

Supplementary Information for “Massively parallel de novo protein design for targeted therapeutics”

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Figures

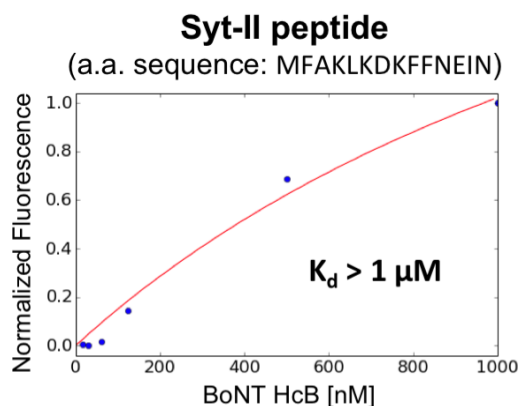


Figure 1. Yeast display titration of BoNT H_CB with a peptide fragment of Syt-II (*Rattus norvegicus*) on yeast display reveals low affinity for the 14 a.a. peptide fragment alone. Literature reported a K_d of ~40 nM for Syt-II towards H_CB; however, this is in presence of flanking regions (a.a. 1-61 and a GST-tag), which are known to be important for high affinity binding, but the details of its structural information is unknown. Without these flanking-elements present, we found that (yeast display system, see Methods) the affinity of Syt-II is drastically reduced to K_d >1 μM. A subset of the amino acids of this binding motif was used to guide the designs of the Bot binders.

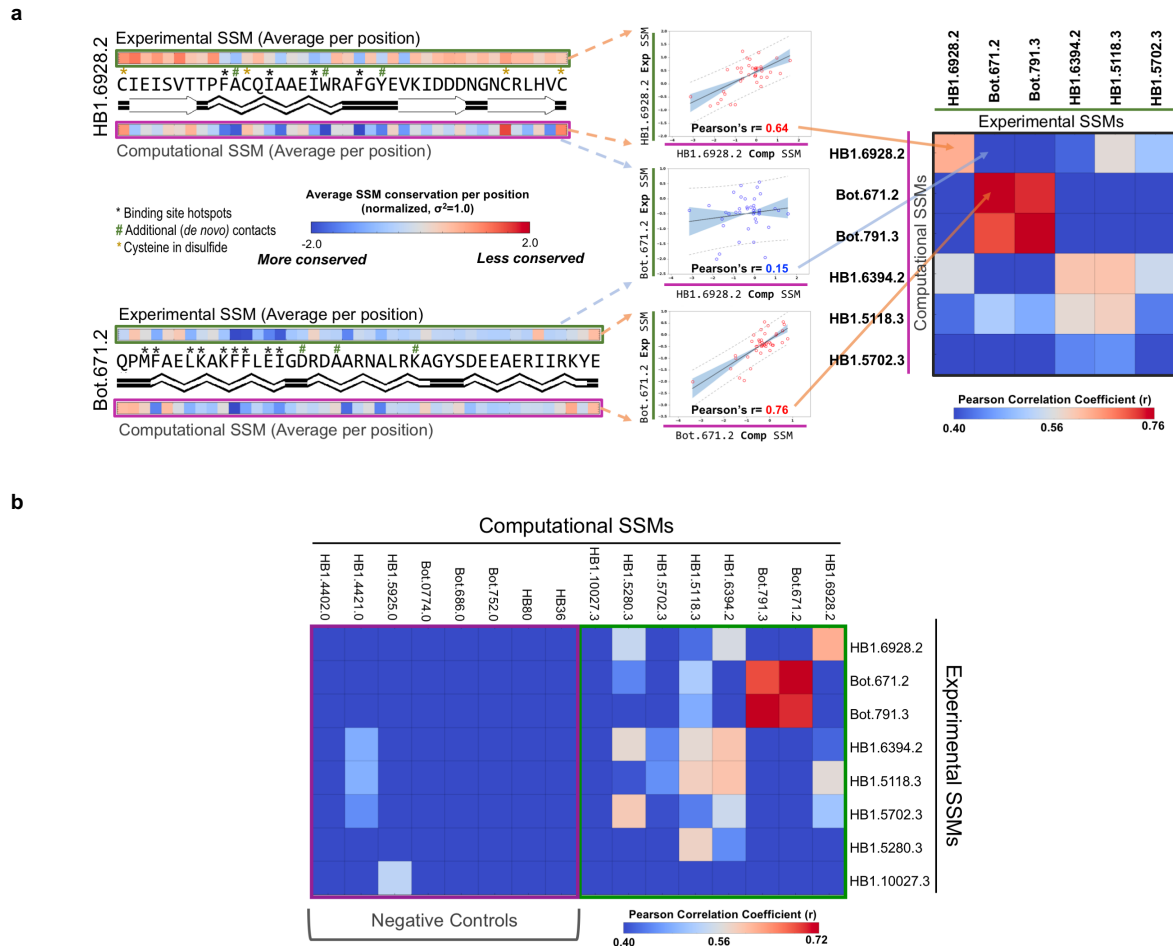


Figure 2. Correlation of experimental and Rosetta-predicted SSM calculations. a) The overall procedure used to calculate the experimental vs. computational (Rosetta) SSM correlation. For the experimental SSMs, each mutant was assigned to a binding category as in Extended Data Fig. 2, and a per-mutation score was calculated as the change in category compared with the designed (native) residue at that position (a positive score indicates a mutation improves binding). We then normalized these values for the whole dataset ($\sigma^2=1$) and calculated the average score of all mutations at each position. For the computational SSMs, we built computational models of each mutant and used them to calculate the change in monomeric and binding energy compared with the original design. We then normalized the monomeric and binding datasets separately ($\sigma^2=1$), summed them to compute an overall score for each mutation (for consistency, we made positive scores indicate improved binding), and again calculated the average of these scores at each position. Finally, we calculated the Pearson correlation between the per-position scores from the computational and experimental SSMs. When the size of the proteins differed, we just considered a 1-1 amino-acid correspondence starting from their first residues and ending at the length of the smallest protein in the pair. **b)** The results of the calculation described in “a” applied to 8 proteins included in the experiments and compared to all of its computational SSMs and 8 negative controls, including 2 natural proteins (previously reported HA binders HB36 and HB80) and 6 other designs chosen randomly. Most of the experimental data correlate better with the computational data for the same protein (i.e. HB1.6928.2, HB1.6394.2, HB1.5118.3 for HB1 designs, and Bot.671.2, Bot.791.3 for Bot designs) than with any other (unrelated) protein, and thus provides an indirect structural validation of the designed model.

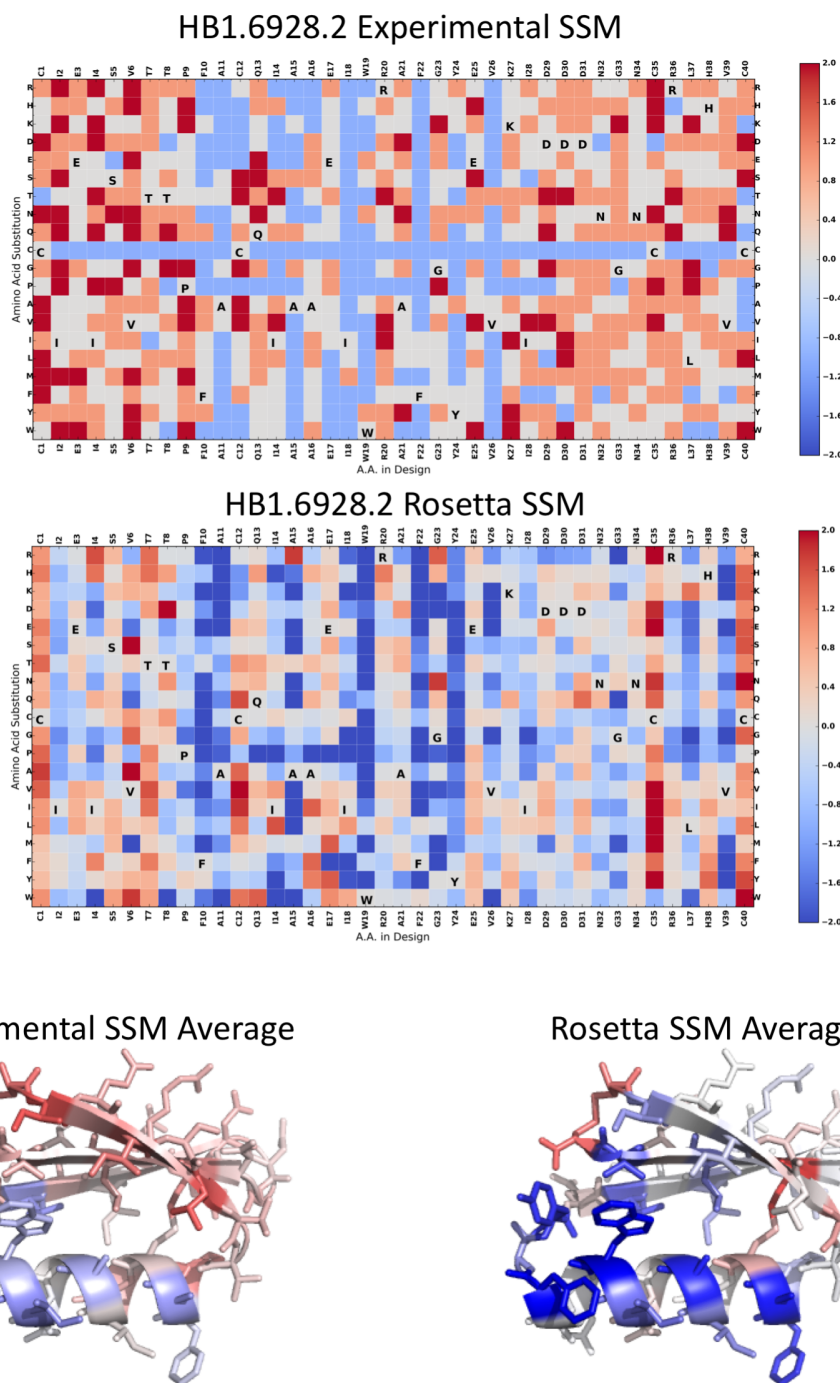
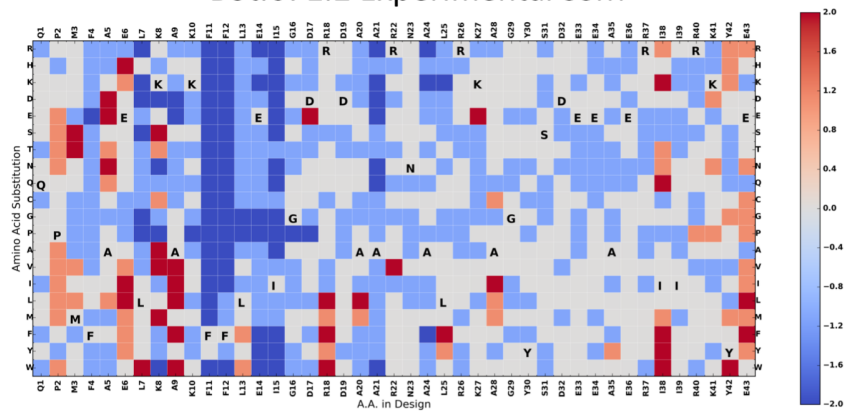
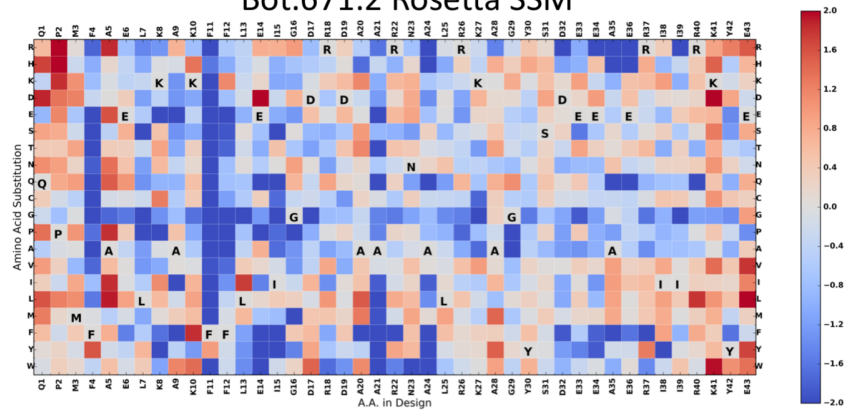


Figure 3a. Experimental and Rosetta-predicted SSM Heatmaps for HB1.6928.2, similar to Supplementary Information Figure 3a. Red indicates mutations that enhance binding, grey indicates mutations equivalent to the designed amino acid, and blue indicates mutations that reduce binding affinity. We show the average mutational effect for each position mapped onto the design model. Computational and experimental values were calculated as described in SI Figure 2. It can be observed there is a good agreement between the computational and experimental SSM, in particular in the core residues and hydrophobic hotspots in the binding helix.

