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G



Assay conc.(nM)

Figure S1. Characterization of ascr#3 imprinting. *Related to Figure 1*.

(A) Percentage of reversal of naive and pre-exposed adult animals performing short reversal, long reversal, or omega turn (See STAR methods). *** indicates different from naive at p<0.001 by one-way ANOVA with Bonferroni's post hoc test. n=80-100 each. Error bars represent SEM. (B) Learning index of males. *** indicates different from hermaphrodites at p<0.001 by Student's t-test. n=60. Error bars represent SEM. (C) Learning index of adult animals that are pre-exposed to, and assayed with ascr#2. n=50 each. Error bars represent SEM. (D) Percentage of reversal of adult animals that are pre-exposed to ascr#3 and assayed with 200 mM glycerol. *** indicates different from naive at 0.001 by one-way ANOVA with Bonferroni's post hoc test. n=60. Error bars represent SEM. (E) Percentage of reversal of wild-type post-dauer animals. The adults are recovered from the dauer stage induced by 600 nM ascr#3 exposure at 25°C. n=30-40 each. Error bars represent SEM. (F) Learning index of 1, 3, or 6 day-old adults. n=30 each. Error bars represent SEM. (G) Learning index of F1 progeny of ascr#3 imprinted animals imprinting. ** indicates different from P0 at p<0.01 by Student's t-test. n=40 each. Error bars represent SEM.











Behavior patterns

Figure S2. Screen of candidate genes with a potential role in ascr#3 imprinting. Related to Figure 2.

(A) Learning index (top) and percent of reversal (bottom) of *egl-4, sra-11, odr-3, odr-7, odr-10, casy-1, ttx-3,* and *tdc-1* mutant animals. *, ** and *** indicate different from WT at p<0.05, 0.01 and 0.001, respectively, by one-way ANOVA with Bonferroni's post hoc test. n=30-260 each. Error bars represent SEM. (B) Percentage of reversal of wild-type and *odr-2* mutants performing short reversal, long reversal, or omega turn. *** indicates different from short reversal at p<0.001 by one-way ANOVA with Bonferroni's post hoc test. n=40-60 each. Error bars represent SEM.



Figure S3. Ca²⁺ transients of the AIB and AVD interneurons in response to ascr#3 exposure. *Related to Figure 3.*

(A-B) Ca^{2+} transients of AIB (A) and AVD (B) in response to 100 nM ascr#3 exposure. The average traces of Ca^{2+} responses to 100 nM ascr#3 (left), heat map of individual Ca^{2+} responses to ascr#3 (bottom), and the maximum value of Ca^{2+} responses to 100 nM ascr#3 (right) in naive (shown in blue) and pre-exposed animals (shown in red) are shown. n=7-12 (naive) and 8 (pre-exposed) each. Error bars represent SEM.



Figure S4. Characterization of the SMB Ca²⁺ responses to ascr#3 exposure. *Related to Figure 4*.

(A) Ca²⁺ transients of the SMB neurons in response to 100 nM ascr#3 in wild-type animals treated with levamisole. The average traces of Ca²⁺ responses to 100 nM ascr#3 (left) and the maximum of Ca²⁺ responses to 100 nM ascr#3 (right) in naive (shown in blue) and pre-exposed animals exposed to ascr#3 (shown in red) and naive animals exposed to S-basal (shown in black) are shown. ** indicates different from naive at p<0.01 by Student's t-test. n=30 (naive and pre-exposed) and 16 (S-basal) each. Error bars represent SEM. (B) Ca²⁺ transients in SMB of animals expressing ADLp::*TeTx*. Wild-type animals without or with an ADLp::*TeTx* transgene are shown in red or black, respectively. ** indicates different from pre-exposed ADLp::*TeTx* transgenic animals at p<0.01 by Student's t-test. n=13 (pre-exposed without ADLp::*TeTx*) and 30 (pre-exposed with ADLp::*TeTx*). Error bars represent SEM.