

Supplementary Information

TGF β 1 reinforces arterial aging in the vascular smooth muscle cell through a long-range regulation of the cytoskeletal stiffness

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The authors declare no conflict of interest

Author Contributions: S.S.A., M.W., and E.G.L. developed the study concept and design. R.M. established and maintained primary VSM cell cultures. W.Z., A.I., J.H., and N.M.B. performed single-cell mechanics and molecular studies. B.C.K. performed single-molecule biophysics studies. W.Z., J.H., and B.C.K. performed data analysis and interpretation. M.W., T.H., and E.G.L. performed data analysis and interpretation, and manuscript preparation. W.Z., B.C.K., M.W., and S.S.A. wrote the manuscript. S.S.A. and E.G.L. directed all studies.

Acknowledgments: This work was supported by funds from the US National Heart, Lung, and Blood Institute (HL107361 to S.S.A.), the Howard Hughes Medical Institute (HHMI) (B.C.K and T.H), and the National Science Foundation Physics Frontier Center (PHY 1430124 to T.H.). S.S.A was also supported by Discovery Award and Catalyst Award from the Johns Hopkins University.

Figure Legends:

Supplementary Figure 1: Cellular models of physiologic arterial aging in VSM. (a) Photomicrographs (X400) of young and old rat aortic wall: elastin fiber (Verhoeff's elastic fiber stain, dark blue) and vascular smooth muscle cells (immunostaining against α -smooth muscle actin, brown color). (b-c) Representative immunofluorescence of smooth muscle cell markers (b, SM-MHC; c, α SMA) in young and old VSM cells. F-actin is stained with phalloidin and nucleus with DAPI. (d) Proliferation (measured by cell count) of young and old VSM cells. Data are presented as Mean \pm SD (n = 3).

Supplementary Figure 2: Dynamics of CSK remodeling in isolated VSM cells. (a) Representative bright field image of isolated VSM cells in culture, showing functionalized RGD-coated ferrimagnetic beads (~ 4.5 μ m in diameter). (b) The trajectories of *spontaneous* bead motions recorded over the course of 5 min. For clarity, only few representative tracings are depicted from the same point of origin (10X objective, n = 20 beads). (c) Computing mean square displacement (MSD) of all beads as function of time t . Data are presented as Mean \pm SE (n = 501-563 individual cell measurements). (d) Corresponding MSD measured at 10s and 300s, respectively.

Supplementary Figure 3: CSK remodeling and stiffness of isolated VSM cells adhered to elastic substrate with varying rigidity. (a, c) Computed MSD of young *vs.* old VSM cells plated on elastic gels (a, 1 kPa 'softer'; c, 8 kPa 'stiffer') as measured by SNTM. Data are presented as Mean \pm SE (n = 9-15 individual cell-wells, comprising ~ 878 -1346 individual beads measurements). (b, d) The elastic modulus g' (stiffness) and the loss modulus g'' (friction)

measured over 5 decades of frequency with MTC in adherent VSM cells on **(b)** ‘softer’ and **(c)** ‘stiffer’ matrices. Data are presented as Geometric Mean \pm SE (n = 67-370 individual cell measurements).

Supplementary Figure 4: Effects of cytochalasin-D and blebbistatin on the stiffness of isolated VSM cells. VSM cells (young and old) were treated for 24 h with or without 5ng/ml TGF β 1, and dynamic changes in cell stiffness in response to **(a)** cytochalasin-D (1 μ M, F-actin inhibitor) and **(b)** blebbistatin (10 μ M, myosin II inhibitor) were measured by MTC. For each individual cell, baseline stiffness was measured for the first 60 s, and after drug addition stiffness was measured continuously for the next 540 s. For each cell, stiffness was normalized to its baseline stiffness before the agonist stimulation. Data are presented as Mean \pm SE (n = 212-321 individual cell measurements per group). **, $P < 0.01$; ****, $P < 0.0001$.

Supplementary Figure 5: This figure displays the full-length gel/blot for Figure 2b shown in the text/Results and Discussion. The identification of **(a)** Smad2/3 and **(b)** p-Smad2/3 bands was based on the expected molecular weight, which was reported in other publications. Panels **a** and **b** are from the same gel/blot. Membrane was cut before probing for Smad2/3 and GAPDH, and re-blotted using p-Smad2/3 antibodies. The blots also show inhibition of TGF β 1-induced Smad2/3 phosphorylation in cells (young and old VSM) treated with TGF β 1 receptor I inhibitors (10 μ M A8301 or 10 μ M GW788388).

Supplementary Figure 6: This figure displays the partial-length cut **(a)** and the full-length **(b)** gels/blots for Figure 4b shown in the text/Results and Discussion using anti-TGF β 1 from Santa

Cruz (SC-146) and R&D Systems (MAB240), respectively. The identification of TGF β 1 bands were based on the expected molecular weight, which were shown in other publications.

Supplementary Figure 7: Integrin profiling in isolated VSM cells. mRNA levels of integrins are expressed relative to young VSM cells. Data are normalized to β -actin and presented as Mean \pm SD (n = 3). #, $P < 0.05$ (untreated vs. TGF β 1 treated for the respective group); *, $P < 0.05$ (young vs. old).

Supplementary Figure 8: Expression of $\alpha_5\beta_1$ and $\alpha_v\beta_3$ in isolated VSM cells. (a) This figure displays the full-length gel/blot showing protein expression levels of $\alpha_5\beta_1$. The identification of α_5 and β_1 subunit bands was based on the expected molecular weight, which was shown in other publications. The same gel/blot was re-blotted using anti-GAPDH antibody for loading control. (b) Immuno-staining of integrin $\alpha_5\beta_1$ and $\alpha_v\beta_3$ expressed in VSM cells. Young VSM cells (with/without 5ng/ml of exogenous TGF β 1) and old VSM cells were stained with an antibody targeting $\alpha_5\beta_1$ or $\alpha_v\beta_3$. Scale bar is 50 μ m. Fluorescence intensities of $\alpha_5\beta_1$ and $\alpha_v\beta_3$ are shown in the bar graph below. ****, $P < 0.0001$.

Supplementary Figure 9: The principle of TGT assay. (a) A schematic of TGT assay. (b) Geometries and estimated tolerances of dsDNA tethers (not drawn in scale).