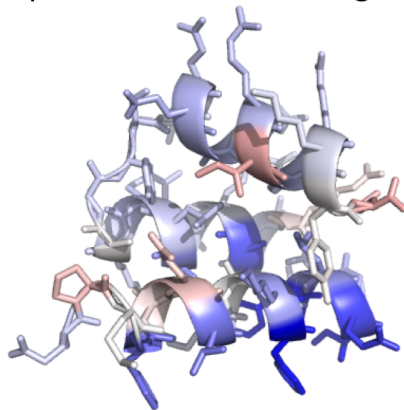
Bot.671.2 Experimental SSM



Bot.671.2 Rosetta SSM



Experimental SSM Average



Rosetta SSM Average

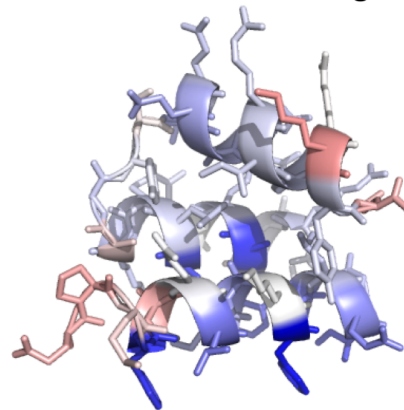
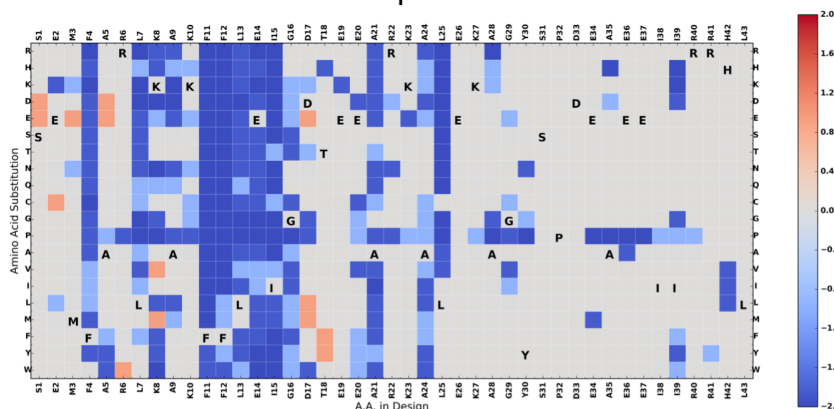
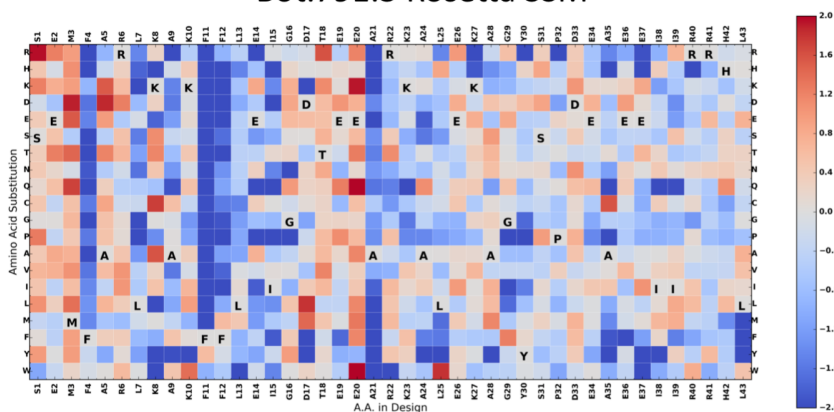


Figure 3b. Experimental and Rosetta-predicted SSM Heatmaps for Bot.671.2, similar to Supplementary Information Figure 3a. Red indicates mutations that enhance binding, grey indicates mutations equivalent to the designed amino acid, and blue indicates mutations that reduce binding affinity. We show the average mutational effect for each position mapped onto the design model. Computational and experimental values were calculated as described in SI Figure 2. It can be observed there is a good agreement between the computational and experimental SSM.

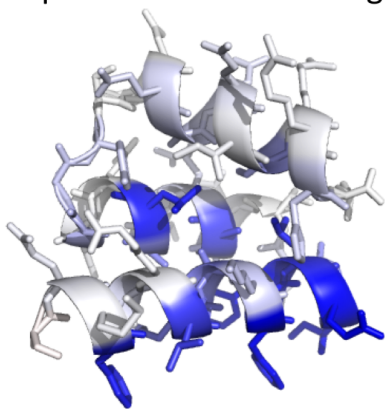
Bot.791.3 Experimental SSM



Bot.791.3 Rosetta SSM



Experimental SSM Average



Rosetta SSM Average

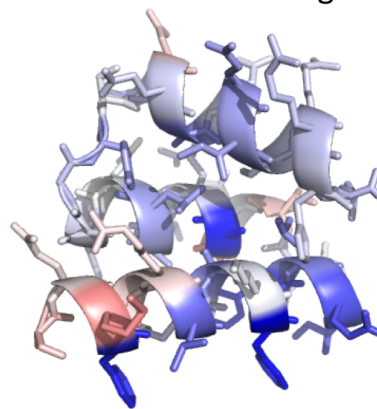


Figure 3c. Experimental and Rosetta-predicted SSM Heatmaps for Bot.791.3, the effect that each point mutation has on binding (experimental SSM) and sum of binding plus monomer stability (Rosetta SSM) is shown (for details see supplementary information Fig. 2). Red indicates mutations that enhance binding, grey indicates mutations equivalent to the designed amino acid, and blue indicates mutations that reduce binding affinity. We show the average mutational effect for each position mapped onto the design model. Computational and experimental values were calculated as described in SI Figure 2. It can be observed there is a good agreement between the computational and experimental SSM.

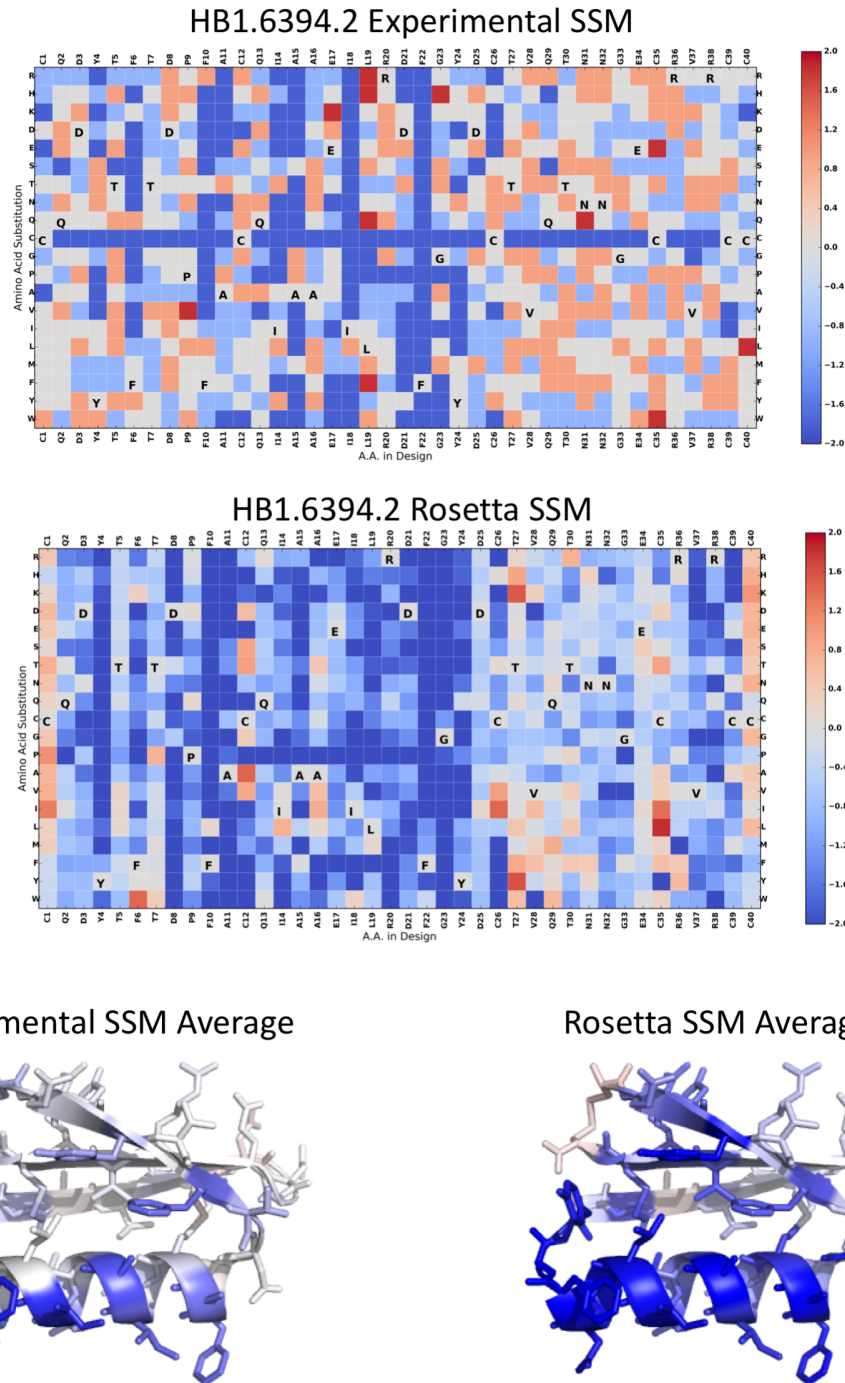
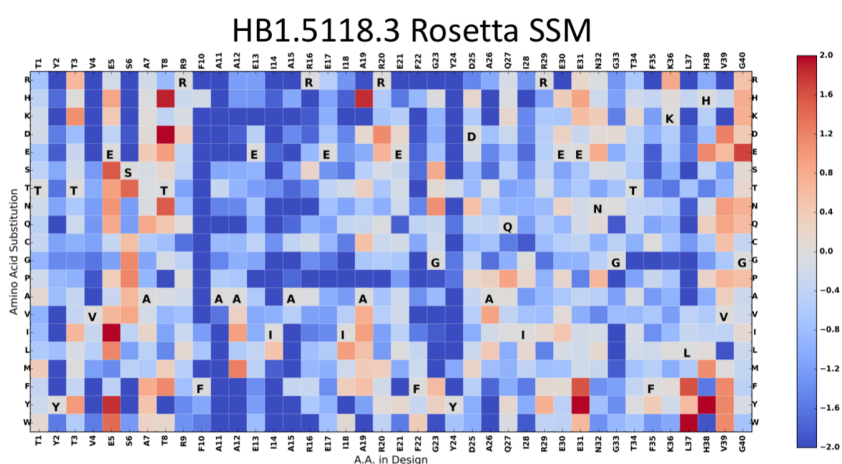
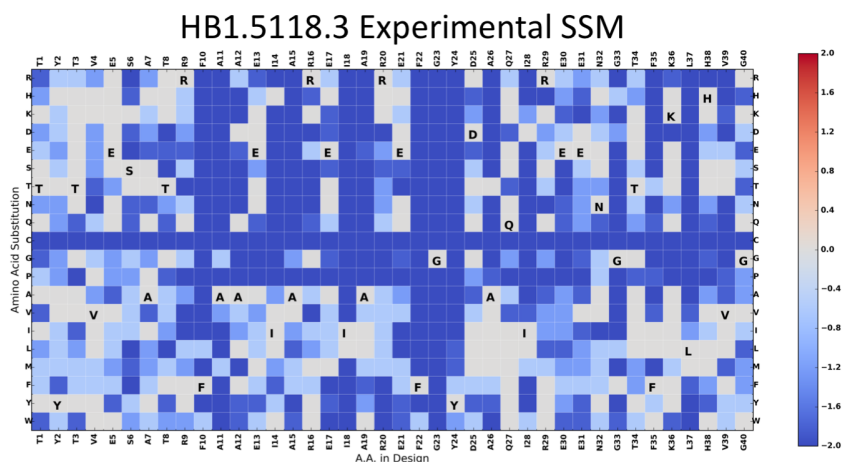
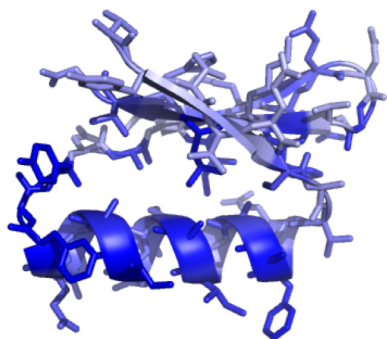


Figure 3d. Experimental and Rosetta-predicted SSM Heatmaps for HB1.6394.2, similar to Supplementary Information Figure 3a. Red indicates mutations that enhance binding, grey indicates mutations equivalent to the designed amino acid, and blue indicates mutations that reduce binding affinity. We show the average mutational effect for each position mapped onto the design model. Computational and experimental values were calculated as described in SI Figure 2. It can be observed there is a good agreement between the computational and experimental SSM.



