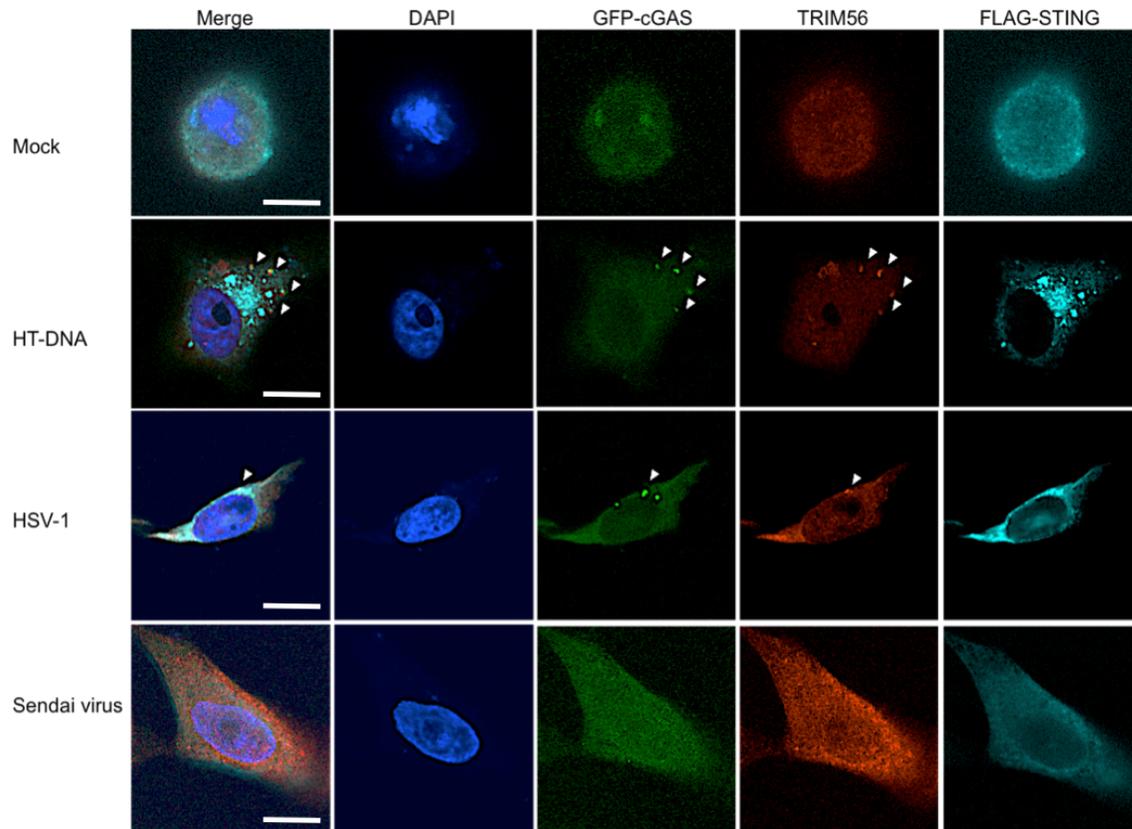
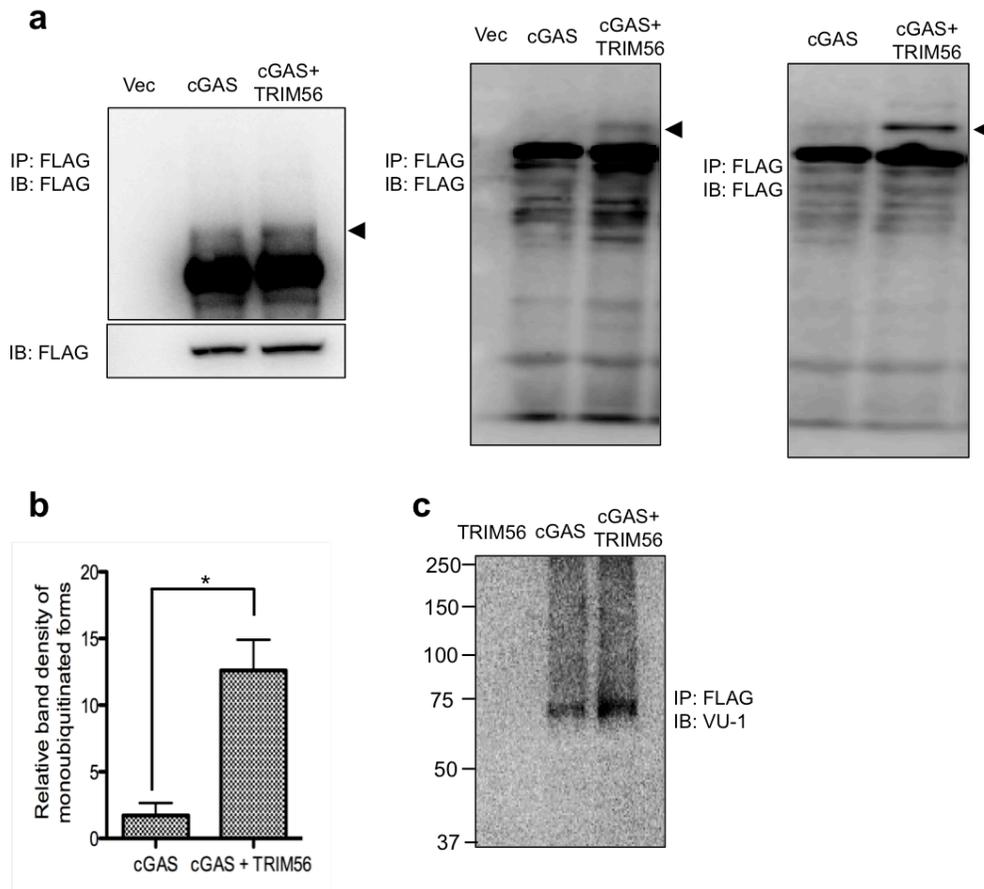