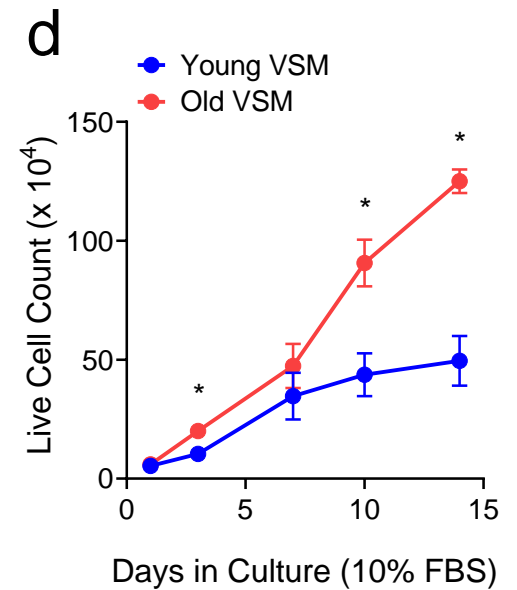
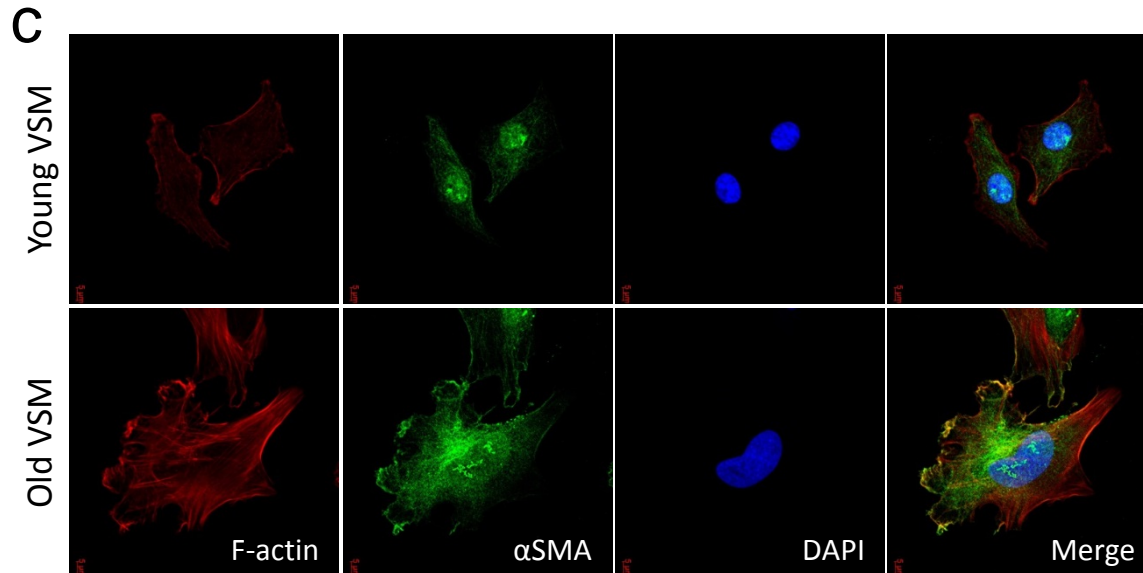
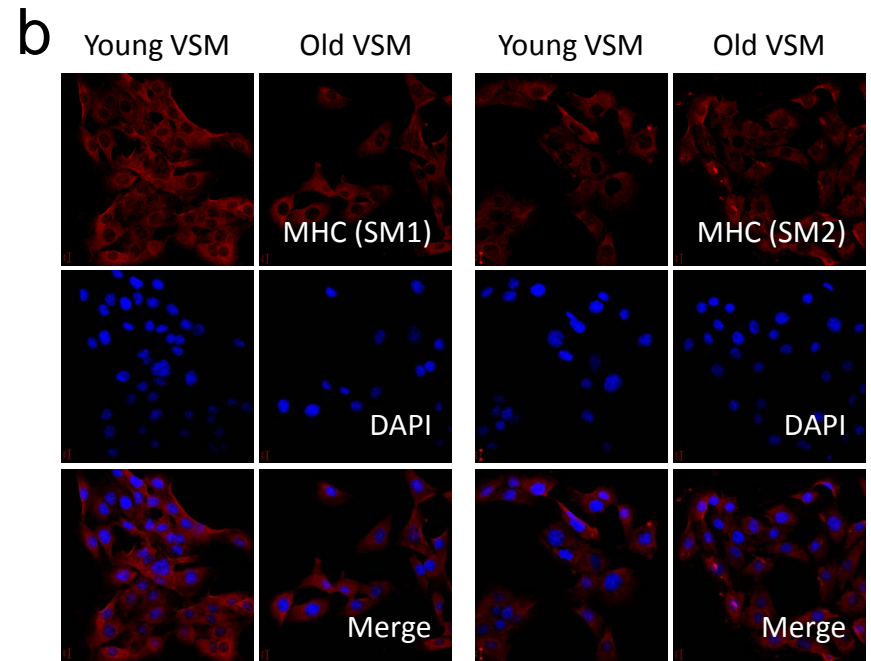
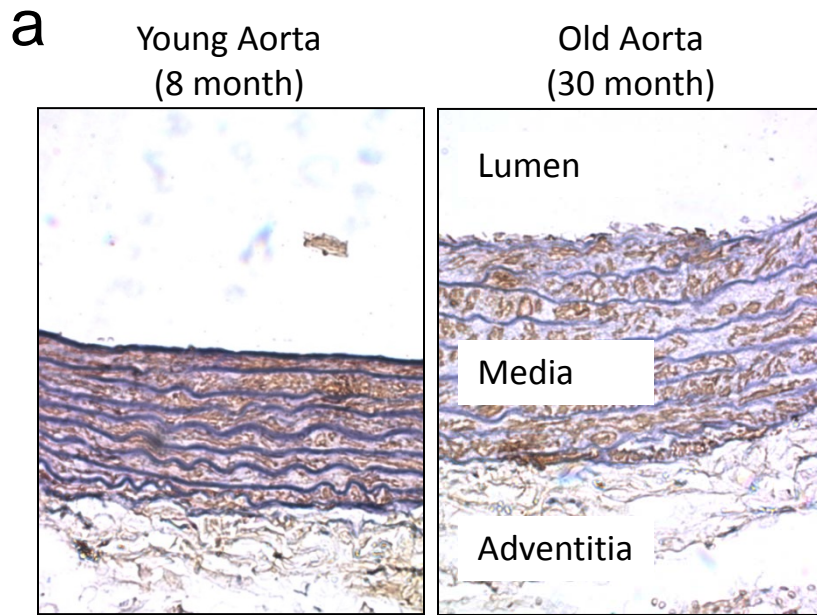
Supplementary Figure 10: Initial cell adhesion on TGT surfaces (incubation time, t = 30 min). (a) Total number of adherent VSM cells on 23 and 54 pN TGT surfaces. The number of adherent cells after 30 min incubation was counted. The TGT assay in each condition was

repeated four times. Data are presented as Mean \pm SE (n = 4). **(b)** Measured projected area of each adherent VSM cell on 23 and 54 pN TGT surfaces (n = 16-92 individual cells). Data are presented as Mean \pm SE. *, $P < 0.05$; **, $P < 0.01$; ***, $P < 0.001$; ****, $P < 0.0001$. **(c)** TGT rupture patterns marked by an individual VSM cell at the ligand-receptor contact area. The dashed lines in each DIC image indicate the periphery of each cell. Scale bar is 25 μ m.

