

SUPPORTING INFORMATION

Table S1: Normalization parameters (Eq. 1 in the main text) for surface area, desolvation, and contact energy terms.

Term	FSA				PSA			
	α	β	X_0	sgn	α	β	X_0	sgn
$\Delta SASA$	5.2300	0.0047	0	+1	5.3700	0.0045	0	+1
E_{des}	8.0990	0.3112	10	+1	8.7069	0.3148	10	+1
E_{AACE18}	6.4403	0.0797	50	-1	5.8480	0.0695	50	-1

Table S2. Top ten template-based docking predictions for dimeric T119/T0917.

No.	Protein model	Template	Rank by F(S)	F(S)	AACE18	TM-score ⁽¹⁾	f_{nat}	i -RMSD, Å ⁽²⁾	CAPRI class ⁽³⁾
1	35 ⁽⁴⁾	3iv7AB	22	0.967	-85.7	0.77	0.62	2.3	A
2	51 ⁽⁵⁾	3iv7AB	6	0.981	-109.6	0.77	0.45	2.4	M
3	67 ⁽⁶⁾	5br4AB	749	0.517	38.8	0.90	0.00	24.9	I
4	71 ⁽⁷⁾	4rflAB	245	0.850	-91.4	0.66	0.12	6.8	I
5	100 ⁽⁸⁾	3iv7AB	49	0.952	-103.2	0.77	0.49	2.0	M
6	154 ⁽⁹⁾	1jq5A ₁ A ₇ ⁽¹³⁾	9	0.975	-83.8	0.76	0.58	3.5	A
7	154 ⁽⁹⁾	3ox4BC	734	0.530	-5.6	0.92	0.00	24.8	I
8	174 ⁽¹⁰⁾	1jq5A ₁ A ₇ ⁽¹³⁾	15	0.969	-76.8	0.74	0.66	3.2	A
9	176 ⁽¹¹⁾	1jq5A ₁ A ₇ ⁽¹³⁾	25	0.965	-78.7	0.75	0.59	3.6	A
10	177 ⁽¹²⁾	1vljAB	49	0.952	-103.2	0.77	0.45	3.3	A

⁽¹⁾ Score of the structural alignment of protein model and one of the template chains.

⁽²⁾ C^α root-mean-square deviation of the ligand interface.

⁽³⁾ I – incorrect; A – acceptable-, M – medium-, H – high-quality model.

⁽⁴⁾ Model 5 of DISTILL server.¹

⁽⁵⁾ Model 4 of Princeton TIGRESS (FLOUDAS) server.²

⁽⁶⁾ Model 5 of GOAL server.³

⁽⁷⁾ Model 4 of HHGG server.⁴

⁽⁸⁾ Model 5 of MULTICOM-CONSTRUCT server.⁵

⁽⁹⁾ Model 3 of SEOK-naïve-assembly server.⁶

⁽¹⁰⁾ Model 2 of YASARA server.⁷

⁽¹¹⁾ Model 4 of YASARA server.

⁽¹²⁾ Model 5 of YASARA server.

⁽¹³⁾ Models 1 and 7 in the PDB biological unit.

Table S3. Top ten template-based docking predictions for trimeric T110/T0860.

Characteristics of the predictions are for the interface obtained by superimposition of target models onto chains A₁ and A₂ of the templates (other interfaces are nearly identical).