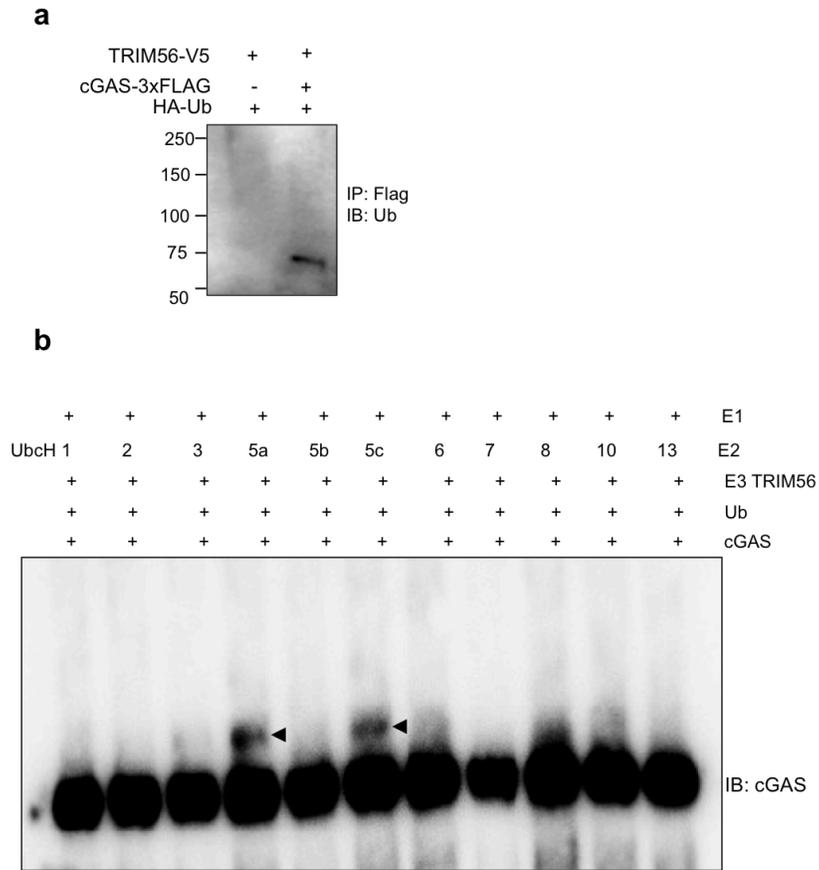
Supplementary Figure 1. TRIM56 directly binds to the cGAS N-terminus and is required for IP-10 induction. (a) Maltose-binding protein (MBP), fusion MBP-cGAS C-terminus (aa 161-522), and fusion MBP-cGAS full-length protein were purified from *E. coli*. TRIM56 was purified from 293T cells. A co-immunoprecipitation assay was conducted using amylose resin. White arrows indicate MBP (left), MBP-cGAS C-terminal 161-522aa (middle) and MBP-cGAS full-length (right), respectively. (b) TRIM56 binds to cGAS, but not STING. 293T cells were co-transfected with TRIM56-V5 and empty vector (EV), cGAS-FLAG, or STING-FLAG. After 48 hours, cGAS or STING co-immunoprecipitate was washed with 500mM NaCl followed by immunoblotting. (c) cGAS-deleted L929 cells with or without TRIM56 deletion were complemented with wild-type cGAS-FLAG (the cell lines generated for Figure 2) before infection with HSV-1ΔICP34.5 (MOI=5). The expression of IP-10 mRNA was measured using real-time PCR. Data (a-b) are representative of two independent experiments. Data (c) are representative of three independent experiments. Error bars indicate mean \pm s.d. of n=3 (c). * P <0.05 versus control using Student's *t*-test (c). Full blots are shown in Supplementary Fig. 10.



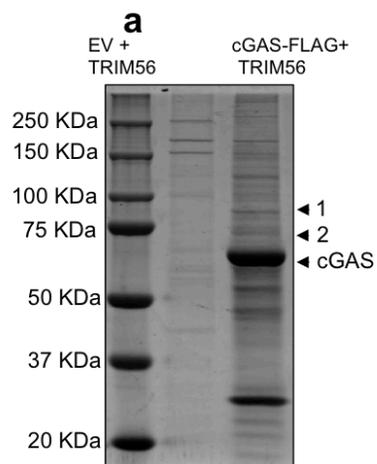
Supplementary Figure 2. TRIM56 and cGAS co-localize at foci in response to DNA stimulation and infection with DNA virus, but not RNA virus. GFP-cGAS and FLAG-STING constructs were exogenously expressed in HeLa Cells for 48 hours before stimulation with herring testis DNA (HT-DNA; 2 μ g/ml), infection with Herpes Simplex Virus 1 (HSV-1), or infection with Sendai virus. Co-localization of cGAS with endogenous TRIM56 was observed at the indicated foci (see arrowhead). Data are representative of three independent experiments. Scale bar, 5 μ M.