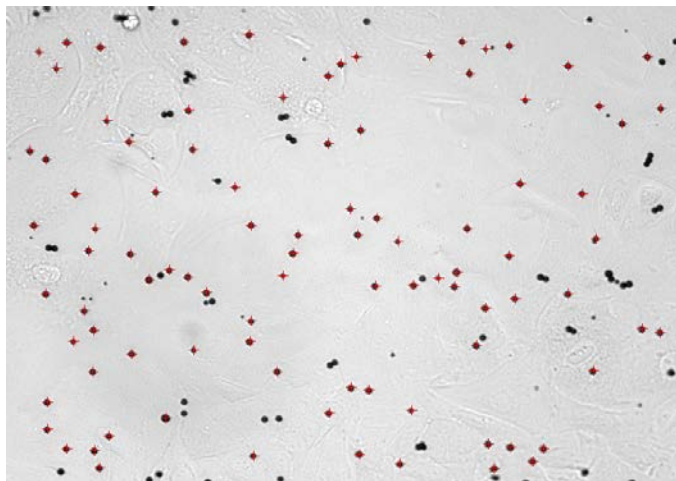
Supplementary Figure 11: Focal adhesion formation on the 54 pN TGT surface. Actin and vinculin were stained in young VSM cells treated for 24 h with or without 5ng/ml TGF β 1. Focal adhesion was produced predominantly at the cell periphery, and corresponded to TGT ruptures shown in the Cy3 channel. TGF β 1 accelerated focal adhesion formation in the young VSM cells, resulting in higher forces above 54 pN at the location of the focal adhesion.

Supplementary Figure 12: TGF β 1-induced stress maps in isolated VSM cells. VSM cells (young and old) were treated for 24 h with or without 5ng/ml TGF β 1, and traction maps were measured with FTTM. For these studies, we tuned the elastic gels to ~20 kPa to mimic physiological stiffness of arterial wall. Computed root mean square (RMS) traction data are presented as Mean \pm SE (n=21-27 individual cells per group). *, $P < 0.05$; ***, $P < 0.001$.

