

Supplementary information

Quantitative analysis of receptor-mediated uptake and pro-apoptotic activity of mistletoe lectin-1 by high content imaging

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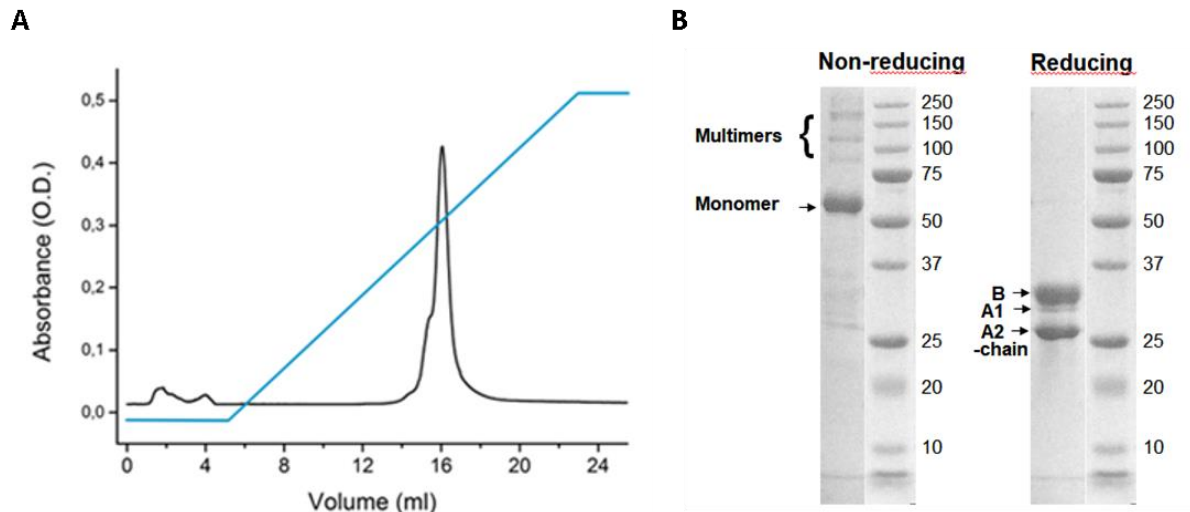


Figure S1 A. FPLC elution profile for ML1 (0.6M NaCl gradient). The main peak represents purified ML1 cytotoxin, with a small left shoulder comprising the variant (A1) of the apoptotic A chain typical for winter mistletoe harvest. B. SDS-PAGE analysis of ML1 under non-reducing and reducing conditions. Monomer ML-1 and multimers of ML-1 are observed under non-reducing conditions. Disruption of disulfide bonds under reducing SDS-PAGE conditions separates the covalently linked A and B chain of ML-1 into 26 kDa and 30 kDa fragments, respectively. A third band was detected at 29 kDa, which is a variant (A1) of the A chain found in winter harvests of the mistletoe plant.

