SUPPLEMENTARY FIGURES

Supplementary Figure 1.

EndoC- β H1 cells were either mock-tranfected (control) or transfected with PolyI:C and analyzed 24 hours later. **(A, B)** Heatmap from global transcriptomic analysis and RT-qPCR data represent up-regulated PRR genes (n=3). **(C)** Heatmap from global transcriptomic analysis indicating that PolyI:C treatment induces the expression of *B2M*. Data from RT-qPCR are the means \pm SD of 3 independent experiments (t-tests; **p<0.01 and ***p<0.001 relative to control.)

Supplementary Figure 2.

EndoC- β H1 cells were either mock-transfected (control) or transfected with PolyI:C and analyzed 24 hours later. β cell disallowed genes (19, 20) from global transcriptomic analysis are listed (n=3; the means \pm SD of 3 independent experiments are listed)

Supplementary Figure 3.

(A-D) EndoC-βH1 cells were either mock-tranfected (control) or transfected with PolyI:C. 24 hours later, cells were stained with Annexin-V and Propidium lodide and FAC-sorted (A, B) (n=3; Representative FACS plot of 3 independent experiment is shown). RNAs were prepared from living Annexin-V^{neg} / Propidium lodide^{neg} cells for RT-qPCR analysis (C, D). Data from FACS analyses and RT-qPCR are the means ± SD of 3 independent experiments (t-tests; *p<0.05, **p<0.01 and ***p<0.001 relative to control.)

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EndoC-βH1 cells were treated with PMA for 8 hours and RNAs were prepared for RT-qPCR analyses of *HES1*, *MYC*, *MAFA* and *SLC30A8* genes expression levels (A). EndoC-βH1 cells were treated with PMA and/or PDTC for 8 hours. RNAs were prepared for RT-qPCR (B) and proteins for Western blot analyses (C).

Data from RT-qPCR and Western blot are the means \pm SD of 3 independent experiments (t-tests or ANOVA with Bonferroni's correction for multiple comparison; *<0.05 and ***p<0.001 relative to control.)

Supplementary Figure 6.

(A) EndoC- β H1 cells were treated for 72 hours with PolyI:C (1-50 μ g/ml) that was added (without transfection) in the culture medium. RNAs were prepared for RT-qPCR (n=3) (B) Conditioned medium from mock-transfected (CTRL) or PolyI:C-transfected EndoC- β H1 cells was treated with or without RNAse-A and added to naive EndoC- β H1 cells. RNAs were prepared 72 hours later, for RT-qPCR (n=3).

Data from RT-qPCR represent the means \pm SD of 3 independent experiments (ANOVA with Bonferroni's correction; ***p<0.001 relative to PolyI:C treatment.)

Supplementary Figure 7.

(A) Scheme of the SOX9 constructs used for EndoC-βH1 transfection. (**B-C**) SOX9 responsive elements were co-transfected with MCS-ires-GFP, SOX9WT-ires-GFP, VP16-ires-GFP or VP16-SOX9ΔTAD-ires-GFP. Luciferase activity was measured 48h later (n=4).

Data from Luciferase assay represent the means ± SD of 4 independent experiments (ANOVA with Bonferroni's correction; **p<0.01 and ***p<0.001 relative to control.)

Supplementary Figure 8.

Venn diagram with genes up-regulated (>2 fold) in EndoC-βH1 cells treated with PolyI:C and following ectopic expression of SOX9/VP16-SOX9ΔTAD and

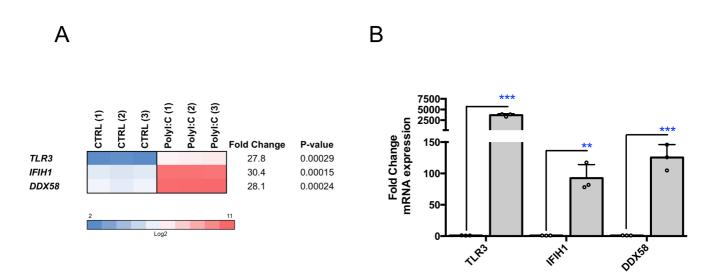
orthologous genes up-regulated (>2 fold) in dedifferentiated β cells from NOD mice (28). In red: genes up-regulated by both PolyI:C and SOX9/VP16-SOX9 Δ TAD; In blue: genes up-regulated in NOD dedifferentiated β cells and by SOX9/VP16-SOX9 Δ TAD; In green: genes up-regulated in all three sets.