Supplementary Figure 3. TRIM56 leads to monoubiquitination of cGAS. (a) In three independent trials, cGAS complexes were pulled down from 293T cells transfected with empty vector, cGAS-FLAG alone, or cGAS-FLAG and TRIM56-V5. Western blot was performed using FLAG antibody. Band shifts (arrowhead) are indicated. (b) For cells overexpressing cGAS with or without TRIM56 co-transfection, the densities of the cGAS band shifts from (a) were quantitated relative to the background. (c) In the ubiquitination assay of Figure 3b, a single band near 75kDa was observed, suggesting preferential monoubiquitination. Data (a-b) show three independent experiments. Error bars indicate mean \pm s.d. of of three independent experiments (b). * $P < 0.05$ versus control using Student's *t*-test (b). Full blots are shown in Supplementary Fig 10.



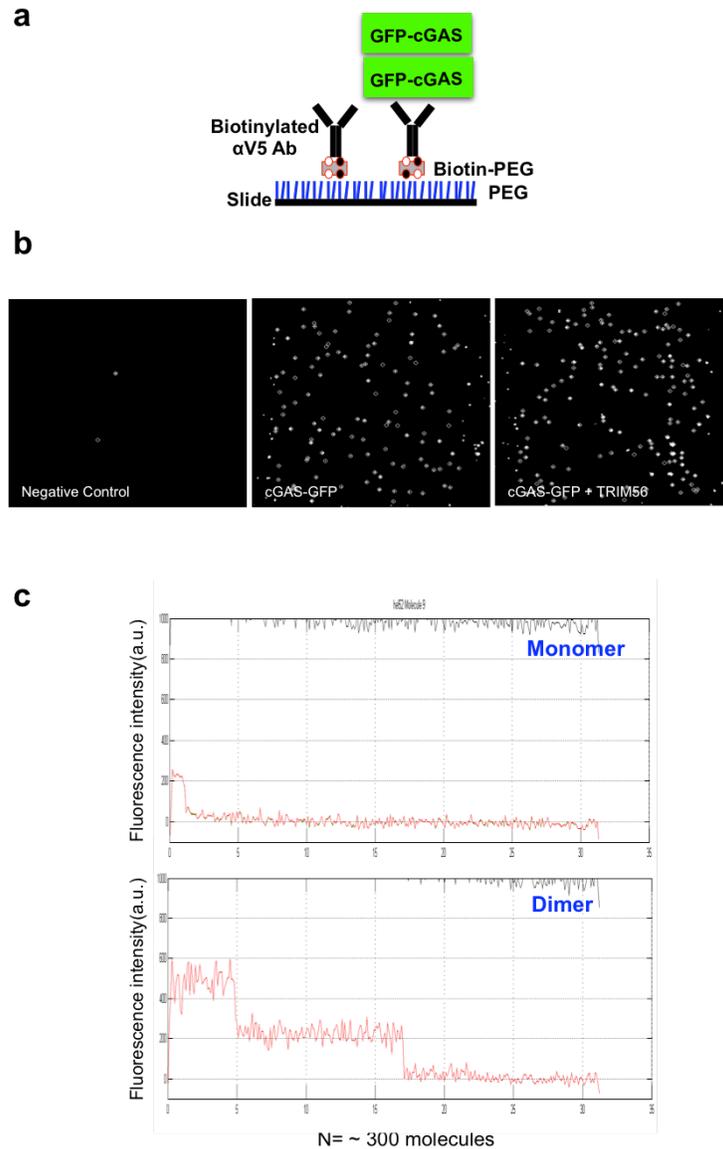
Supplementary Figure 4. TRIM56 triggers the monoubiquitination of cGAS *in vitro*. (a) HEK293T cells were transfected with TRIM56-V5, cGAS-3xFLAG or HA-Ub. 24 hours after transfection, whole cell lysates (WCLs) were used for immunoprecipitation and immunoblotting, as indicated. (b) An *in vitro* ubiquitination assay with the indicated combinations of a mixture of E1, E2 (UbcH; numbers above lanes indicate enzyme variants), E3 (TRIM56), cGAS and Ubiquitin (Ub). Immunoblot of the *in vitro* ubiquitination was detected by cGAS antibody. Data are representative of two independent experiments (b). Full blots are shown in Supplementary Fig. 10.



b

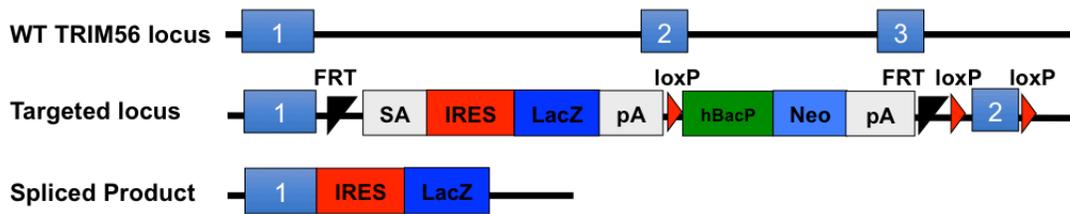
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15794	3	1.44	0.331	13/52	K.EGLPIQGWLGTK#VR.T
17369	2	1.961	0.596	10/18	R.NNGFPIFDK#.L
5068	2	3.189	0.778	20/26	K.NAK#DGNSFQGETWR.L
7304	2	2.68	0.581	14/20	K.EIK#DIDVSVEK.E
8661	3	1.604	0.009	12/44	R.REPFYLVPKNAK#.D

Supplementary Figure 5. Identification of cGAS ubiquitination sites. (a) Purification and Coomassie blue stain of cGAS-FLAG from HEK293T cells transfected with TRIM56 and empty vector (EV), or with TRIM56 and cGAS-FLAG. (b) Summary of mass spectrometry data identifying cGAS ubiquitination sites.