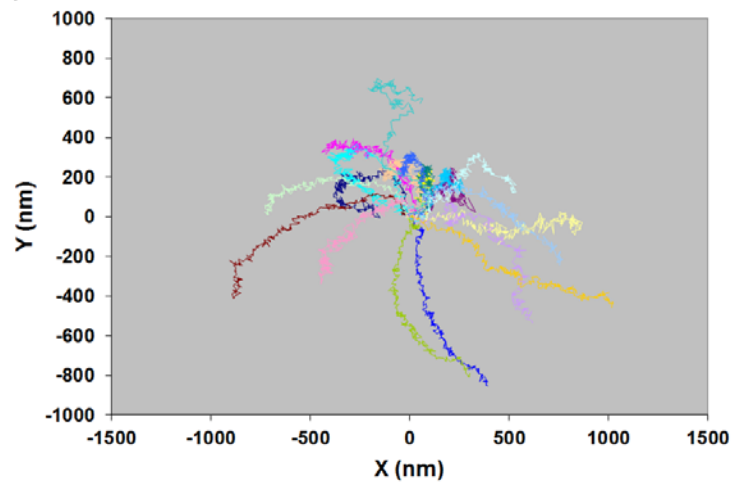
Suppl. Fig. 1



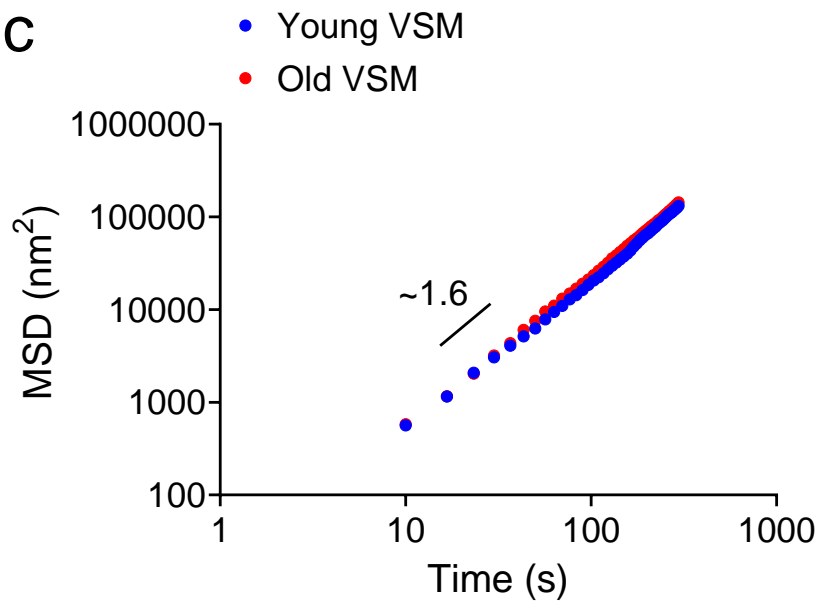
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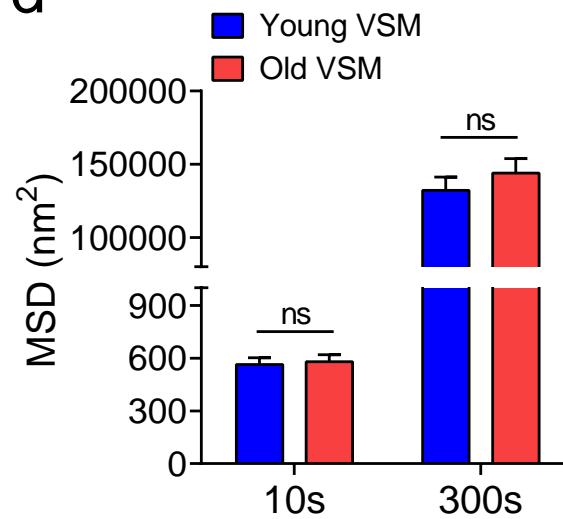
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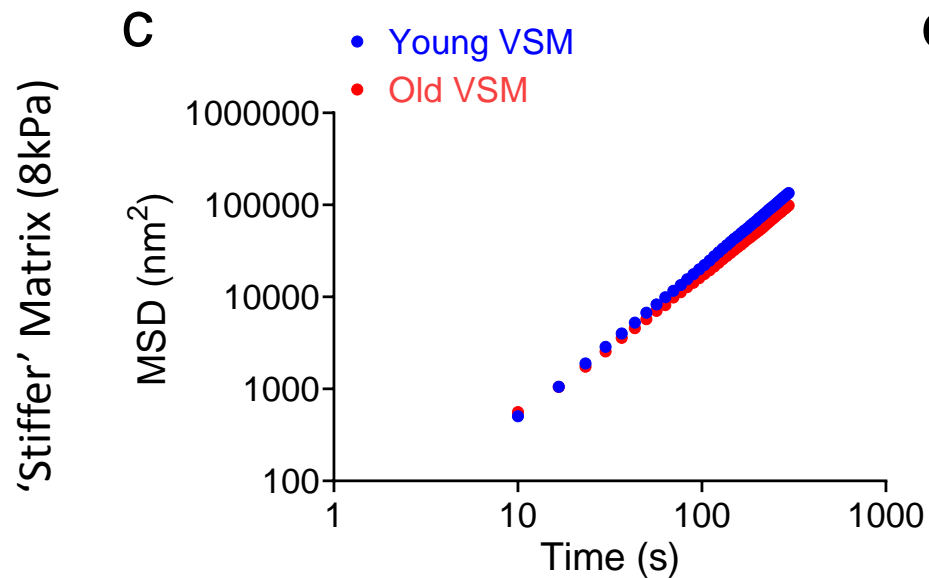
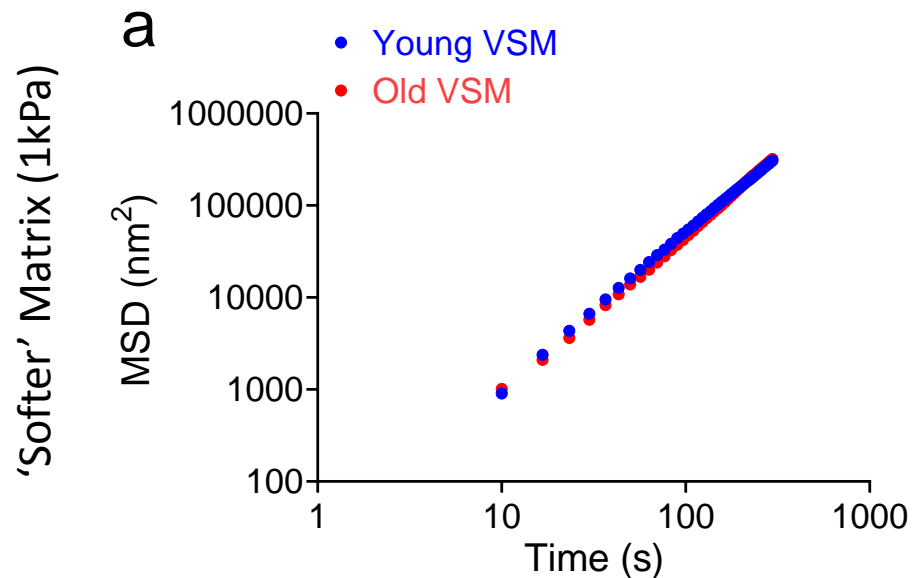
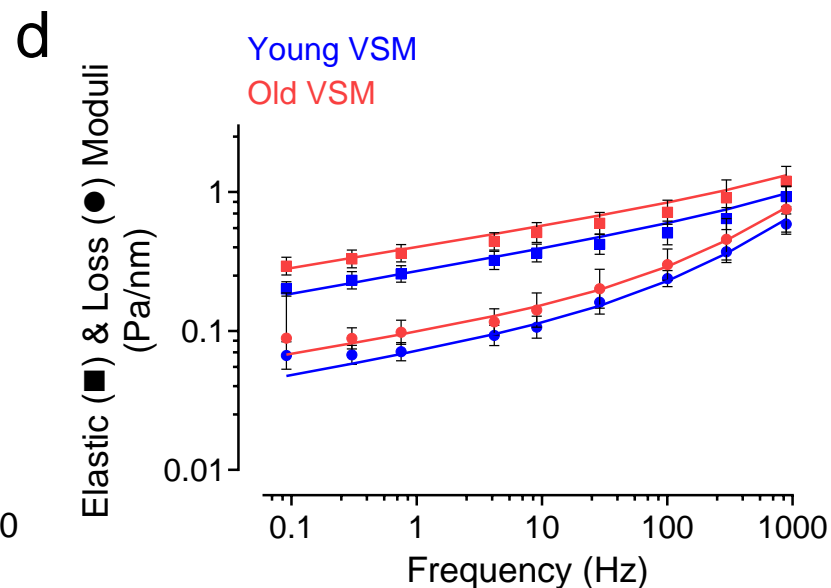
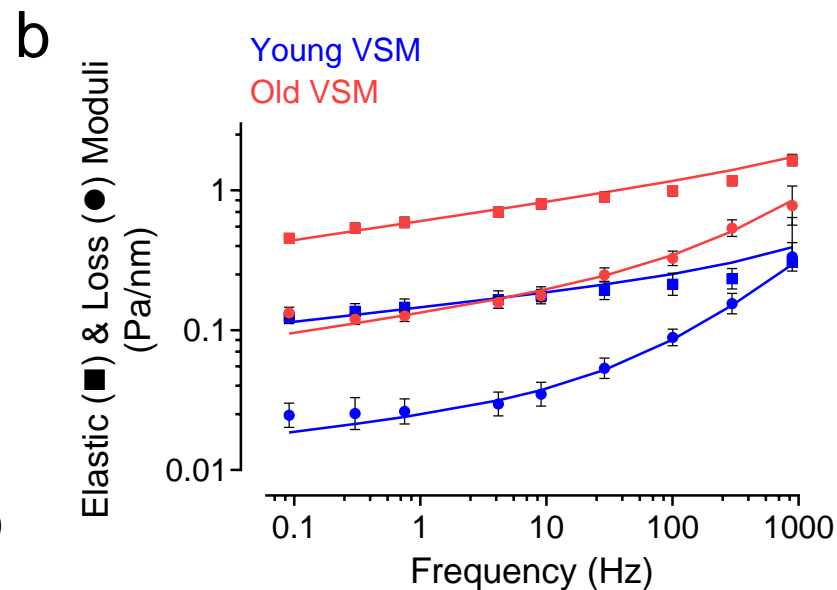


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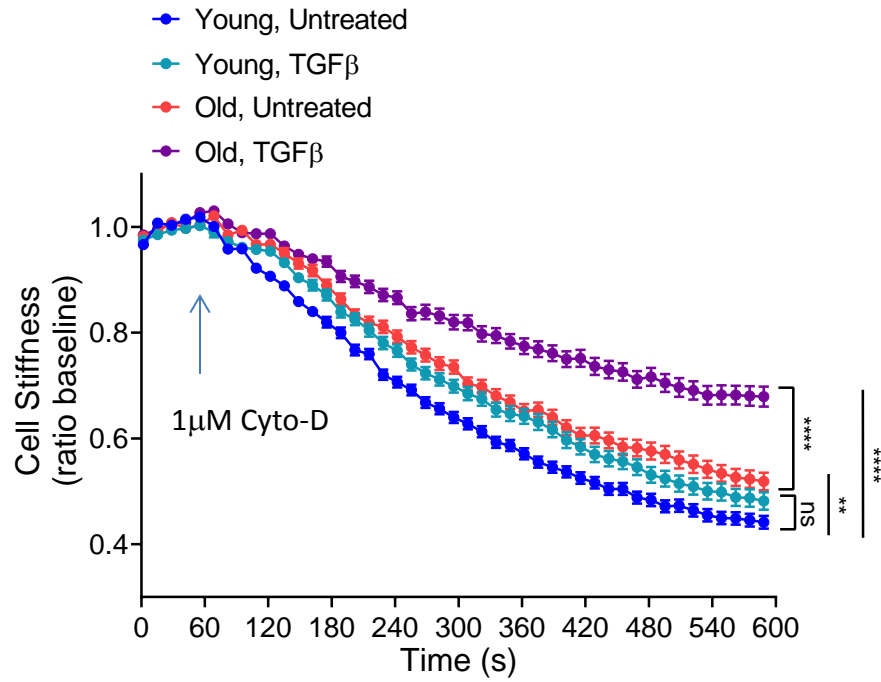