No.	Protein model	Template	F(S)	AACE18	TM-score	f_{nat}	i -RMSD, Å	CAPRI class
1	1 ⁽²⁾	3zpeA ₁ A ₂ A ₃ ⁽¹⁾	0.963	-25.1	0.96	0.31	2.76	M
2	1 ⁽²⁾	4cw8 A ₁ A ₂ A ₃ ⁽¹⁾	0.965	-25.8	0.97	0.29	2.80	A
3	2 ⁽³⁾	3zpe A ₁ A ₂ A ₃	0.957	-19.7	0.97	0.27	2.70	A
4	2 ⁽³⁾	4cw8 A ₁ A ₂ A ₃	0.961	-22.3	0.96	0.29	2.70	A
5	3 ⁽⁴⁾	3zpe A ₁ A ₂ A ₃	0.988	-45.7	0.87	0.58	1.86	M
6	3 ⁽⁴⁾	4cw8A ₁ A ₂ A ₃	0.989	-50.3	0.87	0.58	1.87	M
7	4 ⁽⁵⁾	3zpe A ₁ A ₂ A ₃	0.986	-48.5	0.89	0.62	1.57	M
8	4 ⁽⁵⁾	4cw8 A ₁ A ₂ A ₃	0.988	-51.3	0.88	0.62	1.60	M
9	5 ⁽⁶⁾	3zpe A ₁ A ₂ A ₃	0.991	-39.7	0.98	0.47	2.25	M
10	5 ⁽⁶⁾	4cw8 A ₁ A ₂ A ₃	0.991	-40.8	0.98	0.43	2.28	M

⁽¹⁾ Models of 1, 2 and 3 in the PDB biounit.

⁽²⁾ Model 1 of ZHANG server.⁸

⁽³⁾ Model 2 of ZHANG server.

⁽⁴⁾ Model 1 of BAKER-ROSETTA server.⁹

⁽⁵⁾ Model 5 of BAKER-ROSETTA server.

⁽⁶⁾ Model 1 of HHPRED0 server.¹⁰

Table S4. Top ten template-based docking predictions for trimeric T111/T0867.

Characteristics of the predictions are for the interface obtained by superimposition of target models onto the first two chains of the templates.

No.	Protein model	Template	F(S)	AACE18	TM-score	f_{nat}	i -RMSD, Å	CAPRI class
1	1 ⁽¹⁾	4d0uABC	0.940	-22.7	0.99	0.67	0.92	H
2	1 ⁽¹⁾	4d0uADB	0.832	-24.6	0.97	0.65	0.92	H
3	1 ⁽¹⁾	4d0uACD	0.944	-24.7	0.97	0.02	8.32	I
4	1 ⁽¹⁾	4d1fABE	0.952	-24.9	0.99	0.63	1.00	M
5	1 ⁽¹⁾	4d1fACE	0.954	-25.9	0.87	0.02	8.35	I
6	1 ⁽¹⁾	4d1fAEL	0.804	2.8	0.99	0.00	20.15	I
7	2 ⁽²⁾	4d0uABC	0.977	-26.2	0.96	0.87	0.71	H
8	3 ⁽³⁾	4d0uABC	0.919	-25.0	0.95	0.60	1.19	M
9	4 ⁽⁴⁾	4d0uABC	0.910	-26.1	0.95	0.62	0.96	H
10	5 ⁽⁵⁾	4d0uABC	0.929	-25.7	0.97	0.58	1.01	M

⁽¹⁾ Model 4 of HHGG server.⁴

⁽²⁾ Model 1 of BAKER-ROSETTA servr.⁹

⁽³⁾ Model 1 of GOAL server.³

⁽⁴⁾ Model 3 of SEOK server.⁶

⁽⁵⁾ Model 1 of ZHANG server.⁸

Table S5. Top ten template-based docking predictions for trimeric T112/T0881.

Characteristics of the predictions are for the interface obtained by superimposition of target models onto the first two chains of the templates.