Experimental SSM Average



Rosetta SSM Average

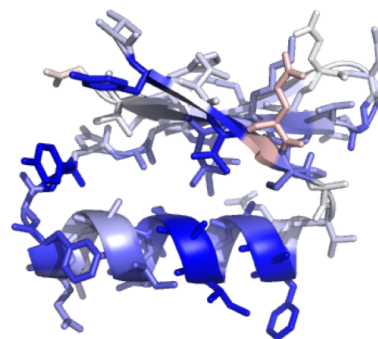
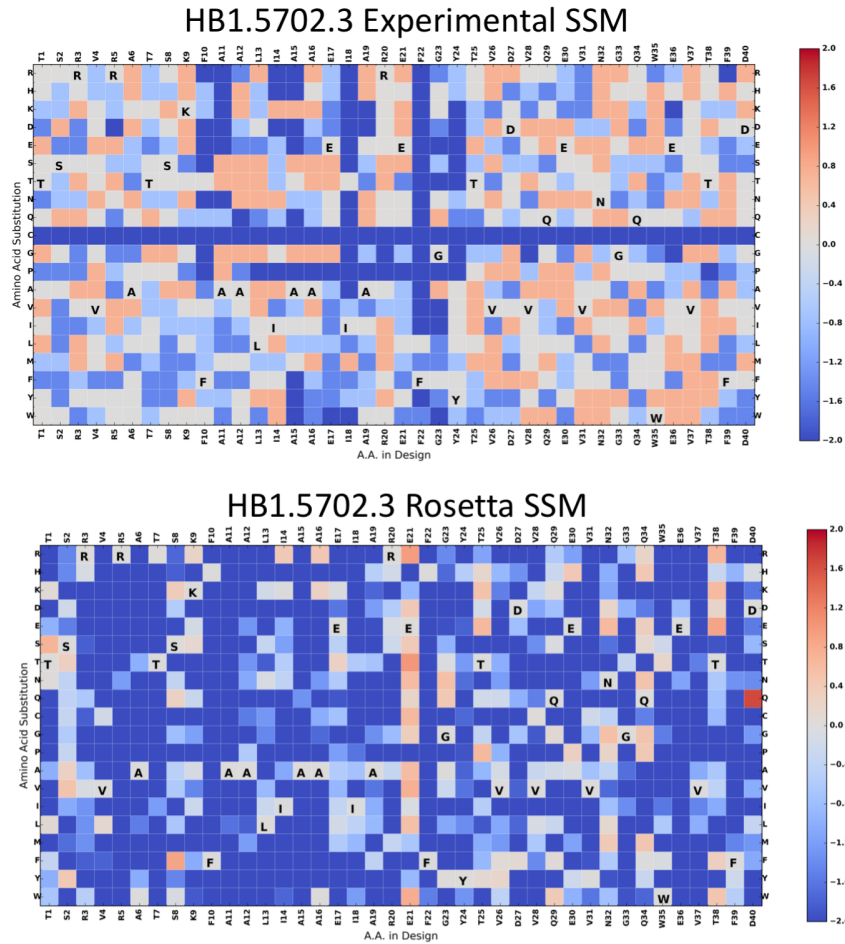
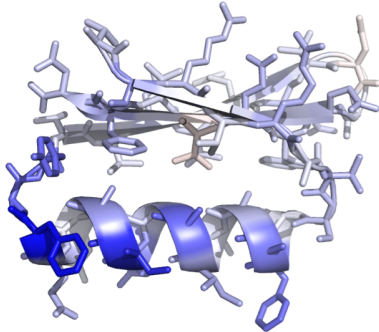


Figure 3e. Experimental and Rosetta-predicted SSM Heatmaps for HB1.5118.3, similar to Supplementary Information Figure 3a. Red indicates mutations that enhance binding, grey indicates mutations equivalent to the designed amino acid, and blue indicates mutations that reduce binding affinity. We show the average mutational effect for each position mapped onto the design model. Computational and experimental values were calculated as described in SI Figure 2. It can be observed there is a good agreement between the computational and experimental SSM. It can be observed there is a good agreement between the computational and experimental SSM, in particular in the core residues and hydrophobic hotspots in the binding helix.



Experimental SSM Average



Rosetta SSM Average

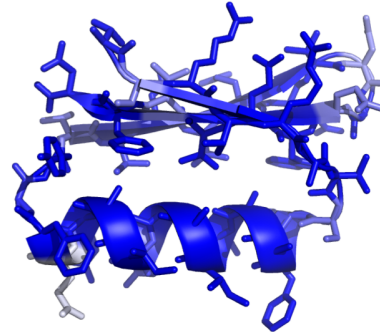


Figure 3f. Experimental and Rosetta-predicted SSM Heatmaps for HB1.5702.3, similar to Supplementary Information Figure 3a. Red indicates mutations that enhance binding, grey indicates mutations equivalent to the designed amino acid, and blue indicates mutations that reduce binding affinity. We show the average mutational effect for each position mapped onto the design model. Computational and experimental values were calculated as described in SI Figure 2.

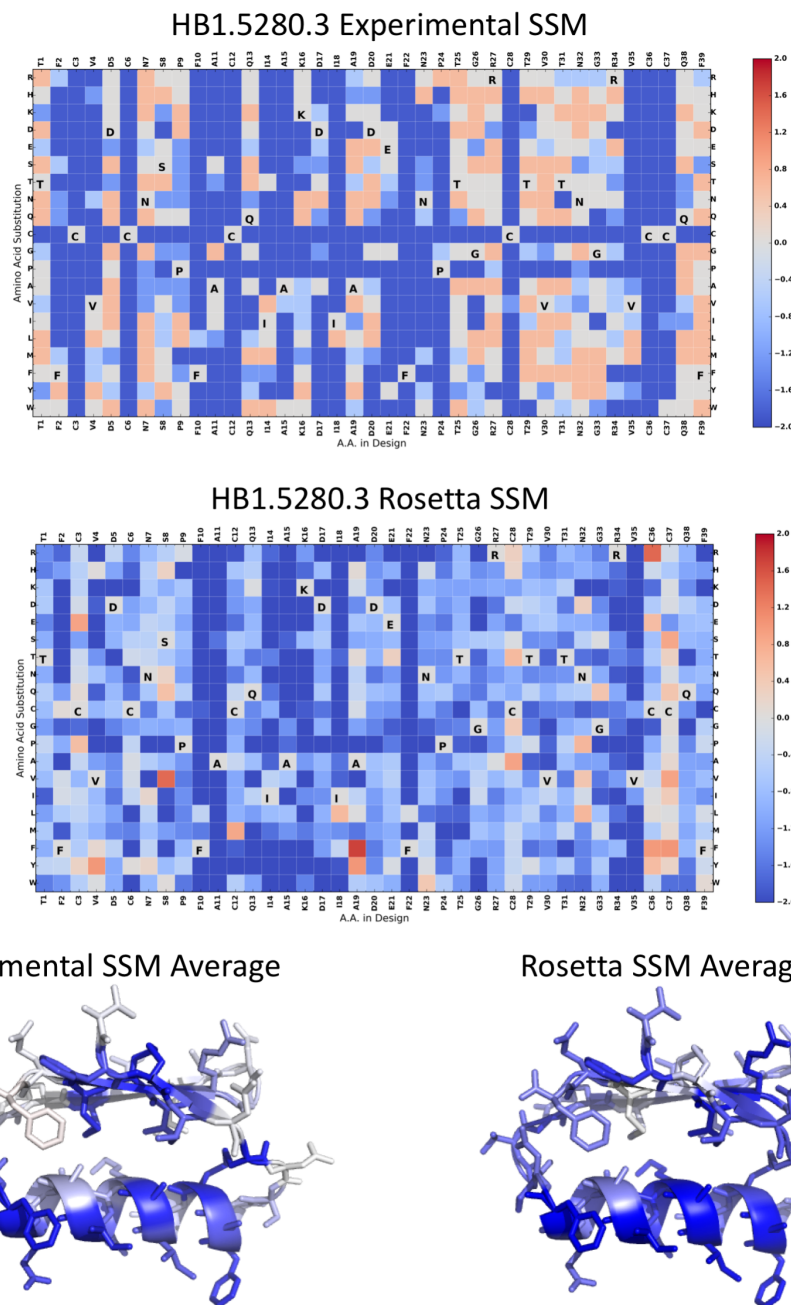
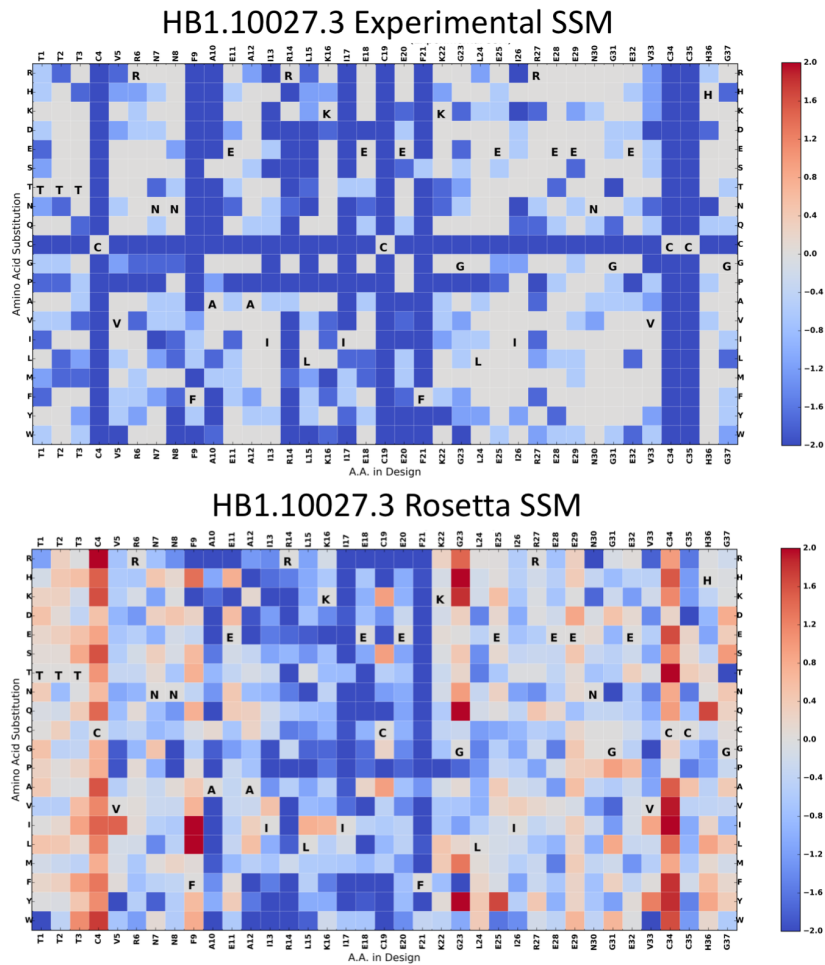
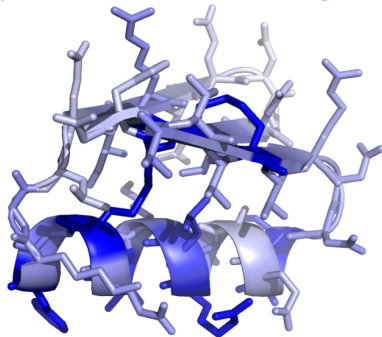


Figure 3g. Experimental and Rosetta-predicted SSM Heatmaps for HB1.5280.3, similar to Supplementary Information Figure 3a. Red indicates mutations that enhance binding, grey indicates mutations equivalent to the designed amino acid, and blue indicates mutations that reduce binding affinity. We show the average mutational effect for each position mapped onto the design model. Computational and experimental values were calculated as described in SI Figure 2. It can be observed there is a good agreement between the computational and experimental SSM. Consistent with the design model, substitutions in the binding and core positions disrupt function; Rosetta fails to capture the details of the disulfide bond which results in a poor agreement with the Rosetta predictions (Supplementary Information Figure 2).



Experimental SSM Average



Rosetta SSM Average

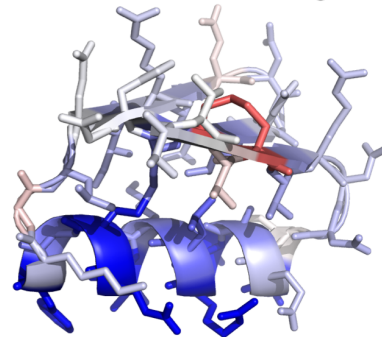


Figure 3h. Experimental and Rosetta-predicted SSM Heatmaps for HB1.10027.3, similar to Supplementary Information Figure 3a. Red indicates mutations that enhance binding, grey indicates mutations equivalent to the designed amino acid, and blue indicates mutations that reduce binding affinity. We show the average mutational effect for each position mapped onto the design model. Computational and experimental values were calculated as described in Supplementary Information Figure 2. Similar to Supplementary figure 3g, substitutions in the binding and core positions are consistent with the design model; the agreement with the Rosetta predictions is poor (Supplementary Information Figure 2) as Rosetta fails to capture the details of the disulfide bonds, and perhaps the experimental SSM lacks dynamic range.