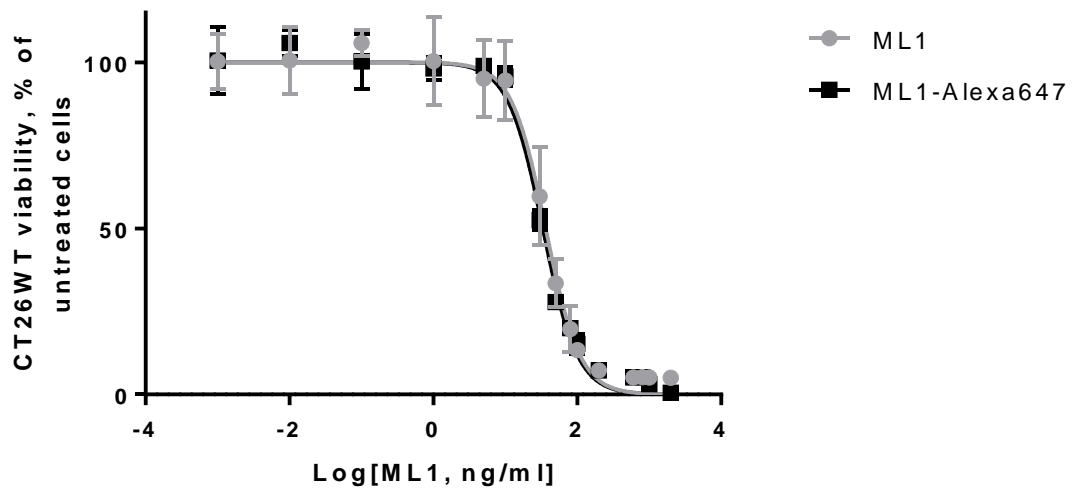


Figure S2. Comparative cytotoxicity of native and Alexa Fluor 647 labeled ML1. Respective IC50 values are 36,6 and 33,2 ng/mL.

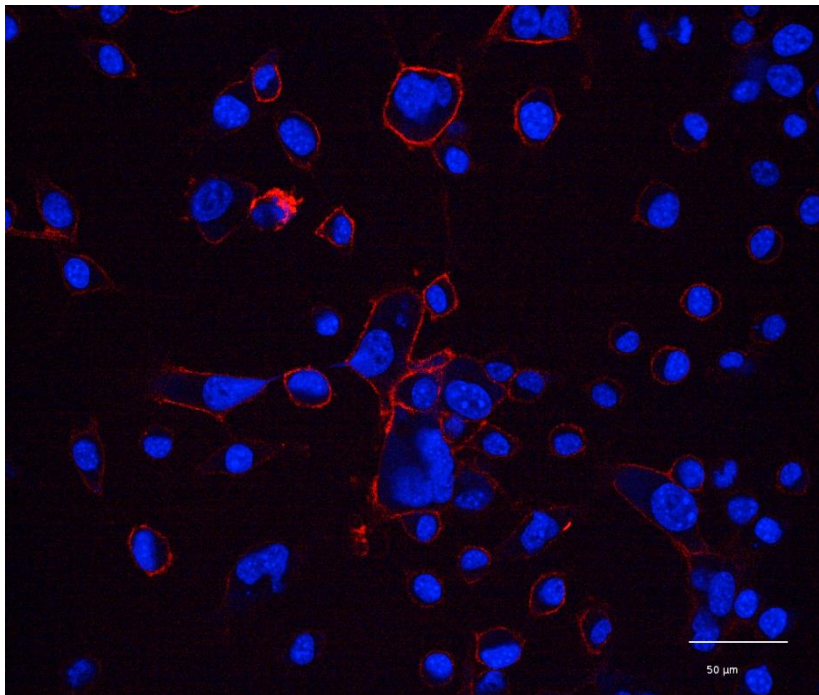


Figure S3. Confocal image of CT26 cells incubated with ML1 at 4°C for 1h (10 µg/mL). Nuclei are stained in blue; size bar – 50 µm.

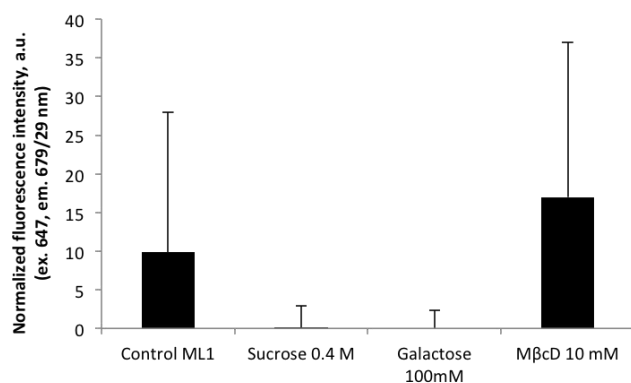
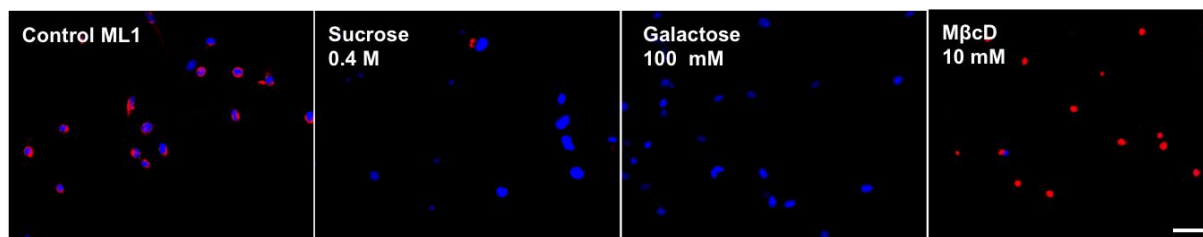