No.	Protein model	Template	F(S)	AACE18	TM-score	f_{nat}	i -RMSD, Å	CAPRI class
1	1 ⁽¹⁾	2iumABC	0.661	-15.03	0.82	0.53	3.99	A
2	1 ⁽¹⁾	2vtwAEF	0.807	-9.56	0.84	0.51	4.27	A
3	2 ⁽²⁾	2vtwAEF	0.829	-15.28	0.92	0.38	4.27	A
4	3 ⁽³⁾	2vtwAEF	0.873	-17.78	0.92	0.26	4.33	A
5	4 ⁽⁴⁾	2vtwAEF	0.885	-14.68	0.93	0.38	4.15	A
6	5 ⁽⁵⁾	2vtwAEF	0.831	-6.83	0.89	0.43	4.32	A
7	6 ⁽⁶⁾	2vtwAEF	0.800	-14.56	0.91	0.45	4.83	I
8	7 ⁽⁷⁾	2vtwAEF	0.778	-7.92	0.89	0.53	4.85	A
9	9 ⁽⁸⁾	2iumABC	0.795	-18.62	0.90	0.55	3.83	A
10	10 ⁽⁹⁾	2vtwAEF	0.796	-12.79	0.90	0.53	4.85	I

⁽¹⁾ Model 1 of BAKER-ROSETTA server.⁹

⁽²⁾ Model 3 of MUFOLD1 server.¹¹

⁽³⁾ Model 3 of MULTICOM-NOVEL server.⁵

⁽⁴⁾ Model 1 of CHUO-U server.¹²

⁽⁵⁾ Model 2 of QUARK server.¹³

⁽⁶⁾ Model 2 of ZHANG server.⁸

⁽⁷⁾ Model 4 of RBO_ALEPH server.¹⁴

⁽⁸⁾ Model 1 of RAPTOR-X server.¹⁵

⁽⁹⁾ Model 2 of SEOK server.⁶

Table S6. Top ten template-based docking predictions for octameric T118/T0906.

Characteristics of the predictions are for the interface obtained by superimposition of target models onto models 2 and 6 of the template biounit (other interfaces are nearly identical).

No.	Protein model	Template	F(S)	AACE18	TM-score	f_{nat}	i -RMSD, Å	CAPRI class
1	1 ⁽²⁾	3t2c ⁽¹⁾	0.998	-72.25	0.97	0.68	0.88	H
2	2 ⁽³⁾		0.996	-52.36	0.96	0.55	0.99	H
3	3 ⁽⁴⁾		0.998	-70.84	0.97	0.61	0.03	H
4	4 ⁽⁵⁾		0.995	-41.39	0.97	0.62	0.93	H
5	5 ⁽⁶⁾		0.997	-72.42	0.94	0.55	1.04	M
6	6 ⁽⁷⁾		0.993	-57.43	0.98	0.59	1.06	M
7	7 ⁽⁸⁾		0.990	-48.84	0.97	0.59	1.06	M
8	8 ⁽⁹⁾		0.998	-58.78	0.98	0.62	1.04	M
9	9 ⁽¹⁰⁾		0.997	-67.39	0.94	0.58	1.03	M
10	10 ⁽¹¹⁾		0.990	-42.93	0.97	0.65	1.23	M

⁽¹⁾ The same template (octameric in biological unit, monomeric in the asymmetric unit) was used for all docking models.

⁽²⁾ Model 5 of BAKER-ROSETTA server.⁹

⁽³⁾ Model 1 of YASARA server.⁷

⁽⁴⁾ Model 1 of QUARK server.¹³

⁽⁵⁾ Model 5 of HHGG server.⁴

⁽⁶⁾ Model 2 of ZHANG server.⁸

⁽⁷⁾ Model 3 of SLBIO server (reference not available).

⁽⁸⁾ Model 1 of TSSPRED2 server (reference not available)

⁽⁹⁾ Model 1 of HHPRED1 server.¹⁰

⁽¹⁰⁾ Model 5 of MULTICOM-CONSTRUCT server.⁵

⁽¹¹⁾ Model 1 of DISTILL server.¹

Table S7. Top ten template-based docking predictions for hetero-dimeric T120/T0921-T0922.