Octet 1:1 binding Fitting vs BoNT HcG

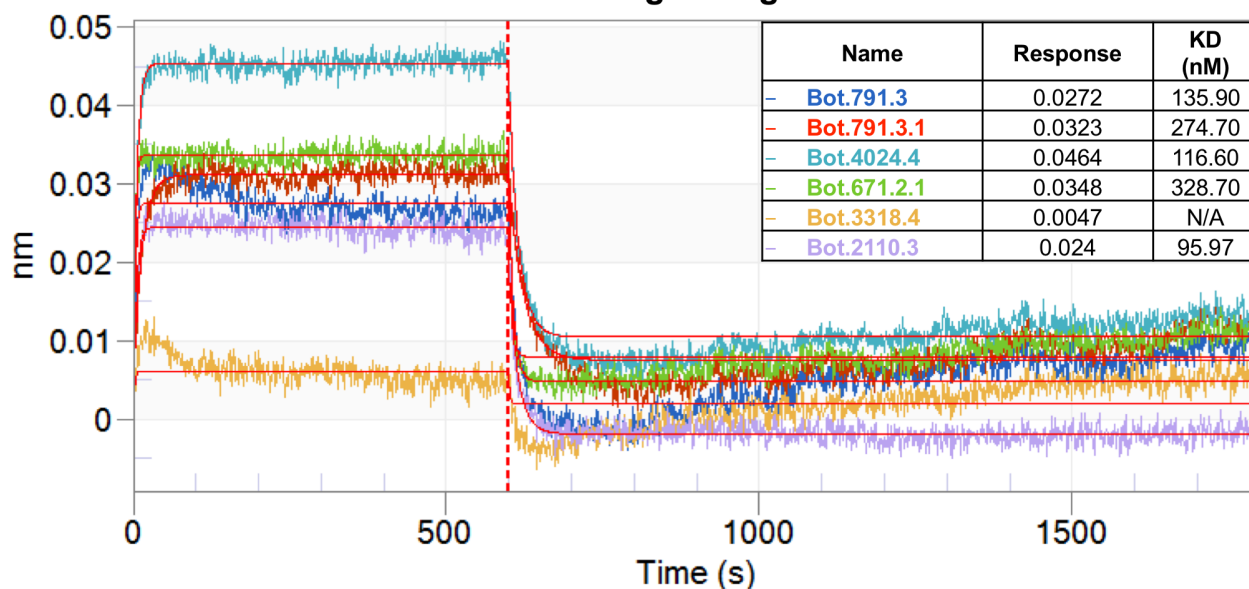


Figure 4. Cross-reactivity binding of Bot designs (BoNT H_cB binders) against BoNT/G (H_cG). The graph shows the results of a biolayer interferometry experiment (OCTET, 1:1 binding model, see Methods) to characterize the cross-reactivity binding of six functional designs against H_cG. The six binders presented can bind BoNT/B with high affinity (<~10 nM, see Main Text). Each line represents the binding and dissociation of an independent design, and the thin red line represents the adjustment of a 1:1 binding model. This demonstrates that 5 of the 6 designs exhibit cross-reactivity against HcG, but with ~10X lower affinity than for HcB (Extended Data Table 1).

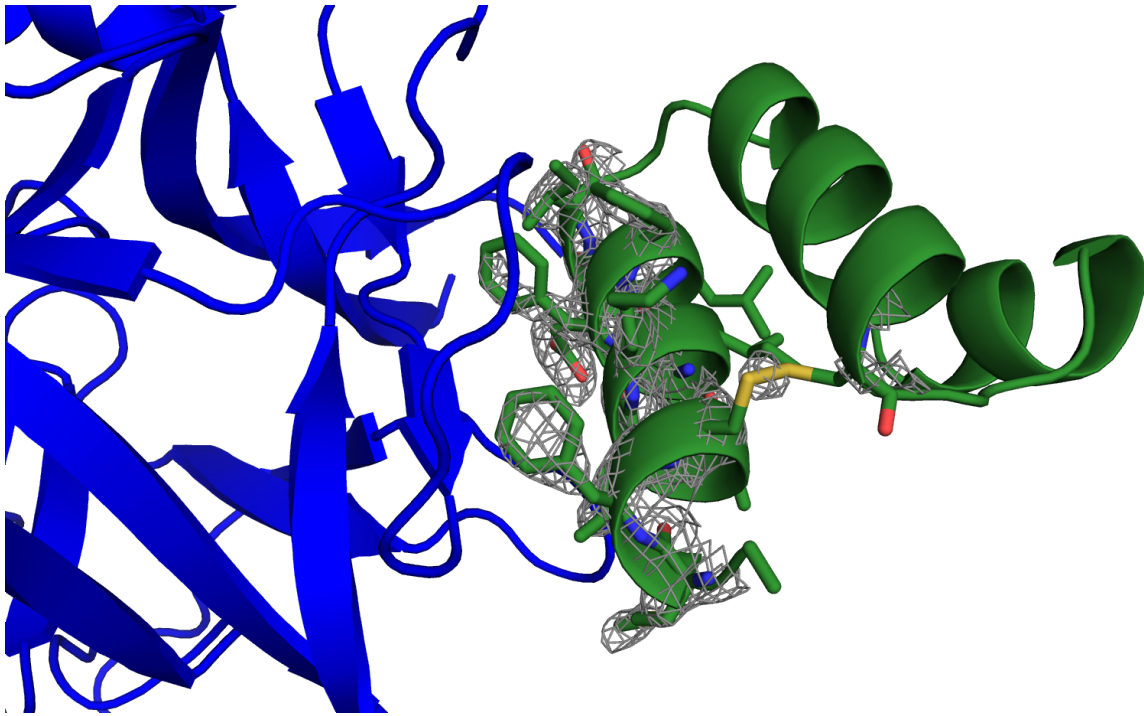


Figure 5. Co-crystal structure with interface side chain density for disulfide containing design Bot.2110.4 in complex with BoNT H_CB (PDB: 5VMR). 2F_o-F_c map for Bot.2110.4 (green) residues 2-15 and disulfide bond (contoured at 1 sigma) shows density for side chains at the interface. The sulfur atoms of the disulfide bond show clear density. BoNT H_CB is shown in blue.

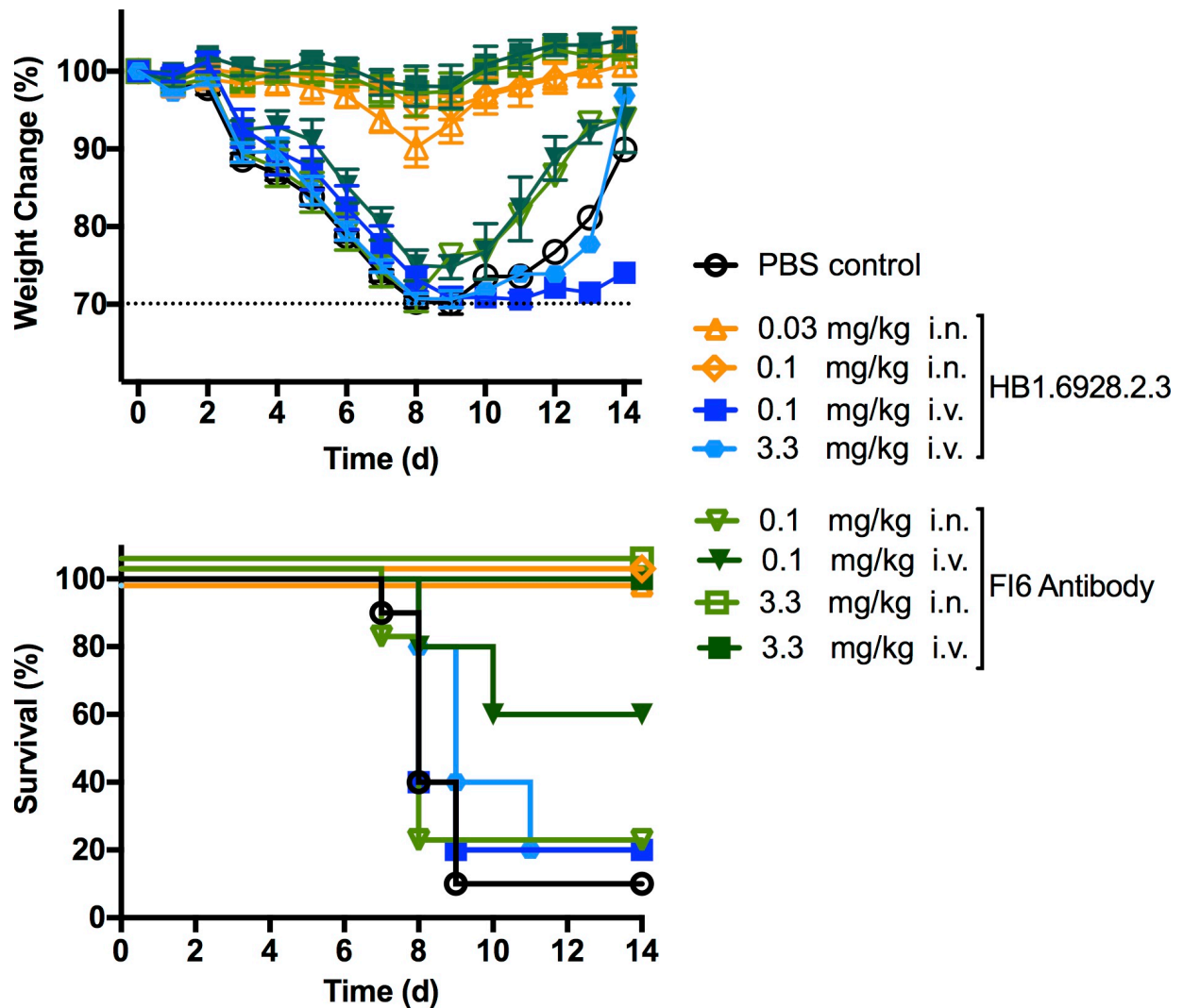


Figure 6. In vivo prophylactic efficiency of intranasal (i.n.) and intravenous (i.v.) administration of the designed miniprotein HB1.6928.2.3 compared in a mass-to-mass basis to the FI6 antibody and a PBS control. Weight change (top) and survival (bottom) of BALB/c mice receiving different doses of HB1.6928.2.3 or the FI6v3 mAb, by the i.n. or i.v. routes, 24 hours prior to challenge with 2MLD50 of H1N1 CA09 virus (n=10, except 0.03mg/kg, n=5, see Main text methods). It is clear that in the i.n route HB1.6928.2.3 is much better neutralizing the virus than FI6 Antibody (mAb Fi6v3). However, HB1.6928.2.3 offers no protection in the i.v. route while FI6 does. It is likely that due to their small size, the mini-protein HB1.6928.2.3 is cleared from blood in a very short time, and is not available anymore to neutralize the virus at the time of challenge. In the top panel the whiskers show ± 1 standard error of the mean (SEM).

Table 1. X-ray data collection and refinement statistics of co-crystal of design H_CB-Bot.671.2 and HCB-Bot.2110.4

	H _C B-Bot.0671.2 (PDB code: 5VID)	H _C B-Bot.2110.4 (PDB code: 5VMR)
Data collection		
Space group	P 1 2 1 1	P 1 2 1 1
Cell dimensions		
a, b, c (Å)	140.89, 87.31, 148.46	38.70, 246.48, 64.36
α, β, γ (°)	90, 114.53, 90	90, 102.90, 90
Resolution (Å)	87.31–2.75 (2.80– 2.75) ^a	19.97–1.95 (1.99– 1.95) ^a
R _{meas}	0.194 (1.099)	0.080 (0.513)
R _{merge}	0.143 (0.803)	0.058 (0.372)
I/σ(I)	8.1 (2.0)	7.7 (2.2)
CC1/2	0.986 (0.701)	0.997 (0.836)
Completeness (%)	100 (100)	99.6 (99.5)
Redundancy	3.8 (3.8)	3.5 (3.5)
No. unique reflections	85598 (4495)	84713 (4433)
Refinement		
Resolution (Å)	67.529–2.75	19.969–1.95
No. reflections	85,554	83967
R _{work} / R _{free}	0.219 / 0.232	0.181 / 0.206
No. atoms		
Protein	18,990	7,733
Ligand/ion	–	–
Water	74	575
Wilson B	45.72	33.14
B factors		
Protein	59.80	46.90
Ligand/ion	–	–
Water	46.30	47.60
R.m.s. deviations		
Bond lengths (Å)	0.01	0.008
Bond angles (°)	1.27	1.111
Ramachandran (%)		
Favored	96.5	96.5
Allowed	3.5	3.5
Outliers	0	0

^a Values in parentheses are for highest-resolution shell.

Table 2. X-ray data collection and refinement statistics of co-crystal of design HB1.6928.2.3

A/PuertoRico/8/1934 HA - HB1.6928.2.3 complex (PDB code: 5VLI)	
Data collection	
Space group	R32
Cell dimensions	
a, b, c (Å)	110.7 110.7 327.4
α , β , γ (°)	90 90 120
Resolution (Å)	47.4-1.80 (1.86-1.80) ^a
Unique reflections	71,919
R _{sym}	8.9 (172.2)
R _{pim}	2.3 (47.0)
I/ σ (I)	27.0 (1.9)
CC _{1/2}	0.973 (0.870)
Completeness (%)	100 (100)
Redundancy	16.1 (14.3)
Refinement	
Resolution (Å)	1.80
No. reflections	71,713
R _{work} / R _{free}	0.182 / 0.213
No. atoms	
Protein (HA)	3934
Glycan	81
Peptide (A13_r33)	308
Water	420
Wilson B	30
B-values	
Protein (HA)	43
Glycan	64
Peptide (A13_r33)	45
Water	49
R.m.s. deviations	
Bond lengths (Å)	0.018
Bond angles (°)	1.45
Ramachandran(%)	
Favored	97.4
Allowed	100
Outliers	0

^a Values in parentheses are for highest-resolution shell.