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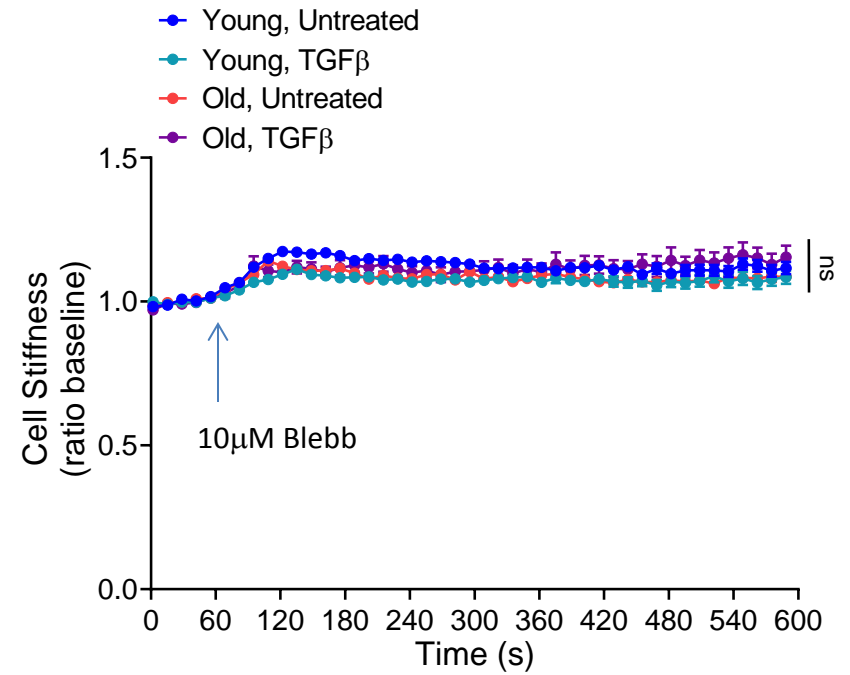