No.	T0921 model	T0922 model	Template	F(S)	AACE18	TM-score ⁽¹⁾	TM-score ⁽²⁾	f_{nat}	i -RMSD, Å	CAPRI class
1	21 ⁽⁴⁾	89 ⁽¹³⁾	4dh2AB	0.765	4.05	0.94	0.98	0.26	3.63	A
2 ⁽³⁾	21 ⁽⁴⁾	89 ⁽¹³⁾	4dh2AB	–	–	–	–	0.22	6.55	I
3	73 ⁽⁵⁾	98 ⁽¹⁴⁾	2cclAB	0.637	-3.92	0.85	0.60	0.26	3.61	A
4	74 ⁽⁶⁾	24 ⁽¹⁵⁾	2cclAB	0.645	-10.24	0.83	0.59	0.33	3.47	M
5	87 ⁽⁷⁾	29 ⁽¹⁶⁾	2cclAB	0.624	-14.41	0.83	0.59	0.26	4.28	A
6	95 ⁽⁸⁾	51 ⁽¹⁷⁾	2cclAB	0.602	-23.20	0.84	0.57	0.36	3.63	M
7	142 ⁽⁹⁾	174 ⁽¹⁸⁾	2cclAB	0.725	-10.99	0.79	0.71	0.32	3.12	M
8	145 ⁽¹⁰⁾	24 ⁽¹⁵⁾	2cclAB	0.667	-15.03	0.80	0.59	0.33	3.41	A
9	169 ⁽¹¹⁾	46 ⁽¹⁹⁾	4uypAB	0.604	-0.33	0.85	0.95	0.03	12.69	I
10	186 ⁽¹²⁾	143 ⁽²⁰⁾	4dh2AB	0.638	-7.57	0.92	0.72	0.24	3.50	A

⁽¹⁾ TM-score of alignment of T0921 model and first template chain.

⁽²⁾ TM-score of alignment of T0922 model and second template chain.

⁽³⁾ Prediction obtained from prediction 1 by manual shift of the T0922 model.

⁽⁴⁾ Model 1 of CHUO-U2 server.¹²

⁽⁵⁾ Model 4 of INTFOLD4 server.¹⁶

⁽⁶⁾ Model 5 of INTFOLD4 server.

⁽⁷⁾ Model 3 of MUFOLD2 server.¹¹

⁽⁸⁾ Model 1 of MULTICOM-CONSTRUCT server.⁵

⁽⁹⁾ Model 2 of RBO-ALEPH server.¹⁴

⁽¹⁰⁾ Model 5 of RBO-ALEPH server.

⁽¹¹⁾ Model 3 of TSSPRED2 server (reference and link are not available).

⁽¹²⁾ Model 5 of ZHOU-SPARKS-X server.¹⁷

⁽¹³⁾ Model 4 of MUFOLD1 server.¹¹

⁽¹⁴⁾ Model 4 of MULTICOM-CONSTRUCT server.⁵

⁽¹⁵⁾ Model 4 of CHUO-U2 server.¹²

⁽¹⁶⁾ Model 4 of CHUO-U server.¹²

⁽¹⁷⁾ Model 4 of FLOUDAS server.²

⁽¹⁸⁾ Model 3 of YASARA server.⁷

⁽¹⁹⁾ Model 1 of FFAS03 server.¹⁸

⁽²⁰⁾ Model 3 of RBO-ALEPH server.¹⁴

Table S8. Top ten template-based (1-6) and free (7-10) docking predictions for homodimeric T114/T0875.

No.	Protein model	Template	F(S)	AACE18	TM-score or Rank ⁽¹⁾	f_{nat}	i -RMSD, Å	CAPRI class
1	1 ⁽²⁾	2fbqA ₁ A ₂	0.056	-83.04	0.34	0.00	9.67	I
2	2 ⁽³⁾	1mscA ₁ A ₂	0.239	-77.24	0.33	0.00	17.27	I
3	4 ⁽⁴⁾	3f13A ₁ A ₂	0.120	-76.98	0.36	0.00	8.80	I
4	7 ⁽⁵⁾	4q95AB	0.832	-1.56	0.78/0.66	0.00	9.88	I
5	8 ⁽⁶⁾	4dpzA ₁ A ₄	0.246	-6.79	0.81	0.00	9.91	I
6	10 ⁽⁷⁾	4dotA ₁ A ₃	0.302	-0.93	0.79	0.00	19.61	I
7	3 ⁽⁸⁾	–	–	-67.54	4759	0.27	15.61	I
8	4 ⁽⁴⁾	–	–	-70.05	7629	0.00	6.64	I
9	6 ⁽⁹⁾	–	–	-96.80	14270	0.04	19.02	I
10	7 ⁽⁵⁾	–	–	-118.38	26793	0.04	8.13	I