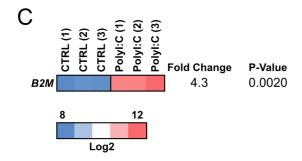
Supplementary Table 1.

Probes sequences

	Forward Sequence (5'>3')	Reverse Sequence (5'>3')	
ACTB	CTGTACGCCAACACAGTGCT	GCTCAGGAGGAGCAATGATC	
CTGF	CAAGGGCCTCTTCTGTGACTT	GGTACACCGTACCACCGAAG	
DDX58	TGACTGGACGTGGCAAAACA	CAGCAACTGAGGTGGCAATC	
DEPTOR	CTTCTGACAGAGAGACGGCA	CCGTCATCCTTTCTAAAGCGG	
G6PC2	CCATGCCTTGAACAGTTCCC	GCCAGATGGACTTCCTGGAC	
HES1	GTCAACACGACACGGGATAAACC	TTTCCAGAATGTCCGCCTTCTCC	
IFIH1	GCATGGAGGAGGAACTGTTGA	CCAGTTTTCTTTCTGCACAATCCT	
IFNA2	AAACCCACAGCCTGGGTAGC	CAGGGATGGTTTCAGCCTTTTGG	
IFNB	GTTGAGAACCTCCTGGCTAATG	GGTAATGCAGAATCCTCCCATAATA	
IL6	CCAGAGCTGTGCAGATGAGT	GGGTCAGGGGTGGTTATTGC	
IL8	AAATCTGGCAACCCTAGTCTG	GTGAGGTAAGATGGTGGCTAAT	
INS	TGTCCTTCTGCCATGGCCCT	TTCACAAAGGCTGCGGCTGG	
INS pre-mRNA	GTGAACCAACACCTGTGCGG	AGGGGCAGCAATGGGCAGTT	
LRRTM2	CCAACTCCCTGCGGACTATC	GCAAATCCATTGCGAGCCAA	
MAFA	ATTCTGGAGAGCGAGAAGTGCCAA	CGCCAGCTTCTCGTATTTCTCCTT	
MAFB	CACCACCTGGAGAATGAGAAG	TTCTCGCACTTGACCTTGTAG	
MAML2	TGGGATAAACGGAGAGCAGC	CATTGGGTCGCTTGCTGTTG	
MYC	GTAGTGGAAAACCAGCAGCC	AGAAATACGGCTGCACCGAG	
NEUROD1	ATTGCACCAGCCCTTCCTTTGATG	TCGCTGCAGGATAGTGCATGGTAA	
NKX6-1	GAAGAGGACGACTACAATAAG	CTGCTGGACTTGTGCTTCT	
PDX1	TACTGGATTGGCGTTGTTTGTGGC	AGGGAGCCTTCCAATGTGTATGGT	
PPIA	ATGGCAAATGCTGGACCCAACA	ACATGCTTGCCATCCAACCACT	
RELA	TGAGCCCACAAAGCCTTATC	ACAATGCCAGTGCCATACA	
SLC2A2	AGCTGCATTCAGCAATTGGACCTG	ATGTGAACAGGGTAAAGGCCAGGA	
SLC30A8	ACAGCCAAGTGGTTCGGAGAGAAA	TTGGGAAACTGACGGTGTGACTGA	
SOX9	TTCACCTACATGAACCCCGC	AAGGTCGAGTGAGCTGTGTG	
TLR3	GCGCTAAAAAGTGAAGAACTGGA	TTGCGTGAAAACACCCTGGA	
TNFA	GATCCCTGACATCTGGAATCTG	GAAACATCTGGAGAGGAAGG	

Supplementary Figure 1.