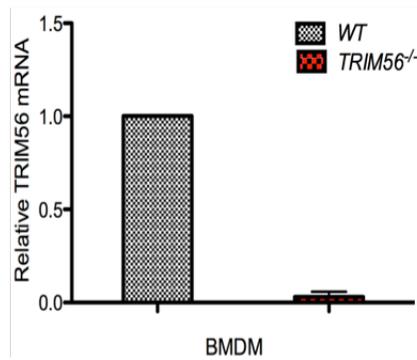


Supplementary Figure 6. GFP-cGAS photobleaching traces. (a) Schematic depiction of GFP-cGAS SiMPull. (b) Representative GFP fluorescence images. (c) Sample GFP time traces depicting one- or two-step photobleaching. For determining the stoichiometry, traces were manually scored for the number of bleaching steps. The graph shows representative photobleaching curves with one- and two-step photobleaching. >300 traces were scored to reliably identify the photobleaching step distribution.

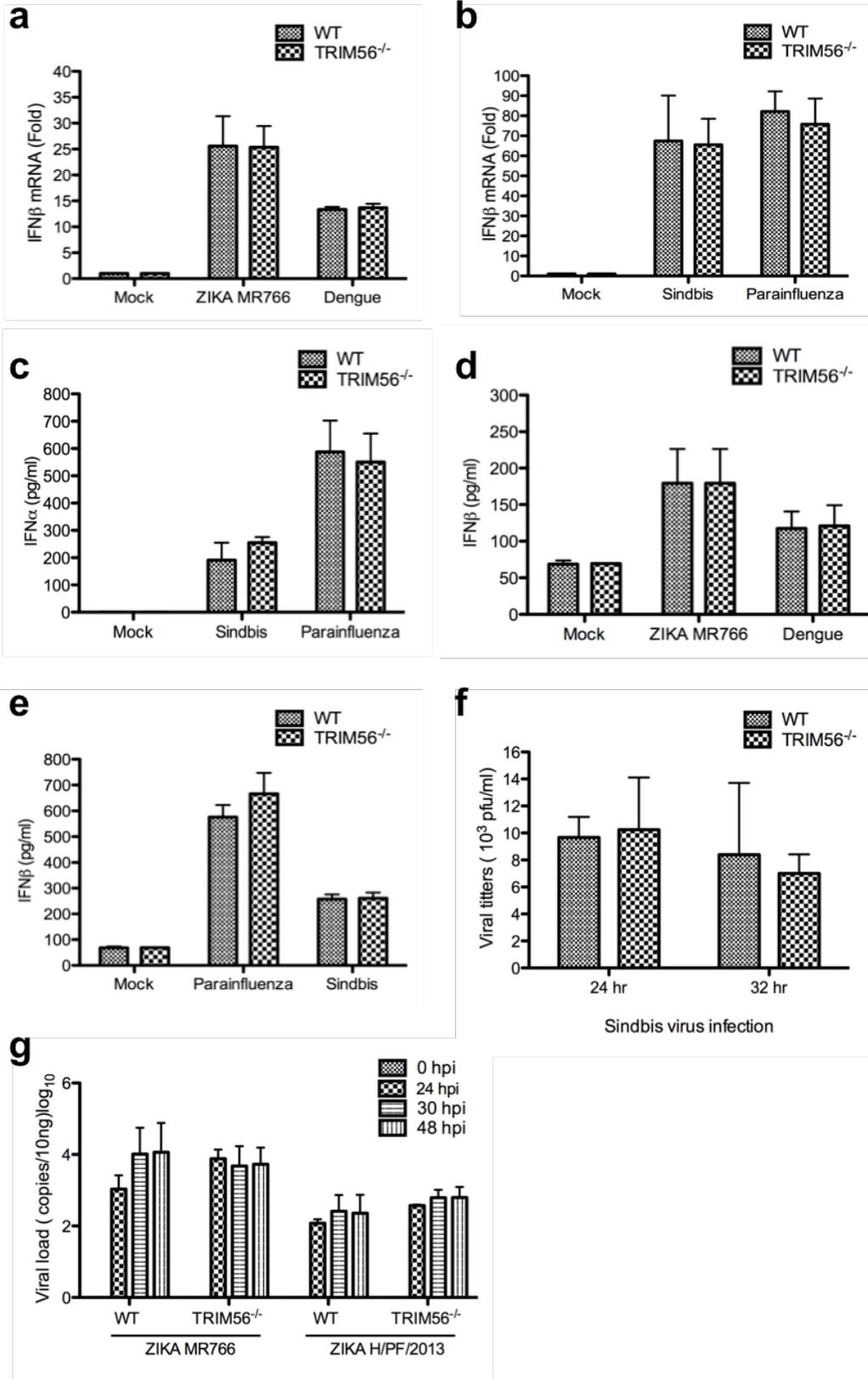
a



b



Supplementary Figure 7. Generation of *TRIM56*^{-/-} mice. (a) Schematic representation of the *TRIM56* WT locus showing its exons (boxes) and introns (lines), and the locus disrupted by the targeting vector. The splice acceptor (SA) sequence in the LacZ cassette interrupts the normal splicing of the *TRIM56* gene, resulting in the loss of *TRIM56* expression. (b) RT-PCR analysis of *TRIM56* mRNA amplified from the total RNA of bone marrow-derived macrophages (BMDMs) from WT or *TRIM56*^{-/-} mice.



