#### **Supplementary Information**

#### **Supplementary Tables**

#### Table S1. miR-125 targets Tom-3`UTR

| miRNA<br>expression<br>plasmid | Renilla /Firefly<br>luciferase ratio<br>( <i>psiCHECK-2</i> ) | Renilla /Firefly<br>luciferase ratio<br>( <i>psiCHECK-2-</i><br><i>Tom- 3'UTR</i> ) | Relative Luciferase<br>levels ( <i>psiCHECK-2-<br/>Tom-3'UTR</i><br>/psiCHECK-2) | Relative<br>Luciferase<br>levels <sup>a</sup> |
|--------------------------------|---|---|--|---|
| <sup>a</sup> none              | $0.17 \pm 0.02$   | $0.24 \pm 0.03$   | $1.41 \pm 0.18$  | $1.00 \pm 0.17$                               |
| miR-125                        | $0.96 \pm 0.44$   | $0.35 \pm 0.01$   | $0.36 \pm 0.01$  | $0.26 \pm 0.02$<br>P=0.0042                   |

<sup>a</sup> Values in the presence of miRNA expressing plasmids were normalized to the downregulation that occurred in the cells with no miRNA-expressing plasmid present.

Firefly luciferase expression is used as an endogenous control where *Tom 3'UTR* was cloned into Renilla luciferase gene. AVE±STDEV values are reported. Two-tailed Student's t-test was used to test for statistical significance.

| Table S2. Expression levels of <i>miR-125, let-7</i> and <i>miR-100</i> in controls and <i>miR-125<sup>LOF</sup></i> |
|--|
| mutants and the efficiency of <i>Tom</i> downregulation by <i>Tom-RNAi</i>   |

| Genotype   | <i>miR-125</i><br>(C <sub>T</sub> )  | 2S<br>(С <sub>т</sub> )   | $^{a}\Delta C_{T}$  | $^{b}\Delta\Delta C_{T}$  | <sup>c</sup> Relative<br><i>miRNA</i> level  |
|--|--|---|---|---|--|
| Control<br>(Oregon x w <sup>1118</sup> )   | 20.51±0.06   | 8.15±0.10   | 12.36±0.06  | 0.00±0.06   | 1.00±0.04  |
| miR-125 <sup>LOF</sup>   | 35.94±0.05   | 7.87±0.12   | 28.07±0.04  | 15.71±0.04  | 0.00±0.00<br>P=2.4E-6  |
| Genotype   | <i>let-7</i><br>(C <sub>T</sub> )  | 2S<br>(С <sub>т</sub> )   | ΔCT   | ΔΔC <sub>T</sub>  | Relative<br><i>miRNA</i> level   |
| Control<br>(Oregon x w <sup>1118</sup> )   | 23.47±0.18   | 8.15±0.10   | 15.32±0.18  | 0.00±0.18   | 1.01±0.13  |
| miR-125 <sup>LOF</sup>   | 23.38±0.22   | 7.87±0.12   | 15.51±0.22  | 0.18±0.22   | 0.88±0.13<br>P=0.311   |
|  |  |   |   |   |  |
| Genotype   | <i>miR-100</i><br>(C <sub>T</sub> )  | 2S<br>(C <sub>T</sub> )   | $\Delta C_{T}$  | $\Delta\Delta C_{T}$  | Relative<br><i>miRNA</i> level   |
| Genotype<br>Control<br>(Oregon x w <sup>1118</sup> )   | <i>miR-100</i><br>(C <sub>T</sub> )<br>21.84±0.11  | $\frac{2S}{(C_{T})}$ 8.15±0.10                                  | ΔC <sub>T</sub><br>13.69±0.11   | ΔΔC <sub>T</sub><br>0.00±0.11                                   | Relative<br>miRNA level  |
| Genotype<br>Control<br>(Oregon x w <sup>1118</sup> )<br>miR-125 <sup>LOF</sup>   | $miR-100 \\ (C_T)$ 21.84±0.11 21.76±0.09   | $2S \\ (C_{T})$ 8.15±0.10 7.87±0.12                             | ΔC <sub>T</sub><br>13.69±0.11<br>13.90±0.08   | ΔΔC <sub>T</sub><br>0.00±0.11<br>0.20±0.06                      | Relative<br>miRNA level           1.00±0.08           0.87±0.05<br>P=0.073   |
| Genotype<br>Control<br>(Oregon x w <sup>1118</sup> )<br>miR-125 <sup>LOF</sup><br>Genotype   | <i>miR-100</i><br>(С <sub>T</sub> )<br>21.84±0.11<br>21.76±0.09<br><i>Тот</i><br>(С <sub>T</sub> ) | $2S \\ (C_{T})$ 8.15±0.10 7.87±0.12 <i>Rpl32</i> (C_{T})        | ΔC <sub>T</sub><br>13.69±0.11<br>13.90±0.08<br>ΔC <sub>T</sub>                          | ΔΔC <sub>T</sub><br>0.00±0.11<br>0.20±0.06<br>ΔΔC <sub>T</sub>  | Relative<br>miRNA level<br>1.00±0.08<br>0.87±0.05<br>P=0.073<br>Relative<br>mRNA level   |
| Genotype<br>Control<br>(Oregon x w <sup>1118</sup> )<br>miR-125 <sup>LOF</sup><br>Genotype<br>Control<br>(Oregon x w <sup>1118</sup> ) | $miR-100 \\ (C_{T})$ 21.84±0.11 21.76±0.09 $Tom \\ (C_{T})$ 21.28±1.94                             | $2S \\ (C_T)$ 8.15±0.10 7.87±0.12 <i>Rpl32</i> (C_T) 15.26±0.08 | ΔC <sub>T</sub><br>$13.69\pm0.11$<br>$13.90\pm0.08$<br>ΔC <sub>T</sub><br>$6.01\pm3.35$ | ΔΔC <sub>T</sub> 0.00±0.11 0.20±0.06 ΔΔC <sub>T</sub> 0.00±0.06 | Relative         miRNA level         1.00±0.08         0.87±0.05         P=0.073         Relative         mRNA level         1.00±0.04 |