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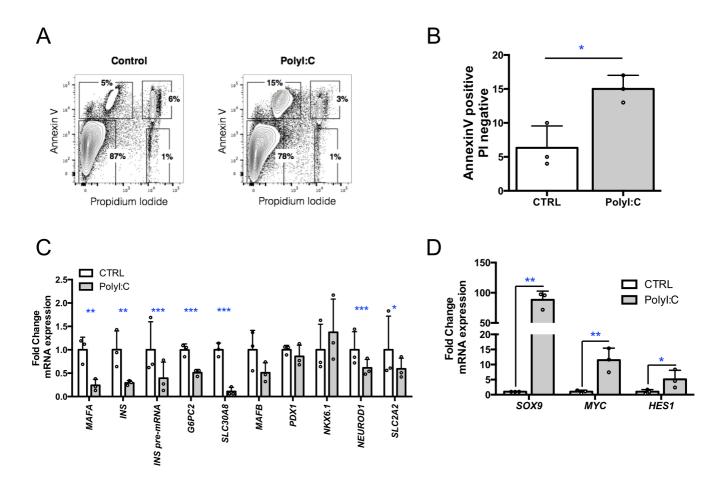
Supplementary Figure 2.

	olyI:C vs CTRL	P-Value
LRIG3	4.04 (±0.28)	0.0002
LMO4	2.28 (±0.04)	0.0003
TPM2	1.74 (±0.27)	0.0206
DAPK2	1.65 (±0.35)	0.0022
GAREM	1.45 (±0.17)	0.0104
RPL36	1.37 (±0.51)	0.0839
ZCCHC24	1.27 (±0.26)	0.0598
PLEC1	1.23 (±0.15)	0.0605
ACOT7	1.19 (±0.12)	0.0526
SMOC2	1.18 (±0.35)	0.2214
ITIH5	1.12 (±0.13)	0.1191
FAM13A	1.12 (±0.19)	0.1886
IGFBP4	1.09 (±0.05)	0.0331
SLC16A1	1.07 (±0.19)	0.2753
MGLL	1.04 (±0.20)	0.3655
ZDHHC9	1.02 (±0.03)	0.1641
COX5A	1.01 (±0.15)	0.4252
NFIB	1.01 (±0.02)	0.2103
HPGD	1.00 (±0.06)	0.4011
MGST1	1.00 (±0.09)	0.4553
RARRES2	0.98 (±0.15)	0.4350
TGM2	0.96 (±0.03)	0.1435
HSD11B1	0.96 (±0.04)	0.1229
GAS6	0.94 (±0.03)	0.0331
PDGFRA	0.91 (±0.18)	0.2788
YAP1	0.91 (±0.12)	0.1761
LDHA	0.88 (±0.19)	0.2256
NDRG2	0.87 (±0.08)	0.0691
IGF1	0.85 (±0.00)	0.0001
TRF	0.84 (±0.07)	0.0383
RASGRP2	0.84 (±0.13)	0.1075
TST	0.80 (±0.05)	0.0118
GAS1	0.80 (±0.14)	0.0771
OLFML1	0.73 (±0.14)	0.0566
FCGRT	0.66 (±0.14)	0.0406

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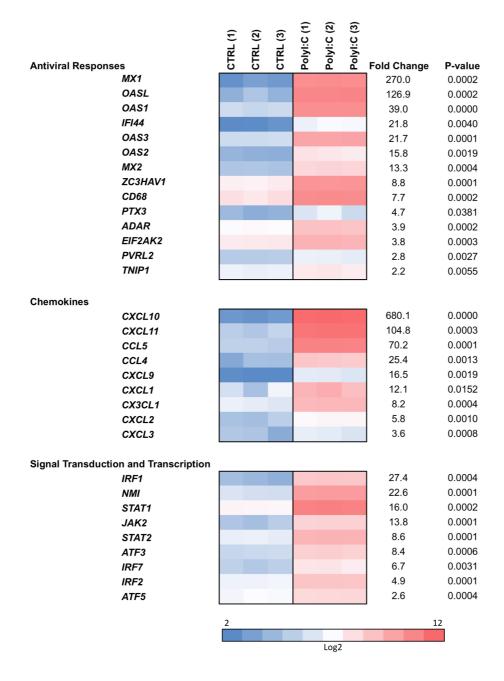
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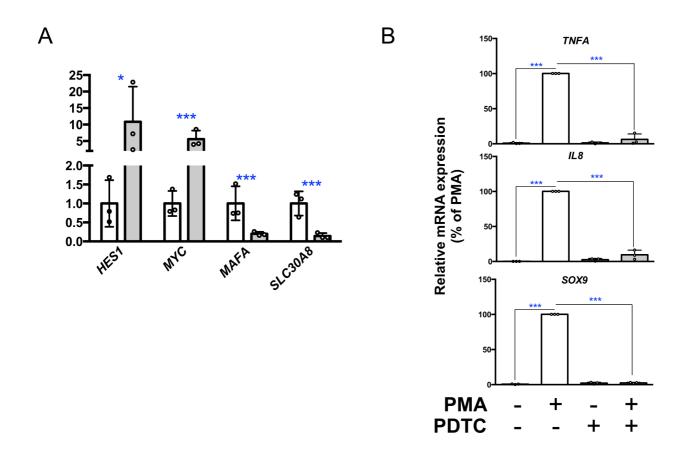
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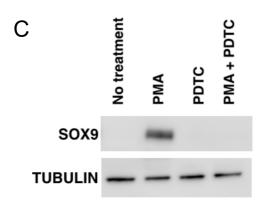