⁽¹⁾ Integer numbers are GRAMM ranking by shape complementarity.

⁽²⁾ Model 1 of BAKER-ROSETTA server.⁹

⁽³⁾ Model 3 of INTFOLD4 server.¹⁶

⁽⁴⁾ Model 4 of SEOK server.⁶

⁽⁵⁾ Model 3 of ZHANG server.⁸

⁽⁶⁾ Model 3 of ATOME2-CBS server.¹⁹

⁽⁷⁾ Model 3 of FFAS03 server.¹⁸

⁽⁸⁾ Model 1 of ZHANG server.

Table S9. Top ten template-based docking predictions for homodimeric T116/T0893.

No.	Protein model	Template	F(S)	AACE18	TM-score ⁽¹¹⁾	f_{nat}	i -RMSD, Å	CAPRI class
1	9 ⁽¹⁾	3d36AB	0.996	-62.09	0.82	0.09	11.48	I
2	21 ⁽²⁾	3a0rA ₁ A ₂	0.969	-105.95	0.72	0.10	15.26	I
3	26 ⁽³⁾	4bixAB	0.837	-94.64	0.75/0.70	0.40	14.47	I
4	44 ⁽⁴⁾	4mpcA ₁ A ₂	0.946	-31.62	0.83	0.00	27.79	I
5	49 ⁽⁵⁾	3a0rA ₁ A ₂	0.965	-85.26	0.75	0.08	15.13	I
6	105 ⁽⁶⁾	4q20AB	0.935	-68.71	0.90/0.75	0.09	15.69	I
7	116 ⁽⁷⁾	4mpcA ₁ A ₂	0.987	-42.27	0.83	0.00	27.24	I
8	120 ⁽⁸⁾	4ctiBD	0.722	-74.04	0.53	0.01	19.04	I
9	140 ⁽⁹⁾	3d36AB	0.998	-71.73	0.84	0.07	11.60	I
10	141 ⁽¹⁰⁾	3d36AB	0.991	-79.13	0.81	0.14	11.68	I

⁽¹⁾ Model 1 of BHAGEERATHH-PLUS server.²⁰

⁽²⁾ Model 2 of DISTILL server.¹

⁽³⁾ Model 2 of FALCON-TOPO server.²¹

⁽⁴⁾ Model 3 of GOAL-COMPLEX server.³

⁽⁵⁾ Model 3 of GOAL server.³

⁽⁶⁾ Model 5 of QUARK server.¹³

⁽⁷⁾ Model 5 of SEOK-ASSEMBLY server.⁶

⁽⁸⁾ Model 4 of SEOK-NAIVE server.⁶

⁽⁹⁾ Model 5 of YASARA server.⁷

⁽¹⁰⁾ Model 1 of ZHANG server.⁸

⁽¹¹⁾ Two values indicate heterodimeric template.

Table S10. Top ten docking predictions for hetero-dimeric T113/T0884-T0885.