<sup>a</sup> - the  $\Delta C_T$  value is determined by subtracting the average  $C_T$  value of endogenous control gene (2S for miRNAs or Rpl32 for mRNA) from the average miRNA or mRNA  $C_T$  value.

<sup>b</sup> - the calculation of  $\Delta\Delta C_T$  involves subtraction by the  $\Delta C_T$  calibrator value ( $\Delta C_T$  value in *OregonR/w*<sup>1118</sup>).

 $^{c}$  - the range is given for relative levels determined by evaluating the expression:  $2^{-\Delta\Delta CT}.$ 

AVE±STDEV values are reported from experiments done in triplicates. Two-tailed Student's ttest was used to test for statistical significance

| Genotype   | Number of<br>CpCs<br>AVE±STDEV | Number of<br>germaria<br>analyzed | Number of<br>GSCs<br>AVE±STDEV | Number of<br>germaria<br>analyzed |
|--|--------------------------------|-----------------------------------|--------------------------------|-----------------------------------|
| $Control$ $(let-7C^{GK1} \times w^{1118})$   | 6.3±0.6                        | 16                                | 2.5±0.5                        | 11                                |
| $miR-125^{LOF}$<br>( $let-7C^{GK1}/let-7C^{K01}$ ; $P\{W8, let-7C^{\Delta miR-125})$ | 8.5±2.3<br>P=0.001             | 44                                | 3.4±1.1<br>P=0.001             | 28                                |
| $Rescue$ $(let-7C^{GK1}/let-7C^{K01}; P\{W8,$ $let-7C^{\Delta miR-125}/Tom^{99})$    | 6.2±0.6<br>P=0.899             | 20                                | 2.6±0.8<br>P=0.4790            | 20                                |
| <i>Tom</i> <sup>99</sup> /+  | 6.1±0.4<br>P=0.085             | 24                                | 2.3±0.8<br>P=0.794             | 27                                |
| ptc>miR-125<br>(UAS-miR-125/ptc-Gal4)  | 5.4±0.7<br>P=0.794             | 21                                | 2.6±0.5<br>P=0.900             | 20                                |
| bab1>miR-125<br>(UAS-miR-125/bab1-Gal4)  | 7.3±1.1<br>P=0.001             | 20                                | 4.1±0.7<br>P=0.001             | 21                                |
| <i>let-7C&gt;miR-125</i><br>( <i>let-7C<sup>GK1</sup>/+;UAS-miR-125/+</i> )          | 7.7±0.9<br>P=0.014             | 31                                | 3.7±1.0<br>P=0.001             | 12                                |

