	10	20	30	40	50	60	70	80	90	100	110	120	130	140
				.			.							
A*02	GACAGCTGCCTTGT	GTGGGACTGAG	AGGCAAGAG	TTGTTCCTGCCC	TTCCCTTTG	IGACTTGAAG	AACCCTGACTI	TGTTTCTGC	AAAGGCACCTG	CATGTGTCTG	TGTTCGTGT	AGGCATAATGT	AGGAGGTG	GGGAG
A*01														
A*03													· · · · · · · · · · · · · · · · · · ·	
A*11														
A*23											A			
A*24											A			
A*26			т								т			
A*29			т								т			
A*30														
A*31			т				г				т			
A*32			т								T			
A*33			т				T				<mark>. T</mark>			
A*68														
	150	160	170	180	190	200	210	220	230	240	250	260	270	280
				.			.							
A*02	ACCACCCCACCCC	ATGTCCACCAT	GACCCTCTT	CCCACGCTGACC	TGTGCTCCC	TCCCCAATCA	TCTTTCCTGT1	CCAGAGAGG	TGGGGCTGAGG	TGTCTCCATC	TCTGTCTCA	ACTTCATGGTG	ACTGAGCT	GTAAC
A*01	.G					T								
A*03														
A*11	G					T								
A*23								G						
A*24								G						
A*26									• • • • • • • • • • • •		c			
A*29						• • • • • • • • • • •			••••••		c			
A*30	•••••		• • • • • • • • • •						• • • • • • • • • • • •					••••
A*31									• • • • • • • • • • • •		c			• • • • •
A*32									••••••		C			• • • • •
A*33	•••••		• • • • • • • • • •	• • • • • • • • • • • • •	•••••	• • • • • • • • • • •		•••••	• • • • • • • • • • •		c	•••••		••••
A*68									• • • • • • • • • • • •					• • • • •
	290	300	310	320	330	340	350	360	370	380	390	400	410	420
				.			.							
A*02	TTCTTCCTTCCCTA	TTAAAATTAGA	ACCTGAGTA	TAAATTTACTTI	CTCAAATTC	TTGCCATGAG	AGGTTGATGAG	TTAATTAAA	GGAGAAGATTC	CTAAAATTTG	AGAGACAAA	ATAAATGGAAC	CATGAGAA	CCTTC
A*01	••••••	• • • • • • • • • • • •	· · · · · · · · · · · · · · · · · · ·	•••••	•••••	•••••	•••••••••	•••••	•••••	•••••	•••••		÷	••••
A*03	•••••	• • • • • • • • • • • •	· · · · T · · · ·	•••••	•••••	•••••	•••••••••	•••••	•••••	•••••	•••••	G	•••••	••••
A*11	•••••	• • • • • • • • • • • • •	•••••	•••••	•••••	•••••	•••••••••••	•••••	••••••	•••••	•••••		••••••	• • • • •
A*23	•••••	• • • • • • • • • • • • •	•••••	•••••	•••••	•••••	••••••	•••••	••••••	•••••	•••••	••••••	•••••	• • • • •
A*24		• • • • • • • • • • • • •	•••••	•••••	•••••	•••••	••••••	•••••	•••••	•••••	•••••	••••••	•••••	• • • • •
A*20 A*20		• • • • • • • • • • • • •	•••••	•••••	•••••	•••••	••••••	•••••	••••••	•••••		••••••	•••••	• • • • •
A*29	•••••	••••••		••••••	•••••	•••••	••••••	•••••	••••••	•••••			•••••	•••••
A*30	•••••	•••••	·····	••••••	•••••	•••••	••••••••••	•••••	••••••••••	•••••		· · · · · · · · · · · · · · · · · · ·	•••••	•••••
¥32		••••••			•••••	•••••				•••••			•••••	•••••
X*32		• • • • • • • • • • • • •							•••••	•••••			•••••	
A*53	•••••	••••••		•••••	•••••	•••••	••••••••••	•••••	••••••••••	•••••		••••••	•••••	•••••
H. 00														• • • • •

Supplemental Figure 1. Two polyadenylation signals (PAS) in the HLA-A 3'UTR. Alignment of 3'UTR sequences of common HLA-A alleles are shown. The 3' UTR was amplified by PCR and sequenced using genomic DNA from individuals homozygous for the indicated allele. Identical nucleotides are shown as dots. Proximal and distal PAS are indicated with the blue and red underlines, respectively. Α.