Figure S4. Confocal images of CT26 cells pre-incubated with uptake inhibitors and then incubated with ML1 for 2h (10 $\mu\text{g}/\text{mL}$) (up); Average ML1 fluorescence (n=4) in the cells quantified with Columbus suite software and normalized by subtraction of baseline cell fluorescence; nuclei are stained in blue, ML1 – red; size bar 50 μm .

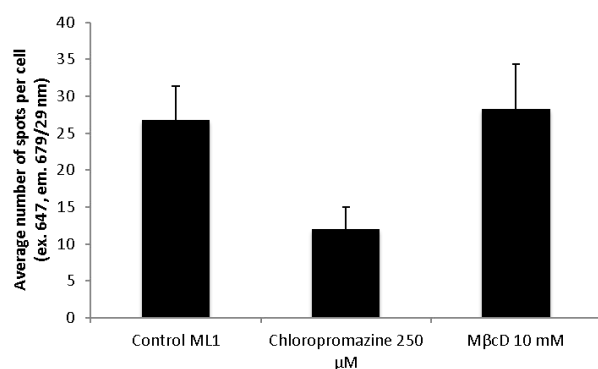
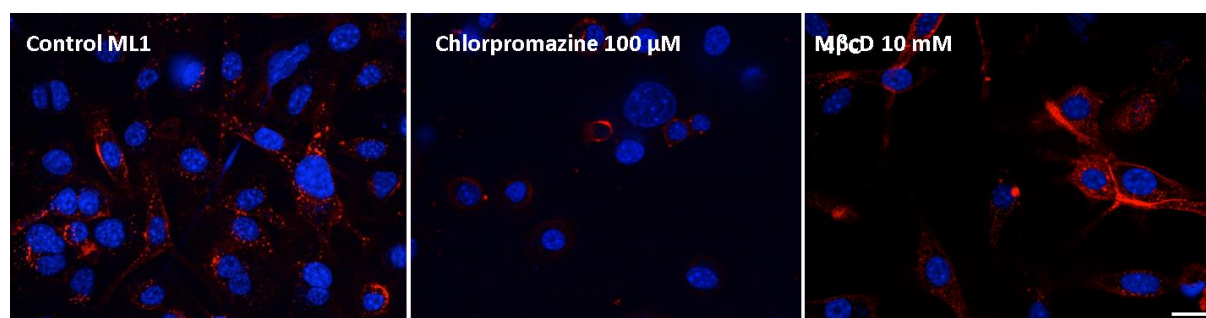


Figure S5. Confocal images of CT26 cells pre-incubated with uptake inhibitors and then incubated with ML1 for 1h (10 $\mu\text{g}/\text{mL}$) (up); Average number of spots per cell (n=3) quantified with Columbus suite software; nuclei are stained in blue, ML1 – red; size bar 20 μm .