Supplementary Figure 9. TRIM56 does not affect the host innate immune response to positive- and negative-sense RNA viruses. (a) Fold change in *IFN β* expression in *WT* and *TRIM56^{-/-}* BMDMs in response to ZIKV African strain MR766 and Dengue virus (MOI=5). (b) Fold change in *IFN β* expression in *WT* and *TRIM56^{-/-}* BMDMs in response to Sindbis virus and Parainfluenza virus (MOI=5). (c) Magnitude of *IFN α* secretion in *WT* and *TRIM56^{-/-}* BMDMs in response to Sindbis virus and Parainfluenza virus (MOI=5). (d) Magnitude of *IFN β* secretion in *WT* and *TRIM56^{-/-}* BMDMs in response to ZIKV MR766 and Dengue virus (MOI=5). (e) Magnitude of *IFN β* secretion in *WT* and *TRIM56^{-/-}* BMDMs in response to Sindbis virus and Parainfluenza virus (MOI=5). (f) Viral titers of *WT* and *TRIM56^{-/-}* BMDMs infected with Sindbis virus. The plaque formation assay was performed on Vero cells. (g) Viral load over time of *WT* and *TRIM56^{-/-}* BMDMs infected with MR766 African lineage ZIKV or H/1N1/2009 Asian lineage ZIKV. Data (a-b, g) are representative of three independent experiments. Data (c-f) are representative of two independent experiments. Error bars indicate mean \pm s.d of n=3 (a-e, g), n=4 (f).

Figure 1b

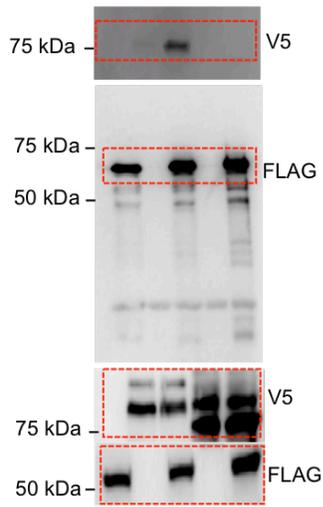


Figure 1c

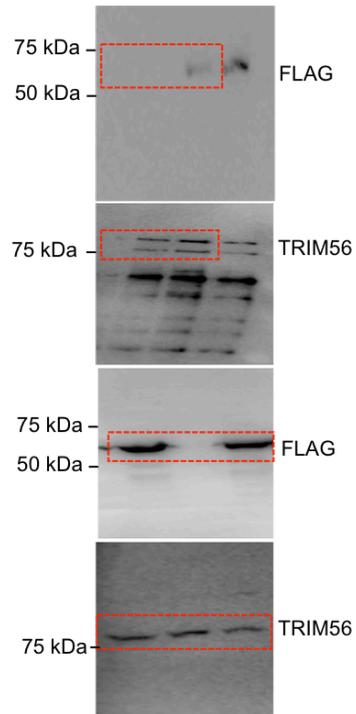
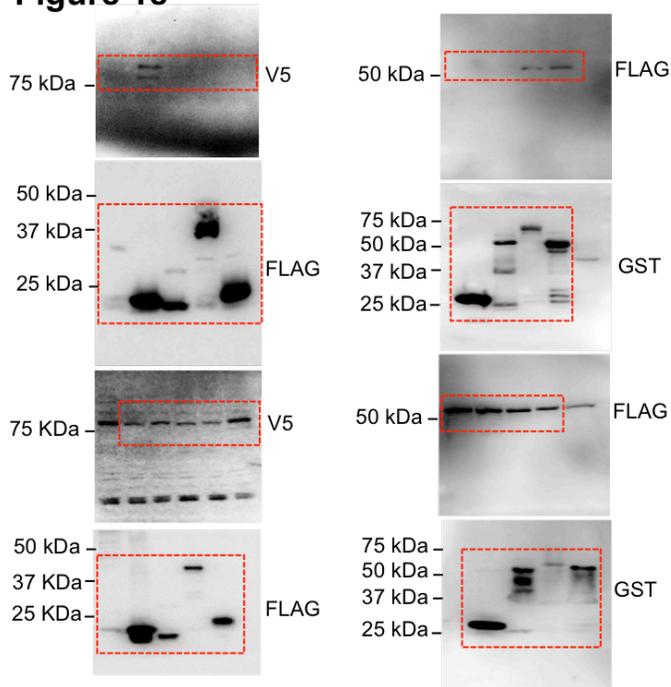


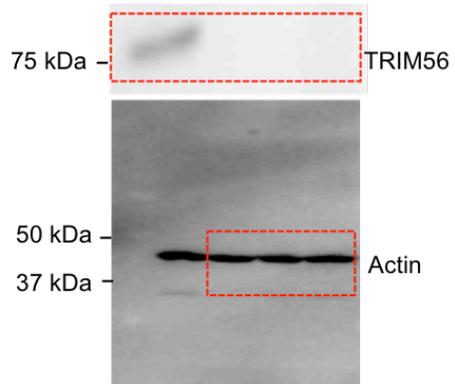
Figure 1e



Supplementary Figure 10. Full-length uncropped western blots.

Uncropped Western blot images for Figure 1. Cropped areas are marked by red box.

Figure 2a



Supplementary Figure 10, cont. Full-length uncropped western blots. Uncropped Western blot images for Figure 2. Cropped areas are marked by red box.