Table 3. DNA sequences of gene pool primers and combination libraries

HB1.5702.2	GGGTCGGCTTCGCATATGACATCTVKAGTTAGARBGACGRBCAAATTT GCTGCGTTGATTGCTGCTGAAATTGCCAGASAATTTGGGTACACAGTT GACGTTCAARWSRHAATGGTVWATGGDDSGTGRYATTTGATCTCGA GGGTGGAGGTTCC
HB1.10027.3	GGGTCGGCTTCGCATATGRCAACTDVGTTGTGTTDNAAACAATTTTGCC GAGGCAATACGTTTGAAGATTGAGTGCGAGTTTAAAGGTTTGVWMAT CRDGAAGAAAATGGTNMAGTTTGCTGCVHNCNBGCTCGAGGGTGGA GGTTCC
HB1.6928.2.3	GGGTCGGCTTCGCATATGKGCNMAGAAMHAAGTBDCACTACTHYGTT CGCTTGTCADYAGCCGCTGAAATCTGGCGTGCGTTCGGCTATRNAGT GAAAATTVHSGWCGATAATGGCAACTGCCGTTTGCACGTTTGTCTCG AGGGTGGAGGTTCC
HB1.6394.2.3	GGGTCGGCTTCGCATATGTGCCAARDSTATVVMTTTACTRACCCATTC GCTTGTCAAATCGCAVBCGAGATATTGAGAGATTTTGGCTACKVCTGC ACTGTCCAAACCMWMVMCGGANAMTGTAGGGTTAGATGCTGTCTCG AGGGTGGAGGTTCC
HB1.5118.3	GGGTCGGCTTCGCATATGVVATATACTGTTGAATCTKBGACCAGATTT GCTGCTGAGATCGCCAGAVWSATCVYAAGARAATTTGGGTACGACGC TCAAATARNAGAGGAAAATGGGACCTTTWHMTTGHWCCTTKGCCTC GAGGGTGGAGGTTCC
HB1.5380.3	GGGTCGGCTTCGCATATGANGTTTTGTGTTVRMTGCHACTCTCCATTT GCATGTCAGDKSGCCAAAGACATCBHCGAMGAGTTTAATCCARVAGG TCGTTGTAAGTAAACCAACGGTAGAGTTTGTGCSRANHCTCGAGGG TGGAGGTTCC
Yeast_fwd_pool_primer	AGTGGTGGAGGAGGCTCTGGTGGAGGCGGTAGCGGAGGCGGAGGGT CGGCTTCGCATATG
Yeast_rev_pool_primer	GATCTCTATTACAAGTCCTCTTCAGAAATAAGCTTTTGTTCGGAACCT CCACCCTCGAG

Appendix A. RosettaScripts' XML used for designing mini-protein scaffolds

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constraints_NtoC="-1.0" blueprint="ehee.bp" />
    <DumpPdb name="dump" fname="pass" tag_time=True/>

    <FastDesign name="fastdes" task_operations="limitchi2,layer_all" scorefxn="sfxn_std"
clear_designable_residues="0" repeats="2" ramp_down_constraints="0" />
    <FastDesign name="fastdes4" task_operations="limitchi2,layer_all" scorefxn="sfxn_std"
clear_designable_residues="0" repeats="4" ramp_down_constraints="0" />

    <ParsedProtocol name="build_dssp1" >
    <Add mover_name="bdr1" />
    <Add mover_name="dssp" />
    <Add filter_name="cs1" />
    <Add filter_name="degree" />
    <Add mover_name="fastdes"/>
    <Add filter_name="cavity_threshold" />
    <Add filter_name="percent_core_SCN" />
    <Add filter_name="AlaCount" />
    <Add filter_name="unsat_hbond" />
    <Add mover_name="dump"/>

    </ParsedProtocol>
    <LoopOver name="lover1" mover_name="build_dssp1" iterations="500" drift="0"
ms_whenfail="FAIL_DO_NOT_RETRY" />
    <ParsedProtocol name="phase1" >
    <Add mover_name="set_ssene1" />
    <Add mover_name="lover1" />
    </ParsedProtocol>

</MOVERS>

```

```
<PROTOCOLS>  
  <Add mover_name="phase1" />  
  
</PROTOCOLS>  
</dock_design>
```

Appendix B. RosettaScripts' XML used for design Influenza of Binders Generation 1/2 (HB1)

```

<dock_design>
  <TASKOPERATIONS>
    <InitializeFromCommandline name="init"/>
    <LimitAromaChi2 name="arochi2"/>
    <IncludeCurrent name="inclcur"/>
    <ExtraRotamersGeneric name="exrot" ex1="1" ex2="1" extrachi_cutoff="1"/>
    <DisallowIfNonnative name="only_native_GPH" disallow_aas="GPH"/>
    <RestrictIdentities name="dont_design_GPH" identities="PRO,HIS" prevent_repacking=0 />
    <OperateOnCertainResidues name="hotspot_norepack">
      <PreventRepackingRLT/>
      <ResiduePDBInfoHasLabel property="HOTSPOT"/>
    </OperateOnCertainResidues>
    <OperateOnCertainResidues name="scaffold_onlyrepack">
      <RestrictToRepackingRLT/>
      <ResiduePDBInfoHasLabel property="SCAFFOLD"/>
    </OperateOnCertainResidues>
    <OperateOnCertainResidues name="context_norepack">
      <PreventRepackingRLT/>
      <ResiduePDBInfoHasLabel property="CONTEXT"/>
    </OperateOnCertainResidues>
    <OperateOnCertainResidues name="context_onlyrepack">
      <RestrictToRepackingRLT/>
      <ResiduePDBInfoHasLabel property="CONTEXT"/>
    </OperateOnCertainResidues>
    <RestrictToInterface name="interface_design_long" jump="1" distance="12.0"/>
    <RestrictToInterface name="interface_design_short" jump="1" distance="8.0"/>
    <SelectBySASA name="core_norepack" mode="sc" state="monomer" probe_radius="2.2" core_asa="20"
surface_asa="40" core="0" boundary="1" surface="1" verbose="1" />
    <RestrictIdentities name=no_glycan identities=AX1 prevent_repacking=1 />
    <RestrictIdentities name=no_CYS identities=CYS prevent_repacking=1 />
    <RetrieveStoredTask name=design_task task_name="design_task" />
  </TASKOPERATIONS>
  <SCOREFXNS>
    <talaris2013_resCons weights="talaris2013">
      <Reweight scoretype="res_type_constraint" weight="1.0"/>
    </talaris2013_resCons>
    <talaris2013_extra weights="talaris2013">
      <Reweight scoretype="fa_elec" weight="1.0"/>
      <Reweight scoretype="hpatch" weight="1.0"/>
      <Set fa_elec_dielectric="1.0" fa_elec_max_dis="15.0" fa_elec_min_dis="3.0"/>
    </talaris2013_extra>
    <talaris2013_cart weights="talaris2013">
      <Reweight scoretype="cart_bonded" weight="1.0"/>
      <Reweight scoretype="pro_close" weight="0.0"/>
    </talaris2013_cart>
  </SCOREFXNS>
  <FILTERS>
    <Geometry name="omega" omega="165" cart_bonded="35" start="491" confidence="0"/>
    <Ddg name="ddg" scorefxn="talaris2013" jump="1" repack="1" repeats="5" threshold="-18.0"
confidence="1"/>
    <ResInInterface name="resInInterface" residues="22" jump_number="1" confidence="0"/>
    <Sasa name="sasa" threshold="1200" confidence="1"/>
    <BuriedUnsatHbonds name="buriedUnsatBonds" scorefxn="talaris2013" jump_number="1" cutoff="9"
confidence="0"/>

```

```

    <ShapeComplementarity name="shapeComplementarity" jump="1" verbose="1" min_sc="0.6"
write_int_area="0" confidence="1"/>
    <InterfaceHoles name="interfaceHoles" jump="1" threshold="200" confidence="0"/>
    <AverageDegree name="averageDegree" threshold="8.3" distance_threshold="8"
task_operations="init,interface_design_short,context_norepack" confidence="0"/>
    <PackStat name="packstat" repeats="5" threshold="0.59" confidence="0"/>
    <ScoreType name="hpatch" scorefxn="talaris2013_extra" score_type="hpatch" threshold="35"
confidence="0"/>
    <ScoreType name="lr_elec" scorefxn="talaris2013_extra" score_type="fa_elec" threshold="1200"
confidence="0"/>
    <ScoreType name="total_score" scorefxn="talaris2013" score_type="total_score" threshold="0"
confidence="0"/>
</FILTERS>
<MOVERS>
  <!-- Epigraft Mover -->
    <StoreTaskMover name=store_design_task task_name="design_task"
task_operations="init,hotspot_norepack,core_norepack,context_onlyrepack,arochi2,inclcur,exrot,only_native_
GPH,dont_design_GPH,no_glycan,no_CYS"/>

    <MotifGraft name="motif_grafting" context_structure="%%CONTEXT%%"
motif_structure="%%MOTIF%%" RMSD_tolerance="1.0" hotspots="%%HOTSPOTS%%" NC_points_RMSD_tolerance="1.0"
clash_score_cutoff="10" clash_test_residue="ALA" combinatorial_fragment_size_delta="0:0"
max_fragment_replacement_size_delta="-2:2" full_motif_bb_alignment="1"
allow_independent_alignment_per_fragment="0" graft_only_hotspots_by_replacement="0"
only_allow_if_N_point_match_aa_identity="0" only_allow_if_C_point_match_aa_identity="0"
revert_graft_to_native_sequence="1" allow_repeat_same_graft_output="1"/>
  <!-- Design Movers -->
    <FavorSequenceProfile name="favor_nativeSeqProfile" scorefxns="talaris2013_resCons"
use_current="1" chain="0" matrix=MATCH weight="0.2"/>
    <PackRotamersMover name="design_graft1" scorefxn="talaris2013_resCons"
task_operations="init,hotspot_norepack,core_norepack,context_onlyrepack,arochi2,inclcur,exrot,only_native_
GPH,dont_design_GPH,no_glycan,no_CYS,scaffold_onlyrepack"/>
    <PackRotamersMover name="design_interface1" scorefxn="talaris2013_resCons"
task_operations="init,hotspot_norepack,core_norepack,context_onlyrepack,arochi2,inclcur,exrot,only_native_
GPH,dont_design_GPH,no_glycan,no_CYS,interface_design_short"/>
  <!-- Minimization Movers -->
    <TaskAwareMinMover name="chain2_kinematic_min_noBB" scorefxn="talaris2013" bb="0" chi="1"
jump="1" task_operations="init,context_norepack,arochi2,inclcur,exrot"/>
    <TaskAwareMinMover name="chain2_kinematic_min_wBB" scorefxn="talaris2013" bb="1" chi="1"
jump="1" task_operations="init,context_norepack,arochi2,inclcur,exrot"/>
    <TaskAwareMinMover name="chain2_cart_min" scorefxn="talaris2013_cart" bb="1" chi="1"
jump="1" cartesian="1" task_operations="init,context_norepack"/>
</MOVERS>
<APPLY_TO_POSE>
</APPLY_TO_POSE>
<PROTOCOLS>

    <Add mover_name="store_design_task"/>
  <!-- Epigraft -->
    <Add mover_name="motif_grafting"/>
  <!-- Favor sequence profile -->
    <Add mover="favor_nativeSeqProfile"/>
  <!-- First Design round -->
    <Add mover_name="design_graft1"/>
    <Add mover_name="design_interface1"/>
    <Add mover_name="design_core1"/>
  <!-- Minimize (kine no BB) -->
    <Add mover_name="chain2_cart_min"/>
    <Add mover_name="chain2_kinematic_min_noBB"/>
    <Add mover_name="chain2_cart_min"/>

```

```
<!-- Second Design round -->
  <Add mover_name="design_graft1"/>
  <Add mover_name="design_interface1"/>
  Add mover_name="design_core2"/>
<!-- Minimize (all BB) -->
  <Add mover_name="chain2_cart_min"/>
  <Add mover_name="chain2_kinematic_min_wBB"/>
  <Add mover_name="chain2_cart_min"/>
<!-- Filter by omega to ensure correct BB geometry and atomic distances -->
  <Add filter="omega"/>
<!-- All the other Filters -->
  <Add filter="ddg"/>
  <Add filter="total_score"/>
  <Add filter="resInInterface"/>
  <Add filter="sasa"/>
  <Add filter="buriedUnsatBonds"/>
  <Add filter="shapeComplementarity"/>
  <Add filter="packstat"/>
  <Add filter="averageDegree"/>
  <Add filter="hpatch"/>
  <Add filter="lr_elec"/>
  <Add filter="interfaceHoles"/>
</PROTOCOLS>
</dock_design>
```

Appendix C. RosettaScripts' XML used for design of BoNT/B Binders (Bot)