# Table S3. Deregulation of miR-125 results in the increased GSC niches that host supernumerary GSCs

One-way ANOVA with post-hoc Tukey HSD was used to test for statistical significance

| Genotype  | Conditions        | Number of<br>CpCs<br>AVE±AVEDEV | Number of<br>germaria<br>analyzed |
|---|-------------------|---------------------------------|-----------------------------------|
| $Control (let-7C^{GK1} x w^{1118})$   |                   | 6.0±0.9                         | 22                                |
| $let-7C^{ts} > miR-125$ $(let-7C^{GK1}/+; UAS-miR-125/tub-Gal80^{ts})$  | 18° C             | 6.0±0.9<br>P=0.900              | 10                                |
| $\begin{array}{c} \textit{let-7C>Tom}^{\textit{RNAi}} \\ \textit{(let-7C}^{\textit{GKI}} / +; \textit{UAS-Tom}^{\textit{RNAi}} / \textit{tub-Gal80}^{\textit{ts}}) \end{array}$ |                   | 6.1±1.2<br>P=0.900              | 16                                |
| $\frac{Control}{(let-7C^{GKts1} x w^{1118})}$   | shifted to        | 5.9±1.2<br>P=0.900              | 17                                |
| $let-7C^{ts} > miR-125$ $(let-7C^{GK1}/+; UAS-miR-125/tub-Gal80^{ts})$  | 29°C as<br>adults | 6.4±1.0<br>P=0.900              | 17                                |
| $\begin{array}{c} let-7C > Tom^{RNAi} \\ (let-7C^{GKI}/+; UAS-Tom^{RNAi}/tub-Gal80^{ts}) \end{array}$   |                   | 5.8±1.3<br>P=0.900              | 18                                |
| $Control (let-7C^{GK1} x w^{1118})$   |                   | 5.8±1.0<br>P=0.900              | 18                                |
| let-7C <sup>ts</sup> >miR-125<br>(let-7C <sup>GK1</sup> /+; UAS-miR-125/tub-Gal80 <sup>ts</sup> )   | 29° C             | 7.8±0.9<br>P=0.002              | 14                                |
| $\begin{array}{c} let-7C > Tom^{RNAi} \\ (let-7C^{GKI}/+; UAS-Tom^{RNAi}/tub-Gal80^{ts}) \end{array}$   |                   | 7.3±0.9<br>P=0.027              | 16                                |
| $Control (let-7C^{GK1} x w^{1118})$   |                   | 5.5±0.5                         | 12                                |
| $\begin{array}{c} let-7C > Tom^{RNAi} \\ (let-7C^{GKI}/+; UAS-Tom^{RNAi}/+) \end{array}$  | 25° C             | 7.1±0.9<br>P=0.006              | 10                                |
| $\begin{array}{c} \textit{let-7C>Tom} \\ \textit{(let-7C}^{GK1}/+; \textit{UAS-Tom/+}) \end{array}$   |                   | 8.2±1.1<br>P=0.001              | 18                                |
| $\begin{array}{c} let-7C > N^{CA} \\ (let-7C^{GKI}/+; UAS-Notch^{CA}/+) \end{array}$  |                   | 7.0±0.9<br>P=0.007              | 12                                |
| $\begin{array}{c} let-7C>N^{RNAi}\\ (let-7C^{GKI}/+; UAS-Notch^{RNAi}/+)\end{array}$  |                   | 6.9±0.7<br>P=0.005              | 16                                |
| $\begin{array}{c} let-7C>Dl\\ (let-7C^{GKl}/+; UAS-Dl/+)\end{array}$  |                   | 7.3±0.8<br>P=0.001              | 17                                |
| $let-7C>Dl^{RNAi} \ (let-7C^{GKI}/+; UAS-Dl^{RNAi}/+)$  |                   | 8.2±1.0<br>P=0.001              | 11                                |
| $\frac{EcR^{LBD}}{(hs-Gal4/UAS-EcR^{LBD})}$ heat shocked for 25' at 37°C at EL3 stage (72h AEL)   |                   | 8.2±1.5<br>P=0.001              | 18                                |