Spontaneous Bead Motions*Forced* Bead Motions

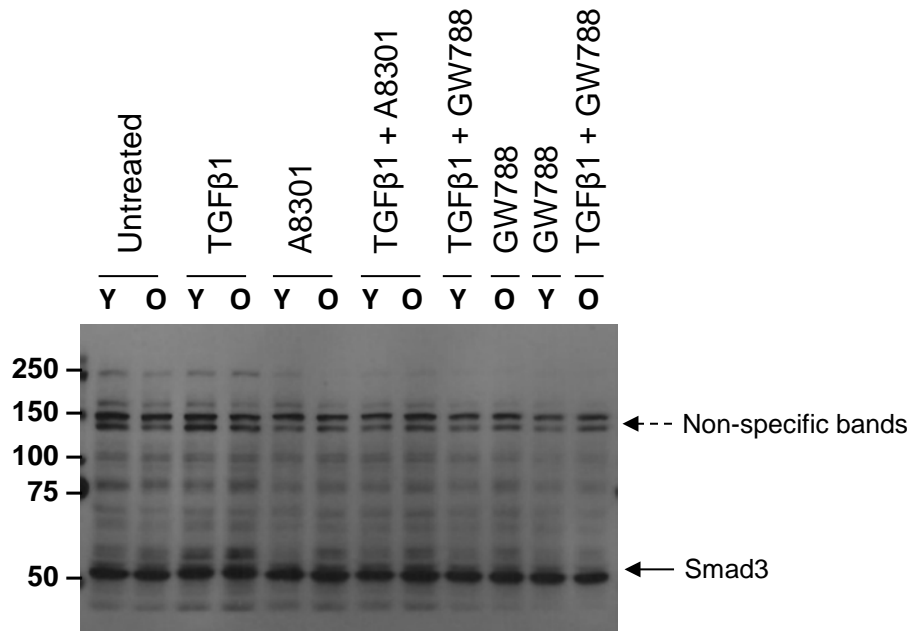
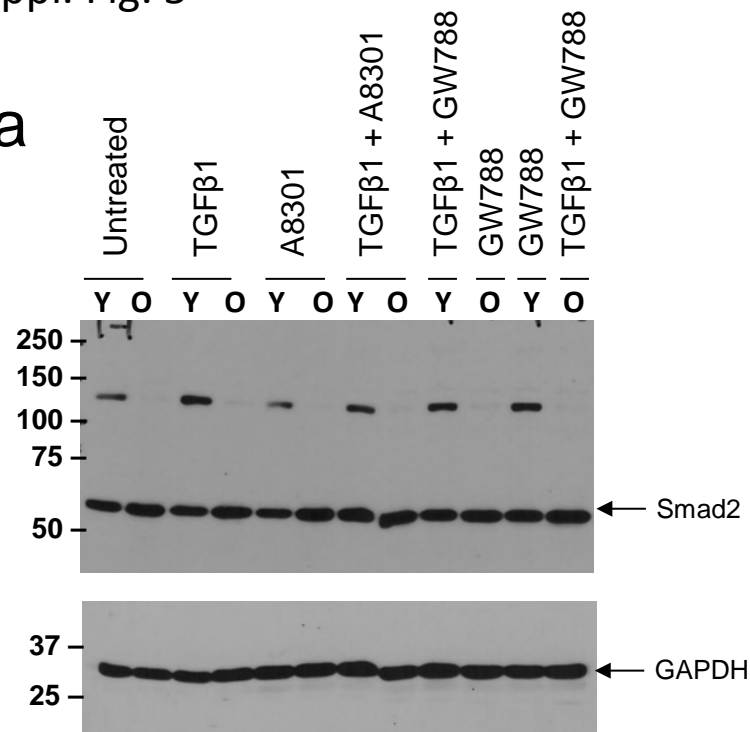
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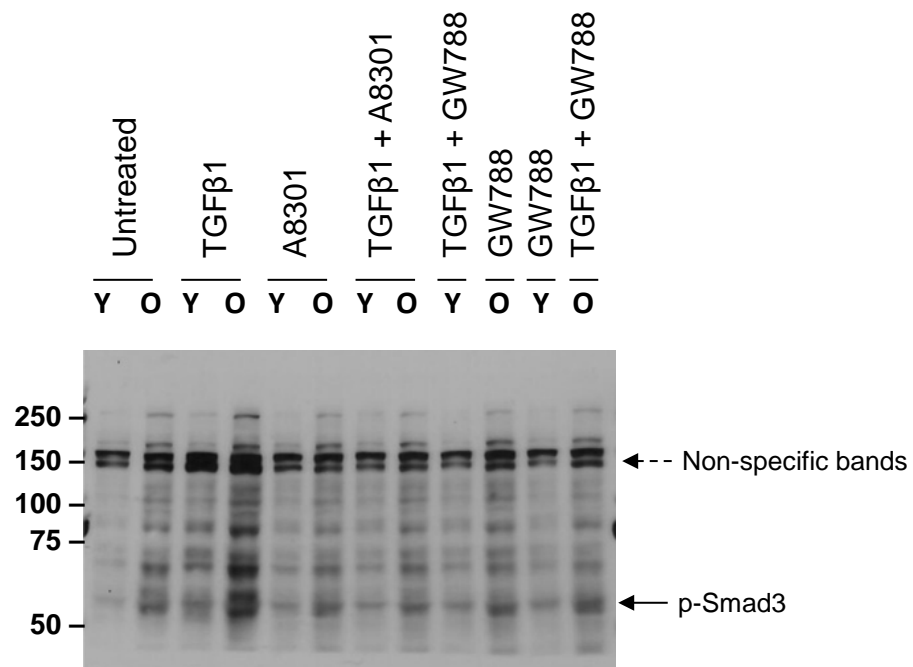
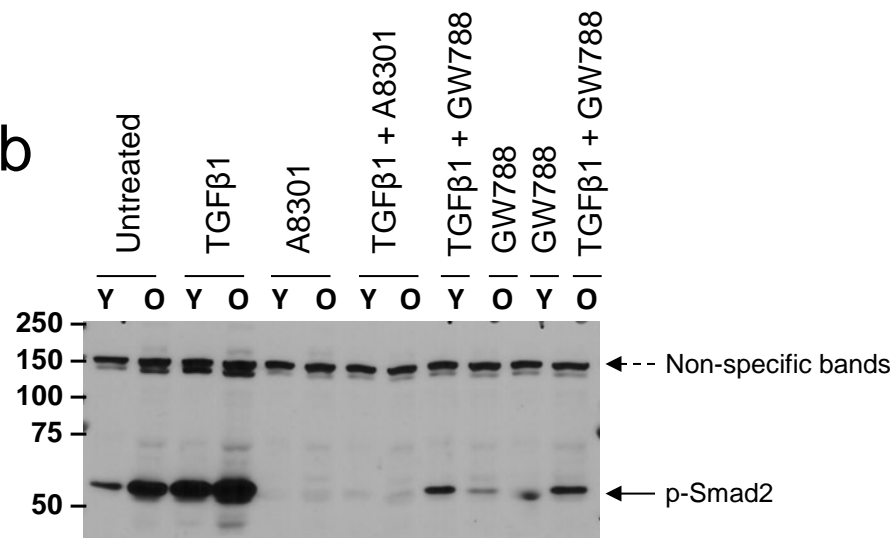
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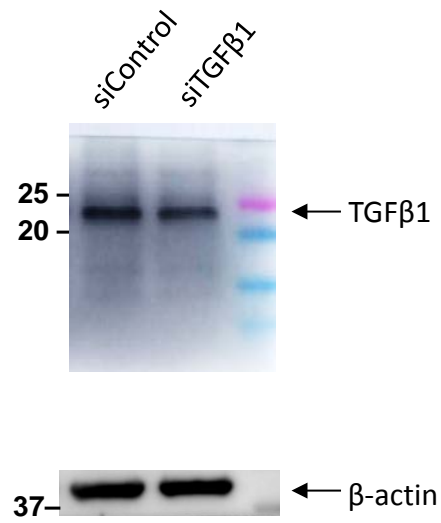
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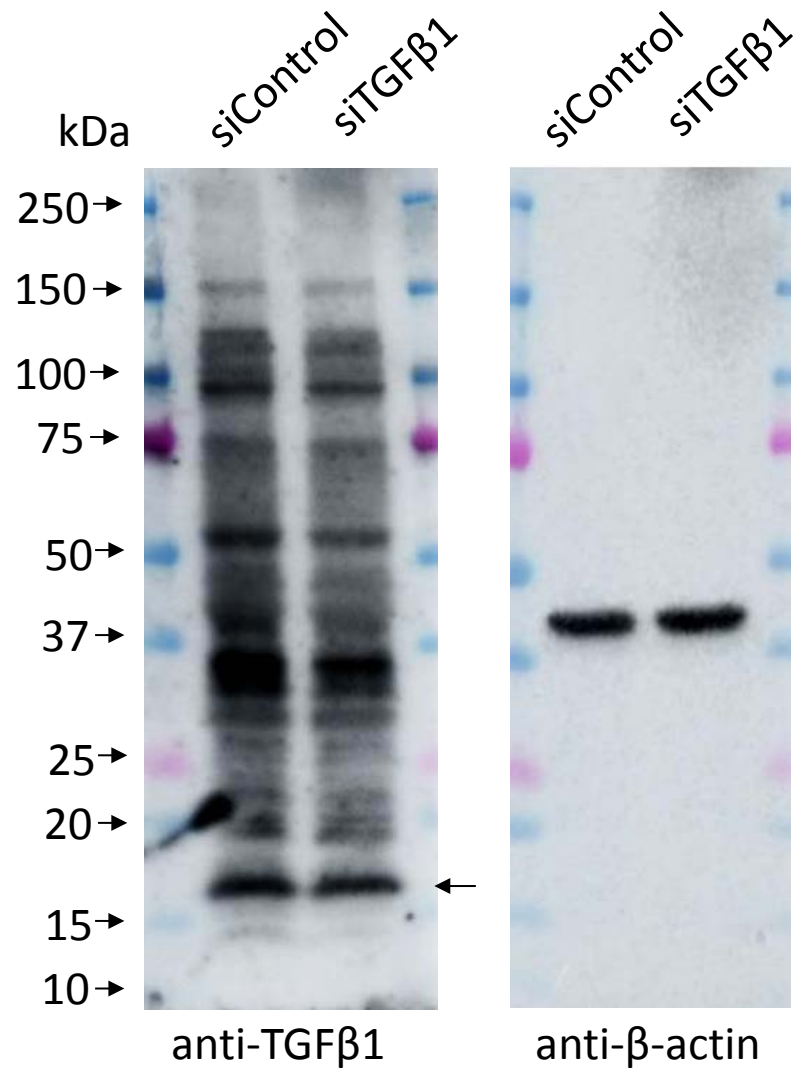
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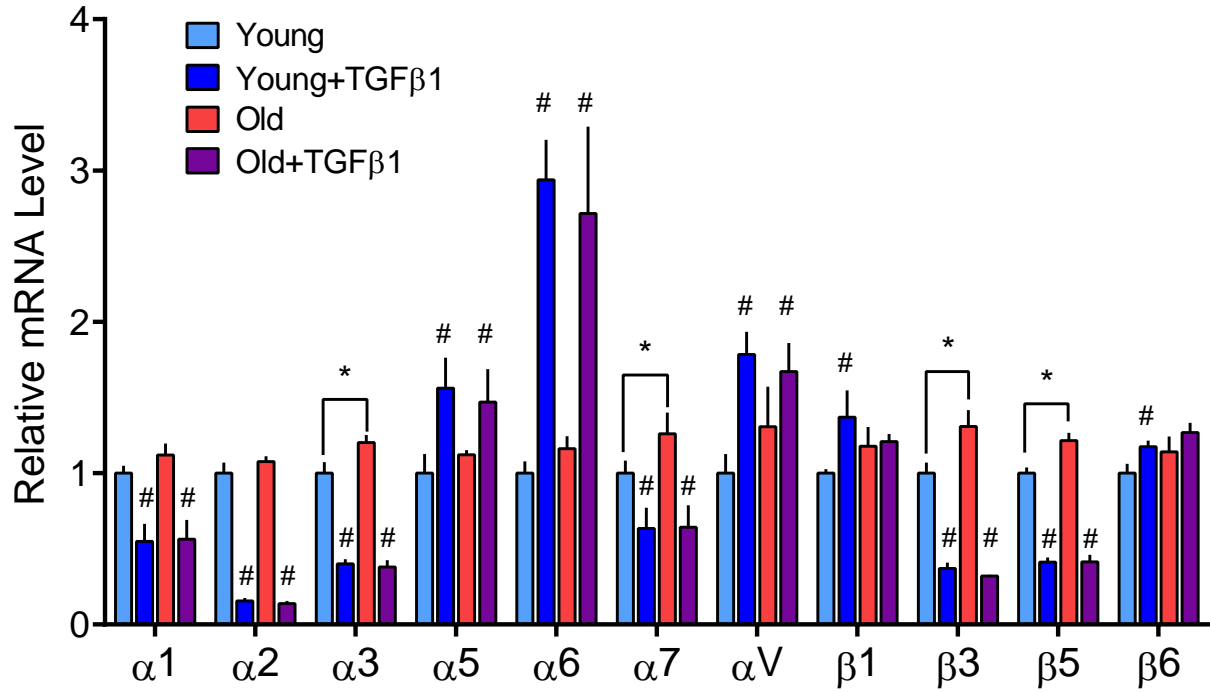
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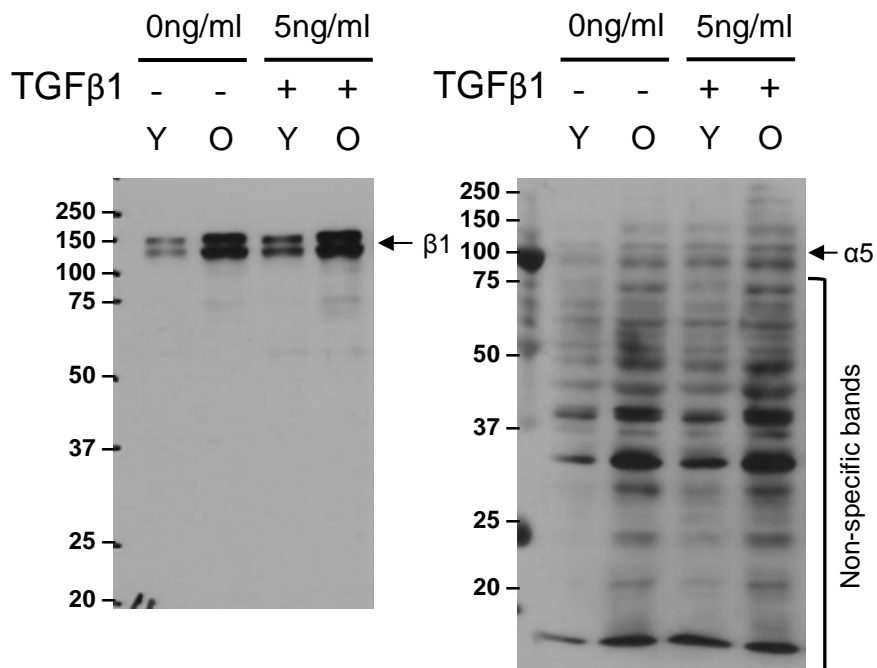
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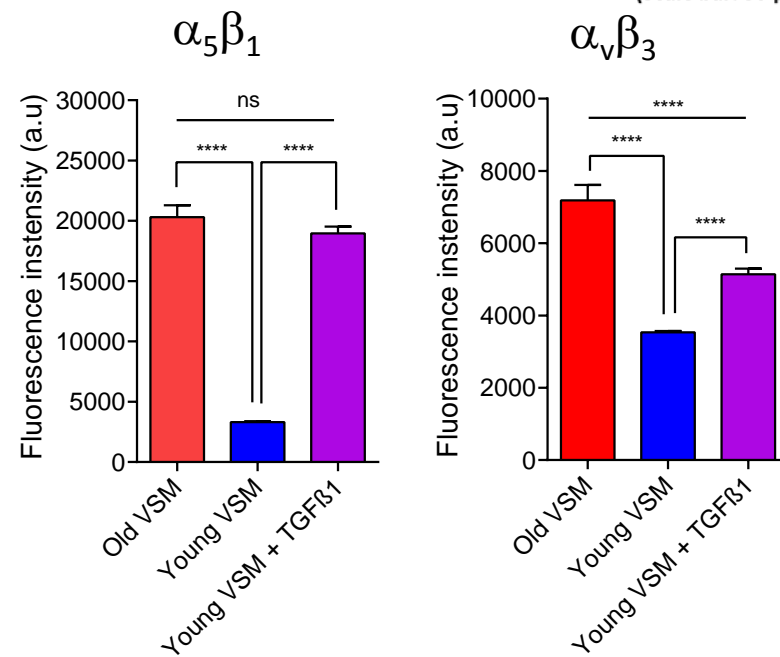
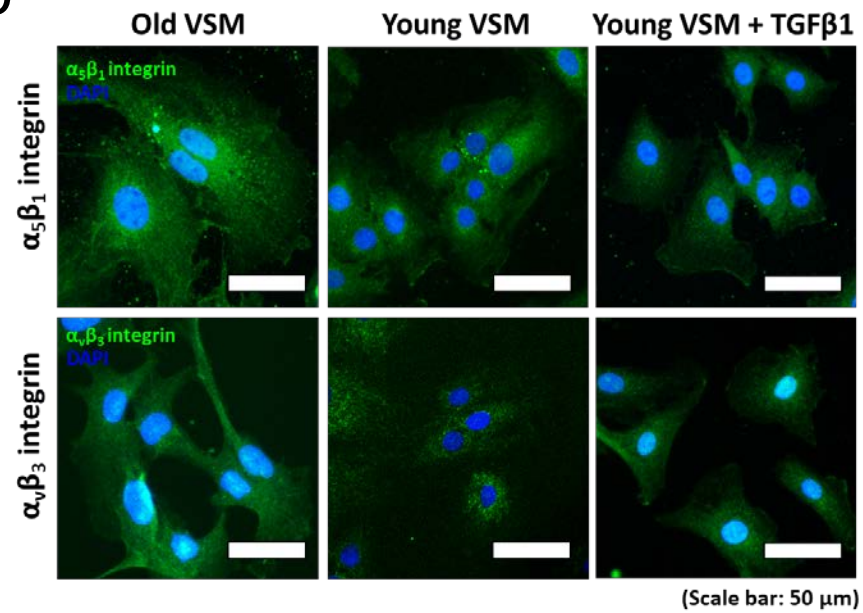
Suppl. Fig. 7

