Supplementary Materials

Genomic Analysis Revealed New Oncogenic Signatures in *TP53* **mutant Hepatocellular Carcinoma**

Venkatesh Kancherla¹, Samir Abdullazade¹, Matthias S Matter¹, Manuela Lanzafame¹, Luca Quagliata¹, Guglielmo Roma², Yujin Hoshida³, Luigi M Terracciano¹, Charlotte KY Ng^{1,4*} and Salvatore Piscuoglio^{1*}

***Correspondence to:**

Dr. Salvatore Piscuoglio [\(Salvatore.Piscuoglio@usb.ch\)](mailto:Salvatore.Piscuoglio@usb.ch)) and **Dr. Charlotte K Y Ng** [\(kiuyancharlotte.ng@usb.ch\),](mailto:kiuyancharlotte.ng@usb.ch)) Institute of Pathology, University Hospital Basel, Schoenbeinstrasse 40, 4031 Basel, Switzerland. Tel: +41613286874; Fax: +41612653194.

- **Supplementary Methods**
- **Supplementary Figures S1 – S4**
- **Supplementary Tables S1 – S5**

Supplementary Methods

Classification of *TP53* **somatic mutations**

TP53 mutations were stratified according to (i) the mutation type as single-nucleotide missense mutations (encompassing missense, and synonymous mutations affecting splice-region) or deleterious mutations (encompassing splice site, nonsense, in-frame, and frameshift mutations); (ii) whether the mutations were within or outside of the DNA-binding domain.

The effects of missense mutations and synonymous mutations affecting splice-regions were predicted using CHASM (Carter et al., 2009), FATHMM (Shihab et al., 2013), VEST (Carter et al., 2013), MutationTaster (Schwarz et al., 2010) and PolyPhen-2 (Adzhubei et al., 2010). Annotation of ClinVar (Landrum et al., 2016), 1000 Genomes (Genomes Project et al., 2015), ESP6500 (Tennessen et al., 2012), ExAC (Lek et al., 2016) and COSMIC (Forbes et al., 2011) allele frequencies were retrieved from cravat.us (Douville et al., 2013). The two synonymous mutations affecting splice-region were included as missense mutations in the current study as both were predicted to be disease causing by MutationTaster (Schwarz et al., 2010).

Oncogenic signatures

Oncogenic signature ("oncosign") classification was performed using the algorithm described by Ciriello *et al.* (Ciriello et al., 2013). The approach to select genomic features as 'selected functional elements' (SFEs) input data was adopted from Ciriello *et al.* (Ciriello et al., 2013). Specifically, from the 76 significantly mutated genes defined by MutSigCV obtained from the cbioportal (Gao et al., 2013), we selected 29 that have previously been reported as cancer genes by any of Cancer Gene Census (Futreal et al., 2004), Kandoth *et al.* (Kandoth et al., 2013), Lawrence *et al.* (Lawrence et al., 2014) or Fujimoto *et al.* (Fujimoto et al., 2012). Somatic mutation data were coded as a binary gene-by-sample matrix, with 1 indicating the presence of one (or more) non-synonymous somatic mutation in a given gene in a given sample. GISTIC peaks were also selected as SFEs, and included 27 amplification and 34 deletion peaks, and were coded as a binary peak-by-sample matrix, with 1 indicating the presence of at least one gene harboring high-level gain/ amplification (copy number state '2' for amplification peaks) or homozygous deletion (copy number state '-2' for deletion peaks). The oncosign algorithm was run with a maximum depth of 1.

Pathway analysis

For Ingenuity Pathway Analysis (IPA), genes of interest were mapped to pathways and networks available in the Ingenuity database and ranked by corrected *P* value (Benjamini– Hochberg multiple correction) as previously described (Piscuoglio et al., 2014;Martelotto et al., 2015). *P<*0.001 was considered significant.