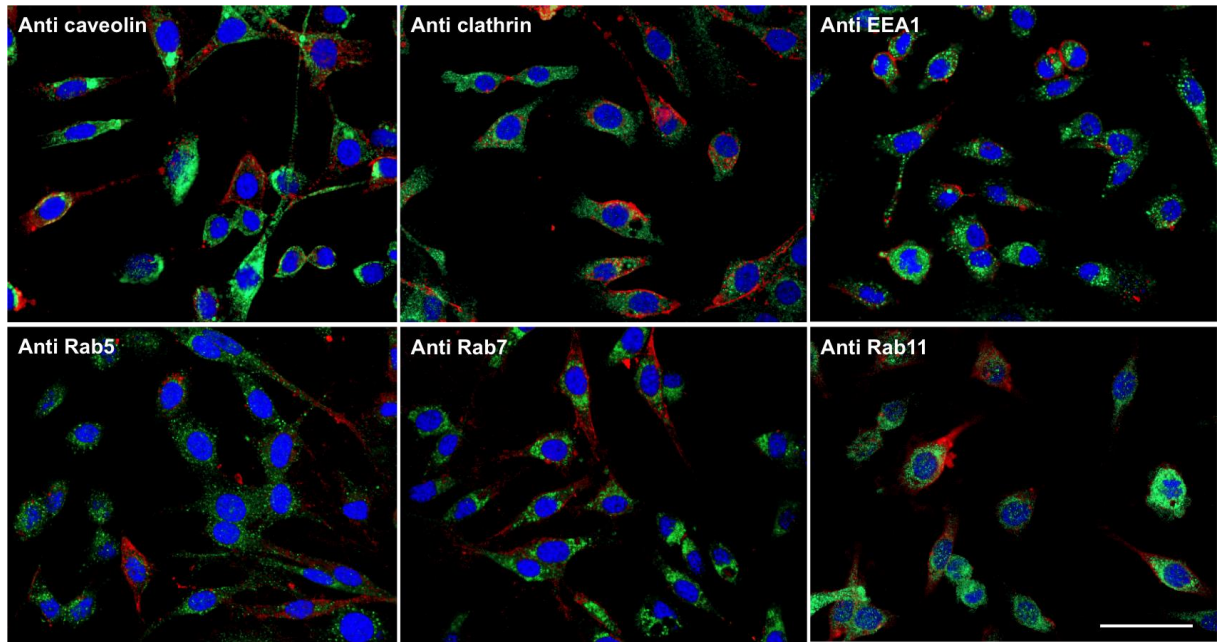


Figure S6. Confocal images of ML1 uptake (10 $\mu\text{g}/\text{mL}$, 2h) and co-staining with endolysosomal pathway specific antibodies; nuclei are stained in blue, ML1 - red, secondary antibody (Anti-Goat conjugated to Alexa Fluor 488) - green; size bar 50 μm .