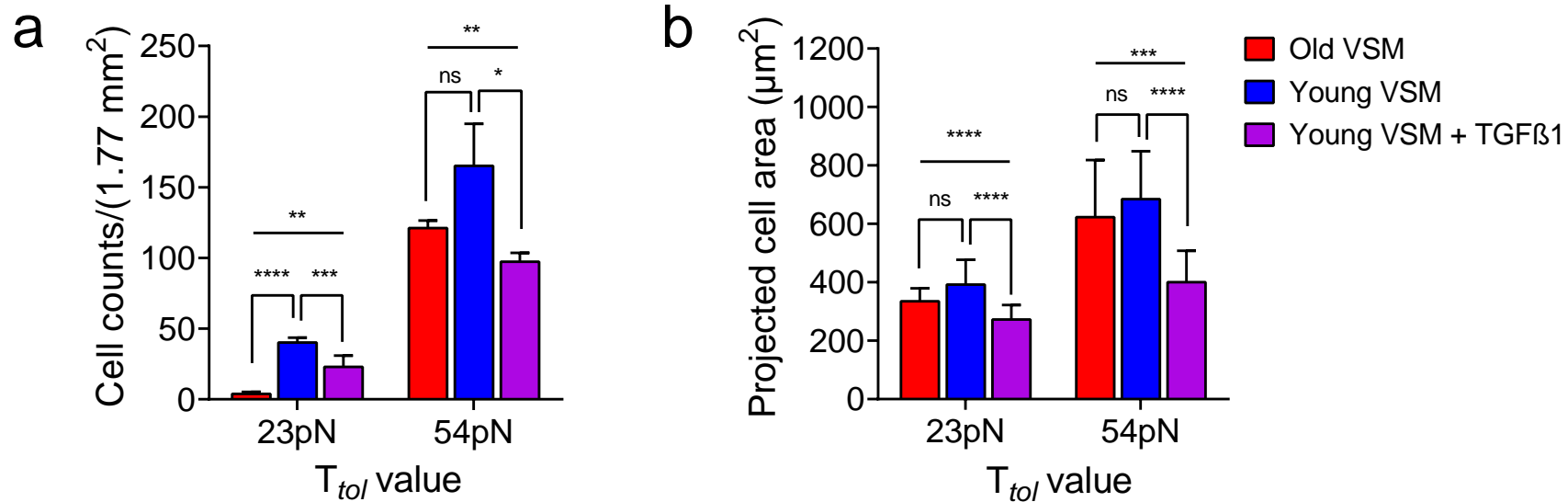
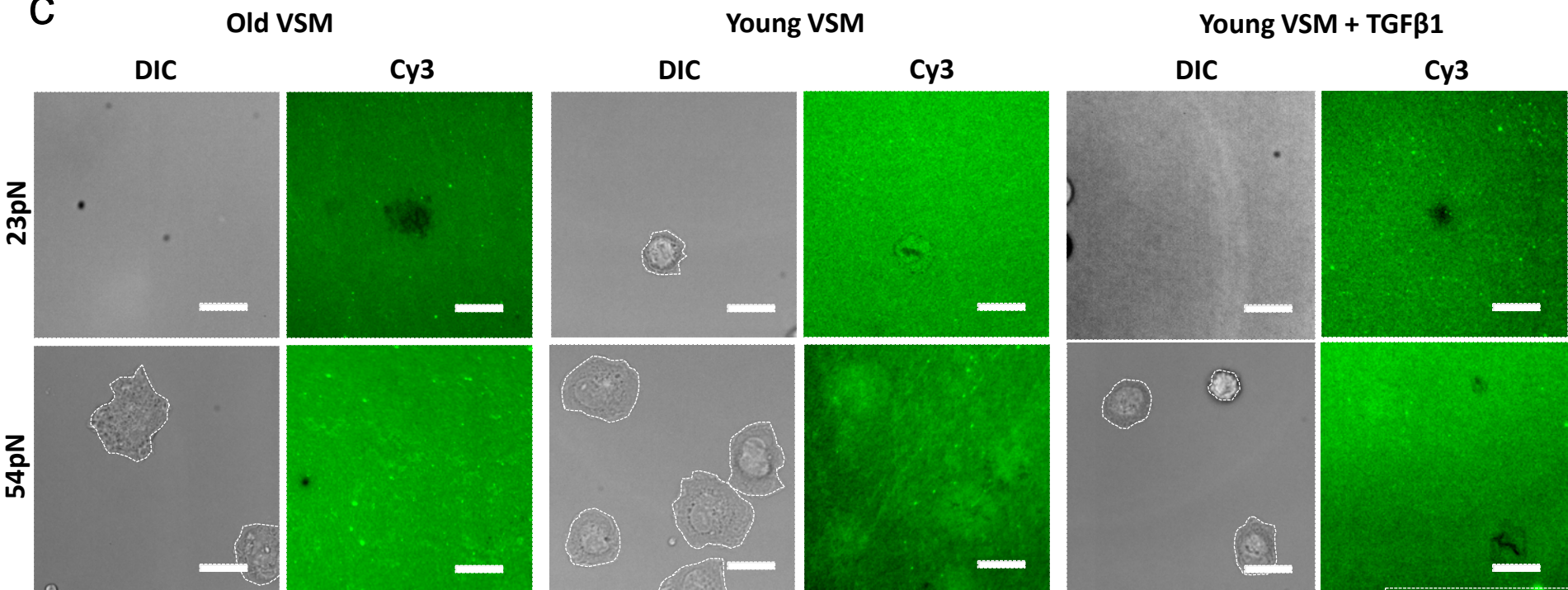