No.	T0885 model	T0884 model	AACE18	Rank	f_{nat}	i -RMSD, Å	CAPRI class
1	1 ⁽¹⁾	8 ⁽⁵⁾	-54.63	21004	0.052	12.72	I
2 ⁽³⁾	1 ⁽¹⁾	29 ⁽⁶⁾	-58.90	19488	0.169	7.28	I
3	2 ⁽²⁾	13 ⁽⁷⁾	-51.22	20900	0.039	11.58	I
4	3 ⁽³⁾	12 ⁽⁸⁾	-51.61	12491	0.104	7.11	I
5	3 ⁽³⁾	12 ⁽⁸⁾	-50.27	24146	0.026	10.52	I
6	3 ⁽³⁾	16 ⁽⁹⁾	-51.17	28463	0.065	8.48	I
7	4 ⁽⁴⁾	13 ⁽⁷⁾	-61.44	5017	0.117	9.76	I
8	4 ⁽⁴⁾	16 ⁽⁹⁾	-50.71	4871	0.078	8.22	I
9	4 ⁽⁴⁾	27 ⁽¹⁰⁾	-50.19	8290	0.130	8.52	I
10	4 ⁽⁴⁾	29 ⁽⁶⁾	-50.90	23951	0.130	7.27	I

⁽¹⁾ Model 1 of BAKER-ROSETTA server.⁹

⁽²⁾ Model 2 of ZHANG server.⁸

⁽³⁾ Model 4 of GOAL-COMPLEX server.³

⁽⁴⁾ Model 1 of QUARK server.¹³

⁽⁵⁾ Model 2 of PCONS-NET server.²²

⁽⁶⁾ Model 5 of MULTICOM-CONSTRUCT server.⁵

⁽⁷⁾ Model 1 of MULTICOM-NOVEL server.⁵

⁽⁸⁾ Model 5 of ZHANG server.

⁽⁹⁾ Model 5 of GOAL-COMPLEX server.

⁽¹⁰⁾ Model 5 of PCONS-NET server.

Table S11. Top ten docking predictions for hetero-tetrameric T117/T0903-T0904.

Characteristics of the predictions are with respect to the interface of chains H and R in the native target 5a7d (blue and yellow in Figure 6A, main text). Predictions with identical docking parameters (columns 2 – 5) differ by arrangement of the truncated parts of the ligand (see main text).

No.	T0903 model	T0904 model	AACE18	Rank	f_{nat}	i -RMSD, Å	CAPRI class
1	15 ⁽¹⁾	1 ⁽²⁾	-87.3	18244	0.556	16.13	I
2	15 ⁽¹⁾	1 ⁽²⁾	-87.3	18244	0.000	16.13	I
3	15 ⁽¹⁾	93 ⁽³⁾	-94.1	66805	0.000	16.15	I
4	15 ⁽¹⁾	93 ⁽³⁾	-94.1	66805	0.641	16.13	I
5	15 ⁽¹⁾	24 ⁽⁴⁾	-35.7	2190	0.578	9.64	I
6	15 ⁽¹⁾	24 ⁽⁴⁾	-35.7	2190	0.000	9.64	I
7	15 ⁽¹⁾	68 ⁽⁵⁾	-65.0	1027	0.000	10.34	I
8	15 ⁽¹⁾	84 ⁽⁶⁾	-55.5	600	0.000	10.95	I
9	15 ⁽¹⁾	68 ⁽⁵⁾	-74.6	480	0.000	21.44	I
10	15 ⁽¹⁾	106 ⁽⁷⁾	-47.0	64/61976 ⁽⁸⁾	0.007	11.01	I

⁽¹⁾ Model 3 of BHAGEERATHH-PLUS server.²⁰

⁽²⁾ Model 1 of ATOME2-CBS server.¹⁹

⁽³⁾ Model 3 of MYPROTEIN-ME server (reference not available).

⁽⁴⁾ Model 4 of DISTILL server.¹

⁽⁵⁾ Model 1 of MUFOLD2 server.¹¹

⁽⁶⁾ Model 4 of MULTICOM-NOVEL server.⁵

⁽⁷⁾ Model 3 of PHYRETOPOALPHA server.²³

⁽⁸⁾ Rank 61976 of the docking prediction consists of two docking predictions with rank 64 between models of receptor 15 and ligand 106.