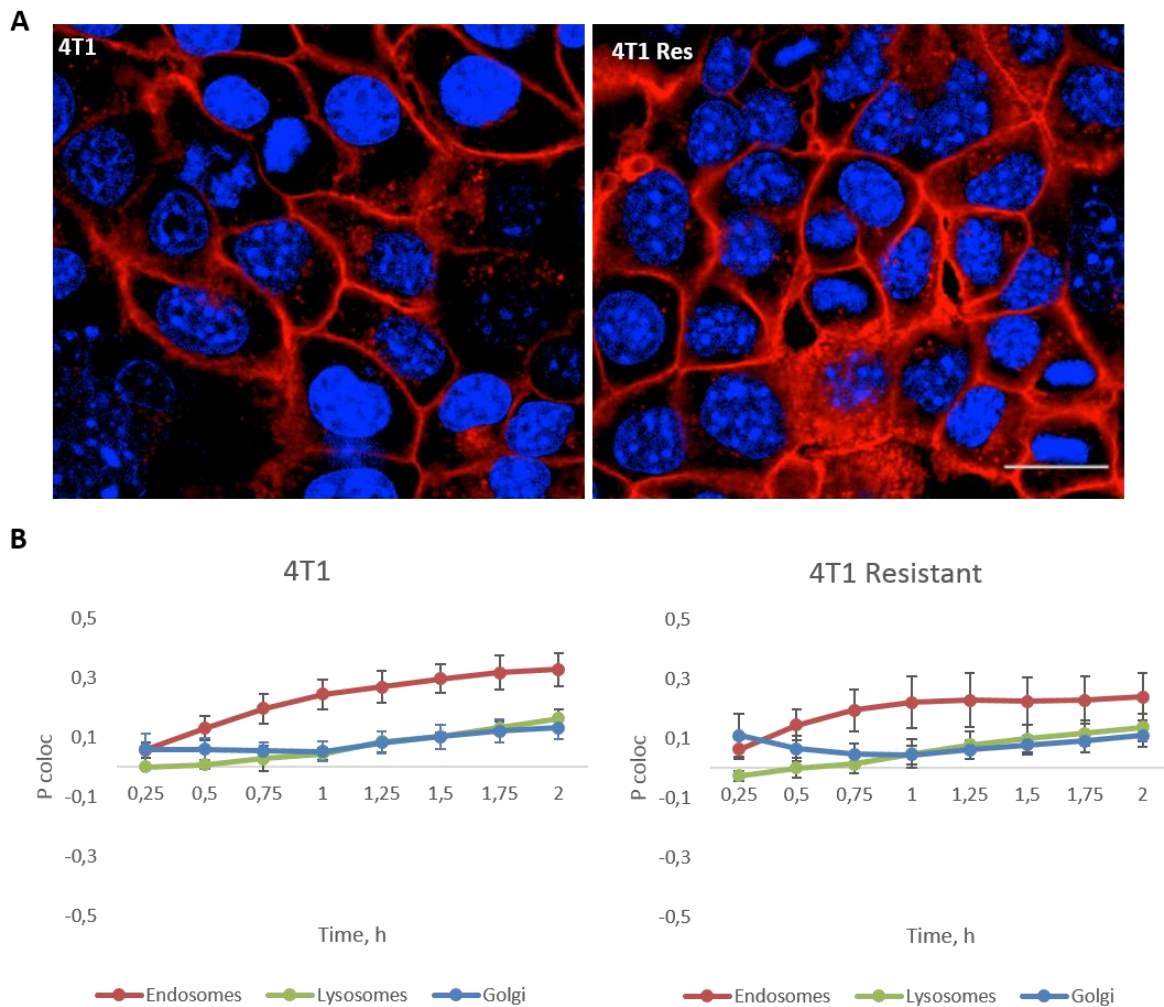


Figure S7. A. Confocal images of ML1 uptake in 4T1 control and doxorubicin resistant cells at 30 min incubation with 10 $\mu\text{g}/\text{mL}$ ML1; nuclei are stained in blue, ML1 – red; size bar 20 μm .; B. Pearson correlation coefficient dynamics of co-localization ML1 with endosomes, lysosomes and Golgi in 4T1 mother and resistant cells.

Supplementary movies:

1 and 2: Time-lapse movies of ML1 induced apoptosis in CT26 cells (1) or untreated CT26 cells in the same experiment (2); nuclei are stained in blue, apoptotic cells in green (CellEvent® live staining), ML1 in red, magnification 60x. The first 8 frames of each movie are taken every 15 min for 2h and the rest 24 frames every 3 hours up to 72h.

3 and 4: Time-lapse movies of ML1 induced apoptosis in 4T1 Res cells (3) or untreated 4T1 Res cells in the same experiment (2); nuclei are stained in blue, apoptotic cells in green (CellEvent® live staining), ML1 in red, magnification 40x. In total 18 frames are taken with imaging every 4 hours up to 72h.

Table S1. Summary of fluorescent labels used for various experiments with living or fixed cells.

Target	Live-cell experiments*	Fixed cells experiments
Nuclei	Hoechst 33342	Hoechst 33342
Lysosomes	LysoTracker Green	Anti-Rab11†
Golgi	BODIPY TR C5 -ceramide complexed to BSA	-
Endosomes early	pHrodo Green Dextran conjugate	Anti-EEA1, Anti-Rab5
Endosomes late	-	Anti-Rab7
Cell membrane/ protein transport	Wheat Germ Agglutinin Oregon Green conjugate	-
Caveolin-mediated uptake	Cholera toxin subunit b Alexa Fluor 488 conjugate	Anti-caveolin
Clathrin-mediated uptake	-	Anti-clathrin
Apoptotic cells	CellEvent caspase-3/7 Green Detection Reagent	CellEvent caspase-3/7 Green Detection Reagent

* All live cell labeling reagents and fluorescent dyes were purchased from Thermo Fisher Scientific, Naarden, The Netherlands.

† Endosomal Marker Antibody Sampler Kit #12666 was purchased from Cell Signaling Technology Europe, B.V., Leiden, The Netherlands.