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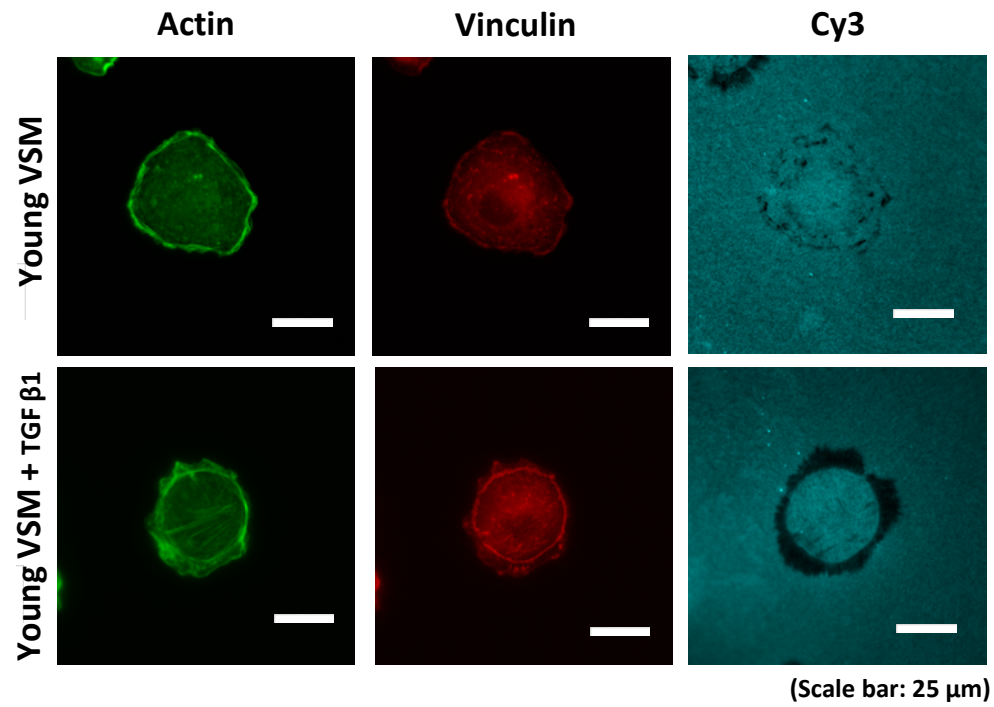


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**c**

(Scale bar: 25 μm)



Suppl. Fig. 12