Statistical Analysis

For statistical analyses comparing copy number profiles, gene-level copy number states (i.e. amplification/high-level gains, gains, losses and homozygous deletions) were compared using Fisher's exact tests corrected for multiple comparisons using the Benjamini-Hochberg method (Piscuoglio et al., 2014). To define genes up-regulated when gained or amplified and genes down-regulated when lost, we applied Mann-Whitney U tests using categorical copy number states (i.e. gain vs. no gain, loss vs. no loss) as the grouping variable and the expression of genes as the dependent variable corrected for multiple comparisons using the Benjamini-Hochberg method.

Supplementary References

- Adzhubei, I.A., Schmidt, S., Peshkin, L., Ramensky, V.E., Gerasimova, A., Bork, P., et al. (2010). A method and server for predicting damaging missense mutations. *Nat Methods* 7**,** 248-249.
- Carter, H., Chen, S., Isik, L., Tyekucheva, S., Velculescu, V.E., Kinzler, K.W., et al. (2009). Cancer-specific high-throughput annotation of somatic mutations: computational prediction of driver missense mutations. *Cancer Res* 69**,** 6660-6667.
- Carter, H., Douville, C., Stenson, P.D., Cooper, D.N., and Karchin, R. (2013). Identifying Mendelian disease genes with the variant effect scoring tool. *BMC Genomics* 14 Suppl 3**,** S3.
- Ciriello, G., Miller, M.L., Aksoy, B.A., Senbabaoglu, Y., Schultz, N., and Sander, C. (2013). Emerging landscape of oncogenic signatures across human cancers. *Nat Genet* 45**,** 1127-1133.
- Douville, C., Carter, H., Kim, R., Niknafs, N., Diekhans, M., Stenson, P.D., et al. (2013). CRAVAT: cancer-related analysis of variants toolkit. *Bioinformatics* 29**,** 647-648.
- Forbes, S.A., Bindal, N., Bamford, S., Cole, C., Kok, C.Y., Beare, D., et al. (2011). COSMIC: mining complete cancer genomes in the Catalogue of Somatic Mutations in Cancer. *Nucleic Acids Res* 39**,** D945-950.
- Fujimoto, A., Totoki, Y., Abe, T., Boroevich, K.A., Hosoda, F., Nguyen, H.H., et al. (2012). Whole-genome sequencing of liver cancers identifies etiological influences on mutation patterns and recurrent mutations in chromatin regulators. *Nat Genet* 44**,** 760- 764.
- Futreal, P.A., Coin, L., Marshall, M., Down, T., Hubbard, T., Wooster, R., et al. (2004). A census of human cancer genes. *Nat Rev Cancer* 4**,** 177-183.
- Gao, J., Aksoy, B.A., Dogrusoz, U., Dresdner, G., Gross, B., Sumer, S.O., et al. (2013). Integrative analysis of complex cancer genomics and clinical profiles using the cBioPortal. *Sci Signal* 6**,** pl1.
- Genomes Project, C., Auton, A., Brooks, L.D., Durbin, R.M., Garrison, E.P., Kang, H.M., et al. (2015). A global reference for human genetic variation. *Nature* 526**,** 68-74.
- Kandoth, C., Mclellan, M.D., Vandin, F., Ye, K., Niu, B., Lu, C., et al. (2013). Mutational landscape and significance across 12 major cancer types. *Nature* 502**,** 333-339.
- Landrum, M.J., Lee, J.M., Benson, M., Brown, G., Chao, C., Chitipiralla, S., et al. (2016). ClinVar: public archive of interpretations of clinically relevant variants. *Nucleic Acids Res* 44**,** D862-868.
- Lawrence, M.S., Stojanov, P., Mermel, C.H., Robinson, J.T., Garraway, L.A., Golub, T.R., et al. (2014). Discovery and saturation analysis of cancer genes across 21 tumour types. *Nature* 505**,** 495-501.
- Lek, M., Karczewski, K.J., Minikel, E.V., Samocha, K.E., Banks, E., Fennell, T., et al. (2016). Analysis of protein-coding genetic variation in 60,706 humans. *Nature* 536**,** 285-291.
- Martelotto, L.G., De Filippo, M.R., Ng, C.K., Natrajan, R., Fuhrmann, L., Cyrta, J., et al. (2015). Genomic landscape of adenoid cystic carcinoma of the breast. *J Pathol* 237**,** 179-189.
- Piscuoglio, S., Ng, C.K., Martelotto, L.G., Eberle, C.A., Cowell, C.F., Natrajan, R., et al. (2014). Integrative genomic and transcriptomic characterization of papillary carcinomas of the breast. *Mol Oncol* 8**,** 1588-1602.
- Schwarz, J.M., Rodelsperger, C., Schuelke, M., and Seelow, D. (2010). MutationTaster evaluates disease-causing potential of sequence alterations. *Nat Methods* 7**,** 575-576.
- Shihab, H.A., Gough, J., Cooper, D.N., Stenson, P.D., Barker, G.L., Edwards, K.J., et al. (2013). Predicting the functional, molecular, and phenotypic consequences of amino acid substitutions using hidden Markov models. *Hum Mutat* 34**,** 57-65.
- Tennessen, J.A., Bigham, A.W., O'connor, T.D., Fu, W., Kenny, E.E., Gravel, S., et al. (2012). Evolution and functional impact of rare coding variation from deep sequencing of human exomes. *Science* 337**,** 64-69.