Figure 3a

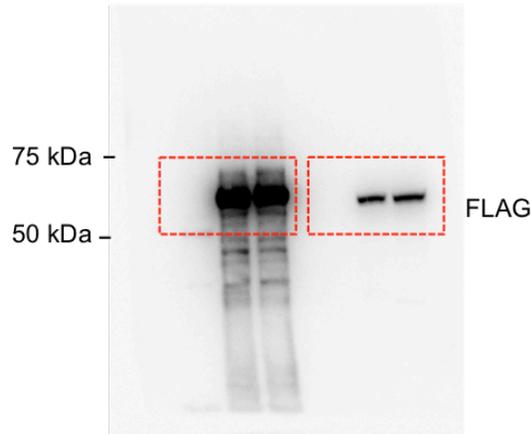


Figure 3b

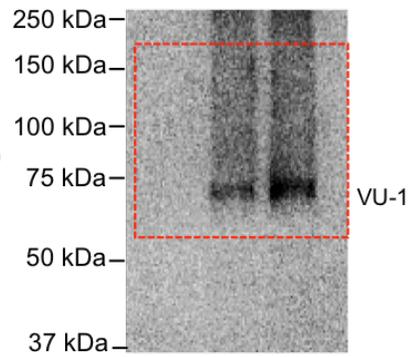


Figure 3c

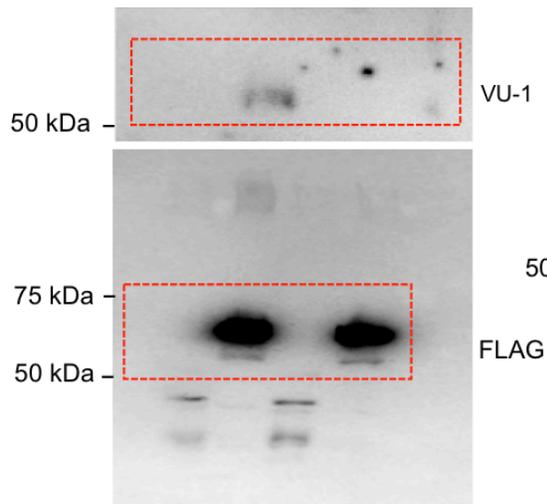
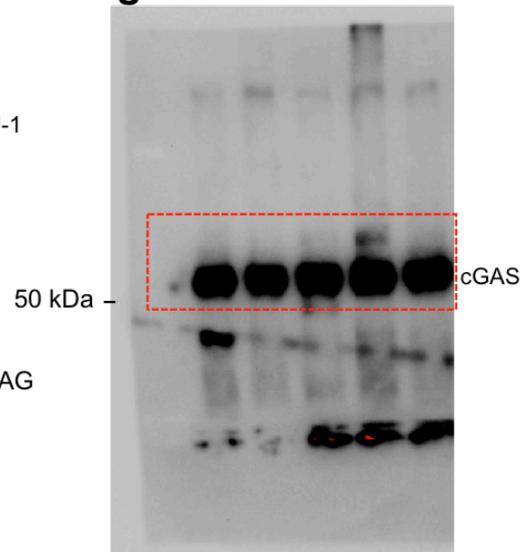


Figure 3d



Supplementary Figure 10, cont. Full-length uncropped western blots. Uncropped Western blot images for Figure 3. Cropped areas are marked by red box.

Figure 4a

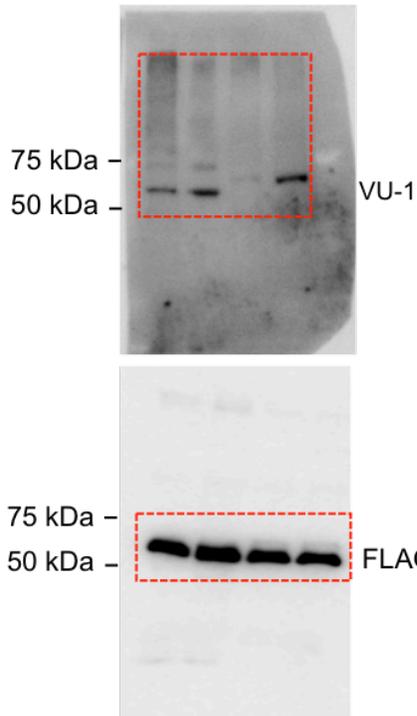
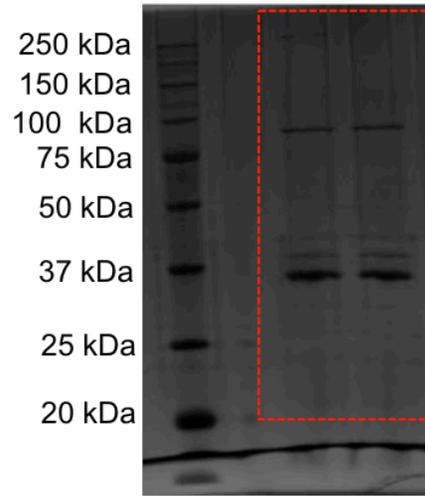
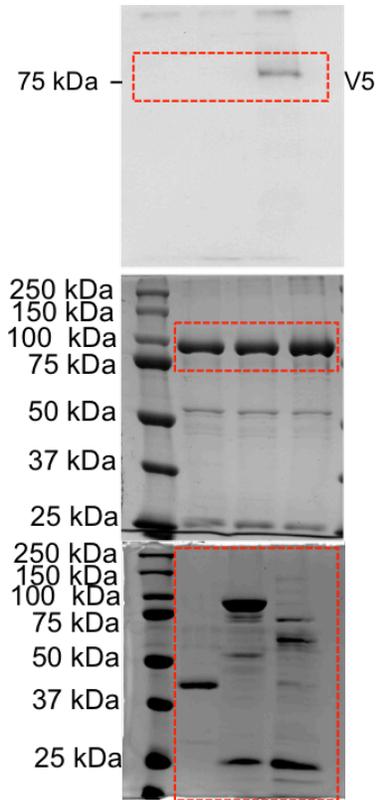