```

<ROSETTASCRIPTS>

  <SCOREFXNS>
    <sfxn_talaris2013 weights="talaris2013" />
  </SCOREFXNS>

  <FILTERS>
    <Geometry name="omega" omega="165" cart_bonded="20" start="115" confidence="0"/>
    <ShapeComplementarity name=sc_filt jump=1 verbose=1 min_sc=0.5 write_int_area=0
confidence=0 />
    <Ddg name=ddG_filt scorefxn=sfxn_talaris2013 jump=1 repack=1 repeats=3 threshold=0.0
confidence=1 />
    <BuriedUnsatHbonds name=buriedUnsatBonds scorefxn=sfxn_talaris2013 jump_number=1 cutoff=9
confidence=0/>
    <Sasa name=sasa threshold=700 hydrophobic=0 polar=0 jump=1 confidence=0 />
    <ScoreType name=total_score scorefxn=sfxn_talaris2013 score_type=total_score threshold=500
confidence=0/>
    <FragmentLookupFilter name="faulty_fragments" lookup_name="source_fragments_4_mer"
store_path="/work/dadriano/Shared/source_fragments_4_mer/" lookup_mode="first" chain="2" threshold="0"
confidence="0" />
  </FILTERS>

  <MOVERS>
    //Here is the mover that generates multiple pose results
    <MotifGraft name="motif_grafting"
      context_structure="./contextStructure.pdb" motif_structure="./motif_2NM1B.pdb"
      RMSD_tolerance="1.0" NC_points_RMSD_tolerance="1.0"
      clash_score_cutoff="5" clash_test_residue="GLY"
      hotspots="2:9:10" combinatorial_fragment_size_delta="0:0"
max_fragment_replacement_size_delta="0:0"
      full_motif_bb_alignment="1" allow_independent_alignment_per_fragment="0"
      graft_only_hotspots_by_replacement="0"
only_allow_if_N_point_match_aa_identity="0"
      only_allow_if_C_point_match_aa_identity="0" revert_graft_to_native_sequence="1"
allow_repeat_same_graft_output="0" />
    //HERE You add all the other design movers
    <MultiplePoseMover name="MPM_design" max_input_poses=10>
    <SELECT>
    </SELECT>
      <ROSETTASCRIPTS>
        <SCOREFXNS>
          <sfxn_talaris2013downAla weights="talaris2013_downAla" >
          </sfxn_talaris2013downAla>
          <sfxn_soft weights="soft_rep" >
          </sfxn_soft>
          <sfxn_soft_cst weights="soft_rep" >
            <Reweight scoretype="coordinate_constraint" weight="2.0"
/>
          </sfxn_soft_cst>
          <sfxn_soft_cart_cst weights="soft_rep" >
            <Reweight scoretype="coordinate_constraint" weight="1.0"
/>
          <Reweight scoretype="cart_bonded" weight="1.0"/>
          <Reweight scoretype="pro_close" weight="0.0"/>
          </sfxn_soft_cart_cst>
          <sfxn_talaris2013_cart_cst weights="talaris2013" >

```

```

        <Reweight scoretype="coordinate_constraint" weight="1.0"
/>
        <Reweight scoretype="cart_bonded" weight="1.0" />
        <Reweight scoretype="pro_close" weight="0.0" />
</sfxn_talaris2013_cart_cst>
<sfxn_talaris2013_cst weights="talaris2013" >
        <Reweight scoretype="coordinate_constraint" weight="1.0"
/>
        </sfxn_talaris2013_cst>
</SCOREFXNS>
<TASKOPERATIONS>
  <InitializeFromCommandline name="init"/>
  <IncludeCurrent name="current" />
  <ConsensusLoopDesign name="disallow_nonnative_loop_sequences"/>
  <RestrictIdentities name="notGPH" identities="PRO,HIS"
prevent_repacking="0" />
  <OperateOnCertainResidues name="hotspot_onlyrepack">
    <RestrictToRepackingRLT/>
    <ResiduePDBInfoHasLabel property="HOTSPOT"/>
  </OperateOnCertainResidues>
  <OperateOnCertainResidues name="scaffold_onlyrepack">
    <RestrictToRepackingRLT/>
    <ResiduePDBInfoHasLabel property="SCAFFOLD"/>
  </OperateOnCertainResidues>
  <OperateOnCertainResidues name="context_norepack">
    <PreventRepackingRLT/>
    <ResiduePDBInfoHasLabel property="CONTEXT"/>
  </OperateOnCertainResidues>
  <RestrictToInterface name="interface_design_long" jump="1"
distance="12.0"/>
  <RestrictToInterface name="interface_design_short" jump="1"
distance="8.0"/>
  <SelectBySASA name="core_norepack" mode="sc" state="monomer"
probe_radius="2.2" core_asa="20" surface_asa="40" core="0" boundary="1" surface="1" verbose="1" />
  <SelectBySASA name="core_design" mode="sc" state="monomer"
probe_radius="2.2" core_asa="20" surface_asa="40" core="1" boundary="0" surface="0" verbose="1" />
  <DisallowIfNonnative name="only_native_GPH" disallow_aas="GPH"/>
  <LimitAromaChi2 name=limitaro /> #avoids extra rotamers
  <RestrictIdentities name=noCYS identities=CYS prevent_repacking=1
/>
  <ProteinInterfaceDesign name=PID repack_chain1=0 repack_chain2=1
design_chain1=0 design_chain2=1 interface_distance_cutoff=10/>
  <LayerDesign name="layer_design" layer=core_boundary_surface
core_E=20 core_H=20 core_L=25 surface_E=50 surface_H=50 surface_L=26 make_pymol_script=0>
    <core>
      <all append="MACFILVWY" exclude="ERKHGQ"/> #PDNST
left normal
    </core>
  </boundary>
      <all append="ACDEIKLNQRVYWF" exclude="HMST"/> #PG
left alone
    </boundary>
  <surface>
      <all append="ACDEHKNQRSTY" exclude="MVILWF" />
#PG left alone
    </surface>
  </LayerDesign>
</TASKOPERATIONS>
<FILTERS>
</FILTERS>

```



```

<MOVERS>
    <AddConstraintsToCurrentConformationMover name=constrainCA
task_operations="current" CA_only=1 />
    <ClearConstraintsMover name=clearConstraints />
    <PackRotamersMover name=design_interface_soft scorefxn=sfxn_soft
task_operations="init,current,limitaro,disallow_nonnative_loop_sequences,noCYS,PID,layer_design,hotspot_on
lyrepack,scaffold_onlyrepack,context_norepack" />
    <PackRotamersMover name=design_interface_scaffold_soft
scorefxn=sfxn_soft
task_operations="init,current,limitaro,disallow_nonnative_loop_sequences,noCYS,PID,layer_design,hotspot_on
lyrepack,context_norepack" />
    <PackRotamersMover name=design_interface_hard
scorefxn=sfxn_talaris2013downAla
task_operations="init,current,limitaro,disallow_nonnative_loop_sequences,noCYS,PID,limitaro,layer_design"
/>
    <PackRotamersMover name=design_interface_scaffold_hard
scorefxn=sfxn_talaris2013downAla
task_operations="init,current,disallow_nonnative_loop_sequences,limitaro,noCYS,PID,layer_design" />
    <TaskAwareMinMover name=min_soft scorefxn=sfxn_soft_cart_cst
bb=0 chi=1 jump=1 cartesian=1 task_operations="init,current,limitaro,hotspot_onlyrepack,context_norepack"
/>
    <TaskAwareMinMover name=min_soft scorefxn=sfxn_soft_cst bb=0
chi=1 jump=1 cartesian=0 task_operations="init,current,limitaro,hotspot_onlyrepack,context_norepack" />
    <TaskAwareMinMover name=min_hard_cart
scorefxn=sfxn_talaris2013_cart_cst bb=0 chi=1 jump=1 cartesian=1
task_operations="init,limitaro,current,hotspot_onlyrepack,context_norepack" />
    <TaskAwareMinMover name=min_hard scorefxn=sfxn_talaris2013_cst
bb=0 chi=1 jump=1 cartesian=0 task_operations="init,current,limitaro,hotspot_onlyrepack,context_norepack"
/>
</MOVERS>
<APPLY_TO_POSE>
</APPLY_TO_POSE>
<PROTOCOLS>
    **DESIGN STEP1>
    <Add mover_name=constrainCA /> #Constraint CAs
    <Add mover_name=design_interface_soft /> #design with soft_rep at
interface
    <Add mover_name=design_interface_scaffold_soft /> #design with
soft_rep at interface and the surrounding scaffold
    <Add mover_name=min_soft_cart /> #Min cartesian
    <Add mover_name=min_soft /> #Min kinematic
    <Add mover_name=min_hard_cart /> #Min cartesian with normal LJ
    <Add mover_name=min_hard /> #Min kinematic with normal LJ
    <Add mover_name=clearConstraints /> #Remove constraints to CAs
    **DESIGN STEP2>
    <Add mover_name=design_interface_soft /> #design once more with
soft_rep at interface
    <Add mover_name=design_interface_scaffold_soft /> #design once
more with soft_rep at interface and the surrounding scaffold
    <Add mover_name=min_soft_cart /> #Min cartesian
    <Add mover_name=min_soft /> #Min kinematic
    <Add mover_name=min_hard_cart /> #Min cartesian with normal LJ
    <Add mover_name=min_hard /> #Min kinematic with normal LJ
</PROTOCOLS>
</ROSETTASCRIPTS>
</MultiplePoseMover>
//HERE You add filters
<MultiplePoseMover name="MPM_filters" max_input_poses=10>
    <SELECT>
        <AndSelector>

```

```

        <Filter filter=omega />
        <Filter filter=faulty_fragments/>
        <Filter filter=sc_filt />
        <Filter filter=ddG_filt />
        <Filter filter=buriedUnsatBonds />
        <Filter filter=sasa />
        <Filter filter=total_score />
    </AndSelector>
</SELECT>
</MultiplePoseMover>
</MOVERS>

<APPLY_TO_POSE>
</APPLY_TO_POSE>

<PROTOCOLS >
//HERE you combine everything together
<Add mover_name=motif_grafting />
<Add mover_name=MPM_design />
<Add mover_name=MPM_filters />
</PROTOCOLS>

</ROSETTASCRIPTS>
```

Appendix D. RosettaScripts' XML used for relaxing mutated models of mini-protein binders (in complex with target) in cases of: SSMs, core to valine, scrambled core, scrambled sequence, disulfided or loops to Gly-Ser

```

<ROSETTASCRIPTS>

  <SCOREFXNS>
    <SFXN_talaris2014_vanilla_cart weights="talaris2014_cart.wts" symmetric=0 />
    <SFXN_beta15_vanilla weights="beta_nov15.wts" symmetric=0 />
    <SFXN_beta15_vanilla_cart weights="beta_nov15_cart.wts" symmetric=0 />
  </SCOREFXNS>

  <TASKOPERATIONS>
    <InitializeFromCommandline name="init"/>
    <IncludeCurrent name="inclcur"/>
    <LimitAromaChi2 name="limitchi2" />
    <RestrictToRepacking name="onlyRepack" />
    <PreventChainFromRepacking name="notChainA" chain=1/>
  </TASKOPERATIONS>

  <MOVERS>
    <SwitchChainOrder name="keep_only_chain_A" chain_order="1"/>
    <SwitchChainOrder name="keep_only_chain_B" chain_order="2"/>
  </MOVERS>

  <FILTERS>
    <ScoreType name="score" scorefxn="SFXN_beta15_vanilla" score_type="total_score"
threshold=0.0 confidence="0" />
    <ResidueCount name="nres" confidence="0" />
    <CalculatorFilter name="score_res" equation="SCORE/NRES" threshold="-3.2" confidence="0">
      <SCORE name="SCORE" filter_name="score" />
      <NRES name="NRES" filter_name="nres" />
    </CalculatorFilter>
    <SSPrediction name="sspred" cmd="/work/dadriano/PROGRAMS/psipred/runpsipred_single"
use_probability="0" use_svm="0" threshold=0.80 confidence="0"/>
    <RmsdSimple name="rmsd_ALL" reference_name="reference_conformation" align="1"
threshold="2.0" confidence="0" />
    <RmsdSimple name="rmsd_A" reference_name="reference_conformation" align="1"
threshold="2.0" confidence="0" chain=1/>
    <RmsdSimple name="rmsd_B" reference_name="reference_conformation" align="1"
threshold="2.0" confidence="0" chain=2/>
    <Ddg name="ddg" scorefxn="SFXN_beta15_vanilla" jump=1 repack=1 repeats=3 threshold="0.0"
confidence="0" />
    <Sasa name="interface_sasa" jump="1" confidence=0/>
    <ShapeComplementarity name="interface_sc" verbose=0 min_sc=0.65 write_int_area=1 jump=1
confidence="0"/>
    <CalculatorFilter name="ddg_100sasa" equation="DDG*hundred/(SASA+small_val)"
threshold="0.0" confidence="0">
      <DDG name="DDG" filter_name="ddg" />
      <SASA name="SASA" filter_name="interface_sasa" />
      <VARA name="hundred" value=100.0 />
      <VARB name="small_val" value=0.0001 />
    </CalculatorFilter>
    <!--Chain A/B Filters-->
    <MoveBeforeFilter name="score_res_chainB" mover="keep_only_chain_B" filter="score_res"
confidence="0"/>
    <MoveBeforeFilter name="sspred_chainB" mover="keep_only_chain_B" filter="sspred"
confidence="0"/>

```

</FILTERS>

<MOVERS>