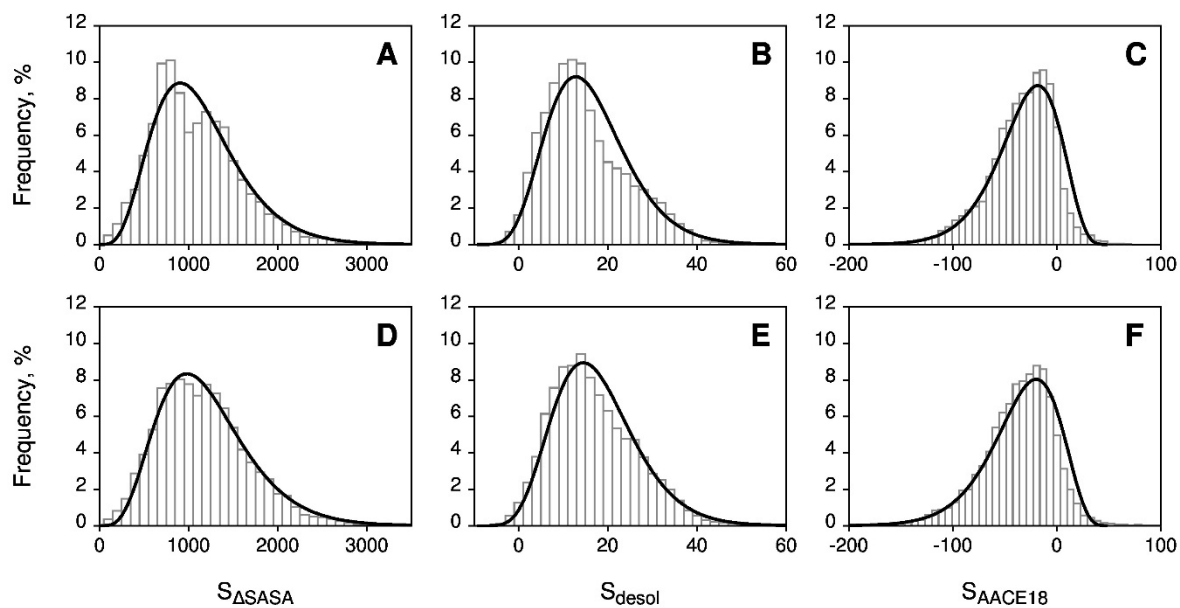


Figure S1. Raw-value histograms of buried surface area (A, D), desolvation energy (B, E) and interface contact potential AACE18 (C, F) for 7,056 FSA (A–C) and 11,400 PSA (D–F) near-native models. The data is fitted by gamma distributions (see main text for details).