Figure 4d

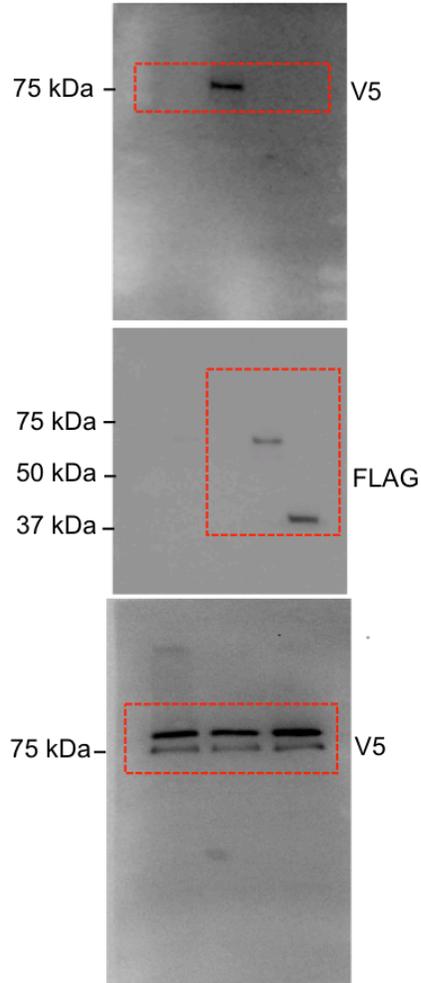


Supplementary Figure 10, cont. Full-length uncropped western blots. Uncropped Western blot images for Figure 4. Cropped areas are marked by red box.

Supplementary Figure 1a



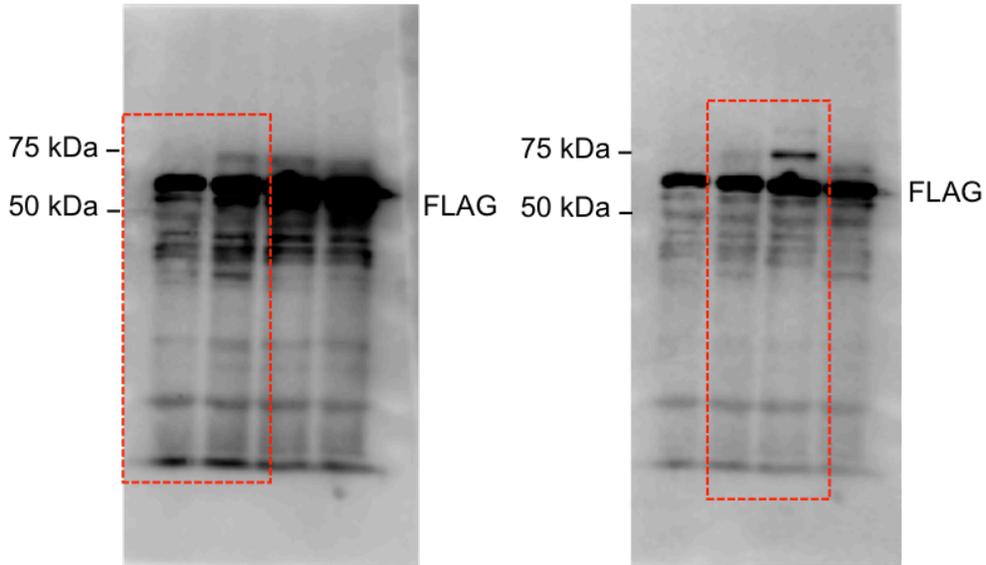
Supplementary Figure 1b



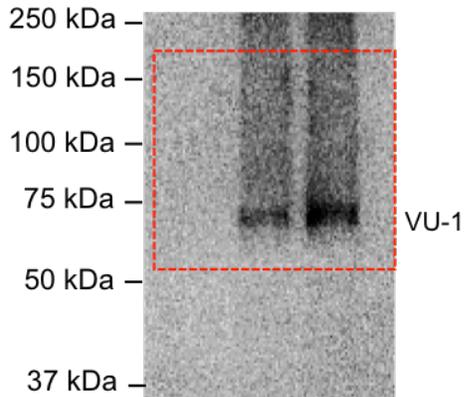
Supplementary Figure 10, cont. Full-length uncropped western blots.

Uncropped Western blot images for Supplementary Figure 1. Cropped areas are marked by red box.

