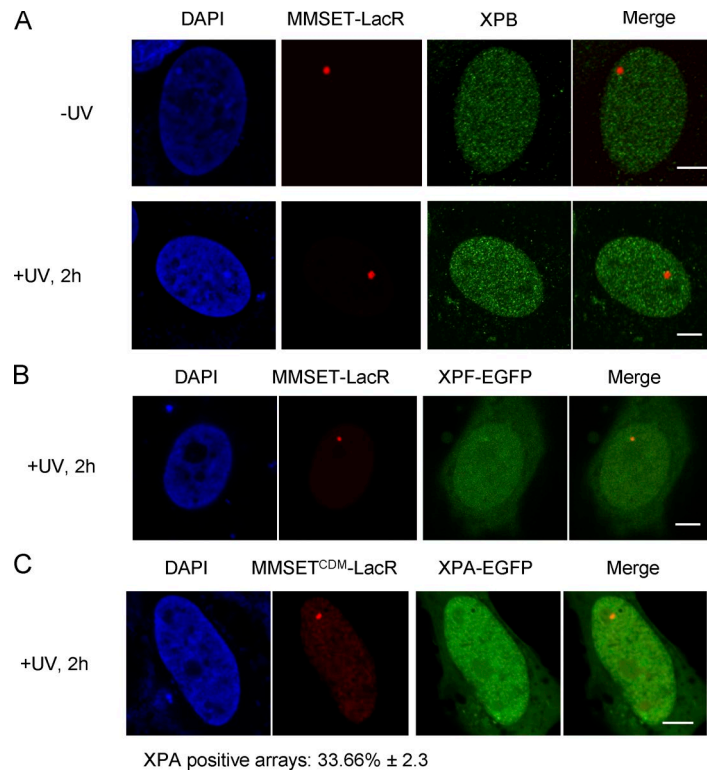


Figure S4. **MMSET-LacR recruits late NER factors to chromatin.** (A) XPB is not recruited to MMSET-LacR-tethered arrays. Immunofluorescence images showing distribution of XPB in cells with MMSET-LacR-tethered arrays, in UV-exposed and unexposed cells. No recruitment was seen in 20 out of 20 cells in both conditions. Bars, 5  $\mu$ m. (B) XPF is weakly recruited to MMSET-LacR-tethered arrays. Immunofluorescence images showing distribution of XPF-EGFP in cells with MMSET-LacR-tethered arrays, in UV exposed cells. Weak recruitment was seen in 15 out of 20 cells. Bar, 5  $\mu$ m. (C) MMSET<sup>CDM</sup>-LacR is unable to recruit XPA. Immunofluorescence images showing the distribution of XPA-EGFP in cells with MMSET<sup>CDM</sup>-LacR-tethered arrays 2 h after UV exposure. Bar, 5  $\mu$ m.

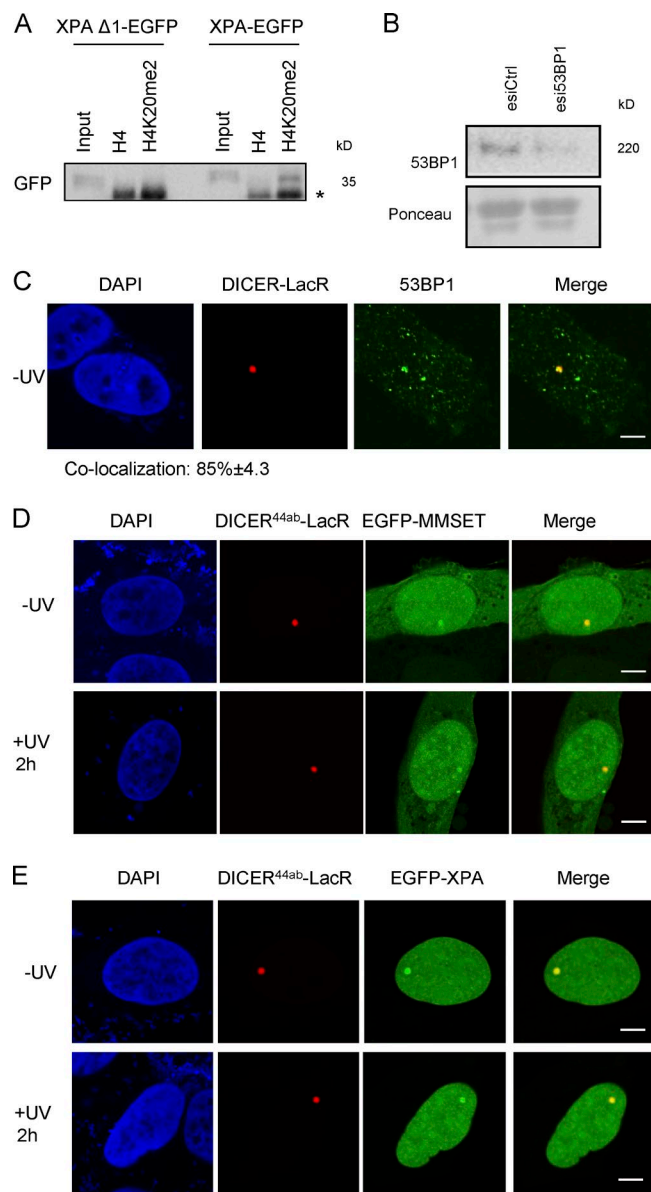


Figure S5. **XPA recruitment to H4K20me2 requires presence of RPA, but is independent of the endoribonuclease activity of DICER.** (A) The RPA32 domain is essential for XPA recruitment to H4K20me2. Pull-downs with H4 and H4K20me2 peptides and protein extracts expressing tagged full-length XPA (XPA-EGFP) or an XPA fusion protein lacking the RPA32 domain (XPA  $\Delta$ 1-EGFP). The asterisk denotes an unspecific band appearing when using the GFP antibody. (B) Western blot showing esiRNA mediated 53BP1 knockdown. Ponceau was used as a loading control. (C) 53BP1 colocalizes with DICER-LacR-tethered arrays in cells unexposed to UV. Bar, 5  $\mu$ m. (D) DICER<sup>44ab</sup> can recruit MMSET to chromatin. Immunofluorescence images showing distribution of EGFP-MMSET in cells with DICER<sup>44ab</sup>-LacR-tethered arrays in both UV-unexposed and exposed cells. Bars, 5  $\mu$ m. (E) DICER<sup>44ab</sup> can recruit XPA to chromatin. Immunofluorescence images showing distribution of EGFP-XPA in cells with DICER<sup>44ab</sup>-LacR tethered arrays, in both UV unexposed and exposed cells. Bars, 5  $\mu$ m.

Provided online are two tables in Excel. Tables S1 and S2 give further information on the plasmids used in this study, as well as the sequences of all si/esirRNAs used in the study.