REFERENCES

1. Bau D, Martin AJ, Mooney C, Vullo A, Walsh I, Pollastri G. Distill: a suite of web servers for the prediction of one-, two- and three-dimensional structural features of proteins. *BMC bioinformatics* 2006;7:402.
2. Khoury GA, Smadbeck J, Kieslich CA, Koskosidis AJ, Guzman YA, Tamamis P, Floudas CA. Princeton_TIGRESS 2.0: High refinement consistency and net gains through support vector machines and molecular dynamics in double-blind predictions during the CASP11 experiment. *Proteins* 2017;85(6):1078-1098.
3. Joo K, Joung I, Lee SY, Kim JY, Cheng QY, Manavalan B, Joung JY, Heo S, Lee J, Nam M, Lee IH, Lee SJ, Lee J. Template based protein structure modeling by global optimization in CASP11. *Proteins* 2016;84:221-232.
4. Meier A, Soding J. Automatic Prediction of Protein 3D Structures by Probabilistic Multi-template Homology Modeling. *Plos Comput Biol* 2015;11(10):e1004343.
5. Cao RZ, Wang Z, Cheng JL. Designing and evaluating the MULTICOM protein local and global model quality prediction methods in the CASP10 experiment. *Bmc Struct Biol* 2014;14:13.
6. Ko J, Park H, Heo L, Seok C. GalaxyWEB server for protein structure prediction and refinement. *Nucleic Acids Res* 2012;40(W1):W294-W297.
7. Krieger E, Vriend G. New Ways to Boost Molecular Dynamics Simulations. *J Comput Chem* 2015;36(13):996-1007.
8. Yang JY, Yan RX, Roy A, Xu D, Poisson J, Zhang Y. The I-TASSER Suite: protein structure and function prediction. *Nat Methods* 2015;12(1):7-8.
9. Simons KT, Bonneau R, Ruczinski I, Baker D. Ab initio protein structure prediction of CASP III targets using ROSETTA. *Proteins* 1999:171-176.
10. Soding J, Biegert A, Lupas AN. The HHpred interactive server for protein homology detection and structure prediction. *Nucleic Acids Res* 2005;33:W244-W248.
11. Zhang JF, Wang QG, Barz BD, He ZQ, Kosztin I, Shang Y, Xu D. MUFOLD: A new solution for protein 3D structure prediction. *Proteins* 2010;78(5):1137-1152.
12. Kanou K, Iwadate M, Hirata T, Terashi G, Umeyama H, Takeda-Shitaka M. FAMSD: A Powerful Protein Modeling Platform that Combines Alignment Methods, Homology Modeling, 3D Structure Quality Estimation and Molecular Dynamics. *Chem Pharm Bull* 2009;57(12):1335-1342.
13. Xu D, Zhang Y. Ab initio protein structure assembly using continuous structure fragments and optimized knowledge-based force field. *Proteins* 2012;80(7):1715-1735.
14. Mabrouk M, Putz I, Werner T, Schneider M, Neeb M, Bartels P, Brock O. RBO Aleph: leveraging novel information sources for protein structure prediction. *Nucleic Acids Res* 2015;43(W1):W343-W348.
15. Wang S, Sun SQ, Li Z, Zhang RY, Xu JB. Accurate De Novo Prediction of Protein Contact Map by Ultra-Deep Learning Model. *PlosS Comput Biol* 2017;13(1):e1005324
16. McGuffin LJ, Atkins JD, Salehe BR, Shuid AN, Roche DB. IntFOLD: an integrated server for modelling protein structures and functions from amino acid sequences. *Nucleic Acids Res* 2015;43(W1):W169-173.
17. Yang Y, Faraggi E, Zhao H, Zhou Y. Improving protein fold recognition and template-based modeling by employing probabilistic-based matching between predicted one-

- dimensional structural properties of query and corresponding native properties of templates. *Bioinformatics* 2011;27(15):2076-2082.
18. Jaroszewski L, Li Z, Cai XH, Weber C, Godzik A. FFAS server: novel features and applications. *Nucleic Acids Res* 2011;39(Web Server issue):W38-44.
 19. Pons JL, Labesse G. @TOME-2: a new pipeline for comparative modeling of protein-ligand complexes. *Nucleic Acids Res* 2009;37(Web Server issue):W485-491.
 20. Jayaram B, Dhingra P, Mishra A, Kaushik R, Mukherjee G, Singh A, Shekhar S. Bhageerath-H: a homology/ab initio hybrid server for predicting tertiary structures of monomeric soluble proteins. *BMC bioinformatics* 2014;15 Suppl 16:S7.
 21. Wang C, Zhang H, Zheng WM, Xu D, Zhu J, Wang B, Ning K, Sun S, Li SC, Bu D. FALCON@home: a high-throughput protein structure prediction server based on remote homologue recognition. *Bioinformatics* 2016;32(3):462-464.
 22. Wallner B, Fang HS, Elofsson A. Automatic consensus-based fold recognition using Pcons, ProQ, and pmodeller. *Proteins* 2003;53:534-541.
 23. Kelley LA, Mezulis S, Yates CM, Wass MN, Sternberg MJE. The Phyre2 web portal for protein modeling, prediction and analysis. *Nat Protoc* 2015;10(6):845-858.