```

    <SavePoseMover name="save_RMSDreference_conformation"
reference_name="reference_conformation"/>
    <AddConstraintsToCurrentConformationMover name="constrainCA" cst_weight=0.5 CA_only=1 />
    <ClearConstraintsMover name="clearAllConstraints" />
    <TaskAwareMinMover name="min_kine_sc_chainB" scorefxn="SFXN_beta15_vanilla" bb=1 chi=1
jump=0 cartesian=0 task_operations="init,inclcur,limitchi2,onlyRepack,notChainA" />
    <TaskAwareMinMover name="min_kine_sc_chainAB" scorefxn="SFXN_beta15_vanilla" bb=0 chi=1
jump=0 cartesian=0 task_operations="init,inclcur,limitchi2,onlyRepack" />
    <TaskAwareMinMover name="min_cart_chainB" scorefxn="SFXN_beta15_vanilla_cart" bb=1 chi=1
jump=1 cartesian=1 tolerance=0.001 max_iter=1000
task_operations="init,inclcur,limitchi2,onlyRepack,notChainA" />
    <FastRelax name="fast_relax_b_beta" scorefxn="SFXN_beta15_vanilla"
task_operations="init,inclcur,limitchi2,onlyRepack,notChainA" >
    <MoveMap name="mappyfd">
        <Chain number=1 chi=0 bb=0/>
        <Chain number=2 chi=1 bb=1/>
        <Jump number=1 setting=1/>
    </MoveMap>
    </FastRelax>
  </MOVERS>

```

<PROTOCOLS>

```

    <Add mover_name="save_RMSDreference_conformation" />
    <Add mover_name="constrainCA" />
    <Add mover_name="min_cart_chainB" />
    <Add mover_name="clearAllConstraints" />

    <Add mover_name="fast_relax_b_beta" />
    <Add mover_name="min_kine_sc_chainB" />
    <Add mover_name="min_kine_sc_chainAB" />
    <Add mover_name="constrainCA" />
    <Add mover_name="min_cart_chainB" />
    <Add mover_name="clearAllConstraints" />

    <Add mover_name="fast_relax_b_beta" />
    <Add filter_name="rmsd_ALL" />
    <Add filter_name="rmsd_A" />
    <Add filter_name="rmsd_B" />
    <Add filter_name="score_res_chainB" />
    <Add filter_name="ddg" />
    <Add filter_name="interface_sasa" />
    <Add filter_name="ddg_100sasa" />
    <Add filter_name="interface_sc" />
    <Add filter_name="sspred_chainB" />

```

</PROTOCOLS>

</ROSETTASCRIPTS>

Appendix E. RosettaScripts XML used for relaxing models of mini-protein binders (in complex with target) for Extended Data Table 1

```

<ROSETTASCRIPTS>

  <SCOREFXNS>
    <SFXN_talaris2014_vanilla_cart weights="talaris2014_cart.wts" symmetric=0 />
    <SFXN_beta15_vanilla weights="beta_nov15.wts" symmetric=0 />
    <SFXN_beta15_vanilla_cart weights="beta_nov15_cart.wts" symmetric=0 />
  </SCOREFXNS>

  <TASKOPERATIONS>
    <InitializeFromCommandline name="init"/>
    <IncludeCurrent name="inclcur"/>
    <LimitAromaChi2 name="limitchi2" />
    <RestrictToRepacking name="onlyRepack" />
    <PreventChainFromRepacking name="notChainA" chain=1/>
  </TASKOPERATIONS>

  <MOVERS>
    <SwitchChainOrder name="keep_only_chain_A" chain_order="1"/>
    <SwitchChainOrder name="keep_only_chain_B" chain_order="2"/>
  </MOVERS>

  <FILTERS>
    <ScoreType name="score" scorefxn="SFXN_beta15_vanilla" score_type="total_score"
    threshold=0.0 confidence="0" />
    <ResidueCount name="nres" confidence="0" />
    <CalculatorFilter name="score_res" equation="SCORE/NRES" threshold="-3.2" confidence="0">
      <SCORE name="SCORE" filter_name="score" />
      <NRES name="NRES" filter_name="nres" />
    </CalculatorFilter>
    <SSPrediction name="sspred" cmd="/work/dadriano/PROGRAMS/psipred/runpsipred_single"
    use_probability="0" use_svm="0" threshold=0.80 confidence="0"/>
    <RmsdSimple name="rmsd_ALL" reference_name="reference_conformation" align="1"
    threshold="2.0" confidence="0" />
    <RmsdSimple name="rmsd_A" reference_name="reference_conformation" align="1"
    threshold="2.0" confidence="0" chain=1/>
    <RmsdSimple name="rmsd_B" reference_name="reference_conformation" align="1"
    threshold="2.0" confidence="0" chain=2/>
    <Ddg name="ddg" scorefxn="SFXN_beta15_vanilla" jump=1 repack=1 repeats=3 threshold="0.0"
    confidence="0" />
    <Sasa name="interface_sasa" jump="1" confidence=0/>
    <ShapeComplementarity name="interface_sc" verbose=0 min_sc=0.65 write_int_area=1 jump=1
    confidence="0"/>
    <CalculatorFilter name="ddg_100sasa" equation="DDG*hundred/(SASA+small_val)"
    threshold="0.0" confidence="0">
      <DDG name="DDG" filter_name="ddg" />
      <SASA name="SASA" filter_name="interface_sasa" />
      <VARA name="hundred" value=100.0 />
      <VARB name="small_val" value=0.0001 />
    </CalculatorFilter>
    <!--Chain A/B Filters-->
    <MoveBeforeFilter name="score_res_chainB" mover="keep_only_chain_B" filter="score_res"
    confidence="0"/>
    <MoveBeforeFilter name="sspred_chainB" mover="keep_only_chain_B" filter="sspred"
    confidence="0"/>
  </FILTERS>

```

```

<MOVERS>
  <SavePoseMover name="save_RMSDreference_conformation"
reference_name="reference_conformation"/>
  <AddConstraintsToCurrentConformationMover name="constrainCA" cst_weight=0.5 CA_only=1 />
  <ClearConstraintsMover name="clearAllConstraints" />
  <TaskAwareMinMover name="min_kine_sc_chainB" scorefxn="SFXN_beta15_vanilla" bb=1 chi=1
jump=0 cartesian=0 task_operations="init,inclcur,limitchi2,onlyRepack,notChainA" />
  <TaskAwareMinMover name="min_kine_sc_chainAB" scorefxn="SFXN_beta15_vanilla" bb=0 chi=1
jump=0 cartesian=0 task_operations="init,inclcur,limitchi2,onlyRepack" />
  <PackRotamersMover name="pack_sc_chainAB" scorefxn="SFXN_beta15_vanilla"
task_operations="init,inclcur,limitchi2,onlyRepack" />
  <TaskAwareMinMover name="min_cart_chainB" scorefxn="SFXN_beta15_vanilla_cart" bb=1 chi=1
jump=1 cartesian=1 tolerance=0.001 max_iter=1000
task_operations="init,inclcur,limitchi2,onlyRepack,notChainA" />
  <FastRelax name="fast_relax_b_beta" scorefxn="SFXN_beta15_vanilla"
task_operations="init,inclcur,limitchi2,onlyRepack,notChainA" >
    <MoveMap name="mappyfd">
      <Chain number=1 chi=0 bb=0/>
      <Chain number=2 chi=1 bb=1/>
      <Jump number=1 setting=1/>
    </MoveMap>
  </FastRelax>
</MOVERS>

<PROTOCOLS>
  <Add mover_name="save_RMSDreference_conformation" />
  <Add mover_name="pack_sc_chainAB" />
  <Add mover_name="constrainCA" />
    <Add mover_name="min_cart_chainB" />
  <Add mover_name="clearAllConstraints" />
  <Add mover_name="pack_sc_chainAB" />
  <Add mover_name="fast_relax_b_beta" />
  <Add mover_name="min_kine_sc_chainB" />
  <Add mover_name="min_kine_sc_chainAB" />
  <Add mover_name="constrainCA" />
    <Add mover_name="min_cart_chainB" />
  <Add mover_name="clearAllConstraints" />
  <Add mover_name="fast_relax_b_beta" />
  <Add filter_name="rmsd_ALL" />
  <Add filter_name="rmsd_A" />
  <Add filter_name="rmsd_B" />
  <Add filter_name="score_res_chainB" />
  <Add filter_name="ddg" />
  <Add filter_name="interface_sasa" />
  <Add filter_name="ddg_100sasa" />
  <Add filter_name="interface_sc" />
  <Add filter_name="sspred_chainB" />
</PROTOCOLS>

</ROSETTASCRIPTS>

```

Appendix F. RosettaScripts XML used to analyze Rosetta Scores and Metrics over mini-protein Monomers

```

<ROSETTASCRIPTS>
  <SCOREFXNS>
    <SFXN7 weights="beta" >
      </SFXN7>
    </SCOREFXNS>
  <TASKOPERATIONS>
    <LayerDesign name="layer_core_SCN" layer="core" pore_radius="2.0" verbose="true"
use_sidechain_neighbors="True" core=4 />
    <LayerDesign name="layer_core_SASA" layer="core" core="20" pore_radius="2.0"
verbose="true" />
    <LayerDesign name="layer_core_boundary_SCN" layer="core_boundary" pore_radius="2.0"
verbose="true" use_sidechain_neighbors="True" />

    <OperateOnCertainResidues name="no_ala_disulf" >
      <RestrictToRepackingRLT />
      <ResidueName3Is name3="ALA" />
    </OperateOnCertainResidues>

    <OperateOnCertainResidues name="no_repack_non-disulf" >
      <PreventRepackingRLT/>
      <ResidueName3Isnt name3="CYS" />
    </OperateOnCertainResidues>
  </TASKOPERATIONS>

  <MOVERS>
    <SwitchChainOrder name="keep_only_chain_B" chain_order="2"/>
  </MOVERS>

  <FILTERS>
    <AverageDegree name="degree_core_SCN" task_operations="layer_core_SCN" confidence="0"
threshold="9.4" />
    <AverageDegree name="degree_core_SASA" task_operations="layer_core_SASA" confidence="0"
threshold="9.4" />
    <AverageDegree name=degree confidence=0 threshold=9.5/>
    <ResidueCount name="res_count_all" max_residue_count="9999" confidence="0"/>
    <ResidueCount name="res_count_core_SCN" task_operations="layer_core_SCN,no_ala_disulf"
max_residue_count="9999" confidence="0"/>
    <ResidueCount name="res_count_core_SASA" task_operations="layer_core_SASA,no_ala_disulf"
max_residue_count="9999" confidence="0"/>
    <ResidueCount name="nres_cys" residue_types="CYS" confidence="0" />
    <ResidueCount name="AlaCount" residue_types="ALA" max_residue_count="9999"
confidence="0"/>
    <TotalSasa name="total_sasa" threshold="1" upper_threshold="100000000000000"
report_per_residue_sasa="False" confidence="0" />
    <CalculatorFilter name="mean_sasa" equation="Tsasa / (rescount2 + 0.01)" threshold="-0.30"
confidence="0" >
      <VAR name="Tsasa" filter="total_sasa"/>
      <VAR name="rescount2" filter="res_count_all"/>
    </CalculatorFilter>
    <CalculatorFilter name="percent_core_SCN" equation="rescount_coreSCN / (rescount3 + 0.01)"
threshold="-0.35" confidence="0" >
      <VAR name="rescount3" filter="res_count_all"/>
      <VAR name="rescount_coreSCN" filter="res_count_core_SCN"/>
    </CalculatorFilter>

```

```

    <CalculatorFilter name="percent_core_SASA" equation="rescount_coreSASA / (rescount4 +
0.01)" threshold="-0.35" confidence="0" >
      <VAR name="rescount4" filter="res_count_all"/>
      <VAR name="rescount_coreSASA" filter="res_count_core_SASA"/>
    </CalculatorFilter>
    <AtomicContactCount name="contact_all" confidence="0" />
    <AtomicContactCount name="contact_core_SCN" task_operations="layer_core_SCN"
confidence="0" />
    <AtomicContactCount name="contact_core_SASA" task_operations="layer_core_SASA"
confidence="0" />
    <ResidueCount name="nres" confidence="0" />
    <ScoreType name="dslf_fa13" scorefxn=SFXN7 score_type=dslf_fa13 threshold=0/>
    <CalculatorFilter name="mean_dslf" equation="dslf / (cyscount+0.01)" threshold="-0.35"
confidence=0>
      <VAR name="dslf" filter="dslf_fa13"/>
      <VAR name="cyscount" filter="nres_cys"/>
    </CalculatorFilter>

    <DisulfideEntropy name="entropy" lower_bound="0" tightness="2" confidence="0"/>
    <TaskAwareScoreType name="dslf_quality_check" task_operations="no_repack_non-disulf"
score_type="dslf_fa13" mode="individual" threshold="-0.27" confidence="0" />
    <ScoreType name="hbond_sfn" scorefxn=SFXN7 score_type=hbond_lr_bb threshold=0/>
    <SSShapeComplementarity name="ss_sc" verbose="0" confidence="0" />
    <SSShapeComplementarity name="helix_sc" verbose="1" loops="0" helices="1" confidence="0"
/>