Supplementary Figure S1: Number of mutations (**A**) and copy number altered genes (**B**) identified in hepatocellular carcinomas stratified according to the *TP53* mutation status. Statistical comparisons were performed using Mann-Whitney U tests. *P*<0.05 was considered statistically significant.

Supplementary Figure S2: Comparative genomic profiling of *TP53*-mutant and *TP53*-wildtype HCCs (**A-D**). Frequency plots and multi-Fisher's exact test comparisons of chromosomal gains and losses in *TP53*-mutant (top) and *TP53*-wild-type (middle) HCCs. The frequency of gains (purple bars) or losses (yellow bars) for each gene is plotted on the *y*-axis, according to their genomic position on the *x*-axis. Inverse Log₁₀values of the Fisher's exact test P values are plotted according to genomic location (*x*-axis) (bottom).

Supplementary Figure S3: Signaling pathways (left) and molecular and cellular functions (right) enriched among genes overexpressed when gained or downregulated when lost in the regions that showed differential CNA frequencies between cases with or without *TP53* mutations using Ingenuity Pathway Analysis (IPA). Log values of the Benjamini-Hochberg corrected *P* value are shown. Dashed lines indicate the significance cut-off ($P = 0.001$).

Supplementary Figure S4: Oncogenic signature subclasses were tested for robustness (**A**) upon removal of 5%, 10%, and 20% of the samples**.** Barplot shows the distribution of cases classified as S1, S2 or S3 based on transcriptomic classification among the four oncogenic signature classes (**B**). The distribution of mutational vs copy number 'selected functional elements' (SFEs) in *TP53*-mutant cases harboring missense (**C**) or deleterious (**D**) somatic *TP53* mutations. The shade of red is proportional to the number of samples for a given (x, y) position. Survival analysis of HCCs sub-classified based on the oncogenic signatures (**E**). Median survival for each group is indicated in parentheses. Statistical comparisons were performed using log-rank tests. *P* < 0.05 was considered statistically significant.

Supplementary Table S1: *In silico* prediction of mutation effect of missense and synonymous mutations affecting splice-regions.

Supplementary Table S2: Detailed list of the TCGA studies included for the comparison of *TP53* mutational spectrum.

Supplementary Table S3: Clinicopathologic features of the 373 HCCs from The Cancer Genome Atlas cohort.

* Patients may have multiple risk factors

** 3 cases were not evaluable.

*** 2 cases were not evaluable.

****1 cases were not evaluable.

*****6 cases were not possible to classified.

Supplementary Table S4: Analyses of *TP53* mutation status sub-divided according to the mutation type and clinicopathologic parameters in the 373 HCCs from The Cancer Genome Atlas cohort. Statistical comparisons were performed using Fisher's exact test or Chi-Squared test. P < 0.05 was considered statistically significant.

Supplementary Table S5: Univariate and multivariate analyses of OS and DFS of TP53-mutant HCCs with clinicopathologic and molecular features.

 \blacksquare

 \blacksquare

HR: Hazard Ratio