Table S4. Deregulation of the steroid-*miR-125*-Tom-Delta-Notch signaling cascade affects GSC niche development

One-way ANOVA with post-hoc Tukey HSD was used to test for statistical significance. All samples were compared to the controls at the same conditions

# Table S5. Steroid-miR-125-Tom-Delta-Notch signaling cascade is important for properGSC niche architecture

|   | Observed niche phenotypes |          |         | Davahaa   | Number of            |  |
|---|---------------------------|----------|---------|-----------|----------------------|--|
| Genotype  | Perfect                   | Enlarged | Ectopic | P-value   | germaria<br>analyzed |  |
| $Control (let-7C^{GK1} x w^{1118})$   | 100%                      | 0%       | 0%      | -         | 38                   |  |
| <i>EcR<sup>LBD</sup></i><br>( <i>hs-Gal4/UAS-EcR<sup>LBD</sup></i> )<br>heat shocked for 25 min at 37°C<br>at EL3 stage (72h AEL) | 31%                       | 25%      | 44%     | P=7.5E-08 | 16                   |  |
| <i>let-7C&gt;miR-125</i><br>( <i>let-7C<sup>GK1</sup>/+;UAS-miR-125/+</i> )   | 50%                       | 50%      | 0%      | P=5.7E-06 | 28                   |  |
| $miR-125^{LOF}$<br>( $let-7C^{GK1}/let-7C^{K01}$ ;<br>$P\{W8, let-7C^{\Delta miR-125}$ )  | 13%                       | 25%      | 63%     | P=1.2E-11 | 24                   |  |
| <i>let-7C&gt;Dl</i><br>( <i>let-7C<sup>GK1</sup></i> /+; <i>UAS-Dl</i> /+)  | 21%                       | 79%      | 0%      | P=1.4E-09 | 19                   |  |
| $\begin{array}{c} let-7C>Delta-RNAi\\ (let-7C^{GK1}/+; UAS-Dl^{RNAi}/+)\end{array}$   | 0%                        | 45%      | 55%     | P=1.1E-30 | 11                   |  |
| <i>let-7C&gt;Tom</i><br>( <i>let-7C<sup>GK1</sup>/+</i> ; <i>UAS-Tom/+</i> )  | 60%                       | 8%       | 32%     | P=3.1E-12 | 25                   |  |
| $\begin{array}{c} let-7C > Tom^{RNAi} \\ (let-7C^{GK1}/+; UAS-Tom^{RNAi}/+) \end{array}$  | 47%                       | 53%      | 0%      | P=7.9E-08 | 19                   |  |
| $\frac{Rescue}{(let-7C^{GKI}/let-7C^{K01};}$ $P\{W8, let-7C^{\Delta miR-125})/Tom^{99}$   | 100%                      | 0%       | 0%      | P=1       | 13                   |  |
| <i>Tom</i> <sup>99</sup> /+   | 100%                      | 0%       | 0%      | P=1       | 24                   |  |

Two-way tables and  $\chi^2$ -test were used to test for statistical significance



### Supplementary Figure 1

#### Figure S1. The strength of Notch signaling controls GSC niche formation

**A-C.** Ectopic activation of Delta leads to the formation of additional CpCs (arrows, E), while upon Notch downregulation ( $N^{lts}$ ), very few CpCs can be observed (arrows, F) when compared to *Control* (D). TFCs are marked by En (red), CpCs are marked by En+Tj (yellow, arrows) and ECs are marked by Tj (green).