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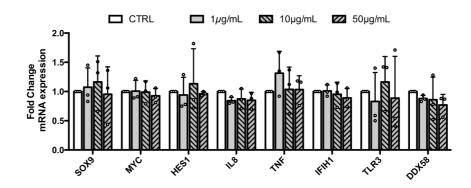
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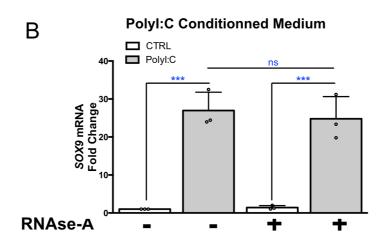
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Supplementary Figure 6.

A





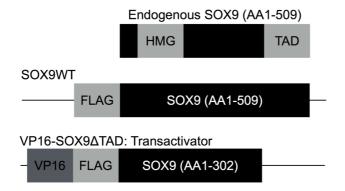
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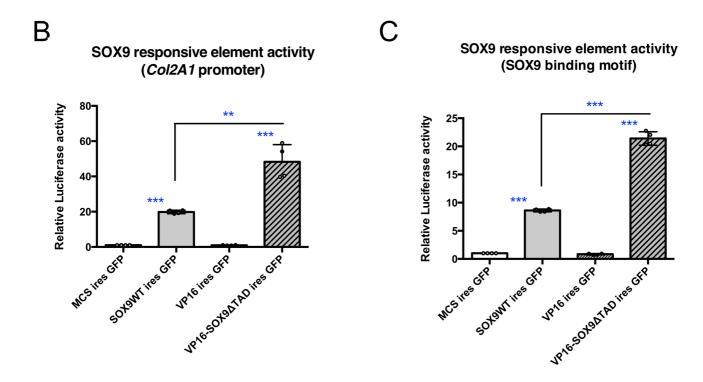
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A



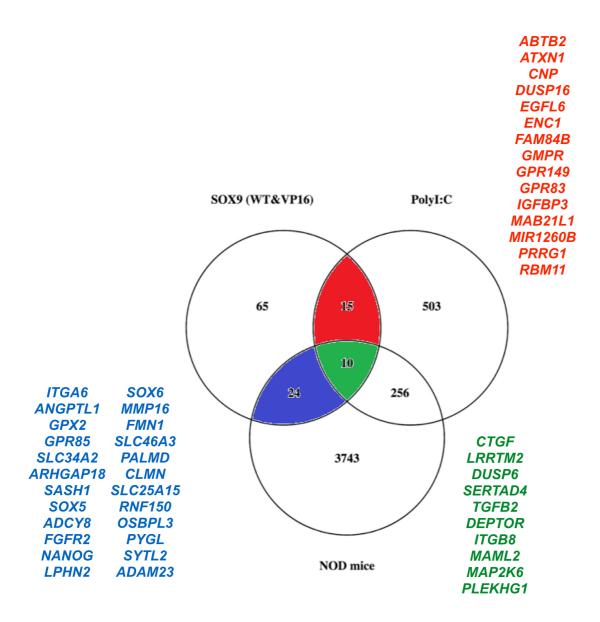


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