Jurkat Lecture 2. HLA-B and –C use a single PAS, and HLA-A transcripts exhibit allele specific patterns of alternative PAS usage consistently across cell types. (A) Alignment of 3'UTR sequences of HLA-A, HLA-B (IMGT-HLA;

http://www.ebi.ac.uk/ipd/imgt/hla/) and HLA-C (60) are shown. Identical nucleotides are shown as dots. Proximal and distal PAS are indicated with blue and red underlines, respectively. The proximal PAS is disrupted in *HLA-B* and *HLA-C* alleles. **(B)** RNA was obtained from peripheral blood CD4+T cells (CD4), CD8+ T cells (CD8), and monocytes (M) of 3 healthy donors with distinct *HLA-A* genotypes noted. Rapid amplification of cDNA ends (3' RACE) of 3'UTRs of *HLA-A* alleles from 3 distinct cell types showed a similar pattern of use of either the proximal or distal PAS. *HLA-B* and *HLA-C* 3'UTRs express only a single isoform owing to absence of the proximal PAS. **(C)** RNA was obtained from Jurkat T cells, transformed B cell lines (BCL), and HEK293 cells. Similar to the lymphocytes, the 3'RACE indicated an allele specific pattern of alternative PAS usage for *HLA-A* transcripts; whereas *HLA-B* and *HLA-C* transcript express a single isoform.

В.



Supplemental Figure 3. Schematic representation of the modifications to the psicheck2 vector. (A) Deletion of the synthetic PolyA signal downstream of *Renilla* luciferase is shown. (B) *Renilla* luciferase sequence was replaced by c-terminal Flag tagged HLA-A ORF [AORF-Flag]. (C) Total cell lysates of HEK293 cells transfected with HLA-AORF-Flag plasmids were used to carry out Western blot analysis with anti-Flag and anti-firefly luciferase antibodies. One of two independent experiments is shown. Differential binding of Syncrip to the long form of *HLA-A* 3'UTR. (D) Coomassie blue stained PAGE gel separating the RBPs that were bound to the long form of *HLA-A* 3'UTR (PASPD), the mutant containing a 25bp deletion immediately downstream of proximal PAS (Δ 300-325), or avidin beads in the absence of any labeled RNA (beads; lane 3). A distinct protein band, which appeared only in the PASPD lane (Band1) and proteins of corresponding molecular weight in the Δ 300-325 lane (Band 2) and the beads only lane (Band 3) were excised and analyzed by MS (E) Syncrip was identified by LC-MS/MS in the RBPs bound to the long form of *HLA-A* 3'UTR (PASPD; Band 1), and no RBPs were detected in the Band 2 from the Δ 300-325 lane or Band 3 from the beads only control.

Supplemental Table I. Sequences of the morpholino-oligonucleotides and primers

Primer	Sequence
MorpholinoOligonucleotides	
Apro1	ACTCAGGTTCTAATTTAATAGGGA
Control Morpholino	CCTCTTACCTCAGTTACAATTTATA
HLA-A 3'UTR sequencing	
HLA-A 3'UTRseqFw	TCTGTGGGATCTGACCAGGTT
HLA-A 3'UTRseqRev	AACACGTGGACTCTGGAAGG
3'RACE	
HLA-A 3'UTR 3'RACE Fw	TGAGAGGCAAGAGTTGTTCCT
UAP	5'GGCCACGCGTCGACTAGTACTTTTTTTTTTTTTTT 3'
Cloning	
psicheckAFw	ATCGCTCGAGTGAGAGGCAAGAGTTGTTCCT
psicheckARev	GCCAGCGGCCGCTGGACTCTGGAAGGTT
A03UTR_325_Rev	GCCAGCGGCCGCTTTATTAAGTAAATTTATACTAAG
HLA-AORFFw	GACTCACTATAGGGAGACCCAAGCTGGCTACGCT
HLA-AORFRev	ACCTTGTACGAGTGCGGGTCTCGAGTTATCACTTATCGTC
qPCR	
HLA-A 3'UTR Rev	AAGAGGGTCATGGTGGACAT
Renilla Fw	AAGAGCGAAGAGGGCGAGAA
Renilla Rev	TGCGGACAATCTGGACGAC
Firefly Fw	CGTGCCAGAGTCTTTCGACA
Firefly Rev	ACAGGCGGTGCGATGAGAC
Gapdh Fw	GGTGAAGGTCGGAGTCAACG
Gapdh Rev	GTTGAGGTCAATGAAGGGGTC

Primer and oligonucleotide sequences used in the study are listed.