    <SSShapeComplementarity name="loop_sc" verbose="1" loops="1" helices="0" confidence="0" />
    <TotalSasa name="exposed_hydrophobics" confidence="0" hydrophobic=True />
    <TotalSasa name="exposed_total" confidence="0"/>
    <TotalSasa name="exposed_polars" confidence="0" polar=True />
    <CalculatorFilter name="fxn_exposed_is_np" equation="exposed / total" threshold="1"
confidence=0>
      <VAR name="total" filter="exposed_total"/>
      <VAR name="exposed" filter="exposed_hydrophobics"/>
    </CalculatorFilter>
    <ScoreType name="total_hydrophobic" scorefxn=TotalHydrophobic threshold=0/>
    <CalculatorFilter name="buried_np" equation="total - exposed" threshold="1" confidence=0>
      <VAR name="total" filter="total_hydrophobic"/>
      <VAR name="exposed" filter="exposed_hydrophobics"/>
    </CalculatorFilter>
    <CalculatorFilter name="buried_over_exposed" equation="buried / (exposed+0.01)"
threshold="1" confidence=0>
      <VAR name="buried" filter="buried_np"/>
      <VAR name="exposed" filter="exposed_hydrophobics"/>
    </CalculatorFilter>
    <CalculatorFilter name="buried_minus_exposed" equation="buried - exposed" threshold="1"
confidence=0>
      <VAR name="buried" filter="buried_np"/>
      <VAR name="exposed" filter="exposed_hydrophobics"/>
    </CalculatorFilter>
    <CalculatorFilter name="bb" equation="hbond / (rescount+0.01)" threshold="-0.23"
confidence=0>
      <VAR name="hbond" filter="hbond_sfn"/>
      <VAR name="rescount" filter="nres"/>
    </CalculatorFilter>
    <CavityVolume name="cavity_volume" />
    <PackStat name="pack" confidence=0 repeats="3" />
    <Holes name="holes" confidence=0/>
    <SSPrediction name="mismatch_probability" confidence="0"
cmd="/work/dadriano/PROGRAMS/psipred/runpsipred_single" use_probability="0" use_svm="0" />
    <BuriedUnsatHbonds name="unsat_hbond" confidence=0 jump_number=0/>

```



```

    <BuriedUnsatHbonds2 name="unsat_hbond2" confidence=0 jump_number=0/>
    <SecondaryStructureHasResidue name="one_core_each" secstruct_fraction_threshold="1.0"
res_check_task_operations="layer_core_SCN" required_restypes="VILMFYW" nres_required_per_secstruct="1"
filter_helix="1" filter_sheet="1" filter_loop="0" min_helix_length="4" min_sheet_length="3"
min_loop_length="1" confidence="0" />
    <SecondaryStructureHasResidue name="two_core_each" secstruct_fraction_threshold="1.0"
res_check_task_operations="layer_core_SCN" required_restypes="VILMFYW" nres_required_per_secstruct="2"
filter_helix="1" filter_sheet="1" filter_loop="0" min_helix_length="4" min_sheet_length="3"
min_loop_length="1" confidence="0" />
    <SecondaryStructureHasResidue name="ss_contributes_core"
secstruct_fraction_threshold="1.0" res_check_task_operations="layer_core_boundary_SCN"
required_restypes="VILMFYW" nres_required_per_secstruct="1" filter_helix="1" filter_sheet="1"
filter_loop="0" min_helix_length="4" min_sheet_length="3" min_loop_length="1" confidence="0" />
    <FragmentLookupFilter name="faulty_fragments_tolerant"
lookup_name="source_fragments_4_mer_tolerant"
store_path="/lab/databases/VALL_clustered/backbone_profiler_database_06032014" lookup_mode="first"
chain="2" threshold="999999" confidence="0" />
    <FragmentLookupFilter name="faulty_fragments" lookup_name="source_fragments_4_mer"
store_path="/lab/databases/VALL_clustered/backbone_profiler_database_06032014" lookup_mode="first"
chain="2" threshold="999999" confidence="0" />
</FILTERS>

<MOVERS>
</MOVERS>

<PROTOCOLS>
    <Add mover_name="keep_only_chain_B" />
    <Add filter_name="res_count_core_SCN" />
    <Add filter_name="res_count_core_SASA" />
    <Add filter_name="percent_core_SCN" />
    <Add filter_name="percent_core_SASA" />
    <Add filter_name="contact_all" />
    <Add filter_name="contact_core_SCN" />
    <Add filter_name="contact_core_SASA" />
    <Add filter_name="degree" />
    <Add filter_name="entropy" />
    <Add filter_name="dslf_quality_check" />
    <Add filter_name="mean_dslf" />
    <Add filter_name="cavity_volume" />
    <Add filter_name="ss_sc" />
    <Add filter_name="helix_sc" />
    <Add filter_name="loop_sc" />
    <Add filter_name="exposed_total" />
    <Add filter_name="exposed_hydrophobics" />
    <Add filter_name="exposed_polars" />
    <Add filter_name="fxn_exposed_is_np" />
    <Add filter_name="holes" />
    <Add filter_name="bb" />
    <Add filter_name="buried_np" />
    <Add filter_name="buried_over_exposed" />
    <Add filter_name="buried_minus_exposed" />
    <Add filter_name="pack" />
    <Add filter_name="mismatch_probability" />
    <Add filter_name="unsat_hbond" />
    <Add filter_name="unsat_hbond2" />
    <Add filter_name="one_core_each" />
    <Add filter_name="two_core_each" />
    <Add filter_name="ss_contributes_core" />
    <Add filter_name="AlaCount" />
</PROTOCOLS>

```

</ROSETTASCRIPTS>

Appendix H. RosettaScripts XML used to analyze Rosetta Scores and Metrics of the complexes mini-protein<->Target

```

<ROSETTASCRIPTS>
  <SCOREFXNS>
    <NOV15 weights="beta_nov15.wts" />
    <VDW weights="empty" >
      <Reweight scoretype="fa_atr" weight=1.0/>
    </VDW>
  </SCOREFXNS>

  <TASKOPERATIONS>
    <ProteinInterfaceDesign name="pack_long" design_chain1="0" design_chain2="0" jump="1"
interface_distance_cutoff="15"/>
    <ProteinInterfaceDesign name="chain2_interface" repack_chain1="0" design_chain2="1"
jump="1" interface_distance_cutoff="8"/>
    <RestrictToInterfaceVector name="restrict_to_interface" jump="1" CB_dist_cutoff="10.0"
nearby_atom_cutoff="5.5" vector_angle_cutoff="75.0" vector_dist_cutoff="9.0"/>
  </TASKOPERATIONS>

  <MOVERS>
    <TaskAwareMinMover name=min scorefxn=NOV15 bb=0 chi=1 task_operations=pack_long/>
    <SwitchChainOrder name=chain1only chain_order="1"/>
    <SwitchChainOrder name=chain2only chain_order="2"/>
  </MOVERS>

  <FILTERS>
    <Ddg name="ddg" threshold=-10 jump=1 repeats=5 repack=1 relax_mover=min confidence=0
scorefxn=NOV15 />
    <Ddg name="ddg_norepack" threshold=-10 jump=1 repeats=1 repack=0 confidence=0
scorefxn=NOV15/>
    <Ddg name="ddg_fa_atr" threshold=-10 jump=1 repeats=5 repack=1 relax_mover=min
confidence=0 scorefxn=VDW />
    <Ddg name="ddg_fa_atr_norepack" threshold=-10 jump=1 repeats=1 repack=0 confidence=0
scorefxn=VDW/>
    <InterfaceHoles name="interface_holes" confidence=0 jump=1/>
    <Sasa name="interface_buried_sasa" confidence=0/>
    <Sasa name="interface_hydrophobic_sasa" confidence=0 hydrophobic=True/>
    <Sasa name="interface_polar_sasa" confidence=0 polar=True/>
    <BuriedUnsatHbonds name="interface_unsat_hbond" confidence=0 jump_number=1/>
    <BuriedUnsatHbonds2 name="interface_unsat_hbond2" confidence=0 jump_number=1/>
    <TotalSasa name="exposed_hydrophobics" confidence="0" hydrophobic=True />
    <MoveBeforeFilter name=exposed_np_chain1 mover=chain1only filter=exposed_hydrophobics/>
    <MoveBeforeFilter name=exposed_np_chain2 mover=chain2only filter=exposed_hydrophobics/>
    <CalculatorFilter name="fxn_np_chain1_buried_approx" equation="(sasa / 2) / (np+0.01)"
threshold="1" confidence="0">
      <VAR name="sasa" filter="interface_hydrophobic_sasa"/>
      <VAR name="np" filter="exposed_np_chain1"/>
    </CalculatorFilter>
    <CalculatorFilter name="fxn_np_chain2_buried_approx" equation="(sasa / 2) / (np+0.01)"
threshold="1" confidence="0">
      <VAR name="sasa" filter="interface_hydrophobic_sasa"/>
      <VAR name="np" filter="exposed_np_chain2"/>
    </CalculatorFilter>
    <CalculatorFilter name="np_chain1_exposed_approx" equation="np - (sasa / 2)" threshold="1"
confidence="0">
      <VAR name="sasa" filter="interface_hydrophobic_sasa"/>
      <VAR name="np" filter="exposed_np_chain1"/>
  </FILTERS>

```

```

    </CalculatorFilter>
    <CalculatorFilter name="np_chain2_exposed_approx" equation="np - (sasa / 2)" threshold="1"
confidence="0">
        <VAR name="sasa" filter="interface_hydrophobic_sasa"/>
        <VAR name="np" filter="exposed_np_chain2"/>
    </CalculatorFilter>
    <CalculatorFilter name="ddg_per_1000sasa" equation="1000 * ddg / (sasa+0.01)"
threshold="1" confidence="0">
        <VAR name="ddg" filter="ddg"/>
        <VAR name="sasa" filter="interface_buried_sasa"/>
    </CalculatorFilter>
    <CalculatorFilter name="ddg_norepack_per_1000sasa" equation="1000 * ddg / (sasa+0.01)"
threshold="1" confidence="0">
        <VAR name="ddg" filter="ddg_norepack"/>
        <VAR name="sasa" filter="interface_buried_sasa"/>
    </CalculatorFilter>
    <CalculatorFilter name="ddg_fa_atr_per_1000sasa" equation="1000 * ddg / (sasa+0.01)"
threshold="1" confidence="0">
        <VAR name="ddg" filter="ddg_fa_atr"/>
        <VAR name="sasa" filter="interface_buried_sasa"/>
    </CalculatorFilter>
    <CalculatorFilter name="ddg_fa_atr_norepack_per_1000sasa" equation="1000 * ddg /
(sasa+0.01)" threshold="1" confidence="0">
        <VAR name="ddg" filter="ddg_fa_atr_norepack"/>
        <VAR name="sasa" filter="interface_buried_sasa"/>
    </CalculatorFilter>
    <CalculatorFilter name="interface_fxn_hydrophobic" equation="hydrophobic / (sasa + 0.01)"
threshold="1" confidence="0">
        <VAR name="hydrophobic" filter="interface_hydrophobic_sasa"/>
        <VAR name="sasa" filter="interface_buried_sasa"/>
    </CalculatorFilter>
    <AverageDegree name="interface_averageDegree" threshold="8.3"
task_operations=chain2_interface confidence="0"/>
    <ShapeComplementarity name="interface_sc" verbose=0 min_sc=0.55 write_int_area=1 jump=1
confidence=0/>
    <AtomicContactCount name="interface_contactcount" task_operations="restrict_to_interface"
confidence="0" />
    </FILTERS>

    <MOVERS>
    </MOVERS>

    <PROTOCOLS>
    <Add filter_name="interface_holes"/>
    <Add filter_name="interface_unsat_hbond" />
    <Add filter_name="interface_unsat_hbond2" />
    <Add filter_name="interface_buried_sasa" />
    <Add filter_name="interface_hydrophobic_sasa" />
    <Add filter_name="interface_polar_sasa" />
    <Add filter_name="interface_fxn_hydrophobic" />
    <Add filter_name="ddg" />
    <Add filter_name="ddg_norepack" />
    <Add filter_name="ddg_per_1000sasa" />
    <Add filter_name="ddg_norepack_per_1000sasa" />
    <Add filter_name="ddg_fa_atr" />
    <Add filter_name="ddg_fa_atr_norepack" />
    <Add filter_name="ddg_fa_atr_per_1000sasa" />
    <Add filter_name="ddg_fa_atr_norepack_per_1000sasa" />
    <Add filter_name="fxn_np_chain1_buried_approx" />
    <Add filter_name="fxn_np_chain2_buried_approx" />

```

```
<Add filter_name="np_chain1_exposed_approx" />  
<Add filter_name="np_chain2_exposed_approx" />  
<Add filter_name="interface_averageDegree" />  
<Add filter_name="interface_contactcount" />  
<Add filter_name="interface_sc" />  
</PROTOCOLS>  
  
</ROSETTASCRIPTS>
```