**D-E.** Expression pattern of the Notch ligand, Delta (DI::GFP) and the Notch receptor (anti-Notch intracellular domain antibodies) in the developing ovary at the second instar larva (L2, D) and early third instar larva (EL3, E) stages. Note that ISCs, which are the somatic cells intermingled with the germline cells (Vasa, blue) co-express Notch and Delta (olive arrows) and are "undecided" with regards to their Notch signaling status.

**F.** Expression pattern of the Notch activity reporter NRE-EGFP in the developing GSC niche unit at the prepupal stage. Note that the posterior TFC (arrowhead) adjacent to the forming CpCs (arrows) shows the reduced Notch activity.



#### Figure S2. *let-7* complex miRNA are expressed in the developing ovary

A. Structure of the *let-7* complex gene region [modified from (Chawla and Sokol, 2012)] with the ecdysone response elements (EcRE, blue).

**B**. Expression of the ecdysone-activated transcription factor BrZ1 (green) during different stages of ovary development. BrZ1 is first detected in the newly formed TFCs at EL3 stage; afterwards, it is broadly expressed in the somatic cells, including TFCs. BrZ1 expression pattern is not altered in *miR-125* mutants.

C. Bar graph represents relative levels of miRNAs from the *let-7C* (*miR-100*, *let-7* and *miR-125*), in controls and *miR-125* mutants (*let-7-C*<sup>K01</sup>/CyO-GFP; *let-7-C*<sup>AmiR-125</sup>).

AVE±STDEV values are reported from the experiments done in triplicates.

**D**. *Tom* mRNA levels in *Control* and upon ubiquitous downregulation of *Tom* (*Act-Gal4/UAS-Tom*<sup>*RNAi*</sup>). Bar graph represents relative levels *Tom* mRNA.

AVE±STDEV values are reported from two biological experiments done in triplicates. Twotailed Student's t-test was used to test for statistical significance: \*\*\*P≤0.001.



#### Figure S3. Expression patterns of the used Gal4 driver lines

**A-C** Expression patterns of the *Gal4* driver lines crossed to *UAS-CD8::GFP* in the pepupal ovaries. *ptc-Gal4* (B) driver is expressed in ISCs, *bab1-Gal4* (C) driver is expressed in TFCs and CpCs, and *let-7C-Gal4* (D) is expressed in solitary TFCs. All three drivers in addition are expressed in SHCs. GFP (B-D, green, B'-D', white), En (red) marks TFC, Tj (blue) marks ESCs and Transient TFCs (TCs). TCs and CpCs express both Tj and En (magenta). Arrowheads point to TCs.





**B.** Deregulation of ecdysone, Delta, Notch or Tom leads to the increased CpC numbers. Box plots represent quantitative analysis of CpCs per germarium. One-way ANOVA with post-hoc Tukey HSD was used to test for statistical significance: \*\*\* $P \le 0.001$ , \*\* $P \le 0.01$ .



#### Figure S5. Delta expression is altered in the posterior TFCs of miR-125 mutants

**A-B**. Developing ovaries of *Control* and *miR-125* mutants of the comparable stage (Prepupa) shows that the expression pattern of Delta in is not globally altered. **C-D**. However, in the posterior TFCs, the membrane Delta levels are not reduced and the Delta-positive vesicle did not appear in *miR-125* mutants in comparison to controls.



#### Figure S6. miR-125 is required for the proper GSC niche establishment

**A**. Examples of control germaria in controls showing typical adult GSC niches (LamC-positive cells outlined by yellow dashed lines), which normally facilitate two or three GSCs (outlined by cyan dashed lines and marked by pMad and the presence of Add-positive spectrosomes). **B**. Loss of *miR-125 (miR-125<sup>LOF</sup>)* leads to the formation of the ectopic niches (dislocated from the anterior tip), which host supernumerary GSCs.

**C**. Upregulation of *miR-125* (*let-7C>miR-125*) leads to the enlarged niches, which also can facilitate supernumerary GSCs.