Supplementary Figure 3a



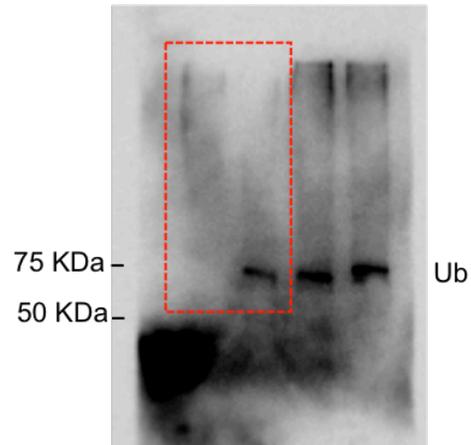
Supplementary Figure 3c



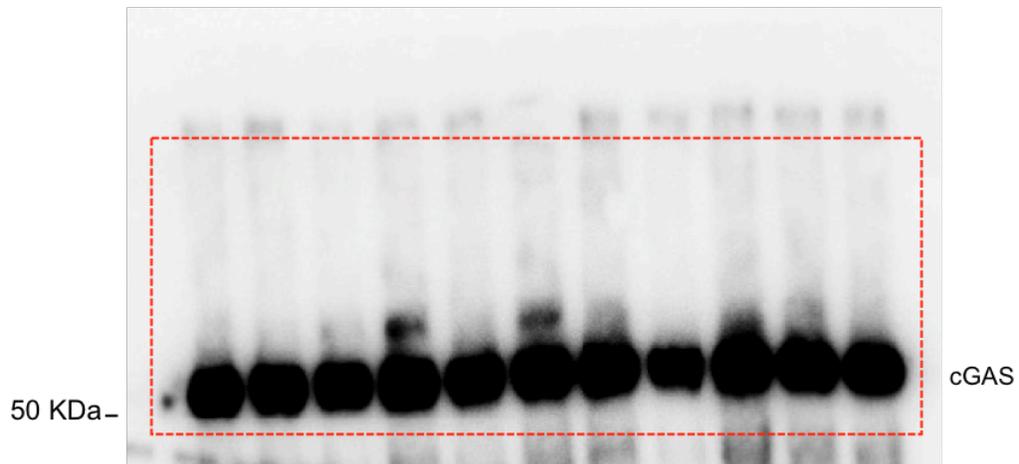
Supplementary Figure 10, cont. Full-length uncropped western blots.

Uncropped Western blot images for Supplementary Figure 3. Cropped areas are marked by red box.

Supplementary Figure 4a



Supplementary Figure 4b



Supplementary Figure 10, cont. Full-length uncropped western blots.

Uncropped Western blot images for Supplementary Figure 4. Cropped areas are marked by red box.

Supplementary Table 1. Primers used for PCR

Gene Name	Direction	Primer sequence (5'-3')
Human TRIM56	Forward	GAGCTCGAGCTGTTTCCCACGGGTCTCGCCCTCC
	Reverse	GATTCTAGAAGTGTCCGGAGA ACG GAC CCG AAA
Mouse TRIM56 genomic PCR#1	Forward	AAGTGGAGGCAGGAAGTTCA
	Reverse	GACACGGTGCTTATGGGTCT
Mouse TRIM56 genomic PCR#2	Forward	CTACCTTTGGCCCTTGACCT
	Reverse	GGCCTTGACAAGATCCAAGA
Mouse TRIM56	Forward	GAAGAATTCGCCACCATGAACTCCAAAGACTCCTCCCCA
	Reverse	GAAACGCGTGCTGCAGGAAT GTTCCAAAGCAAGCAGTT
Mouse TRIM56 Mutant	Forward	AGCGATTTCTAGCCTCTAAAATCTCCCTGGAGCAGTTA
	Reverse	TAACTGCTCCAGGGAGATTTTAGAGGCTAGGAAATCGCT
Mouse cGAS	Forward	GAGCTCGAGGCCACCATGGAAGATCCGCGTAGAAGGACGACG
	Reverse	GGTTCTAGAAAGCTTGTCAAAAATTGGAAA CCCATTATTTCT
McGASK278R	Forward	GAAGTTAAAGAAATCAGAGATATAGATGTCAGTGTG
	Reverse	CACACTGACATCTATATCTCTGATTTCTTTAACTTC
McGASK335R	Forward	GGCTGGCTGGGCACAAGAGTGAGGACCAATCTAAGA
	Reverse	TCTTAGATTGGTCTCACTCTTGTGCCAGCCAGCC
McGASK350R	Forward	TTTTATCTCGTA CC AGGAATGCAAAGGATGGAAAT
	Reverse	ATTTCCATCCTTTGCATTCTGGGTACGAGATAAAA

Supplementary Table 2. shRNA or sgRNA sequence information

shRNA	Target sequence (5'-3')
Human TRIM56#1	GATTTCGAATGGGCAGTGA
Human TRIM56#2	AAGAGAATAGGCTACTGGA
Human TRIM56#3	CACCACCGCCGCTGCTATA
sgRNA	Target sequence (5'-3')
Mouse TRIM56#1	CCAGGACTGTCTGGCACA ACTGG
Mouse TRIM56#2	AGTCCTGGCAATAGGTATGT AGG

Supplementary Table 3. Primers used for real-time PCR

Gene Name	Direction	Primer sequence (5'-3')
Mouse IFN β	Forward	ATGAACTCCACCAGCAGA CAG
	Reverse	ACCACCATCCAGGCGTAGC
Mouse β -actin	Forward	GGCTATGCTCTCCCTCACG
	Reverse	CGCTCGGTCAGGATCTTCAT
Mouse IP-10	Forward	AAGTGCTGCCGTCATTTTCT
	Reverse	GTGGCAATGATCTCAACACG
Mouse TRIM56	Forward	ATGAACTCCAAAGACTCCTCCCA
	Reverse	AGTTGGCTTCCCTGAATGGACGTCTCC